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Chapter 61: Photoreceptor Cell Degeneration in *Abcr*−**/**− **Mice**

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Abstract

Mice harboring a null mutation in *Abca4*/*Abcr* serve as a model of autosomal recessive Stargardt disease. Consistent with the human retinal disorder, deficiency in Abcr is associated with substantial accumulations of lipofuscin pigments in retinal pigment epithelial (RPE) cells. To observe for photoreceptor cell degeneration in these mutant mice, outer nuclear layer (ONL) thickness was measured at 200 μm intervals superior and inferior to the optic nerve head. ONL width in *Abcr^{−/−}* mouse was reduced at 8–9 month and 11 and 13 months relative to *Abcr*^{+/+} mice; thinning was more pronounced centrally and in superior retina. The numbers of photoreceptor nuclei spanning the width of the outer nuclear layer were also reduced. No evidence of age-related ONL thinning was observed in *Abcr*+/+ mice at these ages. We conclude that albino *Abcr*−/− mice exhibit progressive photoreceptor cell loss that is detectable at 8 months of age and that has worsened by 11 and 13 months of age. The measurement of ONL thickness is an established approach to assessing photoreceptor cell integrity and can be used in preclinical studies using *Abcr*−/− mice.

61.1 Introduction

Mutations in *ABCA4* (*ABCR*), the gene encoding the photoreceptor-specific ATP-binding cassette transporter (Sun et al. 1999), are responsible for some types of inherited retinal degeneration including an autosomal recessive form of retinitis pigmentosa, recessive conerod dystrophy and recessive Stargardt disease (Klevering et al. 2004; Maugeri et al. 2000; Shroyer et al. 2001). All of these inherited blinding disorders are characterized by excessive accumulations of autofluorescent lipofuscin in retinal pigment epithelial (RPE) cells. This disease feature is replicated in the *Abcr* null mutant mouse (Weng et al. 1999) wherein levels of the lipofuscin fluorophores A2E and isoA2E are increased several fold (Kim et al. 2004, 2007; Mata et al. 2001; Weng et al. 1999). Even greater increases in another lipofuscin pigment all-*trans*-retinal dimer-phosphatidylethanolamine (atRAL dimer-PE) are observed (Kim et al. 2007). Characterization of the *Abcr*−/− mouse retina also revealed delayed dark adaptation, increased levels of all-*trans*-retinal and elevated phosphatidylethanolamine (Weng et al. 1999). Although at 6 months of age, the numbers of photoreceptor nuclei were found not to be diminished (Mata et al. 2001), it was recently reported that in 11 month old *Abcr*−/− mice fed both control and vitamin A supplemented diet, the numbers of rows of nuclei across the outer nuclear layer was reduced as compared to wild-type mice (Radu et al. 2008).

Several therapeutic strategies aimed at alleviating vision loss in recessive Stargardt disease have been tested in *Abcr*−/− mice. These approaches include vector-based gene therapies (Kong et al. 2008) and the administration of compounds that limit the visual cycle including isotretinoin (Radu et al. 2003), an inhibitor of 11-*cis*-retinol dehydrogenase; the retinoid analog fenretinide that lowers serum vitamin A (Radu et al. 2005); and compounds that

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target RPE65 (Maeda et al. 2008; Maiti et al. 2006). In these pre-clinical studies quantitation of A2E served as the therapeutic outcome measure.

Although HPLC quantitation of the lipofuscin pigment A2E serves as an objective measure of therapeutic efficacy, additional endpoint measures are desirable. Since the measurement of outer nuclear layer thickness is a widely accepted approach to assessing photoreceptor cell integrity (Lavail et al. 1987; Michon et al. 1991), we have undertaken to compare outer nuclear layer (ONL) thickness in age-matched albino *Abcr*−/− and *Abcr*+/+ mice, homozygous for the Leu-450 allele of Rpe65.

61.2 Methods

61.2.1 Animals and Rearing

Albino *Abca4/Abcr* null mutant mice homozygous for Rpe65-Leu450, were generated and genotyped for the *Abcr* null mutation and Rpe65-Leu450Met variant by PCR-amplification of tail DNA as previously reported (Kim et al. 2004). For Rpe65, digestion of the 545-bp product with *Mwo*I restriction enzyme (New England Biolabs), yielded 180- and 365-bp fragments if the sequence corresponded to Leu-450; Met-450 was associated with the undigested 545-bp band; and heterozygous mice exhibited all 3 bands. *Abcr*−/− and *Abcr*+/+ mice were raised under 12-h on-off cyclic lighting with in-cage illuminance of 30–80 lux. Mice were anaesthetized and perfused with 4% paraformaldehyde in phosphate buffered saline. Following enucleation, eyes were immersed in 4% paraformaldehyde for 24 h at 4°C. The proposed research involving animals has been approved by the Institutional Animal Care and Use Committee (IACUC).

