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Fascicular Perineurium Thickness, Size, and Position Affect Model Predictions of Neural Excitation

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Abstract

The number of applications using neural prosthetic interfaces is expanding. Computer models are a valuable tool to evaluate stimulation techniques and electrode designs. Although our understanding of neural anatomy has improved, its impact on the effects of neural stimulation is not well understood. This study evaluated the effects of fascicle perineurial thickness, diameter, and position on axonal excitation thresholds and population recruitment using finite element models and NEURON simulations. The perineurial thickness of human fascicles was found to be $3.0\% \pm 1.0\%$ of the fascicle diameter. Increased perineurial thickness and fascicle diameter increased activation thresholds. The presence of a large neighboring fascicle caused a significant change in activation of a smaller target fascicle by as much as $80\% \pm 11\%$ of the total axon population. Smaller fascicles were recruited at lower amplitudes than neighboring larger fascicles. These effects were further illustrated in a realistic model of a human femoral nerve surrounded by a nerve cuff electrode. The data suggest that fascicular selectivity is strongly dependent upon the anatomy of the nerve being stimulated. Therefore, accurate representations of nerve anatomy are required to develop more accurate computer models to evaluate and optimize nerve electrode designs for neural prosthesis applications.

Keywords

Computer models; functional electrical stimulation; nerve cuff electrode; nerve fascicle; neuroanatomy

I. Introduction

ELECTRICAL stimulation of the nervous system can restore some function in individuals impaired by stroke, spinal cord injury, neural diseases, or trauma [1]. Implantable nerve cuff electrodes can be used to selectively stimulate individual fascicles or fascicular groups, producing selective muscle activation [2]–[4]. Clinically, nerve cuff electrodes have been used to reduce seizures using vagus nerve stimulation [5], restore limited motor function in lower [6] and upper [7] extremities, and restore respiration in patients with spinal cord injury [8]. Despite these successes, there is room for improvement. Computer models can facilitate the evaluation and optimization of nerve cuff electrode designs. They have been used to predict fascicular selectivity of stimulation [2], [9]–[17] and recording [18]–[22] using various electrode configurations.

Previous models examining neural excitation used simplified representations of neural anatomy [2], [9], [11], [12], [14], [19]. Model simplifications included ignoring or fixing the perineurial thickness, (th_p) using constant fascicle diameter (D_f) , and uniformly distributing fascicles within the nerve. The effects of model simplifications on simulation predictions of neural excitation have not been investigated. Neuroanatomical data shows that nerves contain multiple fascicles of varying diameters [23]–[26] and that human nerves have a greater number of fascicles than animal nerves [27]. Additionally, histological studies suggest that a relationship exists between fascicular size and perineurial thickness [23], [28], [29]. The design of clinical peripheral nerve electrodes should account for these factors. Although simulations using realistic nerve anatomy require more computational power and processing time than simplified models, it is feasible to solve them using powerful computers [17].

No standard exists for the th_p of a fascicle. As a result, prior modeling studies have ignored perineurial tissue to decrease the model complexity [30] or used a fixed th_p regardless of the D_f [9], [12]. One study fixed the th_p at 30 μ m for fascicles of 200–800 μ m diameters [12]. Others suggest that th_p is roughly 5% of the D_f [9], [11], [14], although the evidence for this quantity was not found within the cited sources. Even when this relationship was noted, the perineurial thickness in one study was still fixed to 50 μ m for fascicles of 320–870 μ m diameters (producing relative percentages of 5.7%–15.6%), due to limitations of the modeling software [9]. It is unclear how these inconsistencies and variations in representation of the perineurial thickness influence model results.

To further simplify the model, several studies fixed the fascicular size [2], [12], [19], ignoring the physiologic variations in fascicle diameter found within nerves. In these studies, the D_f was fixed to 800 μ m [2], [12] or 617 μ m [19]. Additionally, the fascicles were uniformly spaced within the nerve; another frequent simplification inconsistent with neuroanatomy. These simplifications were made to reduce the number of parameters within the models and to facilitate interpretation of the results.

This study determined the effects of perineurial thickness (th_p) , fascicle diameter (D_f) , and fascicular position on model predictions of nerve stimulation. Computer simulations were used to investigate the hypotheses that 1) increased perineurial thickness increases axonal activation thresholds, 2) increased fascicle diameter increases activation thresholds, and 3) in multifascicular models, perineurial thickness, fascicle diameter, and fascicular position affect axonal recruitment. Understanding how neural anatomy affects fiber recruitment is important for determining the clinical efficacy of neural prostheses and optimizing their design.

II. Methods

A. Overview

An overview of the general methods used in this study is provided first, followed by detailed descriptions of each model created. Results were obtained using a three-step process of 1) creating and solving 3-D finite element method (FEM) models, 2) using results from these FEM models to interpolate voltages along axons, and 3) applying the interpolated voltages to a double-cable axon model to investigate axonal activation thresholds and population recruitment.

Three-dimensional FEM models were designed in Maxwell 3D v 11.1.1 (Ansoft Corporation, Pittsburgh, PA). Generally, FEM models were comprised of neural tissue, a stimulating electrode, and saline. Fascicles were modeled as 60-mm-long dodecahedrons, which simulates the effects of an electric field on a semi-infinite axon [17]. Perineurium

surrounded each fascicle. Tissue conductivities were based on published data (Table I) [12]. In each simulation, a single electrode with one active contact was modeled. The electrode was insulated on all but one side with nonconductive silicone to mimic a peripheral nerve cuff electrode. The stimulating surface of the electrode was $0.5 \text{ mm} \times 0.5 \text{ mm}$, through which a 1 mA cathodic current was applied. Saline surrounded the neural tissue and electrode and had a dimension of $150 \text{ mm} \times 150 \text{ mm}$, $\times 200 \text{ mm}$ a size shown to have little or no affect on the potential along the axon [17]. The edges of the saline volume were defined as sinks to represent a distant ground. Using the built-in solver, the 3-D potential distribution established by the electrode was calculated.

