



Published in final edited form as:

Expert Opin Biol Ther. 2009 January ; 9(1): 21–28. doi:10.1517/14712590802603093.

A Novel Role for the Marrow Microenvironment in Initiating and Sustaining Hematopoietic Disease

Aravind Ramakrishnan, MD and H. Joachim Deeg, MD

Fred Hutchinson Cancer Research Center and the University of Washington, Seattle, WA, 98109

1. Abstract

The marrow microenvironment is composed of a complex network of cells and extra cellular matrix that cooperate to regulate normal hematopoiesis. There is growing evidence that microenvironmental defects can contribute to the pathogenesis of hematologic malignancies. Two basic mechanisms could explain the role of microenvironmental defects in the evolution of hematopoietic neoplasms. There is significant data to support the first mechanism, in which the malignant hematopoietic clone induces reversible functional changes in the microenvironment that result in improved growth conditions for the malignant cells. More recent studies from mouse models have indicated that a second mechanism involving primary microenvironmental defects can also result in malignancy. We will review the role of the microenvironment in inducing and sustaining hematologic malignancies.

Keywords

hematologic malignancy; hematopoiesis; marrow microenvironment; stromal cells

2. Introduction

Hematologic malignancies are thought to arise from a series of genetic abnormalities in a stem or progenitor cell that lead to uncontrolled growth. Although the primary defect leading to disease is thought to occur in a stem or progenitor cell, data from the past few decades have implicated the marrow microenvironment in the pathogenesis of these neoplasms.

The marrow microenvironment consists of a complex structure of both non-hematopoietic and hematopoietic cells, extracellular matrix as well as soluble and membrane bound factors that cooperate to support normal hematopoiesis. It was known as early as the 1960s, based on experiments with the SL/SLd (“steel” mutant) and W/W^v (white spotted mutant) mice that normal hematopoiesis could not occur without a supportive environment.¹ The SL/SLd mouse develops severe anemia due to a lack of kit-ligand on stroma cells and cannot be cured by hematopoietic stem cell transplantation. However, normal hematopoiesis can be restored by implanting a normal spleen in the peritoneum.^{2, 3} In contrast, the W/W^v mouse, which has a similar phenotype, is deficient in c-kit on hematopoietic stem cells and can be cured by stem cell transplantation from a normal donor.^{4, 5} We also know from studies in patients with Hodgkin disease that heavily irradiated areas of the skeleton may no longer be

Corresponding author: H. Joachim Deeg, Member, Clinical Research Division, Fred Hutchinson Cancer Research Center, Professor, University of Washington School of Medicine, 1100 Fairview Avenue N., D1-100, P.O. Box 19024, Seattle, Washington 98109-1024, United States, Phone: (206) 667-5985, Fax: (206) 667-6124, jdeeg@fhcrc.org.

able to support hematopoiesis.⁶ These early studies revealed the crucial role of the microenvironment in normal hematopoiesis.

In vitro studies of the microenvironment over the last several decades have mostly relied on the long-term marrow culture system, first reported by Dexter.⁷ When cells from marrow aspirates are cultured under appropriate conditions, a complex adherent layer of stromal cells forms at the bottom of the flask that recapitulates the in vivo microenvironment and is able to support hematopoiesis for many weeks.^{7, 8} The adherent stromal layer is composed of fibroblast-like cells, adipocytes, endothelial cells, macrophages, lymphocytes, osteoclasts and extracellular matrix.^{9, 10} Although osteoblasts are not always considered as part of the adherent stromal layer of the Dexter culture, they are found in the medullary cavity, and recent studies have revealed that these cells also play a critical role in supporting hematopoiesis.^{11–14} This in vitro system has allowed for the dissection of the components of the microenvironment and the study of the complex contact dependent and contact independent interactions that occur between the stromal compartment and hematopoietic stem cells that regulate stem cell fate decisions.¹⁵

3. The Role of the Microenvironment in Hematologic Malignancies

Extensive research has revealed that the microenvironment also plays a prominent role in sustaining hematologic malignancies.^{16, 17} In many cases this is a reversible functional disturbance caused by interactions of the neoplastic clone with the stromal components. For example, in myeloma, the neoplastic plasma cells communicate with the environment through cell/cell contact as well cytokines to induce functional changes that support the malignant population.¹⁸ The recognition of this dysregulation has led to successful therapeutic targeting of such aberrant signaling in the microenvironment with various drugs including thalidomide and lenalidomide.¹⁹

