

Hypoxia-specific targets in cancer therapy: role of splice variants

Dirk Vordermark

Abstract

Tumour hypoxia is a well known adverse prognostic factor in the treatment of solid tumours. Hypoxia-inducible factor 1 α (HIF-1 α), a transcription factor subunit regulating a large number of hypoxia-responsive genes, is considered an attractive target for novel treatment approaches, due to a frequently reported association between HIF-1 α overexpression and poor outcome in clinical series. This month in *BMC Medicine*, Dales *et al.* report on splice variants of HIF-1 α in fresh frozen tissue samples of early human breast cancer, finding an association of mRNA levels of the variant HIF-1 α^{TAG} with adverse clinical factors (lymph node status, hormone receptor status) and poor metastasis-free survival. This preliminary study addresses the possibility that specific targeting of individual isoforms resulting from alternative splicing may play a role in HIF-1-directed treatment approaches.

See research article: <http://www.biomedcentral.com/1741-7015/8/44>

Background

This month in *BMC Medicine*, Dales and coworkers report on the expression of hypoxia-inducible factor 1 α (HIF-1 α) splice variants in human breast cancer [1]. This work represents an early and preliminary investigation that may become part of a process leading to further individualisation of cancer therapy, specifically addressing the role of hypoxic tumour cells.

Discussion

Low oxygenation of tumour cells is a well known adverse prognostic factor in cancer treatment. It occurs due to (a) rapid tumour growth with resulting long diffusion distances from the nearest blood vessel ('diffusion-limited hypoxia'), as well as (b) the chaotic structure of pathological tumour vessels and resulting inadequate perfusion in part of these vessels ('perfusion-limited hypoxia') [2]. It was established mainly in the 1990 s that a low pretreatment intratumoural partial oxygen pressure (pO₂), as determined by needle electrode measurement, is associated with a poor outcome of treatment, in particular radiotherapy but also surgical treatment, of cervical cancer or head and neck cancer [3,4]. This association has been explained by the reduced ability of ionizing radia-

tion to produce DNA damage in the absence of oxygen as well as, more recently, by an increased potential of hypoxic tumour cells for proliferation, invasion, metastasis and angiogenesis [2].

For decades, investigators have attempted to overcome the treatment resistance of hypoxic tumours in clinical trials, for example by adding so-called 'hypoxic radiosensitiser' drugs to the regimens or introducing hyperbaric oxygen. Although many of the individual trials were negative, a modern meta-analysis confirms the efficacy of hypoxia-directed treatments [5]. While previous strategies were directed at all patients with a given tumour diagnosis, more modern approaches combine (a) the selection of patients with particularly hypoxic tumours and (b) the addition of hypoxia-specific treatment modalities to standard radiotherapy/chemotherapy only in these subgroups.

Determining tumour oxygenation by needle electrode measurements has not been fully accepted in clinical practice and less invasive methods have been proposed: These include the immunohistochemical detection of 'exogenous hypoxia markers' (2-nitroimidazole derivatives such as pimonidazole) injected intravenously before a biopsy, the imaging of hypoxic tumour areas by nuclear medicine methods (for example, F-misonidazole positron emission tomography) or even the measurement of proposed secreted hypoxia markers (for example, osteopontin) in the patient plasma [6-8].

* Correspondence: dirk.vordermark@medizin.uni-halle.de

¹ Department of Radiation Oncology, Martin Luther University Halle-Wittenberg, Halle, Germany

Full list of author information is available at the end of the article

The transcription factor HIF-1 is a central regulator of the physiological or pathophysiological response of mammalian cells to low oxygen levels and has so far been described to regulate hundreds of genes in a hypoxia-dependent manner, many of which are growth factors or involved in cell proliferation, metabolism or vessel formation [9]. HIF-1 is a heterodimer consisting of one of the two α subunits (HIF-1 α or HIF-2 α) and HIF-1 β . Under hypoxic conditions, the oxygen-sensitive subunit HIF-1 α is not degraded via ubiquitylation but rather stabilises, translocates to the nucleus, heterodimerises with constitutively expressed HIF-1 β and binds, in the presence of cofactors, to the hypoxia-responsive elements of HIF-1-regulated genes. HIF-1 is thus assumed to be regulated mostly by protein degradation.