The potential distributions were imported into MATLAB 7.0 (The MathWorks, Inc., Natick, MA) and used to interpolate voltages along each assigned axon inside the fascicle using a 3-D cubic spline [17]. Simulated axon diameters ranged from $4-16 \mu m$, which is within physiological range for mammalian nerves [31], [32]. The positional offset of the Node of Ranvier closest to the z = 0 plane on which the electrode was centered was randomized, falling between no offset and the maximum offset possible: half of the internode length [17].

The interpolated voltages were applied to the McIntyre, Richardson, and Grill (MRG) [33], [34] double-cable axon model in NEURON.¹ This model was chosen because it accurately represents the mammalian axon response to stimulation. NEURON simulations used a binary search to determine the axonal threshold required to generate a propagating action potential for a single pulse of 0.025–0.5 ms duration. The percent activation of a fascicle was determined using simulations of five distinct axon populations. Each population was made up of 20 axons of varying diameter, based on a physiologic diameter distribution from [31], [32], randomly positioned throughout the fascicle. The number of axons in which a propagating action potential occurred during stimulation was divided by the total number of axons in the population. The mean and standard deviation for the activation of the five populations was computed. The activation of an axon or a population of axons was assumed to have no affect on the response of any other axon or population of axons. Statistical significance of the NEURON simulation data was evaluated using Minitab v15 (Minitab Inc., State College, PA) with α set to 0.05.

The subsequent sections provide a more detailed description of the specific modifications made to the general methods to examine the stated hypotheses.

B. Effects of Perineurial Thickness on Activation Thresholds

Before the computational modeling began, we performed a literature search for measurements of the physiologic perineurial thickness. Measurements reported by Sunderland and Bradley [23] for 704 fascicles from 20 human peripheral nerves were analyzed to quantify the relationship between th_p and D_f . In the discussion of [23], the authors suggest that a linear relationship between th_p and D_f exists. To quantify this relationship, we divided each fascicle's th_p by its D_f . The th_p as a percent of the corresponding D_f for individual nerve types and for the entire data set was reported as average \pm one standard deviation. The physiologic measurements of the th_p and D_f were compared to previously used values in modeling studies.

Once the relationship between th_p and D_f was determined, a FEM model was created to investigate effects of th_p on axonal of thresholds. This model consisted of one fascicle with a $D_f 500 \mu m$ and an electrode centered above it [Fig. 1(a)]. One axon was positioned at the

¹Hines, Moore, and Carnevale; http://www.neuron.yale.edu

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center of the fascicle. The electrode-axon distance, d_1 , was 500 μ m. The th_p was varied from 0%–15% of D_f , to determine its effect on axonal threshold.

C. Effects of Fascicle Diameter on Activation Thresholds

FEM models were made to examine the effects of on axonal activation thresholds [Fig. 1(b)]. Fascicular diameters of 100, 500, and 1000 μ m were studied, based on the reported physiologic range for mammalian peripheral nerves [23], [25], [27]. For each model, th_p of 0, 3, 15, and 30 μ m was tested. To account for geometric effects, electrode-perineurium distance, d₂, was held constant at 250 μ m. One axon was positioned inside the fascicle with a constant electrode-axon distance, d₁, of 300 μ m. This axon was not necessarily at the center of the fascicle. The effects of varying both the D_f and th_p were assessed based on axonal threshold.

D. Effects of Fascicles on Neighboring Fascicles

1) Two Fascicles Side-by-Side—Two-fascicle FEM models were designed to determine how neural anatomy affects more complex, multifascicular nerve models [Fig. 1(c)]. The control fascicle (A) had a fixed D_f of 500 μ m and a fixed th_p 15 μ m (a relative thickness of 3%, see Results). Diameter (100, 300, 500, 700, or 1000 μ m) and perineurial thickness (3, 9, 15, 21, or 30 μ m) were varied, at all permutations, in a second fascicle (B). The centers of the two fascicles were a fixed distance of 900 μ m apart, while the distance between the perineurium edges, d₃, varied with B D_f and th_p . The electrode was centered between varied with B and the two fascicles, 800 μ m above the fascicle axis of symmetry. A model containing only fascicle A served as the control. By moving fascicle B relative to fascicle A, the effect of distance between perineurium edges, d₃, was also investigated (d₃ ranged from 25–2000 μ m).

In MATLAB, five distinct populations of axons were randomly distributed throughout fascicle A. These axons were individually simulated in NEURON and stimulated with a pulse ranging from 0.025–0.5 ms in width and 1–1.5 mA in amplitude to achieve recruitment. The percent of axons activated in fascicle A when fascicle B was added to the model was compared against the results obtained from the control-only model, where fascicle B was not included. To allow for direct comparison between control and neighboring fascicle models, the randomized spatial distribution and diameters of the axons in fascicle A were held constant in each simulation. No active interactions between the two fascicles or between multiple axons were simulated.

2) Surrounding Neighbor Effects—The impact of a fascicle oriented around the control fascicle was examined. For this set of simulations, a neighboring fascicle was moved around the target fascicle [Fig. 1(d)]. Fascicle A had a 500 μ m diameter and was centered 1.43 mm below the electrode. Neighboring fascicle B had a variable diameter of 100, 500, or 1000 μ m and was moved around fascicle A in 5° increments, 0° ≤ α ≤ 180°. The *th*_p was modeled at 3% of the D_f for each fascicle. The distance between the edges of the perineurium of A and B, d₃, was 50 μ m. Following the same techniques detailed previously, the percentage of fascicle A axons that propagated an action potential was found in simulations containing only the control fascicle and compared to that obtained from simulations that included a neighboring fascicle.