In other disorders such as chronic myeloid leukemia (CML), clonally derived hematopoietic cells that normally are part of the microenvironment, e.g. macrophages, interact abnormally with other components of the environment to induce functional disturbances that result in a survival advantage of the malignant clone.²⁰ Results of marrow transplantation in animals and humans have clearly demonstrated that the components of the microenvironment that are derived from hematopoietic precursors are replaced by cells of donor origin while stromal cells remain of host origin.^{21–24} Also, the fact that stem cell transplantation is a curative procedure for hematopoietic neoplasms suggests that, in the majority of cases, defects in the microenvironment that contribute to the pathogenesis/pathophysiology of hematologic malignancies are functional disturbances that are reversible.

Until recently, there has been little evidence to support the role of primary stromal abnormalities in the pathogenesis of hematologic neoplasms. There are a few reports of chromosomal abnormalities in stromal cells in patients with myelodysplasia (MDS).^{25–27} However, only recently has there been definitive evidence based on studies in mouse models that abnormal primary stroma function can induce hematologic neoplasia, i.e. evidence for a malignancy-inducing microenvironment.

4. Interactions of Neoplastic Cells with Stromal Elements Can Alter the Microenvironment to Sustain Malignant Cells

The best studied example of this form of microenvironmental dysregulation is multiple myeloma. The pathophysiology of myeloma is determined not only by genetic abnormalities that occur in the clonal plasma cells, but also by the bidirectional complex interactions with the bone microenvironment that lead to the development of skeletal lesions.

Myeloma cells can adhere directly to stroma or extracellular matrix through interactions with numerous adhesion molecules such as VLA-4 and ICAM-1.^{28, 29} These interactions lead to the activation of PI-3 kinase/Akt and NF- κ B pathways.^{30–32} The net result of activation of these pathways is an increase in expression of genes involved in cell proliferation and anti-apoptotic pathways. These interactions also trigger the stromal components to release a variety of cytokines such as IL-6, IGF, IL-1, SDF-1, VEGF,^{33, 34} TNF α , and RANKL to name a few.^{19, 29, 33–35} These factors alter the bone marrow cytokine milieu in the direction of being more supportive to the neoplastic cells. Thus, a paracrine loop is established between the microenvironment and myeloma cells.^{18, 19, 29, 33–35}

The interaction between myeloma cells and stroma is also thought to play a role in the uncoupling between bone formation and bone resorption leading to the development of lytic lesions. The balance between bone formation and resorption is controlled in large part by the balance of two molecules, RANKL and osteoprotegerin (OPG).^{36–38} RANKL activates osteoclastogenesis, whereas OPG functions as a decoy and inhibits the formation of osteoclasts. Wnt signaling regulates the differential activation of these molecules.³⁹ It has recently been shown that myeloma cells inhibit Wnt activation in the microenvironment by the release of soluble DKK1, which causes an increase in the concentrations of RANKL and a decrease of OPG production. This results in increased activation of osteoclasts and bone destruction.⁴⁰ Thus, the microenvironmental dysregulation that occurs in myeloma plays a critical role in the formation of lytic lesions.

This type of functional microenvironmental dysregulation can also be found in myeloproliferative disorders (MPD) and MDS. In chronic idiopathic myelofibrosis (CIMF), the malignant clone induces a polyclonal fibrotic reaction of the stroma that plays a critical role in the pathophysiology of the disease.⁴¹ The mechanism that is currently proposed to explain this scenario is that clonal hematopoietic cells, mainly megakaryocytes and monocyte/macrophages, secrete cytokines that induce a polyclonal stromal reaction that leads to fibrosis. The stromal elements are not part of the malignant clone.⁴² Mouse models have implicated elevated levels of TPO, TGF- β , and low levels of the transcription factor GATA-1 in the pathogenesis of myelofibrosis.^{42–46} In contrast to myeloma, high levels of stromal-derived OPG have been implicated in osteosclerosis.⁴⁷ A similar situation is found in MDS. Studies from our Center have implicated elevated TNF α levels in the microenvironment and increased apoptosis in the marrow.^{48, 49} Engraftment of clonal MDS-derived precursors in NOD/SCID was also dependent on stromal support.⁵⁰

Thus, in a variety of diseases, stroma function is altered by signals from the malignant clone, which apparently serve to support propagation of the clone. It is also evident that agents that target the microenvironmental dysregulation can be very effective in the treatment of these disorders. For example, the immunomodulatory (Imid) family of agents has significant activity in all of these diseases and highlights the importance of targeting the microenvironment.^{51, 52} Other recent findings have led to the identification of new potential targets for therapy in myeloma such as RANKL,⁵³ Wnt signaling and IGF signaling,¹⁹ TNF α signaling in MDS,⁵⁴ and TGF α and PDGF signaling in MPD.⁴² An improved understanding of microenvironmental dysregulation should identify other potentially effective targets for treatment.