61.2.2 Measurement of Outer Nuclear Layer Thickness

Sagittal 6-mm paraffin serial sections of murine retina were prepared and stained with hematoxylin and eosin. Microscopic images were acquired and analyzed using a digital imaging system (Leica Microsystems; Leica Application suite; Welzlar, Germany). For measurement of ONL thickness, two to three sections through the optic nerve head of the left eyes were imaged with a $10 \times$ objective. ONL thickness was measured at 200 μ m intervals superior and inferior to the edge of the optic nerve head along the vertical meridian; ONL width in pixels was converted to microns (1 pixel: 0.92 μm) and data from the three sections were averaged. For groups of *Abcr*−/− and *Abcr*+/+ mice at each age, mean ONL thickness at each position along the vertical meridian was plotted as a function of eccentricity from the optic nerve head (Mittag et al. 1999; Tanito et al. 2005). Values were compared by unpaired t-test or one-way analysis of variance (ANOVA) as appropriate, and significance was assessed at the 0.05 level (GraphPad Software Inc, La Jolla CA).

61.2.3 Counting Photoreceptor Nuclei

The numbers of nuclei extending across the width of the ONL were determined in superior hemiretina at a fixed distance of 600 μm from the edge of the optic disc. Counting was performed using a digital image obtained from one section per eye photographed with a 63 \times objective. Three lines were drawn (5 μm apart) at this position and nuclei traversed by the line were counted by 2 individuals, one of whom was masked to mouse age and genotype. Nuclei counts obtained by the 2 individuals along the three lines were averaged to give a value for each eye. Values were compared by unpaired *t*-test.

61.3 Results

We probed for evidence of photoreceptor cell degeneration in *Abcr*−/− mice at ages 5 months, 8–9 months, 11 and 13 months using standard morphological methods based on

measurement of ONL thickness. Shown in Fig. 61.1 are representative images of hematoxylin and eosin stained superior central retinas obtained with a $10 \times$ objective; images captured at higher magnification $(40 \times$ objective) are in the insets. A difference in ONL width between *Abcr*−/− and *Abcr*+/+ retina was visible at 8–9 months of age (Fig. 61.1).

ONL thicknesses were plotted as a function of distance in 200 μm intervals superior and inferior to the optic nerve head in the vertical plane. Examination of ONL measurements in *Abc*r +/+ at 5 months, 8–9 months (mean age 8.2 months) and 12 months of age revealed that these measurements did not vary as a function of these ages (one-way ANOVA, $p > 0.05$) (Fig. 61.2). Conversely, in *Abcr*−/− mice at 8–9 months (mean age 8.5 months) a decrease in ONL thickness was observed, the thinning being most noticeable in central retina. For example, comparison of *Abcr*−/− and *Abcr*+/+ mice at 8–9 months of age revealed a 15–20% reduction in ONL thickness, $0.2-1.0$ mm superior and inferior to ONH ($p < 0.05$) (Fig. 61.2). In *Abcr*−/− mice at 11 months of age, the difference in ONL thickness was further accentuated, there being a 23–36% decrease at eccentricities of 0.2–1.0 mm in *Abcr*−/[−] relative to $A b c r^{1/4}$ mice ($p < 0.05$) (Fig. 61.2). At 8–9 months of age, the degenerative changes were slightly more distinct in superior retina as compared to inferior retina, the reduction in the inferior hemiretina ranging from 15 to 19%, while the decrease in superior hemiretina was 17–20%. In *Abcr*−/− mice aged 13 months, the thinning of ONL in superior retina was clearly more pronounced, decreases in the 0.2–1.0 mm zones being 54–69% superiorly and $48-62\%$ inferiorly when compared to $A b c r^{+/+}$ mice (age 12 months). Statistically significant differences (*p* < 0.05) between *Abcr*+/+ and *Abcr*−/− mice in terms of ONL thickness were observed at 5 months of age only at 0.2 and 0.4 mm from the ONH inferiorly and 0.2 mm superiorly (Fig. 61.2).