E. Realistic Geometry Based on Cadaveric Nerve

A realistic nerve model was used to illustrate the clinical relevance of our findings. An FEM model of a cadaveric human femoral nerve, based on a histological nerve cross section [24], was chosen as an example of a realistic nerve model. A similar model is currently being used to improve the design of nerve cuff electrodes intended for femoral nerve stimulation

[17]. A pair of neighboring fascicles was chosen to demonstrate the th_p , D_f , and neighboring fascicle effects in a realistic nerve model (Fig. 2). These two fascicles were chosen because of their differences in diameters and functions, which makes selective activation of these fascicles more desired for functional electrical stimulation. Fascicles 1 and 2 had diameters of 200 and 650 μ m, respectively. Their centers were 550 μ m apart.

Four FEM models were generated: 1), 2) a model of each individual fascicle in the pair, 3) a model of both fascicles in the pair, and 4) a model with all fascicles in the nerve cross section. Unlike our previous models, these models also included epineurium around the fascicles. Four perineurial thicknesses were tested, 0% and 3% of the each fascicle diameter, 30 μ m (equal to 15% of fascicle 1 diameter and 4.6% of fascicle 2 diameter), and 50 μ m (equal to 25% of fascicle 1 diameter and 7.7% of fascicle 2 diameter). These perineurial thicknesses were chosen based on thickness used in previous models [9], [12], [30] and our analysis of physiologic perineurial thickness (see Section III). The electrode contact was placed on the inner surface of the cuff and centered between the two fascicles in the chosen pair.

The percent activations of each of the two fascicles alone, with a neighboring fascicle, and with all fascicles in the cross section were compared over a range of recruitment levels (pulse widths of 0.01–0.1 ms). The effects of the D_f and th_p were also assessed based on the computed percent activations.

F. Perineurial Conductivity Sensitivity Analysis

Throughout this study, we used amphibian perineurial conductivity of 0.0021 S/m, which is within the range of 0.0006 to 0.01 S/m used in previous models [9]–[12], [16], [17], [19]. Using the model shown in Fig. 1(c), a perineurial conductivity sensitivity analysis was conducted. Perineurial thickness of 3% of the fascicle diameter was used (see Section III) and its conductivity was varied from 0.0002 to 2 S/m. As perineurial conductivity was varied, fascicle A center axon threshold and the effects of a neighboring fascicle on the percent activation of fascicle A were observed to determine the sensitivity of the models to perineurial conductivity.

III. Results

A. Effects of Perineurial Thickness on Activation Thresholds

Physiologic measurements of th_p and D_f from [23] were plotted (Fig. 3) and their relationship for different nerves was calculated (Table II). The perineurial thickness expressed as a percentage of the fascicle diameter was $3.0\% \pm 1.0\%$ (range 1.3%-8.9%). A linear fit of the data with the intercept fixed at zero was 2.6% ($R^2 = 0.67$). The values used in previous computer modeling studies [9], [12], [15], [19], [22], [30] were also compared to the physiologic perineurium thickness measurements in Fig. 3.

The threshold necessary to activate an axon at the center of the fascicle increased with an increase in the modeled th_p (Fig. 4). For each axon diameter, results were normalized to the activation thresholds when no perineurium was present. Adding a 3% th_p to the model increased the threshold by 14%–51%, depending on axon diameter and pulse width. Increasing th_p had a larger effect on small diameter axons than large ones. These trends were consistent across tested pulse widths.

B. Effects of Fascicle Diameter on Activation Thresholds

Activation thresholds increased with D_f when th_p physiologic (3%) (Fig. 5). With th_p , equal to 3% of D_f increasing from 100 to 500 μ m increased axonal threshold by 28%–60% and

increasing from 500 to 1000 μ m increased axonal threshold by 26%–41%, depending on axon diameter and pulse width. Axonal thresholds increased with increasing th_p , for each modeled D_f and these effects of th_p were more pronounced for larger D_f . Using a nonphysiologic fixed th_p could reverse this trend; without a perineurium, axonal threshold decreased with increasing D_f . This was consistent over the tested pulse widths and axon diameters.

C. Effects of Fascicles on Neighboring Fascicles

1) Two Fascicles Side-by-Side—The percent activation of fascicle A for each permutation of D_f and th_p of fascicle B was compared to that obtained without fascicle B. Fascicle A activation increased as D_f and th_p of fascicle B increased (Fig. 6). Statistically significant increases in activation of A were observed when D_f of B was greater than or equal to D_f , of A (p < Student's *t*-test, as compared to 0% increase in fascicle A activation). The magnitude of the increase in activation depended on the recruitment level of A. With the addition of fascicle B of 1000 μ m diameter and 30 μ m th_p , the percent activation in fascicle A increased from 15% ± 7% to 47% ± 6%; a starting activation level of 80% ± 13% increased to 90% ± 8%. The effect of th_p of fascicle B on activation in fascicle A was not statistically significant. For all pulse widths and amplitudes tested, the activation in fascicle A increased due to the addition of fascicle B, unless the stimulus was large enough to activate 100% of fascicle A.

As fascicle B was moved closer to fascicle A, its affect on fascicle A increased (Fig. 7). For all tested distances between the two fascicles $\leq 700 \,\mu$ m, the addition of a 500 or 1000 μ m diameter fascicle B resulted in a statistically significant increase in activation of fascicle A (p < 0.004). A 100- μ m-diameter neighboring fascicle negligibly affected the control fascicle's activation.

