

Published in final edited form as:

Brain Res. 2010 September 2; 1350: 77–85. doi:10.1016/j.brainres.2010.04.056.

Metabolic Impact Of Sex Hormones On Obesity

Lynda M. Brown¹, Lana Gent², Kathryn Davis², and Deborah J. Clegg^{2,*}

¹Department of Nutrition, University of North Carolina at Greensboro, Greensboro, NC 27412

²Department of Internal Medicine, Touchstone Diabetes Center, University of Texas Southwestern Medical Center, Dallas, TX 75390-8854

Abstract

Obesity and its associated health disorders and costs are increasing. Men and postmenopausal women have greater risk of developing complications of obesity than younger women. Within the brain, the hypothalamus is an important regulator of energy homeostasis. Two of its sub-areas, the ventrolateral portion of the ventral medial nucleus (VL VMN) and the arcuate (ARC) respond to hormones and other signals to control energy intake and expenditure. When large lesions are made in the hypothalamus which includes both the VL VMN and the ARC, animals eat more, have reduced energy expenditure, and become obese. The ARC and the VL VMN, in addition to other regions in the hypothalamus, have been demonstrated to contain estrogen receptors. There are two estrogen receptors, estrogen receptor alpha (ER α) and estrogen receptor beta (ER β). We and others have previously demonstrated that activation of ER α by estrogens reduces food intake and increases body weight. This review focuses on the relative contribution of activation of ER α by estrogens in the ARC and the VL VMN in the regulation of food intake and body weight. Additionally, estrogen receptors have been found in many peripheral tissues including adipose tissue. Estrogens are thought to have direct effects on adipose tissue and estrogens may provide anti-inflammatory properties both in the periphery and the in the central nervous system (CNS) which may protect women from diseases associated with inflammation. Understanding the mechanisms by which estrogens regulate body weight and inflammation will assist in determining potential therapeutic agents for menopausal women to decrease the propensity of diseases associated with obesity.

Keywords

body weight regulation; hypothalamus; inflammation; estrogens; estrogen receptor alpha (ER α); neuropeptides

Introduction

Obesity, resulting from an imbalance between energy intake and expenditure, is now recognized as a serious and global health problem (Steinbaum, 2004; Berthoud and Morrison, 2008). More than 300 million adults worldwide are obese, and in the United

© 2010 Elsevier B.V. All rights reserved

***Corresponding Author:** Correspondence should be addressed to Deborah J. Clegg: Department of Internal Medicine Touchstone Diabetes Center UT Southwestern Medical Center 5323 Harry Hines Blvd., K5.252 Dallas, TX 75390-8854 Phone 214-648-3401 Fax 214-648-8720 deborah.clegg@utsouthwestern.edu.

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

States, over 30% of the population are considered obese and 60% are overweight (Wang et al., 2008). Obese individuals are at an increased risk for developing type II diabetes, hypertension, coronary heart disease and stroke. In the United States obesity is associated with 300,000 premature deaths every year (Allison et al., 1999). The past decade has been attempting to identifying key hormones and other signals that influence body weight.

Males and females differ in terms of how and where they store body fat, the hormones they secrete in proportion to their fat, and the way their brains respond to signals that regulate food intake and body weight (Clegg et al., 2003; Clegg et al., 2006). The focus of this review is to discuss the potential role of estrogens as determinants mediating the sexual dimorphism in body weight regulation. In women, estrogens may be responsible for body weight homeostasis. Reductions in circulating estrogens, which occurs following ovariectomy (OVX) in experimental rodent models, results in increased body adiposity which can be ameliorated by exogenous estradiol-17 β administration (Clegg et al., 2003; Clegg et al., 2006). In women, menopause is has been associated with increased in adiposity and shifts in body fat distribution, which can be altered by exogenous hormone replacement therapy (Samaras et al., 1999; Ryan et al., 2002).

Hypothalamic Control of Body Weight

In an effort to identify targets for therapeutic treatments to reduce food intake and body weight, obesity research has focused on central metabolic and hormonal pathways that control energy homeostasis (Obici, 2009). The hypothalamus is one area within the CNS that has received attention as a region in the brain that controls food intake, energy expenditure and body weight homeostasis. Since the 1930's, several thousand reports have been published investigating the potential role of various hypothalamic nuclei in the regulation of food intake and body weight (York and Bray, 1972; Bray et al., 1982; Bray, 1984). Early interest in the hypothalamus stemmed from findings that dramatic changes in food intake, body weight, and energy homeostasis could be produced by lesioning specific hypothalamic nuclei, such as the ventral medial hypothalamus (VMH) (Hetherington and Ranson, 1940; Rowland et al., 1975; Louis-Sylvestre et al., 1980) or the lateral hypothalamic area (LH) (Anand and Brobeck, 1951; Danguir and Nicolaidis, 1980; Milam et al., 1980). More recently, areas in the hypothalamus such as the ventral medial nucleus (VMN) and arcuate (ARC) nucleus (among others) have been identified as targets for hormonal and neuropeptide signals that can produce marked in energy homeostasis.

Estrogen Receptors and Body Weight Regulation

The classical nuclear estrogen receptor (ER) was cloned in 1985 (Green et al., 1986) and renamed estrogen receptor alpha (ER α) when a second nuclear estrogen receptor (estrogen receptor beta (ER β)), was discovered 10 years later (Kuiper et al., 1996). Both ERs are also expressed in the brain (Simerly et al., 1990; Kuiper et al., 1996; Shughrue et al., 1997; Simonian and Herbison, 1997; Voisin et al., 1997; Osterlund et al., 1998; Mitra et al., 2003; Merchantaler et al., 2004). Stimulation of ERs engages both genomic and non-genomic events that may mediate distinct functions of estrogens. Little is known about the critical intracellular signals that mediate the anorexigenic effects of estrogens. Genomic activation of ERs results from estrogens binding to estrogen response elements (EREs) in promoter regions of target genes (Pappas et al., 1995; Razandi et al., 1999). Additionally, estrogens can also initiate rapid signaling events within minutes via non-genomic mechanisms (Kelly and Levin, 2001). ERs in the plasma membrane are able to activate the MAP-kinase cascade and PI3K pathway, causing a rise in intracellular calcium (Balthazart et al., 2001; Sutter-Dub, 2002). ERs also activate protein kinase B (PKB/Akt) in neurons (Singh, 2001; Ivanova et al., 2002; Wilson et al., 2002). The PI3K/Akt cascade mediates a variety of estrogens'

central actions, including neuronal excitability, neuroprotection, reductions in inflammation, and neurite outgrowth (Vasudevan and Pfaff, 2008), and potentially body weight regulation. Estrogens' activation of ERs results in a range of biological effects which will be described in more detail below.

Male and female mice with a targeted deletion in the ER α gene (ER α KO) have increased adiposity (Heine et al., 2000). The cause of the obesity is unclear; however, overeating does not appear to be essential (Shughrue et al., 1997; Shughrue et al., 1997; Heine et al., 2000; Ohlsson et al., 2000; Vidal et al., 2000; Geary et al., 2001), and the increased adiposity in ER α KO mice is attributed primarily to decreased energy expenditure (Heine et al., 2000; Ohlsson et al., 2000). While estradiol-17 β replacement prevents OVX wildtype mice from developing hyperphagia and obesity, such protective effects are blocked in OVX ER α KO mice (Geary et al., 2001).

Additionally, both estrogen receptors (ERs) are expressed in peripheral tissues, such as adipose tissue, where they may influence adiposity and inflammation. Further supporting a role for ER α in the regulation of body weight, are the findings that increased visceral adiposity has been associated with the XbaI polymorphism of the human ER α gene, in which guanine is substituted for adenine in exon one (Speer et al., 2001; Yamada et al., 2002; Okura et al., 2003). A study of over two thousand Japanese women reported that there was increased fat mass and waist-hip ratios (an index of visceral adiposity) in premenopausal women that had the polymorphism compared to the control cohort of women with the normal genotype (Yamada et al., 2002; Okura et al., 2003; Okura et al., 2003). The polymorphism did not affect adiposity in postmenopausal women or in men. Thus, polymorphisms of the human ER α gene may impair estrogen signaling and lead to increased visceral adiposity and its attendant health risks.