Measurements of ONL thickness agreed with the counts of nuclei spanning the width of the ONL. The numbers of nuclei at 5 months of age in *Abcr*−/− mice were not significantly different than in *Abcr*^{+/+} mice (*Abcr*^{+/+}: 9.8 ± 0.24; *Abcr*^{−/−}: 9.6 ± 0.33, mean ± SEM; *p* > 0.05). Conversely at 8–9 months, the mean number of nuclei in *Abcr*−/− mice was reduced by 22% relative to *Abcr*^{+/+} (*Abcr*^{+/+}: 9.5 ± 0.25; *Abcr*^{−/−}: 7.4 ± 0.39, mean ± SEM; *p* < 0.05).

61.4 Discussion

The *Abcr^{−/−}* mouse is notable for exhibiting an excessive accumulation of the bis-retinoid pigments that constitute the lipofuscin of RPE cells (Kim et al. 2004; 2007; Weng et al. 1999). By morphometric analysis of ONL thickness combined with counting of photoreceptor cell nuclei spanning the ONL, we have demonstrated that albino *Abcr*−/− mice display photoreceptor cell loss that is clearly detectable at 8 months of age and that has worsened by 12 and 13 months of age. Thinning of the ONL was more marked in the superior hemisphere of retina. The amassing of RPE lipofuscin to pronounced levels in *Abcr^{−/−}* mice precedes the loss of photoreceptor cells; for instance by 3 months of age, A2E levels in the mutant mice are approximately 5-fold greater than in *Abcr*+/+ mice (Kim et al. 2007). Indeed it is potentially significant that by 8 months of age, A2E levels appear to reach a plateau.

As compared to some other mouse models of retinal degeneration, the loss of photoreceptors in the *Abcr*−/− mice occurred with later onset. For instance, mice expressing the P23H substitution in rhodopsin, a mutation prevalent in human autosomal dominant retinitis pigmentosa, exhibit substantial reduction in ONL thickness, even at 2 months of age (Naash et al. 1993). Mice carrying a naturally occurring autosomal recessive mutation in *Rpe65* (*Rpe65*rd12) develop a retinal degeneration that is considered to be slowly progressing, yet

In the present study ONL thicknesses in $A b c r^{+/+}$ mice were consistent with previous reports (Kurth et al. 2007); moreover, we did not observe an age-related thinning of ONL in *Abcr*+/+ mice examined between 5 and 12 months. Consistent with this, most studies of wild-type mice have reported age-related photoreceptor cell loss only after 1 year of age. For example, ONL thickness measurements were reported to be the same at 2, 4, 6 and 12 months of age in BALB/cJ, C57BL/6 and C57BL/6-C^{2 J} mice (Bravo-Nuevo et al. 2004; Li et al. 2001), while another study described a 40% decline in rows of photoreceptor nuclei between 2 and 17 months of age (BALB/c mice) (Gresh et al. 2003). Similarly, screening of several in-bred laboratory strains of mice, including BALB/cJ, BALB/cByJ, A/J, NZW/LacJ and 129P3/J, revealed normal retinal morphology at 10–12 months of age but noticeable ONL thinning by 22–24 months (Chang 2008). On the other hand, Danciger et al. presented ONL thickness data that reflected a decline of approximately 6 μm between 6 and 12 months of age in BALB/c mice (Danciger et al. 2003).

In conclusion, we suggest that two measures of therapeutic efficacy are available for preclinical studies of recessive Stargardt disease utilizing the *Abcr*−/− mouse: HPLC quantitation of RPE lipofuscin fluorophores such as A2E and ONL thickness measurement. Both approaches are also translatable to non-invasive endpoint measures in human clinical trials – analysis of fundus autofluorescence in human subjects serves as a measure of RPE lipofuscin while segmentation of the outer retinal complex [ORC: thickness of ONL, inner segments (IS) and outer segments (OS)] in OCT images of the human eye is akin to ONL thickness measurements in mice.

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Fig. 61.1.

Representative light micrographs of *Abcr*+/+ and *Abcr*−/− mouse retinas. Images of inferior hemisphere along the vertical meridian; age 9 months. Insets, higher magnification images obtained in the regions indicated. ONH, optic nerve head; GCL, ganglion cell layer; INL, inner nuclear layer; ONL, outer nuclear layer. Magnification bar, 50 μm

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Fig. 61.2.

Quantification of outer nuclear layer (ONL) thickness in *Abcr*+/+ and *Abcr*−/− mice at age 5 month (**a**), 8–9 months (**b**) and 11–13 months (**c**). Measurements are plotted as a function of distance from the optic nerve head (ONH) in the inferior and superior hemispheres. Mean \pm SEM; numbers of mice presented in *parentheses*