2) Surrounding Neighbor Effects—When the neighboring fascicle was moved around the control fascicle, the location and the D_f of the neighbor affected the percent activation of the control fascicle. Fig. 8 shows the change in percent activation of the control fascicle caused by the addition of the neighboring fascicle. As the relative size of the neighboring fascicle increased, the magnitude of its effect on the activation of the control fascicle increased. For fascicle B diameters of 500 and 1000 μ m, the percentage of axons activated in fascicle A significantly decreased if the neighboring fascicle was positioned at $\alpha < 50^{\circ}$ (p < 0.03, p < 0.02, respectively). This change is representative of over-prediction of activation by the control-only model by as much as $80\% \pm 11\%$ of the total axons in fascicle A. Conversely, the percentage of axons activated in the control fascicle significantly increased if the neighboring fascicle was located at $\alpha > 65^{\circ}$ for 500 μ m fascicle B or $\alpha > 50^{\circ}$ for 1000 μ m for 500 μ m fascicle B (p < 0.03, p < 0.009, respectively). In these positions, the change is representative of under-prediction of activation of fascicle A by the control-only model of as much as $31\% \pm 10\%$ of the total axon population. The under- and over-prediction regions imply that the addition of the neighboring fascicle increased or decreased, respectively, the percent activation of the control fascicle.

D. Realistic Geometry Based on Cadaveric Nerve

Results using realistic neuroanatomy models, shown in Fig. 2, were consistent with results from the previous sections. Simulations showed that the target D_f , the presence of neighboring fascicles, the interaction between the th_p and the interaction between th_p and presence of neighboring fascicles were statistically significant (p < 0.001, ANOVA). In all models, the small fascicle 1 had significantly higher activation levels than the larger fascicle 2 [p < 0.0001, Tukey's *t*-test; Fig. 9(b)]. The addition of all neighboring fascicles in the nerve cross section significantly increased the activation of fascicle 1 as compared to

E. Perineurial Conductivity Sensitivity Analysis

A separate set of models were run to determine the effects of perineurial conductivity (data not shown). Decreasing perineurial conductivity resulted in increased center axon thresholds in fascicle A. However, changing the perineurial conductivity did not change the overall trends of the effects of the neighboring fascicle B on the control fascicle A that were shown in Figs. 6 and 7.

depending on the level of recruitment of the target fascicle [Fig. 9(a) versus (b)].

IV. Discussion

A. Effects of Perineurial Thickness on Activation Thresholds

Histological measurements have shown that perineurial thickness varies with fascicle diameter [23], [28]. Our analysis quantified this relationship and showed that in human peripheral nerves is $3.0\% \pm 1.0\%$ of the D_f As shown in Fig. 3, many studies overlooked this relationship and used constant perineurial thicknesses [9], [12], [22]. Modeling th_p as dependent on D_f is more physiologically accurate than using a constant th_p . The data in this study shows that varying affects the axonal stimulation thresholds in unifascicular computer models (Fig. 4). The presence of the perineurium decreases the gradient of the potential field inside the fascicle [35] and decreases the second difference in extracellular voltage along the axon, thereby increasing stimulation thresholds. These data suggest that ignoring the perineurium, as was done in [30], may result in an underestimation of the activation thresholds by as much as 14%-51%, depending on the axon diameter. Alternatively, using a th_p that is too large, such as 15% of the D_f instead of 3%, may overestimate the activation thresholds by as much as 35%-78%, depending on the axon diameter (Fig. 4). Consequently, modeling nonphysiologic perineurium may lead to over- or under-prediction of fiber recruitment during neural stimulation and inaccurate conclusions when trying to optimize a neural prosthesis.

The effect of perineurium on activation thresholds depends on its relative resistance, which is a function of the thickness and resistivity of the perineurial tissue layer. Therefore, the voltage distributions inside the modeled fascicles and, consequently, the predicted activation thresholds can vary depending on the th_p and the chosen electrical properties of the neural tissue [19], [30]. To our knowledge, perineurial conductivity has not been measured in mammalian nerves. In this study, we varied the th_p and used an amphibian perineurial conductivity of 0.0021 S/m, which is within the range of 0.0006–0.01 S/m used in previous models [9]–[12], [16], [17], [19]. Other studies have used another approach of fixing the th_p and adjusting its conductivity based on D_f [11], [36], which may yield an equivalent resistance of the perineurium. Our results are contingent on the assumption that perineurial conductivity in frogs is similar to that in mammals. Due to its large resistivity relative to that of other neural tissues [35], perineurium causes bending of the electric field lines around fascicles [12]. Therefore, overestimating th_p increases resistance to current flow into the fascicle, decreases the gradient of the potential inside the fascicle and the second difference in voltage along the axon, and ultimately overestimates activation thresholds.

The results of this study suggest that modeling nonphysiologic perineurial thickness has the potential to alter predictions on stimulation selectivity. Several previous studies did not

account for the relationship between th_p and (Fig. 3) [9], [12]. One study used a 30 μ m th_p for fascicles of 200–800 μ m diameters, producing relative percentages of 3.75%–15% [12]. Another study fixed the th_p at 50 μ m, for fascicle diameters of 320–870 μ m, producing relative percentages of 5.7%–15.6% [9]. Based on our results, if the perineurium of the smaller fascicles had been modeled as the physiologic 3% of D_f , more axons in these fascicles would have been activated at lower thresholds. Consequently, modeling physiologic th_p may alter the conclusions drawn from these models. Therefore, computer models designed to investigate neural stimulation selectivity should include physiologic to achieve more precise results.

B. Effects of Fascicle Diameter on Activation Thresholds

Some studies fixed D_f to simplify model complexity [2], [12], [19]. However, mammalian nerves consist of fascicles of varying diameters [23]–[27]. This simplification ignores the effect of D_f on activation thresholds. Since th_p is related to D_f (Fig. 3), fixing D_f also fixes th_p We showed that D_f significantly affects axonal threshold. Using physiologic th_p small fascicles had lower activation thresholds than large fascicles (Fig. 5). These results match those seen experimentally where the small common peroneal nerve fascicle consistently had a lower activation threshold than the larger tibial nerve fascicle [11]. Increasing D_f increases resistance to current flow into the fascicle due to the increasing th_p and through the fascicle due to the increasing amount of transverse endoneurium resistance, which may explain our findings that larger fascicles have higher thresholds.