CNS ERs and Body Weight Regulation

Due to the findings that targeted deletion in the ER α gene (ER α KO) have increased adiposity (Heine et al., 2000), ER α may be important in the regulation of body weight. ER α is ubiquitously expressed in rodent brains, but the physiologically relevant sites of ER α in the regulation of food intake and energy expenditure have not been identified. ER α is expressed in the ventrolateral portion of the VMN, the ARC, the medial preoptic area (MPOA), and the paraventricular nuclei (PVN) (Simerly et al., 1990; Simonian and Herbison, 1997; Voisin et al., 1997; Osterlund et al., 1998; Wilkinson et al., 2002; Shima et al., 2003; Merchenthaler et al., 2004). ER β is found in the same hypothalamic nuclei as ER α , but ER β expression is significantly reduced relative to ER α .

In earlier attempts to determine the effects of estrogens on food intake and body weight in hypothalamic regions, intranuclear microinjections of estradiol benzoate were performed (Wade and Zucker 1970). However, due to the difficulty in precisely placing cannulae or producing lesions in small but complex hypothalamic regions, findings obtained from these studies are somewhat controversial. For example, earlier studies showed that estradiol-17 β implanted in the PVN decreases food intake and body weight in OVX rats (Butera and Beikirch, 1989) in the absence of any signs of peripheral estrogenic stimulation. Additionally, the anorexigenic effects of subcutaneous estradiol-17 β are blunted in rats with PVN lesions (Butera et al., 1992). However, the subsequent studies failed to reproduce these phenotypes in rats with PVN implants (Hrupka et al., 2002). Effects of estrogen in the MPOA are also controversial, with only one report showing an anorexigenic response of estradiol-17 β (Dagnault and Richard, 1997), whereas several other groups showed that estradiol-17 β implanted in this nucleus has no effect on feeding (Butera and Beikirch, 1989). The ARC and VMH are nearby structures in the ventral hypothalamus. Earlier

microinjection studies may not have been able to rigorously distinguish these two regions, and therefore failed to provide consistent results (Butera and Beikirch, 1989). However, we have reported that site-specific reductions of ER α in the VL VMN result in increased adiposity, no change in food intake, and suppression of energy expenditure, implicating VL VMN ER α in energy homeostasis (Musatov et al., 2007). Additionally, the hindbrain (caudal NTS) might be another site of estrogens' regulation of food intake due to the recent findings of Thammacharoen where they placed an estradiol-containing haemostatic cloth which decreased food intake in OVX rats (Thammacharoen et al., 2008).

Following OVX, the oft-observed consequence is that a rat becomes transiently hyperphagic and gains weight, mainly as fat (Wade, 1972; Drewett, 1973; Leshner and Collier, 1973; Blaustein and Wade, 1976; Wade and Gray, 1979; Wade et al., 1985; Kemnitz et al., 1989; Wallen et al., 2001). However, an important but often forgotten aspect of OVX is that there is not an obligatory increase of food intake. OVX rats that are pair-fed to estradiol-17 β -treated rats, however, increase body weight, gaining as much as *ad libitum*-fed OVX rats (Mueller and Hsiao, 1980). Additionally, if a female rat is fattened prior to removal of the ovaries, she simply remains fat after the operation without the necessity of overeating; i.e., the OVX (and presumably lack of estrogens) dictated that the rat carry more fat, but did not dictate that the rat become hyperphagic (Bartness and Waldbillig, 1984).

Estrogens Interact with Leptin

First described in 1994 (Zhang et al., 1994), leptin has proven to be a key metabolic protein with actions throughout the body. Leptin provides a powerful catabolic signal to the brain, resulting in inhibition of food intake and increasing energy expenditure (Ahima et al., 1999; Elias et al., 1999; Elmquist et al., 1999; Schwartz et al., 2000; Woods et al., 2000; Morton et al., 2003; Seeley and Woods, 2003; Balthasar et al., 2004; Schwartz and Porte, 2005). Leptin is secreted from adipose tissue in direct proportion to fat content, and it crosses the blood-brain barrier to interact with leptin receptors in the hypothalamus and brainstem (Tartaglia et al., 1995; Ahima et al., 1996; Seeley et al., 1996; Ahima et al., 1999; Elias et al., 1999; Morton et al., 2003; Schwartz and Porte, 2005). Although there are several splice variants of the leptin receptor, the long form (termed *leprb*) is the critical variant for regulating energy balance (Chen et al., 1996). *Leprb* are localized in several brain areas including the VMN and the ARC, and *leprb* are colocalized with several neuropeptides believed to be involved in controlling food intake and reproduction (Van Dijk et al., 1996; Elmquist et al., 1997; Elmquist et al., 1998). Leptin has the ability to activate or inhibit hypothalamic neurons (Elmquist et al., 1998; Elmquist et al., 1998; Elmquist et al., 1998). Thus, leptin is ideally situated to link metabolic status and brain function. It has been reported that *leprb* expression in the ARC is co-localized with ER α (Diano et al., 1998), and estrogens have been reported to regulate the expression of *leprb* mRNA in the ARC (Bennett et al., 1999), possibly via an ERE on the leptin receptor gene (Lindell et al., 2001). The extensive hypothalamic co-localization of these two receptors suggests a closely coupled interaction between these peripheral signals in the regulation of energy homeostasis.

Leptin levels are higher in females, even before puberty, compared with males, and these levels are independent of differences in body composition (Shimizu et al., 1997; Demerath et al., 1999; Wu-Peng et al., 1999). After puberty, estrogens and testosterone further modulate leptin synthesis and secretion via sex steroid receptor-dependent transcriptional mechanisms (Machinal et al., 1999).

Estrogens may be modulators of leptin's catabolic action in the brain. Higher levels of estrogens have been associated with increased leptin sensitivity (Ainslie et al., 2001; Clegg et al., 2003; Clegg et al., 2006). Although circulating leptin levels do not change appreciably

during the estrous cycle, ARC *leprb* expression is highest during estrous and metestrous and is inversely correlated with neuropeptide Y (NPY) mRNA expression (Bennett et al., 1999). OVX lowers sensitivity to central leptin when compared to intact females, and this can be restored by estradiol-17 β replacement (Clegg et al., 2006). Analogously, exogenous estradiol-17 β administration to male rats will increase sensitivity to central leptin (Clegg et al., 2006). The differences in leptin sensitivity caused by the presence or absence of estrogens may occur downstream of *leprb* transcription and translation. Additionally, females have greater c-Fos (a marker of neuronal activation) and pSTAT3 (a marker of leptin receptor activation) immunoreactivity in the ARC than males following intra-third ventricular leptin (unpublished data), which suggests that enhanced leptin sensitivity. Therefore, there may be a threshold for estrogens required to enhance the central sensitivity to leptin.

It should be noted that OVX greatly reduces, but does not eliminate the most physiologically active estrogen, estradiol-17 β , since adrenal androgens and other steroids can be aromatized to estrogens in fat and other tissue. Additionally, there is a second form of estradiol, estradiol-17 α , which can be synthesized in peripheral tissue (Finkelstein et al., 1981) and in some brain areas (MacLusky et al., 1994) from aromatization of androstenedione by cytochrome P450 aromatase. Recent reports demonstrated that estradiol-17 α has binding affinity for the ER α receptor and that levels increase in the hypothalamus of OVX females (Toran-Allerand, 2005; Toran-Allerand et al., 2005). Peripheral and central administration estradiol-17 α decreases food intake and body weight of OVX rats (Donohoe et al., 1984; Butera et al., 1990). These findings suggest that estradiol-17 α may modulate estrogen regulation of energy balance and, further, that even following OVX, estradiol-17 α may activate ER α .

Leptin resistance may also occur in other states such as pregnancy (Ladyman and Grattan, 2005). These authors demonstrated that pregnant rats treated with leptin had impaired activation of pSTAT3 and reduced *leprb* mRNA in the VMH when compared to non-pregnant females. However, there were no changes in leptin activation of pSTAT3 in the ARC of pregnant females, nor were there changes in *leprb* in pregnant versus non-pregnant females. Pregnancy is a state of relative deficiency of estrogens, and these data may support the idea that low levels of estrogens is associated with reduced leptin sensitivity.