5. Hematopoietic Components of the Microenvironment Dysregulate Stromal Function

Many hematologic malignancies are derived from an abnormal stem cell. Thus, they generate clonally-derived progeny including monocytes/macrophages that can lead to a

dysfunctional microenvironment. This was first demonstrated in CML. Bhatia et al. showed that stroma derived from patients with CML did not provide optimal support for normal hematopoietic cells.²⁰ In contrast, growth of CML cells on CML-derived stroma was significantly better, suggesting that the microenvironment in CML was more supportive for the malignant clone. Using fluorescent activated cell sorting (FACS) and fluorescent in situ hybridization (FISH), it was determined that stromal macrophages were all bcr-abl positive and were directly responsible for the selective advantage of clonal bcr-abl cells to proliferate through a contact-dependent mechanism.²⁰

A similar scenario exists in MDS and MPD. Using FISH in MDS patients with cytogenetic markers, we determined that the percentage of clonally marked monocytes closely approximates the percentage of abnormal cells on routine marrow cytogenetics. We have also determined that these cells contribute to the high levels of TNF α in the microenvironment.⁵⁵ Furthermore, the clonally-derived MDS monocytes respond abnormally to stromal signals. For example, MDS monocytes fail to upregulate MMP9 expression when exposed to stromal signals.⁵⁶ The inducible MMP9 levels were inversely correlated with marrow cellularity. MMP9 has been implicated in the cleavage of SDF1 from the microenvironment and may facilitate the egress of hematopoietic cells from the marrow to the peripheral blood.^{57, 58} Based on our data, one could speculate that lack of inducible MMP9 levels in MDS monocytes could contribute to the hypercellularity often seen in this disease. In MPD, clonally-derived megakaryocytes and macrophages are thought to play a principle role in the pathogenesis of the fibrotic reaction by secreting cytokines such as PDGF, FGF and TGF α .^{41, 42}

6. Primary Stromal Defects Can Induce Hematologic Disease

Until recently, little was known about the potential of primary microenvironmental defects in the induction of hematopoietic diseases. There have been some reports of chromosomal abnormalities noted in stromal cells from patients with hematologic neoplasms, and in some cases, the same abnormality was found in both the stroma and the malignant clone.^{25, 59} However, these studies must be interpreted with caution as they did not fully account for clonal macrophage contamination, which can often resemble fibroblasts in cultures.⁶⁰ There have also been numerous conflicting reports on stromal function as either being normal or abnormal in vitro. The fact that hematopoietic cell transplantation is curative in many of the disorders under discussion indicates that intrinsic stroma function is intact or that alterations are reversible in the majority of patients. However, there have been reports of patients who are unable to achieve engraftment despite numerous attempts at stem cell transplantation⁶¹ as well as cases of donor cell-derived leukemia,⁶²⁻⁶⁴ and one may speculate that these patients represent groups that do indeed have an underlying stromal defect.

Only in the past few years, three mouse models have been described which show that primary stroma abnormalities can induce a malignancy in the hematopoietic compartment. In the first model, conditional loss of I κ B α , the inhibitor of NF κ B, resulted in a disorder similar to chronic myelomonocytic leukemia (CMML) with components of MDS and MPD, which in turn resulted in the death of mice within a week of birth. These findings could not be replicated when I κ B α was conditionally deleted in just the myeloid population; thus, constitutive activation of NF κ B in myeloid cells did not lead to malignancy. However, it was not clear from this study whether the loss of I κ B α was necessary in both the microenvironment and stem cell compartments to develop the disease.⁶⁵

Similarly, Walkely et al. demonstrated that conditional deletion of the Retinoblastoma gene (RB) resulted in a myeloproliferative disorder in mice. They also showed that this was a

result of interactions between myeloid cells and the microenvironment. The defect had to be present in both hematopoietic cells and the microenvironment to initiate disease.⁶⁶

The final model, reported by the same group, may be the most compelling. In this report, deletion of the Retinoic Acid Receptor γ (RAR γ) in mice resulted in a chronic myeloproliferative disorder. Transplant studies revealed that RAR γ -hematopoietic cells functioned normally when transplanted into normal mice. However, transplantation of normal hematopoietic cells into the RAR γ -microenvironment resulted in a myeloproliferative disorder in the transplanted cells. TNF α was implicated in the pathogenesis of this MPD as the disorder was partially abrogated when TNF α null stem cells were transplanted into the RAR γ -microenvironment.⁶⁷ These studies therefore showed that a single microenvironmental defect was sufficient to generate a myeloproliferative disorder.