Using a fixed th_p resulted in different activation thresholds than when the perineurium was modeled as 3% of the D_f (Fig. 5). When no perineurium was modeled ($th_p = 0 \mu m$) axons in larger fascicles had lower thresholds than those in smaller fascicles. This trend reversal further demonstrates the importance of modeling physiologic th_p and D_f We believe that the model geometry is the underlying cause for the trend reversal in activation thresholds. The distance between the electrode and the axon and the amount of endoneurium between the electrode and axon were held constant. Therefore, larger fascicle diameters resulted in an increased amount of endoneurium below the location of the axon. Due to the anisotropy of the endoneurium, as the fascicle diameter increases, there will be an increased resistance to current flow through the fascicle as compared to along the fascicle. Current paths will be altered as th_p and endoneurial size/volume change.

As shown in this and previous studies, large fascicles are more difficult to stimulate than small fascicles [11], [12]. Therefore, models looking at fascicle selectivity during neural stimulation should include the physiologic sizes of the fascicles for more accurate results. Otherwise, if all fascicles were modeled with equal diameter, selectivity predictions may not match experimental outcomes. These results further illustrate the importance of modeling physiologic around fascicles because fixing the th_p may alter axonal thresholds and consequently selectivity measures of fascicles of different sizes.

C. Effects of Fascicles on Neighboring Fascicles

The effect of one fascicle on its neighboring fascicle has not been previously documented. As demonstrated in Figs. 6–8, fascicular activation of the target fascicle was altered by the location, th_p , and D_f of neighboring fascicles. This trend was true even when the neighboring fascicle was located behind the target fascicle, relative to the electrode. The change in activation level of the target fascicle was more pronounced when the neighboring fascicle size and th_p exceeded that of the target. As shown in Fig. 7, increased distance between fascicles reduced the magnitude of this effect, but the effect was still significant for physiologic distances of 95 ± 47 μ m (range of 28–174 μ m) between a fascicle and its closest neighbor in a femoral nerve cross section. Therefore, within this range of physiologic

distances, neighboring fascicles are expected to influence the fascicular stimulation selectivity of nerve cuff electrodes.

One possible mechanism for this neighboring fascicle effect could be due to current being shunted away from the large fascicle towards its smaller, less resistive neighbor, which alters the voltage distributions. As expected, there is an increase in impedance of the fascicles caused by increased D_f and th_p . Because current follows the path of least resistance, more current may enter a smaller fascicle due to the relatively high resistance of the larger neighboring fascicle, resulting in a larger percentage of activated axons within the smaller fascicle. The effect may be more prominent when fascicles have a large disparity in size due to a larger mismatch in their resistances.

Gradients in the extracellular field potentials determine the axon activation levels; therefore, additional insight may be gained from examining the field distributions directly. Our functional approach of showing the neighboring fascicle effects on the percent activation of the target fascicle was chosen because, in general, previous modeling studies [12], [17] used activation levels, rather than raw field distributions or activating functions, to make conclusions about stimulation selectivity. Activation levels are also used to predict muscle forces and joint moments in biomechanical models that may follow neural stimulation modeling. Future studies should explore the potential field gradients to determine if current shunting is the mechanism behind the observed neighboring fascicle effects.

Studies that use uniformly distributed fascicles of equal diameter [2], [12], [19] minimize the effects neighboring fascicles have on each other. Fixing the fascicle diameter may lead to inaccuracies in model predictions because the effects of target and neighboring fascicle diameters on activation thresholds are ignored. It is possible that using these types of simplifications would yield predictions that do not match experimental results.

In an effort to simplify computationally intensive models, some investigators may also choose to exclude fascicles from the modeling space. Most likely, the fascicles that would be eliminated from the modeling space would be those that did not fall between the stimulating contact and the fascicle of interest. If a small amount of error can be tolerated, then these simulations indicate that it may be acceptable to remove fascicles from the simulation space if they are much smaller in diameter than the fascicle of interest. However, if the removed fascicles are the same size or larger than the fascicle of interest, even if they do not fall between the stimulating contact and the fascicle of interest, the effect on the percentage of axons that propagate an action potential in a target fascicle can be nontrivial. Subdividing a nerve model or excluding certain fascicles may yield inaccurate predictions of fascicle activation levels if the neighboring fascicle effects are not taken into account. Modeling the physiologic nerve anatomy would be the most accurate way to account for the effects of each fascicle on its neighbors.

D. Realistic Geometry Based on Cadaveric Nerve

Neural anatomy directly impacts clinical peripheral nerve stimulation, especially where the desired outcome is selective activation of a specific fascicle or a group of fascicles. Our results showed that axonal thresholds are affected by the D_f , th_p , and position of not only the target fascicle but also of its neighboring fascicles. Demonstration of these effects in a multifascicular model of a human nerve specimen (Fig. 2) being used to design and optimize clinical electrodes to restore standing and walking [17] shows the relevance of these findings.

Simulations of the realistic nerve model (Fig. 2) showed that increasing the th_p resulted in decreased recruitment of the target fascicles, as predicted by the idealized model in Fig. 4.

The fascicle diameter effects, shown in Fig. 5, were also seen in the realistic model, where the large fascicle required higher stimulus amplitudes for activation than the small fascicle (Fig. 9). However, due to the geometry of the nerve cross section, more axons in the large fascicle were located further from the electrode, which could also explain the lower levels of activation of the large fascicle as compared to the small fascicle. Additionally, the percent activation of the small fascicle (fascicle 1) increased due to the addition of just one large neighboring fascicle (fascicle 2) and further increased due to the addition of the remaining fascicles in the nerve cross section, reaffirming the trends seen in idealized models (Figs. 6–8).