Estrogen interacts with the Melanocortin System

The ARC has been well-investigated with regard to central leptin action (Woods and Seeley, 2000; Williams et al., 2001). *Leprb* is expressed by two populations of ARC neurons, those expressing proopiomelanocortin (POMC) (Cheung et al., 1997; Thornton et al., 1997) and those expressing NPY and agouti-related peptide (NPY/AgRP neurons) (Baskin et al., 1999). NPY is an effective anabolic peptide. Central administration of NPY potently increases food intake and decreases energy expenditure and fat oxidation (Chavez et al., 1995; Levin, 1999; Cone et al., 2001; Herzog, 2003). ARC neurons co-express NPY mRNA and *leprb* protein. Leptin administration decreases, while lack of leptin or leptin signaling increases, NPY (and AgRP) mRNA, demonstrating that leptin is a critical determinant of ARC NPY functioning (Baskin et al., 1999). POMC neurons secrete the catabolic melanocortin neuropeptide, α melanocyte stimulating hormone (α MSH), which acts in the PVN and lateral hypothalamus on melanocortin 3 and melanocortin 4 (MC3/MC4) receptors to produce a profound catabolic effect by reducing food intake and increasing energy expenditure (Elmqvist et al., 1998; Elias et al., 1999; Elmquist et al., 1999; Elias et al., 2000). If administered chronically, MC3/4 agonists reduce body weight and adiposity (Pierroz et al., 2002). Leptin stimulates POMC neurons to synthesize and release α MSH (Seeley et al., 1997; Korner et al., 1999). AgRP is an antagonist at MC3/MC4 receptors, and

its administration increases food intake. Hence, within the ARC, leptin elicits a powerful catabolic affect by activating α MSH and simultaneously inhibiting anabolic NPY/AgRP production and release. This produces reduced feeding and increased energy expenditure (Elmquist et al., 1999). Recently, NPY neurons in the ARC were not found to co-express ER α (Olofsson et al., 2009); whereas we and others have found that POMC neurons do co-express ER α (Simonian et al., 1999). Because the *leprb* gene contains an ERE (Lindell et al., 2001), this provides a critical potential mechanistic link between estrogens' activity and that of leptin on ARC POMC neurons.

A wealth of data support a role for NPY in the regulation of food intake and body weight, e.g., (Herzog, 2003). Consistent with a role of estrogens in the regulation of food intake and body weight, estrogen suppresses NPY release. OVX increases and estradiol-17 β treatment decreases levels of NPY mRNA in the ARC (Baskin et al., 1995; Ainslie et al., 2001). Additionally, food deprivation increases ARC NPY mRNA levels less in estradiol-17 β treated OVX rats than in vehicle-treated OVX rats (Baskin et al., 1995). Lastly, chronic estradiol-17 β treatment decreases NPY levels and NPY release in the PVN (Bonavera et al., 1994).

POMC levels are also responsive to gonadal steroids. POMC mRNA levels fluctuate over the course of the estrous cycle, with the most dramatic changes on the day of proestrus (Wise et al., 1990; Bohler et al., 1991; Korner et al., 1999; Slamberova et al., 2004). POMC neurons express ER α , and OVX decreases POMC mRNA, which is reversed by estradiol-17 β replacement (Pelletier et al., 2007). Lower POMC levels are also observed in ER α knockout mice (Hirosawa et al., 2008). Estrogens activate POMC neurons partly via PI3K-mediated mechanisms (Qiu et al., 2003; Malyala et al., 2008). Estradiol-17 β administration rapidly increases activity at incoming excitatory synapses onto POMC neurons, enhancing miniature excitatory postsynaptic current recorded from POMC-GFP neurons (Gao et al., 2007). These synaptic rearrangements in POMC neurons tightly parallel the effects of estrogens on food intake, energy expenditure and body weight (Gao et al., 2007). Collectively, these findings suggest that ER α in POMC neurons may contribute to energy homeostasis and the effects of estrogens.

Estrogen Regulates Adiposity

The accumulation of fat centrally (intra-abdominal/visceral adipose tissue) has emerged as a risk factor for the metabolic syndrome (Kannel et al., 1991; Lee et al., 2009). Estrogens promote the accumulation of subcutaneous fat (Krotkiewski et al., 1983); however, visceral fat varies inversely with levels of estrogens (Bouchard et al., 1993). The loss of estrogens with menopause is associated with an increase in intra-abdominal fat accrual (Poehlman et al., 1995; Lee et al., 2009). OVX rats gain fat, specifically visceral fat with no change of subcutaneous fat (Clegg et al., 2006). Furthermore, peripheral or central administration of estradiol-17 β to OVX rats changes their body fat distribution to mirror that of intact females. Altering the sex hormone milieu in males with estradiol-17 β administration increases subcutaneous fat deposition (Clegg et al., 2006). An important implication from these findings is that estrogens may be critical determinates of body fat distribution.

Progesterone and androgen receptors (PR and AR) as well as ERs are expressed in adipose tissues (Crandall et al., 1998). Subcutaneous adipose tissue has higher concentrations of ERs and PR; however, visceral adipose tissue has higher concentrations of AR (Lu et al., 1998). Additionally, subcutaneous adipose tissue has few AR, and estrogens down-regulates AR expression in subcutaneous fat (Bjorntorp, 1997). Adipose tissue-specific AR knockout mice have increased intra-adipose estrogens, which leads to increased subcutaneous obesity and hyperleptinemia (Yu et al., 2008). The sexual dimorphism in adipose tissue distribution may

partially explain the greater risk for the metabolic syndrome in men compared with premenopausal women.

Estrogen Decreases Inflammation

Obesity is now recognized as a global health problem as previously mentioned, and it may be a result of increased consumption of high fat (HF) diets. Free fatty acids (FFAs), particularly saturated fatty acids, increase inflammation by activating toll-like receptor 4 (TLR4) (Shi et al., 2006). It is increasingly evident that chronic activation of pro-inflammatory pathways may be at least partly responsible for obesity-induced insulin resistance and diabetes (Kahn and Flier, 2000; Wellen and Hotamisligil, 2003; Wellen and Hotamisligil, 2005). For example, the pro-inflammatory cytokines such as tumor necrosis factor-alpha (TNF α), interleukin-6 (IL-6) and C-reactive protein are elevated in individuals with insulin resistance and diabetes (Shoelson et al., 2007; de Luca and Olefsky, 2008; Wang et al., 2008), and elevated in muscle and liver with HF diets (Shi et al., 2006). Suppression of pro-inflammatory responses represents a promising strategy to combat obesity and associated disorders. ERs are expressed in monocytes and macrophages, and estrogens activate these cells (Vegeto et al., 2001; Pozzi et al., 2006). Female rats and mice are relatively protected from HF-induced obesity, insulin resistance and inflammatory responses (Dhar et al., 2004; Gallou-Kabani et al., 2007; Payette et al., 2009). Recent studies have shown that estradiol-17 β may play a role in reducing the inflammatory response in adipose, cardiovascular, and neural systems (Turgeon et al., 2006; hisletti et al., 2005).

In vitro studies have demonstrated estradiol-17 β -activated ER α decreases the number of pro-inflammatory cytokines (Vegeto et al., 2001; Vegeto et al., 2003). The anti-inflammatory properties of estradiol-17 β can be partially explained by the ability of ERs to act as transcriptional repressors by inhibiting the activity of nuclear factor kappa B (NF κ B) through protein-protein interactions between agonist-bound ERs and activated NF κ B subunits (Stein and Yang, 1995; Ghisletti et al., 2005; Kalaitzidis and Gilmore, 2005). Estrogens' inhibitory effect on NF κ B function is not fully understood and may be target selective (Harris et al., 2003; Chadwick et al., 2005; Kalaitzidis and Gilmore, 2005; Wise et al., 2009). The PI3K pathway is also implicated in the anti-inflammatory effects of estrogens. For example, estradiol-17 β blocks LPS-induced NF κ B nuclear translocation in macrophages, an effect that involves the activation of PI3K (Ghisletti et al., 2005). Similarly, estradiol-17 β decreases vascular leukocyte accumulation after an ischemia-reperfusion injury (Simoncini et al., 2000). These effects are blocked by PI3K inhibitors (Simoncini et al., 2000).