7. EXPERT OPINION

Evidence from research conducted over the last few decades has clearly implicated abnormalities of the marrow microenvironment in the pathophysiology of hematologic malignancies. In the past, abnormalities in the microenvironment were thought to be generated via interactions with the clonal hematologic disorder and that underlying stromal function was normal. Thus in general, treatment strategies have been focused on the eradication of the stem or progenitor cell from which the malignancy arose.

However, recent evidence suggests that focusing therapeutic strategies on the microenvironmental abnormalities can be extremely effective. The Imid family of agents has changed the treatment paradigm in diseases such as myeloma and MDS and highlighted the importance of targeting the microenvironment.^{51, 68} It may also very well be that other agents such as hypomethylating agents that have activity in a number of myeloid disorders act through modifying the microenvironment. In the solid tumor literature, tumor associated stroma can acquire aberrant methylation patterns due either to direct contact with or via factors secreted by the malignant cells.^{69, 70} Thus, in diseases such as MDS, hypomethylating agents may impart their beneficial effects in part by acting on the stroma as well as the malignant clone. This may partially explain why responses to hypomethylating agents are not always correlated with reactivation of tumor suppressor genes in hematopoietic cells.^{71, 72} Further studies focusing on the stroma will be necessary to answer these questions.

The fact that there are components of the microenvironment that are derived from the hematopoietic clone does have therapeutic implications in the setting of stem cell transplantation. With the recent advances in reduced-intensity conditioning regimens, older patients are able to undergo transplant with low-intensity conditioning regimens.⁷³ These regimens may not be effective at eradicating the abnormal macrophages, and the new stem cells may arrive in an environment that still provides an advantage to the malignant clone. This may partially explain the higher rate of relapses in patients undergoing stem cell transplants with low-intensity conditioning regimens for diseases such as MDS.^{74–76} Further research is needed to determine if targeting these hematopoietic-derived microenvironmental components during conditioning will benefit transplant outcomes.

Also of interest are the recent reports of abnormalities in the stroma that lead to malignancies of the hematopoietic compartment. Although historically, hematologic malignancies are thought to arise from a stem or progenitor cell abnormality, there may be groups of patients that have a primary stromal defect leading to the hematologic malignancy. Mouse models have implicated stromal abnormalities in RAR γ , RB and I κ B α in the development of MPDs. Further research is necessary in patients with MPDs to determine if

these genes are dysregulated in the stromal compartment. One may also speculate that there will be patients with abnormalities in both the stem cell and stromal compartments.

If primary stromal defects are identified in humans and implicated in the initiation of malignancy, this clearly will have great impact on the treatment strategies offered to patients. These patients would not be good candidates for allogeneic hematopoietic stem cell transplantation as this modality cannot correct a stromal abnormality; instead, therapies targeted at correcting the underlying stromal compartment would be necessary. Only further study of the stromal compartment in patients with these disorders will reveal whether primary stromal abnormalities exist.

Acknowledgments

Supported in part by grants HL082941 and HL036444.

