BRITISH PHARMACOLOGICAL BPS SOCIETY

British Journal of Pharmacology (2010), 159, 1595–1597 © 2010 The Authors Journal compilation © 2010 The British Pharmacological Society All rights reserved 0007-1188/10 www.brjpharmacol.org

## COMMENTARY

## A<sub>2b</sub> adenosine receptors can change their spots

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Recently, a central role for the A<sub>2b</sub> adenosine receptor in a variety of cardiovascular functions including inflammation, erectile function, coronary artery dilation, asthma and cardioprotection has been demonstrated. Despite this evidence, the low-affinity A<sub>2b</sub> adenosine receptor is still poorly understood. This receptor appears to be very promiscuous in its coupling. In most tissues, it couples to G<sub>s</sub> much like its cousin, the A<sub>2a</sub> adenosine receptor, but in mast cells and now, most recently, in cardiac fibroblasts, the A<sub>2b</sub> receptor also couples to G<sub>q</sub>. Because of its low affinity, this receptor was originally thought unlikely to play any important physiological role. But the sensitivity of A<sub>2b</sub> adenosine receptors can be greatly increased by interaction with protein kinase C (PKC) making this receptor, under various conditions, both an activator and a target of PKC. We have recently documented a third coupling involving G<sub>i</sub>. This plasticity and versatility of A<sub>2b</sub> adenosine receptors position them as potential triggers of signalling in multiple signalling cascades in many physiological responses, making this a most interesting receptor indeed. British Journal of Pharmacology (2010) 159, 1595–1597; doi:10.1111/j.1476-5381.2010.00668.x

This article is a commentary on Feng et al., pp. 1598–1607 of this issue. To view this paper visit http://dx.doi.org/10.1111/j.1476-5381.2009.00558.x

Keywords: A<sub>2b</sub> adenosine receptor; G protein; protein kinase C

Abbreviations: IL-6, interleukin-6; NECA, 5'-(N-ethylcarboxamido) adenosine

Adenosine is a purine nucleoside that is widely distributed in all tissues and body fluids. Adenosine agonists and antagonists are being targeted for treatment of a variety of clinical conditions. These actions are attributed to binding of the purine to four distinct receptors: A<sub>1</sub>, A<sub>2a</sub>, A<sub>2b</sub> and A<sub>3</sub>. Although all four receptors have been characterized pharmacologically and have been cloned, our familiarity and appreciation of the A<sub>2b</sub> adenosine receptor is still very limited because selective agonists and antagonists have only recently become available. And what we think we know about A<sub>2b</sub> adenosine receptors may not be accurate.

All four receptors have been found in cardiac tissue, which is a mixed tissue composed of cardiomyocytes, fibrous tissue and fibroblasts, vascular endothelial and smooth muscle cells, mast cells, etc. A1 and A3 adenosine receptors have been shown to be present in cardiomyocytes (Grdeń et al., 2005). A<sub>2a</sub> and A<sub>2b</sub> adenosine receptors are both present in the coronary vasculature (Mubagwa and Flameng, 2001). Message for A<sub>2a</sub> receptor has been identified in cardiomyocytes (Xu et al., 1996). The presence of A<sub>2b</sub> receptors in cardiomyocytes has been controversial (Liang and Haltiwanger, 1995; Morrison et al., 2002; Grdeń et al., 2005; Yang et al., 2006), although we have detected A<sub>2b</sub> receptor message in both rabbit and rat ventricular myocytes (unpublished observation). Given this new evidence that A<sub>2b</sub> adenosine receptors were expressed in cardiomyocytes, we were very surprised when A<sub>2a</sub> receptors, but not A<sub>2b</sub> receptors, were found on the sarcolemma in single cardiomyocytes in which cAMP was monitored (Xin et al., 2009).

Binding of agonists to A<sub>1</sub> and A<sub>3</sub> adenosine receptors results in activation and cleavage of G<sub>i</sub> leading to a decrease in cAMP and activation of PKC (Mubagwa and Flameng, 2001), whereas A<sub>2a</sub> and A<sub>2b</sub> receptors stimulate adenylyl cyclase and increase cAMP production by liberating components of G<sub>s</sub> (Mubagwa and Flameng, 2001). The A<sub>2a</sub> and A<sub>2b</sub> adenosine receptors were first identified by their differential ability to stimulate cAMP production in brain slices at low  $(0.1-1 \,\mu\text{M})$ and high (>10 µM) adenosine concentrations (Schulte and Fredholm, 2003). Thus the A<sub>2b</sub> receptor became known as the low-affinity receptor. Using human embryonic kidney (HEK) 293 cells overexpressing human A<sub>2b</sub> receptors Linden et al. (1999) found that A<sub>2b</sub> receptors coupled not only to G<sub>s</sub> but also to Gq/11 leading to activation of phospholipase C and mobilization of calcium in the transfected kidney cells. Although G<sub>s</sub> coupling mediated vasodilation, the physiological significance of Gq/11 coupling was demonstrated in the

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Received 5 August 2009; revised 28 October 2009; accepted 17 November 2009

degranulation of mast cells. Thus the  $A_{2b}$  adenosine receptor could trigger at least two distinct signalling cascades.