Conclusion

When women enter menopause, they have a dramatically increased risk for developing obesity, type II diabetes and the metabolic syndrome. Hormone replacement therapy (HRT) could be seen as a way to reduce these risks and, in fact, recent data suggest that pharmacological estrogens can reverse the progression of metabolic diseases. However, due to ubiquitous expression of ERs, especially in peripheral tissues, and to complex intracellular events coupled to ERs ("genomic" and "non-genomic" actions), the metabolic benefits provided by HRT are often associated with increased risk of heart disease and breast cancer. Obviously, one solution to this dilemma would be to target only the ERs involved in energy balance and to develop estrogen-like drugs that only initiate intracellular events that produce metabolic benefits without unwanted side effects. Therefore, future research should focus on identifying the critical brain sites where ERs regulate body weight homeostasis and delineate the intracellular signaling pathways that are required for estrogens' actions. Additionally, understanding the functional role of ERs in the periphery

may reveal new pharmacological targets for the beneficial actions of therapeutic estrogens without the deleterious side effects.

Acknowledgments

Funding: This work was supported by NIH DK 073689 (DJC).

References

- Ahima RS, Kelly J, Elmquist JK, Flier JS. Distinct physiologic and neuronal responses to decreased leptin and mild hyperleptinemia. *Endocrinology*. 1999; 140:4923–4931. [PubMed: 10537115]
- Ahima RS, Prabakaran D, Mantzoros C, Qu D, Lowell B, Flier-Maratos E, Flier JS. Role of leptin in the neuroendocrine response to fasting. *Nature*. 1996; 382:250–252. [PubMed: 8717038]
- Ainslie DA, Morris MJ, Wittert G, Turnbull H, Proietto J, Thorburn AW. Estrogen deficiency causes central leptin insensitivity and increased hypothalamic neuropeptide Y. *Int J Obes Relat Metab Disord*. 2001; 25:1680–1688. [PubMed: 11753591]
- Allison DB, Fontaine KR, Manson JE, Stevens J, VanItallie TB. Annual deaths attributable to obesity in the United States. *Jama*. 1999; 282:1530–1538. [PubMed: 10546692]
- Anand BK, Brobeck JR. Hypothalamic control of food intake in rats and cats. *Yale Journal of Biology and Medicine*. 1951; 24:123–140. [PubMed: 14901884]
- Balthasar N, Coppari R, McMinn J, Liu SM, Lee CE, Tang V, Kenny CD, McGovern RA, Chua SC Jr, Elmquist JK, Lowell BB. Leptin receptor signaling in POMC neurons is required for normal body weight homeostasis. *Neuron*. 2004; 42:983–991. [PubMed: 15207242]
- Balthazart J, Baillien M, Ball GF. Phosphorylation processes mediate rapid changes of brain aromatase activity. *J Steroid Biochem Mol Biol*. 2001; 79:261–277. [PubMed: 11850233]
- Bartness TJ, Waldbillig RJ. Dietary self-selection in intact, ovariectomized, and estradiol-treated female rats. *Behav Neurosci*. 1984; 98:125–137. [PubMed: 6538090]
- Baskin DG, Breininger JF, Schwartz MW. Leptin receptor mRNA identifies a subpopulation of neuropeptide Y neurons activated by fasting in rat hypothalamus. *Diabetes*. 1999; 48:828–833. [PubMed: 10102700]
- Baskin DG, Norwood BJ, Schwartz MW, Koerker DJ. Estradiol inhibits the increase of hypothalamic neuropeptide Y messenger ribonucleic acid expression induced by weight loss in ovariectomized rats. *Endocrinology*. 1995; 136:5547–5554. [PubMed: 7588307]
- Bennett PA, Lindell K, Wilson C, Carlsson LM, Carlsson B, Robinson IC. Cyclical variations in the abundance of leptin receptors, but not in circulating leptin, correlate with NPY expression during the oestrous cycle. *Neuroendocrinology*. 1999; 69:417–423. [PubMed: 10364693]
- Berthoud HR, Morrison C. The brain, appetite, and obesity. *Annu Rev Psychol*. 2008; 59:55–92. [PubMed: 18154499]
- Bjorntorp P. Hormonal control of regional fat distribution. *Human Reproduction*. 1997; 12(Suppl 1): 21–25. [PubMed: 9403318]
- Blaustein JD, Wade GN. Ovarian influences on the meal patterns of female rats. *Physiol Behav*. 1976; 17:201–208. [PubMed: 1033580]
- Bohler HC Jr, Tracer H, Merriam GR, Petersen SL. Changes in proopiomelanocortin messenger ribonucleic acid levels in the rostral periaqueductal region of the female rat during the estrous cycle. *Endocrinology*. 1991; 128:1265–1269. [PubMed: 1999146]
- Bonavera JJ, Dube MG, Kalra PS, Kalra SP. Anorectic effects of estrogen may be mediated by decreased neuropeptide-Y release in the hypothalamic paraventricular nucleus. *Endocrinology*. 1994; 134:2367–2370. [PubMed: 8194462]
- Bouchard C, Despres JP, Mauriege P. Genetic and nongenetic determinants of regional fat distribution. *Endocr Rev*. 1993; 14:72–93. [PubMed: 8491156]
- Bray GA. Hypothalamic and genetic obesity: an appraisal of the autonomic hypothesis and the endocrine hypothesis. *International Journal of Obesity*. 1984; 8(supplement 1):119–137. [PubMed: 6398803]

- Bray GA, Sclafani A, Novin D. Obesity-inducing hypothalamic knife cuts: effects on lipolysis and blood insulin levels. *Am J Physiol.* 1982; 243:R445–449. [PubMed: 7051866]
- Butera PC, Beikirch RJ. Central implants of diluted estradiol: independent effects on ingestive and reproductive behaviors of ovariectomized rats. *Brain Res.* 1989; 491:266–273. [PubMed: 2765887]
- Butera PC, Beikirch RJ, Willard DM. Changes in ingestive behaviors and body weight following intracranial application of 17 alpha-estradiol. *Physiol Behav.* 1990; 47:1291–1293. [PubMed: 2395935]
- Butera PC, Willard DM, Raymond SA. Effects of PVN lesions on the responsiveness of female rats to estradiol. *Brain Res.* 1992; 576:304–310. [PubMed: 1515922]
- Chadwick CC, Chippari S, Matelan E, Borges-Marcucci L, Eckert AM, Keith JC Jr, Albert LM, Leathurby Y, Harris HA, Bhat RA, Ashwell M, Trybulski E, Winneker RC, Adelman SJ, Steffan RJ, Harnish DC. Identification of pathway-selective estrogen receptor ligands that inhibit NF-kappaB transcriptional activity. *Proc Natl Acad Sci U S A.* 2005; 102:2543–2548. [PubMed: 15699342]
- Chavez M, van Dijk G, Arkies BJ, Woods SC. Third ventricular insulin infusion attenuates NPY-induced feeding at the level of the paraventricular nucleus. *Obesity Research.* 1995; 3:335s.
- Chen H, Charlat O, Tartaglia LA, Woolf EA, Weng X, Ellis SJ, Lakey ND, Culpepper J, Moore KJ, Breitbart RE, Duyk GM, Tepper RI, Morgenstern JP. Evidence that the diabetes gene encodes the leptin receptor: identification of a mutation in the leptin receptor gene in db/db mice. *Cell.* 1996; 84:491–495. [PubMed: 8608603]
- Cheung CC, Clifton DK, Steiner RA. Proopiomelanocortin neurons are direct targets for leptin in the hypothalamus. *Endocrinology.* 1997; 138:4489–4492. [PubMed: 9322969]
- Clegg DJ, Benoit SC, Barrera JG, Woods SC. Estrogen Mediates Body Fat Distribution and Brain Sensitivity to Adiposity Signals. *Diabetes.* 2003; 52(supplement 1)
- Clegg DJ, Benoit SC, Fisher ME, Barrera JG, Seeley RJ, Woods SC. Sex hormones determine body fat distribution and sensitivity to adiposity signals. *Appetite.* 2003; 40:324.
- Clegg DJ, Brown LM, Woods SC, Benoit SC. Gonadal hormones determine sensitivity to central leptin and insulin. *Diabetes.* 2006; 55:978–987. [PubMed: 16567519]
- Clegg DJ, Riedy CA, Smith KA, Benoit SC, Woods SC. Differential sensitivity to central leptin and insulin in male and female rats. *Diabetes.* 2003; 52:682–687. [PubMed: 12606509]
- Cone RD, Cowley MA, Butler AA, Fan W, Marks DL, Low MJ. The arcuate nucleus as a conduit for diverse signals relevant to energy homeostasis. *Int J Obes Relat Metab Disord.* 2001; 25(Suppl 5):S63–67. [PubMed: 11840218]
- Crandall DL, Busler DE, Novak TJ, Weber RV, Kral JG. Identification of estrogen receptor beta RNA in human breast and abdominal subcutaneous adipose tissue. *Biochem Biophys Res Commun.* 1998; 248:523–526. [PubMed: 9703958]
- Dagnault A, Richard D. Involvement of the medial preoptic area in the anorectic action of estrogens. *Am J Physiol.* 1997; 272:R311–317. [PubMed: 9039023]
- Danguir J, Nicolaidis S. Cortical activity and sleep in the rat lateral hypothalamic syndrome. *Brain Res.* 1980; 185:305–321. [PubMed: 7357431]
- de Luca C, Olefsky JM. Inflammation and insulin resistance. *FEBS Lett.* 2008; 582:97–105. [PubMed: 18053812]
- Demerath EW, Towne B, Wisemandle W, Blangero J, Chumlea WC, Siervogel RM. Serum leptin concentration, body composition, and gonadal hormones during puberty. *Int J Obes Relat Metab Disord.* 1999; 23:678–685. [PubMed: 10454100]
- Despres JP. Abdominal obesity as important component of insulin-resistance syndrome. *Nutrition.* 1993; 9:452–459. [PubMed: 8286886]
- Dhar MS, Sommardahl CS, Kirkland T, Nelson S, Donnell R, Johnson DK, Castellani LW. Mice heterozygous for *Atp10c*, a putative amphipath, represent a novel model of obesity and type 2 diabetes. *J Nutr.* 2004; 134:799–805. [PubMed: 15051828]
- Diano S, Kalra SP, Horvath TL. Leptin receptor immunoreactivity is associated with the Golgi apparatus of hypothalamic neurons and glial cells. *J Neuroendocrinol.* 1998; 10:647–650. [PubMed: 9744481]