In clinical practice, selective stimulation of fascicles with different functions may be desired. Based on the D_f , th_p , and neighboring fascicle effects, selective stimulation of a large fascicle that has a small neighbor becomes more challenging, while selective activation of a small fascicle with large neighbor becomes easier. In the specific fascicle pair used in this study, the percent activation of the small fascicle increased by as much as $17\% \pm 6\%$ due to the presence of neighboring fascicles with physiologic th_p . This change could have significant clinical impact depending on the functions of each fascicle and on the application of the neuroprosthesis.

E. Sensitivity Analyses

The conductivity of mammalian perineurium is currently unknown. A sensitivity analysis on the effects of perineurial conductivity showed that predicted axonal activation thresholds varied with perineurial conductivity but the overall trends in the effects of neighboring fascicles did not. This suggests that the effects of neighboring fascicles observed in this study are robust.

The presence of epineurium did not affect the trends in the results observed in this study. The realistic nerve model in Fig. 2 included epineurial tissue around the fascicles. Although the magnitudes of the effects of D_f and th_p changed with the presence of the epineurium, the trends observed in the saline models carried over to this realistic model containing epineurium.

Based on further sensitivity analyses, the electrode-axon distance did not appear to affect the trends shown in this study. Axonal activation thresholds increase with an increase in electrode-axon distance [37]. The electrode-axon distance was varied in these models by moving the fascicle and the axon relative to the electrode. As expected, the effects of th_p and of D_f on axon thresholds reported in our study decreased with an increase in electrode-axon distance, however overall trends remained unchanged (data not shown). This suggests that if the model contains an electrode sufficiently far away from the nerve, the th_p and D_f of the fascicle of interest may not have as significant of an impact on the model results as compared to when the electrode is within close proximity of the fascicle. However, in neural prostheses that use nerve cuff electrodes, the electrode-axon distance is limited by the dimensions of the cuff. Due to the proximity of the cuff to the nerve, the th_p and D_f on the model prediction of excitation. Therefore, th_p and D_f should be included at their physiological sizes for computer models of nerve cuff electrodes.

V. Conclusion

Neural anatomy significantly affects model predictions of activation thresholds and recruitment levels during nerve stimulation. Increased perineurial thickness and fascicle diameter result in predictions of increased stimulation thresholds of peripheral nerve axons. The fascicular diameter, perineurial thickness, and relative position of one fascicle affect the

recruitment levels of its neighboring fascicle in models of neural stimulation. These effects result in smaller fascicles being recruited first and therefore being easier to selectively activate. Since perineurial thickness is dependent on fascicle diameter and fascicle size varies in humans, physiologic values and accurate representations of the nerve of interest should be used to develop computer models to evaluate and design electrodes for neural prosthesis applications. Improved accuracy of modeling results has the potential to improve neural prostheses designs and ultimately the predictions of clinical efficacy of neural prostheses.

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References

- Peckham PH, Knutson JS. Functional electrical stimulation for neuromuscular applications. Annu. Rev. Biomed. Eng 2005;vol. 7:327–360. [PubMed: 16004574]
- [2]. Leventhal DK, Durand DM. Subfascicle stimulation selectivity with the flat interface nerve electrode. Ann. Biomed. Eng 2003;vol. 31:643–652. [PubMed: 12797613]
- [3]. Tyler DJ, Durand DM. Functionally selective peripheral nerve stimulation with a flat interface nerve electrode. IEEE Trans. Neural Syst. Rehabil. Eng Dec.;2002 vol. 10(no. 4):294–303.
 [PubMed: 12611367]
- [4]. Tarler MD, Mortimer JT. Selective and independent activation of four motor fascicles using a four contact nerve-cuff electrode. IEEE Trans. Neural Syst. Rehabil. Eng Jun.;2004 vol. 12(no. 2): 251–257. [PubMed: 15218938]
- [5]. Binnie CD. Vagus nerve stimulation for epilepsy: A review. Seizure 2000;vol. 9:161–169. [PubMed: 10775511]
- [6]. Hansen M, Haugland M, Sinkjaer T, Donaldson N. Real time foot drop correction using machine learning and natural sensors. Neuromodulation 2002;vol. 5:41–53.
- [7]. Polasek KH, Hoyen HA, Keith MW, Tyler DJ. Human nerve stimulation thresholds and selectivity using a multi-contact nerve cuff electrode. IEEE Trans. Neural Syst. Rehabil. Eng Mar.;2007 vol. 15(no. 1):76–82. [PubMed: 17436879]
- [8]. Creasey G, Elefteriades J, DiMarco A, Talonen P, Bijak M, Girsch W, Kantor C. Electrical stimulation to restore respiration. J. Rehabil. Res. Develop 1996;vol. 33:123–132.
- [9]. Deurloo KE, Holsheimer J, Bergveld P. Fascicular selectivity in transverse stimulation with a nerve cuff electrode: A theoretical approach. Neuromodulation 2003;vol. 6:258–269.
- [10]. Veltink PH, van Veen BK, Struijk JJ, Holsheimer J, Boom HB. A modeling study of nerve fascicle stimulation. IEEE Trans. Biomed. Eng Jul.;1989 vol. 36(no. 7):683–692. [PubMed: 2744792]
- [11]. Koole P, Holsheimer J, Struijk JJ, Verloop AJ. Recruitment characteristics of nerve fascicles stimulated by a multigroove electrode. IEEE Trans. Rehabil. Eng Mar.;1997 vol. 5(no. 1):40–50.
 [PubMed: 9086384]