8. References

1. Russell ES. Hereditary anemias of the mouse: a review for geneticists (Review). *Advances in Genetics* 1979;20:357–459. [PubMed: 390999]
2. McCulloch EA, Siminovitch L, Till JE, et al. The cellular basis of the genetically determined hemopoietic defect in anemic mice of genotype Sl-Sld. *Blood* 1965;26:399–410. [PubMed: 5317869]
3. Copeland NG, Gilbert DJ, Cho BC, et al. Mast cell growth factor maps near the steel locus on mouse chromosome 10 and is deleted in a number of steel alleles. *Cell* 1990;63:175–83. [PubMed: 1698554]
4. Williams DE, Eisenman J, Baird A, et al. Identification of a ligand for the c-kit proto-oncogene. *Cell* 1990;63:167–74. [PubMed: 1698553]
- *5. Huang E, Nocka K, Beier DR, et al. The hematopoietic growth factor KL is encoded by the Sl locus and is the ligand of the c-kit receptor, the gene product of the W locus. *Cell* 1990;63:225–33. Original report on KL as a ligand for c-kit. [PubMed: 1698557]
6. Gerdes AJ, Storb R. The repopulation of irradiated bone marrow by the infusion of stored autologous marrow. *Radiology* 1970;94:441–5. [PubMed: 4904901]
- **7. Dexter TM, Allen TD, Lajtha LG. Conditions controlling the proliferation of haemopoietic stem cells in vitro. *J Cell Physiol* 1977;91:335–44. Classic reference for in vitro hematopoiesis. [PubMed: 301143]
- *8. Gartner SM, Kaplan HS. Long term culture of human bone marrow cells. *Proc Natl Acad Sci USA* 1980;77:4756–9. Development of long-term culture systems. [PubMed: 6933522]
9. Mayani H, Guilbert LJ, Janowska-Wieczorek A. Biology of the hemopoietic microenvironment (Review). *Eur J Haematol* 1992;49:225–33. [PubMed: 1473584]
10. Greenberger JS. The hematopoietic microenvironment (Review). *Critical Reviews in Oncology-Hematology* 1991;11:65–84.
11. Calvi LM, Adams GB, Weibrecht KW, et al. Osteoblastic cells regulate the haematopoietic stem cell niche. *Nature* 2003;425:841–6. [PubMed: 14574413]
- *12. Zhang J, Niu C, Ye L, et al. Identification of the haematopoietic stem cell niche and control of the niche size. *Nature* 2003;425:836–41. The niche concept is “growing up”. [PubMed: 14574412]
13. Taichman RS. Blood and bone: two tissues whose fates are intertwined to create the hematopoietic stem-cell niche (Review). *Blood* 2005;105:2631–9. [PubMed: 15585658]
14. Visnjic D, Kalajzic Z, Rowe DW, et al. Hematopoiesis is severely altered in mice with an induced osteoblast deficiency. *Blood* 2004;103:3258–64. [PubMed: 14726388]
15. Chabannon C, Torok-Storb B. Stem cell-stromal cell interactions. *Curr Top Microbiol Immunol* 1992;177:123–36. [PubMed: 1379138]
16. Duhrsen U, Hossfeld DK. Stromal abnormalities in neoplastic bone marrow diseases (Review). *Ann Hematol* 1996;73:53–70. [PubMed: 8774614]