- Donohoe TP, Stevens R, Johnson NJ, Barker S. Effects of stereoisomers of estradiol on food intake, body weight and hoarding behavior in female rats. *Physiol Behav.* 1984; 32:589–592. [PubMed: 6484012]
- Drewett RF. Sexual behaviour and sexual motivation in the female rat. *Nature.* 1973; 242:476–477. [PubMed: 4735606]
- Elias CF, Aschkenasi C, Lee C, Kelly J, Ahima RS, Bjorbaek C, Flier JS, Saper CB, Elmquist JK. Leptin differentially regulates NPY and POMC neurons projecting to the lateral hypothalamic area. *Neuron.* 1999; 23:775–786. [PubMed: 10482243]
- Elias CF, Kelly JF, Lee CE, Ahima RS, Drucker DJ, Saper CB, Elmquist JK. Chemical characterization of leptin-activated neurons in the rat brain [In Process Citation]. *J Comp Neurol.* 2000; 423:261–281. [PubMed: 10867658]
- Elmquist JK, Ahima RS, Elias CF, Flier JS, Saper CB. Leptin activates distinct projections from the dorsomedial and ventromedial hypothalamic nuclei. *Proceedings of the National Academy of Sciences USA.* 1998; 95:741–746.
- Elmquist JK, Ahima RS, Maratos-Flier E, Flier JS, Saper CB. Leptin activates neurons in ventrobasal hypothalamus and brainstem. *Endocrinology.* 1997; 138:839–842. [PubMed: 9003024]
- Elmquist JK, Bjorbaek C, Ahima RS, Flier JS, Saper CB. Distributions of leptin receptor mRNA isoforms in the rat brain. *J Comp Neurol.* 1998; 395:535–547. [PubMed: 9619505]
- Elmquist JK, Elias CF, Saper CB. From lesions to leptin: Hypothalamic control of food intake and body weight. *Neuron.* 1999; 22:221–232. [PubMed: 10069329]
- Elmquist JK, Maratos-Flier E, Saper CB, Flier JS. Unraveling the central nervous system pathways underlying responses to leptin. *Nature Neuroscience.* 1998; 1:445–450.
- Finkelstein M, Weidenfeld J, Ne'eman Y, Samuni A, Mizrahi Y, Ben-Uzilio R. Comparative studies of the aromatization of testosterone and epitestosterone by human placental aromatase. *Endocrinology.* 1981; 108:943–947. [PubMed: 7460853]
- Gallou-Kabani C, Vige A, Gross MS, Boileau C, Rabes JP, Fruchart-Najib J, Jais JP, Junien C. Resistance to high-fat diet in the female progeny of obese mice fed a control diet during the periconceptual, gestation, and lactation periods. *Am J Physiol Endocrinol Metab.* 2007; 292:E1095–1100. [PubMed: 17164437]
- Gao Q, Mezei G, Nie Y, Rao Y, Choi CS, Bechmann I, Leranath C, Toran-Allerand D, Priest CA, Roberts JL, Gao XB, Mobbs C, Shulman GI, Diano S, Horvath TL. Anorectic estrogen mimics leptin's effect on the rewiring of melanocortin cells and Stat3 signaling in obese animals. *Nat Med.* 2007; 13:89–94. [PubMed: 17195839]
- Geary N, Asarian L, Korach KS, Pfaff DW, Ogawa S. Deficits in E2-dependent control of feeding, weight gain, and cholecystokinin satiation in ER-alpha null mice. *Endocrinology.* 2001; 142:4751–4757. [PubMed: 11606440]
- Ghisletti S, Meda C, Maggi A, Vegeto E. 17beta-estradiol inhibits inflammatory gene expression by controlling NF-kappaB intracellular localization. *Mol Cell Biol.* 2005; 25:2957–2968. [PubMed: 15798185]
- Green S, Walter P, Kumar V, Krust A, Bornert JM, Argos P, Chambon P. Human oestrogen receptor cDNA: sequence, expression and homology to v-erb-A. *Nature.* 1986; 320:134–139. [PubMed: 3754034]
- Harris HA, Albert LM, Leathurby Y, Malamas MS, Mewshaw RE, Miller CP, Kharode YP, Marzolf J, Komm BS, Winneker RC, Frail DE, Henderson RA, Zhu Y, Keith JC Jr. Evaluation of an estrogen receptor-beta agonist in animal models of human disease. *Endocrinology.* 2003; 144:4241–4249. [PubMed: 14500559]
- Heine PA, Taylor JA, Iwamoto GA, Lubahn DB, Cooke PS. Increased adipose tissue in male and female estrogen receptor-alpha knockout mice. *Proc Natl Acad Sci U S A.* 2000; 97:12729–12734. [PubMed: 11070086]
- Herzog H. Neuropeptide Y and energy homeostasis: insights from Y receptor knockout models. *Eur J Pharmacol.* 2003; 480:21–29. [PubMed: 14623347]
- Hetherington R, Ranson S. Hypothalamic lesions and adiposity in the rat. *Anat Rec.* 1940; 78:149–172.