HIF-1 α protein itself and HIF-1-regulated proteins, for example, carbonic anhydrase IX (CA IX) have been studied by immunohistochemistry in paraffin-embedded tumour material as potential 'endogenous hypoxia markers'. Due to the availability of such material, a large number of retrospective analyses of series with long-term clinical outcome were published and, despite some concern about the reproducibility of HIF-1 α staining, significant associations between a strong expression of such HIF-1-related proteins and poor prognosis was seen in the majority of studies on a wide range of solid tumour entities [10]. Such observations were also made in breast cancer [11,12], the topic now studied by Dales *et al.*

Despite its negative prognostic relevance, tumour hypoxia has also been discussed as an opportunity for tumour-specific treatment approaches, for low pO₂ levels as found in solid tumours very rarely occur in normal tissues [13]. Therefore, a prognostically and mechanistically relevant gene or gene product such as HIF-1 α or downstream genes may serve at the same time as an indicator of hypoxic treatment resistance (and therefore be used for patient selection) and as a therapeutic target (in a group thus selected). Inhibitors of the HIF-1 pathway have been grouped into inhibitors of transcription (for example, topoisomerase 2 inhibitors), inhibitors of translation (for example, topotecan, taxanes, epidermal growth factor receptor (EGFR)-targeting agents), inhibitors of DNA binding and transactivation (for example, chetomin) and promoters of degradation (for example, farnesyl transferase inhibitors) [14]. HIF-1 inhibitors have been shown *in vitro* and *in vivo* to reduce angiogenesis and tumour growth and enhance radiosensitivity [15-18]. Several HIF-1 inhibitors are now in early clinical trials.

The findings published by Dales *et al.* [1] are interesting for the further development of HIF-1 targeting approaches. The authors describe the presence of different HIF-1 α splice variants in human breast cancer and non-malignant tissue samples. Splice variants result from

alternative splicing, a process by which the exons of the RNA produced by transcription of a primary gene are reconnected in multiple ways, resulting in different mRNAs which may be translated into different protein isoforms. By performing real-time quantitative PCR of fresh frozen tissue, the authors demonstrate that the mRNA levels of a specific splice variant termed HIF-1 α^{TAG} are associated with positive lymph node status, high tumour grade and negative oestrogen and progesterone status, as well as poor metastasis-free survival (on univariate analysis) in early breast cancer.

Although this study has a number of limitations (small sample size, limited information on patient and treatment characteristics, selection of two subgroups with good and poor metastasis-free survival rather than a homogenous cohort) and failed to demonstrate an independent prognostic role (on multivariate analysis) of HIF-1 α splice variant expression, follow-up studies may advance our understanding of the role of alternative splicing in the identification of prognostic variables and therapeutic targets. In *in vitro* tumour models, small interfering RNAs (siRNAs) directed against wild-type vs individual splice variants of genes relevant for treatment resistance have produced specific effects on clonogenicity and radiosensitivity of human tumour cells [19].

Conclusions

The data from Dales *et al.* suggest that if splice variants detectable in clinical tumour samples can reliably be related to clinical endpoints, targeting approaches directed specifically at these variants may play a role in the further individualisation of cancer treatment.

Competing interests

The author declares that they have no competing interests.

Acknowledgements

This work was supported by Deutsche Forschungsgemeinschaft (VO 871/2-3).

Author Details

Department of Radiation Oncology, Martin Luther University Halle-Wittenberg, Halle, Germany

Received: 23 March 2010 Accepted: 12 July 2010

Published: 12 July 2010

References

1. Dales JP, Beaufile N, Silvy M, Picard C, Pauly V, Pradel V, Formisano-Tréziny C, Bonnier P, Giusiano S, Charpin C, Gabert J: **Hypoxia inducible factor 1 α (HIF-1 α) splice variants: potential prognostic biomarkers in breast cancer.** *BMC Med* 2010, **8**:44.
2. Ruan K, Song G, Ouyang G: **Role of hypoxia in the hallmarks of human cancer.** *J Cell Biochem* 2009, **107**:1053-1062.
3. Nordmark M, Bentzen SM, Rudat V, Brizel D, Lartigau E, Stadler P, Becker A, Adam M, Molls M, Dunst J, Terris DJ, Overgaard J: **Prognostic value of tumor oxygenation in 397 head and neck tumors after primary radiation therapy. An international multi-center study.** *Radiother Oncol* 2005, **77**:18-24.
4. Fyles A, Milosevic M, Hedley D, Pintilie M, Levin W, Manchul L, Hill RP: **Tumor hypoxia has independent predictor impact only in patients with node-negative cervix cancer.** *J Clin Oncol* 2002, **20**:680-687.