- [12]. Choi AQ, Cavanaugh JK, Durand DM. Selectivity of multiple-contact nerve cuff electrodes: A simulation analysis. IEEE Trans. Biomed. Eng Feb.;2001 vol. 48(no. 2):165–172. [PubMed: 11296872]
- [13]. Meier JH, Rutten WL, Zoutman AE, Boom HB, Bergveld P. Simulation of multipolar fiber selective neural stimulation using in-trafascicular electrodes. IEEE Trans. Biomed. Eng Feb.; 1992 vol. 39(no. 2):122–134. [PubMed: 1612615]
- [14]. Lertmanorat Z, Durand DM. Extracellular voltage profile for reversing the recruitment order of peripheral nerve stimulation: A simulation study. J. Neural Eng 2004;vol. 1:202–211. [PubMed: 15876640]
- [15]. Hennings K, Arendt-Nielsen L, Christensen SS, Andersen OK. Selective activation of smalldiameter motor fibres using exponentially rising waveforms: A theoretical study. Med. Biol. Eng. Comput 2005;vol. 43:493–500. [PubMed: 16255432]
- [16]. Deurloo KE, Holsheimer J, Bergveld P. The effect of sub-threshold prepulses on the recruitment order in a nerve trunk analyzed in a simple and a realistic volume conductor model. Biol. Cybern 2001;vol. 85:281–291. [PubMed: 11592625]
- [17]. Schiefer MA, Triolo RJ, Tyler DJ. A model of selective activation of the femoral nerve with a flat interface nerve electrode for a lower extremity neuroprosthesis. IEEE Trans. Neural Syst. Rehabil. Eng Apr.;2008 vol. 16(no. 2):195–204. [PubMed: 18403289]
- [18]. Meier JH, Rutten WL, Boom HB. Extracellular potentials from active myelinated fibers inside insulated and noninsulated peripheral nerve. IEEE Trans. Biomed. Eng Sep.;1998 vol. 45(no. 9): 1146–1153. [PubMed: 9735564]
- [19]. Perez-Orive J, Durand DM. Modeling study of peripheral nerve recording selectivity. IEEE Trans. Rehabil. Eng Sep.;2000 vol. 8(no. 3):320–329. [PubMed: 11001512]
- [20]. Tesfayesus W, Yoo P, Moffitt M, Durand DM. Blind source separation of nerve cuff recordings. Proc. IEEE Eng. Med. Biol. Soc. Conf Sep.;2004 vol. 1:95–98.
- [21]. Yoo PB, Durand DM. Selective recording of the canine hypoglossal nerve using a multicontact flat interface nerve electrode. IEEE Trans. Biomed. Eng Aug.;2005 vol. 52(no. 8):1461–1469.
 [PubMed: 16119242]
- [22]. Rahal M, Taylor J, Donaldson N. The effect of nerve cuff geometry on interference reduction: A study by computer modeling. IEEE Trans. Biomed. Eng Jan.;2000 vol. 47(no. 1):136–138. [PubMed: 10646289]
- [23]. Sunderland S, Bradley KC. The perineurium of peripheral nerves. Anat. Rec 1952;vol. 113:125– 141. [PubMed: 14933806]
- [24]. Gustafson, KJ., et al. Fascicular anatomy of the human femoral nerve: Implications for standing neural prostheses utilizing nerve cuff electrodes. Proc. 34th Annu. NIH Neural Prosthesis Workshop; 2003;
- [25]. Stewart JD. Peripheral nerve fascicles: Anatomy and clinical relevance. Muscle Nerve 2003;vol. 28:525–541. [PubMed: 14571454]
- [26]. Gustafson KJ, Zelkovic PF, Feng AH, Draper CE, Bodner DR, Grill WM. Fascicular anatomy and surgical access of the human pudendal nerve. World J. Urol 2005;vol. 23:411–418. [PubMed: 16333625]
- [27]. Gustafson, KJ.; Artis, E. A comparative analysis of fundamental nerve fascicular anatomy: Physiological limits and implications for nerve cuff electrode design. presented at the NIH NINDS Neural Interfaces Workshop; Bethesda, MD. 2005;
- [28]. Burkel WE. The histological fine structure of perineurium. Anat. Rec 1967;vol. 158:177–189.[PubMed: 6039587]
- [29]. Lowry A, Wilcox D, Masson EA, Williams PE. Immunohis-tochemical methods for semiquantitative analysis of collagen content in human peripheral nerve. J. Anat 1997;vol. 191(pt. 3):367–374. [PubMed: 9418993]
- [30]. Szlavik RB, de Bruin H. The effect of anisotropy on the potential distribution in biological tissue and its impact on nerve excitation simulations. IEEE Trans. Biomed. Eng Sep.;2000 vol. 47(no. 9):1202–1210. [PubMed: 11008421]
- [31]. Garven HS, Gairns FW, Smith G. The nerve fibre populations of the nerves of the leg in chronic occlusive arterial disease in man. Scott. Med. J 1962;vol. 7:250–265. [PubMed: 13897116]

- [32]. Behse F. Morphometric studies on the human sural nerve. Acta Neurol. Scand. Suppl 1990;vol. 132:1–38. [PubMed: 2239145]
- [33]. McIntyre CC, Richardson AG, Grill WM. Modeling the excitability of mammalian nerve fibers: Influence of afterpotentials on the recovery cycle. J. Neurophysiol 2002;vol. 87:995–1006. [PubMed: 11826063]
- [34]. Richardson AG, McIntyre CC, Grill WM. Modelling the effects of electric fields on nerve fibres: Influence of the myelin sheath. Med. Biol. Eng. Comput 2000;vol. 38:438–446. [PubMed: 10984943]
- [35]. Vuckovic A, Struijk JJ, Rijkhoff NJ. Influence of variable nerve fibre geometry on the excitation and blocking threshold. A simulation study. Med. Biol. Eng. Comput 2005;vol. 43:365–374. [PubMed: 16035225]
- [36]. Goodall EV, Kosterman LM, Holsheimer J, Struijk JJ. Modeling study of activation and propagation delays during stimulation of peripheral nerve fibers with a tripolar cuff electrode. IEEE Trans. Rehabil. Eng Sep.;1995 vol. 3:272–282.
- [37]. Grill WM Jr. Modeling the effects of electric fields on nerve fibers: Influence of tissue electrical properties. IEEE Trans. Biomed. Eng Aug.;1999 vol. 46(no. 8):918–928. [PubMed: 10431456]



Fig. 1.