- Hirosawa M, Minata M, Harada KH, Hitomi T, Krust A, Koizumi A. Ablation of estrogen receptor alpha (ERalpha) prevents upregulation of POMC by leptin and insulin. *Biochem Biophys Res Commun.* 2008; 371:320–323. [PubMed: 18439911]
- Hrupka BJ, Smith GP, Geary N. Hypothalamic implants of dilute estradiol fail to reduce feeding in ovariectomized rats. *Physiol Behav.* 2002; 77:233–241. [PubMed: 12419399]
- Ivanova T, Mendez P, Garcia-Segura LM, Beyer C. Rapid stimulation of the PI3-kinase/Akt signalling pathway in developing midbrain neurones by oestrogen. *J Neuroendocrinol.* 2002; 14:73–79. [PubMed: 11903815]
- Kahn BB, Flier JS. Obesity and insulin resistance. *J Clin Invest.* 2000; 106:473–481. [PubMed: 10953022]
- Kalaitzidis D, Gilmore TD. Transcription factor cross-talk: the estrogen receptor and NF-kappaB. *Trends Endocrinol Metab.* 2005; 16:46–52. [PubMed: 15734144]
- Kannel WB, Cupples LA, Ramaswami R, Stokes J, Kreger BE, Higgins M. Regional obesity and risk of cardiovascular disease; the Framingham study. *J Clin Epidemiol.* 1991; 44:183–190.
- Kelly MJ, Levin ER. Rapid actions of plasma membrane estrogen receptors. *Trends Endocrinol Metab.* 2001; 12:152–156. [PubMed: 11295570]
- Kemnitz JW, Gibber JR, Lindsay KA, Eisele SG. Effects of ovarian hormones on eating behaviors, body weight, and gluoregulation in rhesus monkeys. *Horm Behav.* 1989; 23:235–250. [PubMed: 2663699]
- Korner J, Chua SC Jr, Williams JA, Leibel RL, Wardlaw SL. Regulation of hypothalamic proopiomelanocortin by leptin in lean and obese rats. *Neuroendocrinology.* 1999; 70:377–383. [PubMed: 10657730]
- Krotkiewski M, Bjorntorp P, Sjostrom L, Smith U. Impact of obesity on metabolism in men and women. Importance of regional adipose tissue distribution. *J Clin Invest.* 1983; 72:1150–1162. [PubMed: 6350364]
- Kuiper GG, Enmark E, Peltö-Huikko M, Nilsson S, Gustafsson JA. Cloning of a novel receptor expressed in rat prostate and ovary. *Proc Natl Acad Sci U S A.* 1996; 93:5925–5930. [PubMed: 8650195]
- Ladyman SR, Grattan DR. Suppression of leptin receptor messenger ribonucleic acid and leptin responsiveness in the ventromedial nucleus of the hypothalamus during pregnancy in the rat. *Endocrinology.* 2005; 146:3868–3874. [PubMed: 15905318]
- Lee CG, Carr MC, Murdoch SJ, Mitchell E, Woods NF, Wener MH, Chandler WL, Boyko EJ, Brunzell JD. Adipokines, inflammation, and visceral adiposity across the menopausal transition: a prospective study. *J Clin Endocrinol Metab.* 2009; 94:1104–1110. [PubMed: 19126626]
- Leshner AI, Collier G. The effects of gonadectomy on the sex differences in dietary self-selection patterns and carcass compositions of rats. *Physiol Behav.* 1973; 11:671–676. [PubMed: 4748063]
- Levin BE. Arcuate NPY neurons and energy homeostasis in diet-induced obese and resistant rats. *Am J Physiol.* 1999; 276:R382–387. [PubMed: 9950915]
- Lindell K, Bennett PA, Itoh Y, Robinson IC, Carlsson LM, Carlsson B. Leptin receptor 5' untranslated regions in the rat: relative abundance, genomic organization and relation to putative response elements. *Mol Cell Endocrinol.* 2001; 172:37–45. [PubMed: 11165038]
- Louis-Sylvestre J, Larue-Achagiotis C, Le Magnen J. Oral induction of the insulin hyper-responsiveness in rats with ventromedial hypothalamic lesions. *Horm Metab Res.* 1980; 12:671–676. [PubMed: 7009367]
- Lu SF, McKenna SE, Cologer-Clifford A, Nau EA, Simon NG. Androgen receptor in mouse brain: sex differences and similarities in autoregulation. *Endocrinology.* 1998; 139:1594–1601. [PubMed: 9528939]
- Machinal F, Dieudonne MN, Leneuve MC, Pecquery R, Giudicelli Y. In vivo and in vitro ob gene expression and leptin secretion in rat adipocytes: evidence for a regional specific regulation by sex steroid hormones. *Endocrinology.* 1999; 140:1567–1574. [PubMed: 10098489]
- MacLusky NJ, Walters MJ, Clark AS, Toran-Allerand CD. Aromatase in the cerebral cortex, hippocampus, and mid-brain: ontogeny and developmental implications. *Mol Cell Neurosci.* 1994; 5:691–698. [PubMed: 7704444]

- Malyala A, Zhang C, Bryant DN, Kelly MJ, Ronnekleiv OK. PI3K signaling effects in hypothalamic neurons mediated by estrogen. *J Comp Neurol.* 2008; 506:895–911. [PubMed: 18085586]
- Merchenthaler I, Lane MV, Numan S, Dellovade TL. Distribution of estrogen receptor alpha and beta in the mouse central nervous system: in vivo autoradiographic and immunocytochemical analyses. *J Comp Neurol.* 2004; 473:270–291. [PubMed: 15101093]
- Milam KM, Stern JS, Storlien LH, Keesey RE. Effect of lateral hypothalamic lesions on regulation of body weight and adiposity in rats. *Am J Physiol.* 1980; 239:R337–343. [PubMed: 7435604]
- Mitra SW, Hoskin E, Yudkovitz J, Pear L, Wilkinson HA, Hayashi S, Pfaff DW, Ogawa S, Rohrer SP, Schaeffer JM, McEwen BS, Alves SE. Immunolocalization of estrogen receptor beta in the mouse brain: comparison with estrogen receptor alpha. *Endocrinology.* 2003; 144:2055–2067. [PubMed: 12697714]
- Morton GJ, Niswender KD, Rhodes CJ, Myers MG Jr, Blevins JE, Baskin DG, Schwartz MW. Arcuate nucleus-specific leptin receptor gene therapy attenuates the obesity phenotype of Koletsky (fa(k)/fa(k)) rats. *Endocrinology.* 2003; 144:2016–2024. [PubMed: 12697710]
- Mueller K, Hsiao S. Estrus- and ovariectomy-induced body weight changes: evidence for two estrogenic mechanisms. *J Comp Physiol Psychol.* 1980; 94:1126–1134. [PubMed: 7193690]
- Musatov S, Chen W, Pfaff DW, Mobbs CV, Yang XJ, Clegg DJ, Kaplitt MG, Ogawa S. Silencing of estrogen receptor alpha in the ventromedial nucleus of hypothalamus leads to metabolic syndrome. *Proc Natl Acad Sci U S A.* 2007; 104:2501–2506. [PubMed: 17284595]
- Obici S. Minireview: Molecular targets for obesity therapy in the brain. *Endocrinology.* 2009; 150:2512–2517. [PubMed: 19372196]
- Ohlsson C, Hellberg N, Parini P, Vidal O, Bohlooly M, Rudling M, Lindberg MK, Warner M, Angelin B, Gustafsson JA. Obesity and disturbed lipoprotein profile in estrogen receptor-alpha-deficient male mice. *Biochem Biophys Res Commun.* 2000; 278:640–645. [PubMed: 11095962]
- Okura T, Koda M, Ando F, Niino N, Ohta S, Shimokata H. Association of polymorphisms in the estrogen receptor alpha gene with body fat distribution. *Int J Obes Relat Metab Disord.* 2003; 27:1020–1027. [PubMed: 12917706]
- Okura T, Koda M, Ando F, Niino N, Tanaka M, Shimokata H. Association of the mitochondrial DNA 15497G/A polymorphism with obesity in a middle-aged and elderly Japanese population. *Hum Genet.* 2003; 113:432–436. [PubMed: 12905068]
- Olofsson LE, Pierce AA, Xu AW. Functional requirement of AgRP and NPY neurons in ovarian cycle-dependent regulation of food intake. *Proc Natl Acad Sci U S A.* 2009; 106:15932–15937. [PubMed: 19805233]
- Osterlund M, Kuiper GG, Gustafsson JA, Hurd YL. Differential distribution and regulation of estrogen receptor-alpha and -beta mRNA within the female rat brain. *Brain Res Mol Brain Res.* 1998; 54:175–180. [PubMed: 9526077]
- Pappas TC, Gametchu B, Watson CS. Membrane estrogen receptors identified by multiple antibody labeling and impeded-ligand binding. *FASEB J.* 1995; 9:404–410. [PubMed: 7896011]
- Payette C, Blackburn P, Lamarche B, Tremblay A, Bergeron J, Lemieux I, Despres JP, Couillard C. Sex differences in postprandial plasma tumor necrosis factor-alpha, interleukin-6, and C-reactive protein concentrations. *Metabolism.* 2009
- Pelletier G, Li S, Luu-The V, Labrie F. Oestrogenic regulation of proopiomelanocortin, neuropeptide Y and corticotrophin-releasing hormone mRNAs in mouse hypothalamus. *J Neuroendocrinol.* 2007; 19:426–431. [PubMed: 17388940]
- Pierroz DD, Ziotopoulou M, Ungsuan L, Moschos S, Flier JS, Mantzoros CS. Effects of acute and chronic administration of the melanocortin agonist MTII in mice with diet-induced obesity. *Diabetes.* 2002; 51:1337–1345. [PubMed: 11978628]
- Poehlman ET, Toth MJ, Gardner AW. Changes in energy balance and body composition at menopause: a controlled longitudinal study. *Ann Intern Med.* 1995; 123:673–675. [PubMed: 7574222]
- Pozzi S, Benedusi V, Maggi A, Vegeto E. Estrogen action in neuroprotection and brain inflammation. *Ann N Y Acad Sci.* 2006; 1089:302–323. [PubMed: 17261778]