5. Overgaard J: **Hypoxic radiosensitization: adored and ignored.** *J Clin Oncol* 2007, **25**:4066-4074.
6. Hoogsteen IJ, Lok J, Marres HA, Takes RP, Rijken PF, van der Kogel AJ, Kaanders JH: **Hypoxia in larynx carcinomas assessed by pimonidazole binding and the value of CA-IX and vascularity as surrogate markers of hypoxia.** *Eur J Cancer* 2009, **45**:2904-2916.
7. Rischin D, Hicks RJ, Fisher R, Binns D, Corry J, Porceddu S, Peters LJ: **Trans-Tasman Radiation Oncology Group Study 98.02. Prognostic significance of [18F]-misonidazole positron emission tomography-detected tumor hypoxia in patients with advanced head and neck cancer randomly assigned to chemoradiation with or without tirapazamine: a substudy of Trans-Tasman Radiation Oncology Group Study 98.02.** *J Clin Oncol* 2006, **24**:2098-2104.
8. Overgaard J, Eriksen JG, Nordsmark M, Alsner J, Horsman MR, Danish Head and Neck Cancer Study Group: **Plasma osteopontin, hypoxia, and response to the hypoxia sensitizer nimorazole in radiotherapy of head and neck cancer: results from the DAHANCA 5 randomised double-blind placebo-controlled trial.** *Lancet Oncol* 2005, **6**:757-764.
9. Semenza GL: **Hypoxia-inducible factor 1 (HIF-1) pathway.** *Sci STKE* 2007, **407**:cm8.
10. Bache M, Kappler M, Said HM, Staab A, Vordermark D: **Detection and specific targeting of hypoxic regions within solid tumors: current preclinical and clinical strategies.** *Curr Med Chem* 2008, **15**:322-338.
11. Brennan DJ, Jirstrom K, Kronblad A, Milikan RC, Landberg G, Duffy MJ, Ryden L, Gallagher WM, O'Brien SL: **CA IX is an independent prognostic marker in premenopausal breast cancer patients with one to three positive lymph nodes and a putative marker of radiation resistance.** *Clin Cancer Res* 2006, **12**:6421-6431.
12. Bos R, van der Groep P, Greijer AE, Shvarts A, Meijer S, Pinedo HM, Semenza GL, van Diest PJ, van der Wall E: **Levels of hypoxia-inducible factor-1 α independently predict prognosis in patients with lymph node negative breast carcinoma.** *Cancer* 2003, **97**:1573-1581.
13. Brown JM: **The hypoxic cell: a target for selective cancer therapy - eighteenth Bruce Cain Memorial Award lecture.** *Cancer Res* 1999, **59**:5863-5870.
14. Koh MY, Spivak-Kroizman TR, Powis G: **Inhibiting the hypoxia response for cancer therapy: the new kid on the block.** *Clin Cancer Res* 2009, **15**:5945-5946.
15. Dewhirst MW: **Intermittent hypoxia furthers the rationale for hypoxia-inducible factor-1 targeting.** *Cancer Res* 2007, **67**:854-855.
16. Kung AL, Zabludoff SD, France DS, Friedmann SJ, Tanner EA, Vieira A, Cornell-Kennon S, Lee J, Wang B, Wang J, Memmert K, Naegeli HU, Petersen F, Eck MJ, Bair KW, Wood AW, Livingston DM: **Small molecule blockade of transcriptional coactivation of the hypoxia-inducible factor pathway.** *Cancer Cell* 2004, **6**:33-43.
17. Williams KJ, Telfer BA, Xenaki D, Sheridan MR, Desbaillets I, Peters HJ, Honess D, Harris AL, Dachs GU, van der Kogel A, Stratford IJ: **Enhanced response to radiotherapy in tumours deficient in the function of hypoxia-inducible factor-1.** *Radiother Oncol* 2005, **75**:89-98.
18. Staab A, Loeffler J, Said HM, Diehlmann D, Katzer A, Beyer M, Fleischer M, Schwab F, Baier K, Einsele H, Flentje M, Vordermark D: **Effects of HIF-1 inhibition by chetomin on hypoxia-related transcription and radiosensitivity in HT 1080 human fibrosarcoma cells.** *BMC Cancer* 2007, **7**:213.
19. Kappler M, Rot S, Taubert H, Greither T, Bartel F, Dellas K, Hänsgen G, Trott KR, Bache M: **The effects of knockdown of wild-type survivin, survivin-2B or survivin-delta3 on the radiosensitization in soft-tissue sarcoma cells in vitro under different oxygen conditions.** *Cancer Gene Ther* 2007, **14**:994-1001.

Pre-publication history

The pre-publication history for this paper can be accessed here:
<http://www.biomedcentral.com/1741-7015/8/45/prepub>

doi: 10.1186/1741-7015-8-45

Cite this article as: Vordermark, Hypoxia-specific targets in cancer therapy: role of splice variants *BMC Medicine* 2010, **8**:45

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