17. Scadden DT. The stem cell niche in health and leukemic disease (Review). *Bailliere's Best Practice in Clinical Haematology* 2007;20:19–27.
18. Podar K, Richardson PG, Hideshima T, et al. The malignant clone and the bone-marrow environment (Review). *Bailliere's Best Practice in Clinical Haematology* 2007;20:597–612.
19. Mitsiades CS, Mitsiades NS, Munshi NC, et al. The role of the bone microenvironment in the pathophysiology and therapeutic management of multiple myeloma: interplay of growth factors, their receptors and stromal interactions (Review). *Eur J Cancer* 2006;42:1564–73. [PubMed: 16765041]
- *20. Bhatia R, McGlave PB, Dewald GW, et al. Abnormal function of the bone marrow microenvironment in chronic myelogenous leukemia: Role of malignant stromal macrophages. *Blood* 1995;85:3636–45. An important contribution to the question as to whether the “microenvironment” is part of the disease process. [PubMed: 7780147]
- **21. Simmons PJ, Przepiora D, Thomas ED, Torok-Storb B. Host origin of marrow stromal cells following allogeneic bone marrow transplantation. *Nature* 1987;328:429–32. The definitive paper documenting that the marrow stroma remains of host origin, even after hematopoietic cell transplantation. [PubMed: 2886914]
22. Perkins S, Fleischman RA. Hematopoietic microenvironment. Origin, lineage, and transplantability of the stromal cells in long-term bone marrow cultures from chimeric mice. *J Clin Invest* 1988;81:1072–80. [PubMed: 3350965]
23. Lennon JE, Micklem HS. Stromal cells in long-term murine bone marrow culture: FACS studies and origin of stromal cells in radiation chimeras. *Exp Hematol* 1986;12:287–92. [PubMed: 3516716]
24. Laver J, Jhanwar SC, O'Reilly R, Castro-Malaspina H. Host origin of the human hematopoietic microenvironment following allogeneic bone marrow transplantation. *Blood* 1987;70:1966–8. [PubMed: 3315046]
25. Blau O, Hofmann WK, Baldus CD, et al. Chromosomal aberrations in bone marrow mesenchymal stroma cells from patients with myelodysplastic syndrome and acute myeloblastic leukemia. *Exp Hematol* 2007;35:221–9. [PubMed: 17258071]
26. Flores-Figueroa E, Gutierrez-Espindola G, Montesinos JJ, et al. In vitro characterization of hematopoietic microenvironment cells from patients with myelodysplastic syndromes. *Leuk Res* 2002;26:677–86. [PubMed: 12008086]
- *27. Marcondes AM, Mhyre AJ, Stirewalt DL, et al. Dysregulation of IL-32 in myelodysplastic syndrome and chronic myelomonocytic leukemia modulates apoptosis and impairs NK function. *PNAS* 2008;105:2865–70. A potential role for a novel cytokine in stroma and NK cell function in MDS. [PubMed: 18287021]
28. Uchiyama H, Barut BA, Chauhan D, et al. Characterization of adhesion molecules on human myeloma cell lines. *Blood* 1992;80:2306–14. [PubMed: 1421401]
29. Uchiyama H, Barut BA, Mohrbacher AF, et al. Adhesion of human myeloma-derived cell lines to bone marrow stromal cells stimulates interleukin-6 secretion. *Blood* 1993;82:3712–20. [PubMed: 8260708]
- *30. Mitsiades CS, Mitsiades N, Poulaki V, et al. Activation of NF-kappaB and upregulation of intracellular anti-apoptotic proteins via the IGF-1/Akt signaling in human multiple myeloma cells: therapeutic implications. *Oncogene* 2002;21:5673–83. An important contribution to our understanding of the interactions of stroma and hematopoietic cells in multiple myeloma. [PubMed: 12173037]
31. Hideshima T, Chauhan D, Richardson P, et al. NF-kappa B as a therapeutic target in multiple myeloma. *J Biol Chem* 2002;277:16639–47. [PubMed: 11872748]
32. Hideshima T, Nakamura N, Chauhan D, Anderson KC. Biologic sequelae of interleukin-6 induced PI3-K/Akt signaling in multiple myeloma. *Oncogene* 2001;20:5991–6000. [PubMed: 11593406]
33. Gupta D, Treon SP, Shima Y, et al. Adherence of multiple myeloma cells to bone marrow stromal cells upregulates vascular endothelial growth factor secretion: therapeutic applications. *Leukemia* 2001;15:1950–61. [PubMed: 11753617]