- Qiu J, Bosch MA, Tobias SC, Grandy DK, Scanlan TS, Ronnekleiv OK, Kelly MJ. Rapid signaling of estrogen in hypothalamic neurons involves a novel G-protein-coupled estrogen receptor that activates protein kinase C. *J Neurosci.* 2003; 23:9529–9540. [PubMed: 14573532]
- Razandi M, Pedram A, Greene GL, Levin ER. Cell membrane and nuclear estrogen receptors (ERs) originate from a single transcript: studies of ERalpha and ERbeta expressed in Chinese hamster ovary cells. *Mol Endocrinol.* 1999; 13:307–319. [PubMed: 9973260]
- Rowland N, Meile MJ, Nicolaidis S. Metering of intravenously infused nutrients in VMH lesioned rats. *Physiol Behav.* 1975; 15:443–448. [PubMed: 815926]
- Ryan AS, Nicklas BJ, Berman DM. Hormone replacement therapy, insulin sensitivity, and abdominal obesity in postmenopausal women. *Diabetes Care.* 2002; 25:127–133. [PubMed: 11772913]
- Samaras K, Hayward CS, Sullivan D, Kelly RP, Campbell LV. Effects of postmenopausal hormone replacement therapy on central abdominal fat, glycemic control, lipid metabolism, and vascular factors in type 2 diabetes: a prospective study. *Diabetes Care.* 1999; 22:1401–1407. [PubMed: 10480500]
- Schwartz MW, Porte D Jr. Diabetes, obesity, and the brain. *Science.* 2005; 307:375–379. [PubMed: 15662002]
- Schwartz MW, Woods SC, Porte DJ, Seeley RJ, Baskin DG. Central nervous system control of food intake. *Nature.* 2000; 404:661–671. [PubMed: 10766253]
- Seeley R, Yagaloff K, Fisher S, Burn P, Thiele T, van DG, Baskin D, Schwartz M. Melanocortin receptors in leptin effects. *Nature.* 1997; 390:349. [PubMed: 9389472]
- Seeley RJ, van Dijk G, Campfield LA, Smith FJ, Burn P, Nelligan JA, Bell SM, Baskin DG, Woods SC, Schwartz MW. Intraventricular leptin reduces food intake and body weight of lean rats but not obese Zucker rats. *Horm Metab Res.* 1996; 28:664–668. [PubMed: 9013738]
- Seeley RJ, Woods SC. Monitoring of stored and available fuel by the CNS: implications for obesity. *Nat Rev Neurosci.* 2003; 4:901–909. [PubMed: 14595401]
- Shi H, Kokoeva MV, Inouye K, Tzameli I, Yin H, Flier JS. TLR4 links innate immunity and fatty acid-induced insulin resistance. *J Clin Invest.* 2006; 116:3015–3025. [PubMed: 17053832]
- Shima N, Yamaguchi Y, Yuri K. Distribution of estrogen receptor beta mRNA-containing cells in ovariectomized and estrogen-treated female rat brain. *Anat Sci Int.* 2003; 78:85–97. [PubMed: 12828421]
- Shimizu H, Shimomura Y, Nakanishi Y, Futawatari T, Ohtani K, Sato N, Mori M. Estrogen increases in vivo leptin production in rats and human subjects. *J Endocrinol.* 1997; 154:285–292. [PubMed: 9291839]
- Shoelson SE, Herrero L, Naaz A. Obesity, inflammation, and insulin resistance. *Gastroenterology.* 2007; 132:2169–2180. [PubMed: 17498510]
- Shughrue P, Scrimo P, Lane M, Askew R, Merchenthaler I. The distribution of estrogen receptor-beta mRNA in forebrain regions of the estrogen receptor-alpha knockout mouse. *Endocrinology.* 1997; 138:5649–5652. [PubMed: 9389555]
- Shughrue PJ, Lane MV, Merchenthaler I. Comparative distribution of estrogen receptor-alpha and -beta mRNA in the rat central nervous system. *J Comp Neurol.* 1997; 388:507–525. [PubMed: 9388012]
- Shughrue PJ, Lubahn DB, Negro-Vilar A, Korach KS, Merchenthaler I. Responses in the brain of estrogen receptor alpha-disrupted mice. *Proc Natl Acad Sci U S A.* 1997; 94:11008–11012. [PubMed: 9380750]
- Simerly RB, Chang C, Muramatsu M, Swanson LW. Distribution of androgen and estrogen receptor mRNA-containing cells in the rat brain: an in situ hybridization study. *J Comp Neurol.* 1990; 294:76–95. [PubMed: 2324335]
- Simoncini T, Hafezi-Moghadam A, Brazil DP, Ley K, Chin WW, Liao JK. Interaction of oestrogen receptor with the regulatory subunit of phosphatidylinositol-3-OH kinase. *Nature.* 2000; 407:538–541. [PubMed: 11029009]
- Simonian SX, Herbison AE. Differential expression of estrogen receptor alpha and beta immunoreactivity by oxytocin neurons of rat paraventricular nucleus. *J Neuroendocrinol.* 1997; 9:803–806. [PubMed: 9419830]