This receptor fickleness was confirmed by Feng et al. (2009) in the current edition of the British Journal of Pharmacology. They examined production of the proinflammatory cytokine interleukin-6 (IL-6) by mouse cardiac fibroblasts following exposure to 5'-(N-ethylcarboxamido) adenosine (NECA), a potent, albeit not selective, A<sub>2b</sub> receptor agonist. Interestingly these fibroblasts expressed all four adenosine receptor subtypes. In response to NECA these cells produced significant amounts of IL-6, which was not mimicked by selective A1, A2a or A3 receptor agonists. And only the selective A2b receptor antagonist MRS 1754 could interfere with the response to NECA. Furthermore silencing of A<sub>2b</sub> receptors with siRNA also suppressed NECA-induced IL-6 secretion. Clearly IL-6 production by cardiac fibroblasts was greatly influenced by stimulation of A<sub>2b</sub> receptors. When Feng et al. (2009) studied the downstream signalling they found involvement of G<sub>q</sub> and not G<sub>s</sub>. Pretreatment with either the cAMP-competitive analog Rp-cAMPS or 1 of two protein kinase A (PKA) inhibitors, H-89 or KT 5720, before exposure to NECA failed to block NECAinduced IL-6 production. Nor could a cAMP analogue trigger IL-6 release. Therefore, the G<sub>s</sub>-cAMP-PKA pathway was not involved in the regulation of IL-6 production by A<sub>2b</sub> receptors. Although the authors did not directly test for G<sub>q</sub> coupling, it was presumed to be responsible. These data confirm the plasticity of A<sub>2b</sub> adenosine receptors, and demonstrate their complicated biology. This evidence of Gq coupling of A2b receptors is probably as important as the observation that the heart can be its own source of inflammatory cytokines.

Interest in adenosine in the heart was stimulated when we found that adenosine released by a transient period of ischaemia (preconditioning) was a trigger for a signalling cascade that resulted in protection of myocardium from a subsequent more prolonged coronary occlusion (Liu et al., 1991). Although A<sub>1</sub> adenosine receptor agonists were powerful preconditioning agents, they had little clinical value. For treatment of patients with acute myocardial infarction, an intervention was needed that could be applied at the time of reperfusion. We first reported that A<sub>2b</sub> receptors were critical to signalling at the onset of reperfusion in the preconditioned heart (Philipp et al., 2006). Thus both ischaemic preconditioning (Solenkova et al., 2006) and post-conditioning (repeated episodes of very brief coronary reocclusion in the first minutes of reperfusion) (Philipp et al., 2006) were aborted if A<sub>2b</sub> receptors were blocked at reperfusion. But these observations posed a significant theoretical quandary. Under basal conditions, the extracellular concentration of adenosine ranges from 30 to 300 nM. Although the concentration increases substantially during ischaemia to 1-4 µM, it rarely exceeds 10  $\mu$ M (Schulte and Fredholm, 2003). Yet the K<sub>i</sub> of A<sub>2b</sub> receptors for adenosine may be as high as 24 µM (Gao and Jacobson, 2007). Therefore, how could endogenous adenosine released from ischaemic heart muscle bind significantly to A<sub>2b</sub> receptors? And why only in preconditioned hearts? In the study by Feng et al. in which the end point was IL-6 production, they also found that A<sub>2b</sub> receptors activated PKC. However, in investigations of cardioprotection in which the end point was anatomical infarct size, Philipp et al. (2006) showed that in the cardioprotective signal transduction pathway, PKC is actually upstream of A<sub>2b</sub> adenosine receptors, the reverse of the sequence reported by Feng et al. in cardiac fibroblasts. It is well known that PKC activity can sensitize A<sub>2b</sub> receptor signalling, although the mechanism has never been elucidated nor has a physiological function been attributed to it (Feoktistov and Biaggioni, 1997). We (Kuno et al., 2007) tested whether this sensitization phenomenon might be involved in preconditioning. We found that a subthreshold dose of NECA, which, by itself could not increase phosphorylation of the survival kinases Akt and ERK in rabbit myocardium could activate them if the PKC activator phorbol 12-myristate 13-acetate (PMA) had been previously administered. This same effect was reproduced by a preconditioning cycle of ischaemia/reperfusion that would also have activated endogenous PKC. Therefore, we proposed that, in a preconditioned heart, PKC increased the sensitivity of A<sub>2b</sub> receptor signalling to the point that endogenous adenosine released during ischaemia could now bind to A<sub>2b</sub> receptors at reperfusion and initiate protective intracellular signalling. This observation again emphasized the versatility of A<sub>2b</sub> receptors and justified the impression that this adenosine receptor could play an important physiological role in the myocardium.

Most recently Kuno et al. (2009) further examined A<sub>2b</sub> receptor coupling to survival kinases in HEK 293 cells stably transfected with human A<sub>2b</sub> receptors. As in the heart NECA caused a dose-dependent phosphorylation of Akt and ERK. Furthermore NECA increased cAMP consistent with the previously reported G<sub>s</sub>-coupling of A<sub>2b</sub> receptors. Yet when G<sub>s</sub> was inhibited with cholera toxin or the G<sub>s</sub> antagonist NF449, the ability of NECA to phosphorylate Akt and ERK was actually enhanced. On the other hand, Pertussis toxin abolished NECA-triggered kinase phosphorylation indicating either a G<sub>i</sub> or G<sub>o</sub> coupling. These studies have now been repeated with the A<sub>2b</sub> adenosine receptor selective agonist BAY 60-6583, and the observations are identical (submitted for publication). So here we have yet a third coupling pathway for cardiac  $A_{2b}$  receptors. The biology of the  $A_{2b}$  receptor is much more complicated than that of the other adenosine receptors and may be the key to a host of clinically relevant therapies. Unlike the leopard, A<sub>2b</sub> adenosine receptors can change their spots.

Of course, additional questions must still be addressed. Where are the  $A_{2b}$  receptors located if they are not on the sarcolemma? Are they associated with some intracellular organelle? Are they functional? Does the cardioprotective signalling cascade occur in the cardiomyocyte, or does  $A_{2b}$  receptor signalling occur on outer cell membranes of another tissue, perhaps vascular endothelium, with production of a diffusible messenger that then targets cardiomyocytes?

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