34. Hideshima T, Chauhan D, Schlossman R, et al. The role of tumor necrosis factor alpha in the pathophysiology of human multiple myeloma: therapeutic applications. *Oncogene* 2001;20:4519–27. [PubMed: 11494147]
35. Ferlin M, Noraz N, Hertogh C, et al. Insulin-like growth factor induces the survival and proliferation of myeloma cells through an interleukin-6-independent transduction pathway. *Br J Haematol* 2000;111:626–34. [PubMed: 11122111]
36. Lacey DL, Timms E, Tan HL, et al. Osteoprotegerin ligand is a cytokine that regulates osteoclast differentiation and activation. *Cell* 1998;93:165–76. [PubMed: 9568710]
37. Simonet WS, Lacey DL, Dunstan CR, et al. Osteoprotegerin: a novel secreted protein involved in the regulation of bone density. *Cell* 1997;89:309–19. [PubMed: 9108485]
38. Yasuda H, Shima N, Nakagawa N, et al. Osteoclast differentiation factor is a ligand for osteoprotegerin/osteoclastogenesis-inhibitory factor and is identical to TRANCE/RANKL. *PNAS* 1998;95:3597–602. [PubMed: 9520411]
39. Suda T, Takahashi N, Udagawa N, et al. Modulation of osteoclast differentiation and function by the new members of the tumor necrosis factor receptor and ligand families (Review). *Endocr Rev* 1999;20:345–57. [PubMed: 10368775]
40. Qiang YW, Chen Y, Stephens O, Brown N, Chen B, Epstein J, et al. Myeloma-derived dickkopf-1 disrupts wnt-regulated osteoprotegerin and RANKL production by osteoblasts: A potential mechanism underlying osteolytic bone lesions in multiple myeloma. *Blood* 2008;112:196–207. [PubMed: 18305214]
- *41. Tefferi A. Myelofibrosis with myeloid metaplasia. (Review). *N Engl J Med* 2000;342:1255–65. An excellent, comprehensive overview. [PubMed: 10781623]
42. Chagraoui H, Wendling F, Vainchenker W. Pathogenesis of myelofibrosis with myeloid metaplasia: Insight from mouse models (Review). *Bailliere's Best Practice in Clinical Haematology* 2006;19:399–412.
43. Chagraoui H, Komura E, Tulliez M, et al. Prominent role of TGF-beta 1 in thrombopoietin-induced myelofibrosis in mice. *Blood* 2002;100:3495–503. [PubMed: 12393681]
44. Vannucchi AM, Bianchi L, Cellai C, et al. Development of myelofibrosis in mice genetically impaired for GATA-1 expression (GATA-1(low) mice). *Blood* 2002;100:1123–32. [PubMed: 12149188]
- *45. Vannucchi AM, Bianchi L, Paoletti F, et al. A pathobiologic pathway linking thrombopoietin, GATA-1, and TGF-beta1 in the development of myelofibrosis. *Blood* 2005;105:3493–501. Solidifying the pathophysiologic concept of myelofibrosis. [PubMed: 15665119]
46. Villeval JL, Cohen-Solal K, Tulliez M, et al. High thrombopoietin production by hematopoietic cells induces a fatal myeloproliferative syndrome in mice. *Blood* 1997;90:4369–83. [PubMed: 9373248]
47. Chagraoui H, Tulliez M, Smayra T, et al. Stimulation of osteoprotegerin production is responsible for osteosclerosis in mice overexpressing TPO. *Blood* 2003;101:2983–9. [PubMed: 12506018]
48. Kerbauy DMB, Deeg HJ. Apoptosis and antiapoptotic mechanisms in the progression of myelodysplastic syndrome (Review). *Exp Hematol* 2007;35:1739–46. [PubMed: 17976524]
- *49. Stirewalt DL, Mhyre AJ, Marcondes M, et al. Tumour necrosis factor-induced gene expression in human marrow stroma: clues to the pathophysiology of MDS? *Br J Haematol* 2007;140:444–53. Aiming at characterizing signals in stroma that may be dependent upon the hematopoietic clone. [PubMed: 18162123]
50. Benito AI, Bryant E, Loken MR, et al. NOD/SCID mice transplanted with marrow from patients with myelodysplastic syndrome (MDS) show long-term propagation of normal but not clonal human precursors. *Leuk Res* 2003;27:425–36. [PubMed: 12620294]
51. Sokol L, List AF. Immunomodulatory therapy for myelodysplastic syndromes. *Int J Hematol* 2007;86:301–5. [PubMed: 18055335]
52. Barlogie B, Tricot G, Anaissie E. Thalidomide in the management of multiple myeloma (Review). *Semin Oncol* 2001;28:577–82. [PubMed: 11740812]
53. Schwarz EM, Ritchlin CT. Clinical development of anti-RANKL therapy (Review). *Arthritis Research & Therapy* 2007;9 (Suppl 1):S7. [PubMed: 17634146]