- Simonian SX, Spratt DP, Herbison AE. Identification and characterization of estrogen receptor alpha-containing neurons projecting to the vicinity of the gonadotropin-releasing hormone perikarya in the rostral preoptic area of the rat. *J Comp Neurol.* 1999; 411:346–358. [PubMed: 10404258]
- Singh M. Ovarian hormones elicit phosphorylation of Akt and extracellular-signal regulated kinase in explants of the cerebral cortex. *Endocrine.* 2001; 14:407–415. [PubMed: 11444439]
- Slamberova R, Hnaticzuk OC, Vathy I. Expression of proopiomelanocortin and proenkephalin mRNA in sexually dimorphic brain regions are altered in adult male and female rats treated prenatally with morphine. *J Pept Res.* 2004; 63:399–408. [PubMed: 15140157]
- Speer G, Cseh K, Fuszek P, Dworak O, Vargha P, Takacs I, Nagy Z, Lakatos P. [The role of estrogen receptor, vitamin D receptor and calcium receptor genotypes in the pathogenesis of colorectal cancer]. *Orv Hetil.* 2001; 142:947–951. [PubMed: 11392075]
- Stein B, Yang MX. Repression of the interleukin-6 promoter by estrogen receptor is mediated by NF-kappa B and C/EBP beta. *Mol Cell Biol.* 1995; 15:4971–4979. [PubMed: 7651415]
- Steinbaum SR. The metabolic syndrome: an emerging health epidemic in women. *Progress in Cardiovascular Disorders.* 2004; 46:321–326.
- Sutter-Dub MT. Rapid non-genomic and genomic responses to progestogens, estrogens, and glucocorticoids in the endocrine pancreatic B cell, the adipocyte and other cell types. *Steroids.* 2002; 67:77–93. [PubMed: 11755172]
- Tartaglia LA, Dembski M, Weng X, Deng N, Culpepper J, Devos R, Richards GJ, Campfield LA, Clark FT, Deeds J, Muir C, Sanker S, Moriarty A, Moore KJ, Smutko JS, Mays GG, Woolf EA, Monroe CA, Tepper RI. Identification and expression cloning of a leptin receptor, OB-R. *Cell.* 1995; 83:1263–1271. [PubMed: 8548812]
- Thammacharoen S, Lutz TA, Geary N, Asarian L. Hindbrain administration of estradiol inhibits feeding and activates estrogen receptor-alpha-expressing cells in the nucleus tractus solitarius of ovariectomized rats. *Endocrinology.* 2008; 149:1609–1617. [PubMed: 18096668]
- Thornton JE, Cheung CC, Clifton DK, Steiner RA. Regulation of hypothalamic proopiomelanocortin mRNA by leptin in ob/ob mice. *Endocrinology.* 1997; 138:5063–5067. [PubMed: 9348241]
- Toran-Allerand CD. Estrogen and the brain: beyond ER-alpha, ER-beta, and 17beta-estradiol. *Ann N Y Acad Sci.* 2005; 1052:136–144. [PubMed: 16024756]
- Toran-Allerand CD, Tinnikov AA, Singh RJ, Nethrapalli IS. 17alpha-estradiol: a brain-active estrogen? *Endocrinology.* 2005; 146:3843–3850. [PubMed: 15947006]
- Turgeon JL, Carr MC, Maki PM, Mendelsohn ME, Wise PM. Complex actions of sex steroids in adipose tissue, the cardiovascular system, and brain: Insights from basic science and clinical studies. *Endocr Rev.* 2006; 27:575–605. [PubMed: 16763155]
- Van Dijk G, Thiele TE, Donahey JC, Campfield LA, Smith FJ, Burn P, Bernstein IL, Woods SC, Seeley RJ. Central infusions of leptin and GLP-1-(7–36) amide differentially stimulate c-FOS in the rat brain. *Am J Physiol.* 1996; 271:R1096–1100. [PubMed: 8898006]
- Vasudevan N, Pfaff DW. Non-genomic actions of estrogens and their interaction with genomic actions in the brain. *Front Neuroendocrinol.* 2008; 29:238–257. [PubMed: 18083219]
- Vegeto E, Belcredito S, Etteri S, Ghisletti S, Brusadelli A, Meda C, Krust A, Dupont S, Ciana P, Chambon P, Maggi A. Estrogen receptor-alpha mediates the brain antiinflammatory activity of estradiol. *Proc Natl Acad Sci U S A.* 2003; 100:9614–9619. [PubMed: 12878732]
- Vegeto E, Bonincontro C, Pollio G, Sala A, Viappiani S, Nardi F, Brusadelli A, Viviani B, Ciana P, Maggi A. Estrogen prevents the lipopolysaccharide-induced inflammatory response in microglia. *J Neurosci.* 2001; 21:1809–1818. [PubMed: 11245665]
- Vidal O, Lindberg MK, Hollberg K, Baylink DJ, Andersson G, Lubahn DB, Mohan S, Gustafsson JA, Ohlsson C. Estrogen receptor specificity in the regulation of skeletal growth and maturation in male mice. *Proc Natl Acad Sci U S A.* 2000; 97:5474–5479. [PubMed: 10805804]
- Voisin DL, Simonian SX, Herbison AE. Identification of estrogen receptor-containing neurons projecting to the rat supraoptic nucleus. *Neuroscience.* 1997; 78:215–228. [PubMed: 9135102]
- Wade GN. Gonadal hormones and behavioral regulation of body weight. *Physiol Behav.* 1972; 8:523–534. [PubMed: 4556652]
- Wade GN, Gray JM. Gonadal effects on food intake and adiposity: a metabolic hypothesis. *Physiology and Behavior.* 1979; 22:583–593. [PubMed: 379889]

- Wade GN, Gray JM, Bartness TJ. Gonadal influences on adiposity. *Int J Obes.* 1985; 9(Suppl 1):83–92. [PubMed: 4066126]
- Wallen WJ, Belanger MP, Wittnich C. Sex hormones and the selective estrogen receptor modulator tamoxifen modulate weekly body weights and food intakes in adolescent and adult rats. *J Nutr.* 2001; 131:2351–2357. [PubMed: 11533278]
- Wang P, Mariman E, Renes J, Keijzer J. The secretory function of adipocytes in the physiology of white adipose tissue. *J Cell Physiol.* 2008; 216:3–13. [PubMed: 18264975]
- Wang Y, Beydoun MA, Liang L, Caballero B, Kumanyika SK. Will all Americans become overweight or obese? estimating the progression and cost of the US obesity epidemic. *Obesity (Silver Spring).* 2008; 16:2323–2330. [PubMed: 18719634]
- Wellen KE, Hotamisligil GS. Obesity-induced inflammatory changes in adipose tissue. *J Clin Invest.* 2003; 112:1785–1788. [PubMed: 14679172]
- Wellen KE, Hotamisligil GS. Inflammation, stress, and diabetes. *J Clin Invest.* 2005; 115:1111–1119. [PubMed: 15864338]
- Wilkinson HA, Dahllund J, Liu H, Yudkovitz J, Cai SJ, Nilsson S, Schaeffer JM, Mitra SW. Identification and characterization of a functionally distinct form of human estrogen receptor beta. *Endocrinology.* 2002; 143:1558–1561. [PubMed: 11897716]
- Williams G, Bing C, Cai XJ, Harrold JA, King PJ, Liu XH. The hypothalamus and the control of energy homeostasis: different circuits, different purposes. *Physiol Behav.* 2001; 74:683–701. [PubMed: 11790431]
- Wilson ME, Liu Y, Wise PM. Estradiol enhances Akt activation in cortical explant cultures following neuronal injury. *Brain Res Mol Brain Res.* 2002; 102:48–54. [PubMed: 12191493]
- Wise PM, Scarbrough K, Weiland NG, Larson GH. Diurnal pattern of proopiomelanocortin gene expression in the arcuate nucleus of proestrous, ovariectomized, and steroid-treated rats: a possible role in cyclic luteinizing hormone secretion. *Mol Endocrinol.* 1990; 4:886–892. [PubMed: 2233745]
- Wise PM, Suzuki S, Brown CM. Estradiol: a hormone with diverse and contradictory neuroprotective actions. *Dialogues Clin Neurosci.* 2009; 11:297–303. [PubMed: 19877497]
- Woods SC, Schwartz MW, Baskin DG, Seeley RJ. Food intake and the regulation of body weight. *Annual Review of Psychology.* 2000; 51:255–277.
- Woods SC, Seeley RJ. Adiposity signals and the control of energy homeostasis. *Nutrition.* 2000; 16:894–902. [PubMed: 11054594]
- Wu-Peng S, Rosenbaum M, Nicolson M, Chua SC, Leibel RL. Effects of exogenous gonadal steroids on leptin homeostasis in rats. *Obes Res.* 1999; 7:586–592. [PubMed: 10574518]
- Yamada Y, Ando F, Niino N, Ohta S, Shimokata H. Association of polymorphisms of the estrogen receptor alpha gene with bone mineral density of the femoral neck in elderly Japanese women. *J Mol Med.* 2002; 80:452–460. [PubMed: 12110951]
- York D, Bray G. Dependence of hypothalamic obesity on insulin, the pituitary and the adrenal gland. *Endocrinology.* 1972; 90:885–894. [PubMed: 4258778]
- Yu IC, Lin HY, Liu NC, Wang RS, Sparks JD, Yeh S, Chang C. Hyperleptinemia without obesity in male mice lacking androgen receptor in adipose tissue. *Endocrinology.* 2008; 149:2361–2368. [PubMed: 18276764]
- Zhang Y, Proenca R, Maffei M, Barone M, Leopold L, Friedman JM. Positional cloning of the mouse obese gene and its human homologue. *Nature.* 1994; 372:425–432. [PubMed: 7984236]