FEMs used to examine effects of fascicle perineurial thickness, diameter, and position on model predictions of axonal thresholds and recruitment. The fascicle, perineurium, and electrode were encased in a saline volume in all models shown.



Fig. 2.

Model of a femoral nerve cross section inside a nerve cuff electrode adapted from [17], [24]. Fascicles 1 and 2 were chosen to investigate the effects of perineurium thickness, fascicle size, and neighboring fascicle effects on fascicular activation. Perineurial thickness was varied between 0% and 3% of fascicle diameter, $30 \,\mu\text{m}$, and $50 \,\mu\text{m}$. Fascicles were surrounded by epineurium which was encased in saline and stimulating electrode was placed in an insulating cuff.



Fig. 3.

Physiologic thickness of human perineurium versus the fascicle diameter for individual fascicles. The shaded area represents the relationship between human perineurial thickness and fascicle diameter of $3.0\% \pm 1.0\%$, based on the raw data reported by Sunderland and Bradley [23]. The values used in previous computer modeling studies are presented for comparison: \bullet Deurloo *et al.* [9], \blacktriangle Choi *et al.* [12], \blacksquare Hennings *et al.* [15], \blacksquare Perez-Orive and Durand [19], \blacktriangle Rahal *et al.* [22], and \diamondsuit Szlavik and de Bruin [30].



Fig. 4.

Axonal threshold increased with increasing perineurial thickness and decreasing axon diameter. The axon was positioned at the center of a 500- μ m-diameter fascicle. This data was obtained at 0.075 ms pulse width; however, trends were consistent for pulse widths ranging from 0.025–0.5 ms.



Fig. 5.

Cathodic activation thresholds increased with fascicle diameter when perineurial thickness was physiologic (3%, solid line). Thresholds increased with increasing perineurial thickness for each tested diameter. Not including perineurium resulted in under-prediction of thresholds. Trends shown in this plot of thresholds for a 10- μ m-diameter axon stimulated for 0.025 ms were consistent over all simulated pulse widths (0.025–0.5 ms) and axon diameters (4–15 μ m).



Fig. 6.

Fascicle A activation increased as fascicle B diameter and perineurial thickness increased. Increase in activation was computed by subtracting the activation of fascicle A alone (500 μ m diameter, 15 μ m perineurium thickness) from activation of fascicle A with fascicle B. Fascicle B diameter was varied between 100–1000 μ m and its perineurial thickness (*th_p*) was varied between 3–30 μ m. The percent activation of fascicle A when no neighbor was present was 15% ± 7% and increased to as high as 47% ± 6% when a neighboring fascicle was added to the model (0.1 ms, -1 mA stimulation). The increase in activation of fascicle A due to the addition of a neighboring fascicle was significant for fascicle B diameters > 300 μ m, at all perineurium thicknesses (*p* < 0.005).



Fig. 7.

Decreased distance between the fascicle perineurium edges resulted in increased effect of the neighboring fascicle on activation of the control fascicle. Control fascicle A had a constant diameter of 500 μ m, while neighboring fascicle B diameter was varied between 100–1000 μ m. The perineurium thickness was modeled as 3% of the fascicle diameter. The addition of a 500 or 1000 μ m diameter fascicle B resulted in a significant increase in activation of fascicle A for all distances 700 μ m (p < 0.004, Student's *t*-test, as compared to 0% increase in fascicle A activation). The shaded region represents the physiologic range of distances (28–174 μ m) between neighboring fascicles in the human femoral nerve cross section.



Fig. 8.

When the neighboring fascicle was moved around the control fascicle, the location and the diameter of the neighbor changed the percent activation of the control fascicle. The amount of change was computed by subtracting the percent activation of fascicle A alone from fascicle A with a neighboring fascicle present (standard deviation of the change in activation shown with the shaded region around each curve). The neighbor effects were significant at all locations except at $\alpha = 50^{\circ}$ for a 1000- μ m neighbor and at $50^{\circ} < \alpha < 65^{\circ}$ for a 500- μ m neighbor. The effects of a 100- μ m-diameter neighbor were negligible.



Fig. 9.

Fascicle diameter and the addition of neighboring fascicles significantly changed the activation of fascicles 1 and 2 in the realistic nerve model shown in Fig. 2. The percent activation of fascicles 1 and 2 alone, with neighbor, and with all fascicles in the cross section were compared at two recruitment levels (a) $20 \ \mu$ s and (b) $30 \ \mu$ s pulse widths and 0.1 mA stimulation amplitude. The smaller fascicle 1 (ABC) was recruited first and was significantly more activated than the larger fascicle 2 (DEF), at all perineurium thicknesses (b). With perineurium included, the addition of neighboring fascicles significantly changed the activation of the small fascicle 1 but not the large fascicle 2. The addition of perineurium to the model containing all fascicles resulted in significant changes in activation of fascicle 1 (a).

TABLE I

Conductivities Used for Modeling [12]

Material	Conductivity $(\Omega^{-1} \cdot m^{-1})$
Perineurium	0.0021
Endoneurium	
longitudinal	0.57
transverse	0.083
Epineurium	0.083
Saline	2.0

TABLE II

Relationship Between th_p and D_f Categorized by Nerve Type

	Total	Sciatic	Medial Popliteal	Lateral Popliteal	Ulnar	Radial	Median
th_p as % of $D_f(\mu \pm \sigma)$	3.0 ± 1.0	3.3 ± 0.7	2.6 ± 0.4	2.8 ± 0.6	2.6 ± 0.8	3.0 ± 0.9	3.3 ± 1.3
# specimens	20	1	1	1	4	2	11
# fasciles	704	32	51	39	107	189	286