54. Deeg HJ, Gotlib J, Beckham C, et al. Soluble TNF receptor fusion protein (etanercept) for the treatment of myelodysplastic syndrome: A pilot study. *Leukemia* 2002;16:162–4. [PubMed: 11840280]
55. Deeg HJ, Beckham C, Loken MR, et al. Negative regulators of hemopoiesis and stroma function in patients with myelodysplastic syndrome. *Leuk Lymphoma* 2000;37:405–14. [PubMed: 10752992]
- *56. Iwata M, Pillai M, Ramakrishnan A, et al. Reduced expression of inducible gelatinase B/matrix metalloproteinase-9 in monocytes from patients with myelodysplastic syndrome: correlation of inducible levels with the percentage of cytogenetically marked cells and with marrow cellularity. *Blood* 2007;109:85–92. The macrophage as a major factor in the dysregulation of the microenvironment in patients with myelodysplastic syndrome. [PubMed: 16954500]
57. Heissig B, Hattori K, Dias S, et al. Recruitment of stem and progenitor cells from the bone marrow niche requires MMP-9 mediated release of kit-ligand. *Cell* 2002;109:625–37. [PubMed: 12062105]
58. McQuibban GA, Butler GS, Gong JH, et al. Matrix metalloproteinase activity inactivates the CXC chemokine stromal cell-derived factor-1. *J Biol Chem* 2001;276:43503–8. [PubMed: 11571304]
59. Flores-Figueroa E, Montesinos JJ, Flores-Guzman P, et al. Functional analysis of myelodysplastic syndromes-derived mesenchymal stem cells. *Leuk Res* 2008;32:1407–16. [PubMed: 18405968]
60. Deeg HJ. Marrow stroma in MDS: culprit or bystander? *Leuk Res* 2002;26:687–8. [PubMed: 12008087]
61. Marsh JC, Harhalakis N, Dowding C, et al. Recurrent graft failure following syngeneic bone marrow transplantation for aplastic anaemia. *Bone Marrow Transplant* 1989;4:581–5. [PubMed: 2790337]
- *62. Witherspoon RP, Schubach W, Neiman P, et al. Donor cell leukemia developing six years after marrow grafting for acute leukemia. *Blood* 1985;65:1172–4. Does the microenvironment have “leukemogenic” potential? [PubMed: 2986742]
63. Lawler M, Locasciulli A, Longoni D, et al. Leukaemic transformation of donor cells in a patient receiving a second allogeneic bone marrow transplant for severe aplastic anaemia (Review) [erratum appears in *Bone Marrow Transplant*. 2002 May;29(9):805]. *Bone Marrow Transplant* 2002;29:453–6. [PubMed: 11919737]
64. McCann SR, Lawler M, Gardiner N, et al. Donor leukemia following allogeneic bone marrow transplantation. *Leukemia* 1994;8 (Suppl 1):S133–S135. [PubMed: 8152280]
- *65. Rupec RA, Jundt F, Rebholz B, et al. Stroma-mediated dysregulation of myelopoiesis in mice lacking IkBa. *Immunity* 2005;22:479–91. Dysregulation of TNF α as a major factor in the evolution of a myeloproliferative/myelodysplastic clone. [PubMed: 15845452]
66. Walkley CR, Shea JM, Sims NA, et al. Rb regulates interactions between hematopoietic stem cells and their bone marrow microenvironment. *Cell* 2007;129:1081–95. [PubMed: 17574022]
67. Walkley CR, Olsen GH, Dworkin S, et al. A microenvironment-induced myeloproliferative syndrome caused by retinoic acid receptor gamma deficiency. *Cell* 2007;129:1097–110. [PubMed: 17574023]
68. Melchert M, List A. The thalidomide saga (Review). *International Journal of Biochemistry and Cell Biology* 2007;39:1489–99. [PubMed: 17369076]
69. Hanson JA, Gillespie JW, Grover A, et al. Gene promoter methylation in prostate tumor-associated stromal cells. *J Natl Cancer Inst* 2006;98:255–61. [PubMed: 16478744]
70. Fiegl H, Millinger S, Goebel G, et al. Breast cancer DNA methylation profiles in cancer cells and tumor stroma: association with HER-2/neu status in primary breast cancer. *Cancer Res* 2006;66:29–33. [PubMed: 16397211]
71. Raj K, John A, Ho A, et al. CDKN2B methylation status and isolated chromosome 7 abnormalities predict responses to treatment with 5-azacytidine. *Leukemia* 2007;21:1937–44. [PubMed: 17611569]
72. Yang AS, Doshi KD, Choi SW, et al. DNA methylation changes after 5-aza-2'-deoxycytidine therapy in patients with leukemia. *Cancer Res* 2006;66:5495–503. [PubMed: 16707479]
73. Baron F, Storb R. Current roles for allogeneic hematopoietic cell transplantation following nonmyeloablative or reduced-intensity conditioning. *Clinical Advances in Hematology & Oncology* 2005;3:799–813. [PubMed: 16258490]

74. Alyea EP, Kim HT, Ho V, et al. Impact of conditioning regimen intensity on outcome of allogeneic hematopoietic cell transplantation for advanced acute myelogenous leukemia and myelodysplastic syndrome. *Biol Blood Marrow Transplant* 2006;12:1047–55. [PubMed: 17067911]
75. Martino R, Iacobelli S, Brand R, et al. Retrospective comparison of reduced-intensity conditioning and conventional high-dose conditioning for allogeneic hematopoietic stem cell transplantation using HLA-identical sibling donors in myelodysplastic syndromes. *Blood* 2006;108:836–46. [PubMed: 16597592]
76. Scott BL, Sandmaier BM, Storer B, et al. Myeloablative vs nonmyeloablative allogeneic transplantation for patients with myelodysplastic syndrome or acute myelogenous leukemia with multilineage dysplasia: a retrospective analysis. *Leukemia* 2006;20:128–35. [PubMed: 16270037]