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## Integration of Diverse Research Methods to Analyze and Engineer Ca<sup>2+</sup>-Binding Proteins: From Prediction to Production

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### Abstract

In recent years, increasingly sophisticated computational and bioinformatics tools have evolved for the analyses of protein structure, function, ligand interactions, modeling and energetics. This includes the development of algorithms to recursively evaluate side-chain rotamer permutations, identify regions in a 3D structure that meet some set of search parameters, calculate and minimize energy values, and provide high-resolution visual tools for theoretical modeling. Here we discuss the interdependency between different areas of bioinformatics, the evolution of different algorithm design approaches, and finally the transition from theoretical models to real-world design and application as they relate to Ca<sup>2+</sup>-binding proteins. Within this context, it has become evident that significant pre-experimental design and calculations can be modeled through computational methods, thus eliminating potentially unproductive research and increasing our confidence in the correlation between real and theoretical models. Moving from prediction to production, it is anticipated that bioinformatics tools will play an increasingly significant role in research and development, improving our ability to both understand the physiological roles of Ca<sup>2+</sup> and other metals and to extend that knowledge to the design of function-specific synthetic proteins capable of fulfilling different roles in medical diagnostics and therapeutics.

### Keywords

Calcium-binding; EF-hand; prediction; algorithm; design; statistics; graph theory; machine learning

## 1. INTRODUCTION

The importance of Ca<sup>2+</sup> in biological systems has been well-established, yet research continues to identify new and diverse functions associated with this metal. Calcium's biological role is mainly fulfilled by its interaction with different classes of Ca<sup>2+</sup>-binding proteins (CaBPs) [1–3]. Intracellularly, calcium binding to trigger proteins such as calmodulin and troponin C with helix-loop-helix motifs results in Ca<sup>2+</sup>-induced conformational change which in turn mediates the activity of different cellular processes [4]. Buffer proteins such as calbindin D9k and parvalbumin are essential to maintain proper calcium homeostasis. Conversely, many extracellular CaBPs, such as metabotropic glutamate receptors (mGluRs) and calcium sensing

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receptors, have weak  $\text{Ca}^{2+}$ -binding affinities ( $\sim\text{mM}$ ) yet are essential for extracellular calcium signaling and cell-cell communication [5,6].

Currently the availability of sophisticated computational methods and resources has become indispensable as research tools. These methods may be directed at the prediction and identification of  $\text{Ca}^{2+}$ -binding sites; determining the function of CaBPs in a biological system; tracing or predicting the evolutionary path of CaBPs; analyzing the effects of competing, toxic metals on CaBP functions; and engineering binding sites for  $\text{Ca}^{2+}$  or other metal ions either for pure research or for various biotechnological applications, including medical imaging, drug delivery and cancer therapies.

As with any computational approach, the analyses of known data provide an important foundation for developing input parameters. For studies of CaBPs, this includes the structural, physical and chemical properties associated with  $\text{Ca}^{2+}$ -binding proteins in both the bound (holo) and unbound (apo) states. Fortunately, the increasing availability of bioinformatics tools such as databases, programs and algorithms have provided researchers with the means to statistically evaluate large samples [7–10].

In this review we begin with a discussion of current knowledge related to (1) structural parameters associated with  $\text{Ca}^{2+}$ -binding (i.e., ligand number and type, bond angles, geometric configurations and charge distributions) and (2) dynamic properties including conformational change and charge interactions. Other important dynamic properties, specifically affinity, selectivity, energy minimization and molecular dynamics, have been excluded from this review due to space limitations.

As the data presented here were largely obtained through extensive statistical analyses or mathematical models, we then discuss their significance with respect to the development of algorithms designed to predict or identify either  $\text{Ca}^{2+}$ -binding proteins or specific binding sites, followed by an overview and comparison of the algorithms. Finally, we discuss specific examples detailing experimental results that were obtained as a result of pre-experimental application of algorithms to design or predict  $\text{Ca}^{2+}$ -binding sites.

## 2. DEFINING $\text{Ca}^{2+}$ -BINDING THROUGH STATISTICAL ANALYSES

### a. Distribution of Ligand Number and Types

In broad terms, metal-binding sites are characterized by a central shell of hydrophilic ligands to chelate the ion, with a surrounding shell of hydrophobic residues [11,12]. The centers of high hydrophobic contrast, where ions are located, are typically solvent inaccessible, low-dielectric cavities that enhance electrostatic metal-ligand interactions [13–15]. Binding ligand atoms for common biologically-important metals (e.g. –  $\text{Fe}^{3+}$ ,  $\text{Mg}^{2+}$ ,  $\text{Zn}^{2+}$ ,  $\text{Ca}^{2+}$ ,  $\text{Mn}^{2+}$ ) are mainly oxygen, nitrogen and sulfur from sidechain groups, and oxygen from mainchain carbonyls [13,14,16–23]. Although nitrogen is frequently considered a potential  $\text{Ca}^{2+}$  ligand based on small molecule interactions [21–23], it is apparent that interactions between sulfur and nitrogen from proteins with  $\text{Ca}^{2+}$  are negligible. According to an explanation of the orbital chemistry relevant to ligand binding, as well as an understanding of how metal-ligand binding relates to polarizability of different classes of Lewis acid metals, the preference of  $\text{Ca}^{2+}$  for oxygen can be understood from its hard Lewis acid definition describing less covalent interaction with a more rigid ionic radius [24].

The coordination of  $\text{Ca}^{2+}$ -binding utilizes various types of oxygen atoms from sidechain carboxyl groups (Asp, Glu), carboxamide groups (Asn, Gln), hydroxyl groups (Ser, Thr), mainchain carbonyl oxygen atoms from most residues, cofactors, and water molecules. The majority of all  $\text{Ca}^{2+}$ -binding ligands originate from turn/loop regions, rather than helix and

sheet regions [12,25,26]. This distribution persists even after EF-Hand sites are deliberately excluded [26].

The coordination number is frequently reported in the range of 3–8 ligand atoms [27–29]. However, certain classes of  $\text{Ca}^{2+}$ -binding sites exhibit preferences with respect to residue type and coordination number. In EF-Hand binding sites [26,30–32], certain residues appear consistently at specific positions within the loop (Fig. (1a)), with some variations apparent based on the extent of binding sites analyzed. Collectively these include: Asp at position 1, Asp or Asn at position 3, Gly at position 6, Ile, Val or Leu at position 8, and Glu at position 12 [33–35].

A recent statistical analysis of the structural characteristics of both EF-Hand and Non-EF-Hand proteins conducted in our laboratory revealed significant structural diversity in Non-EF-Hand proteins [28]. The dataset used for our analysis was comprised of 1605 binding sites from 558 Protein Data Bank (PDB) files with resolution  $R \leq 2.0 \text{ \AA}$ , based on a cutoff distance of  $3.5 \text{ \AA}$  as the maximum ligand distance for oxygen and nitrogen, and following removal of all structures with 90% or greater sequence homology. The coordination numbers reported were similar to previous studies indicating an average between 6–7 ligands [26,32]. Non-EF-Hand sites utilize lower coordination numbers ( $6 \pm 2$  vs.  $7 \pm 1$ ), fewer protein ligands ( $4 \pm 2$  vs.  $6 \pm 1$ ) and more water ligands ( $2 \pm 2$  vs.  $1 \pm 0$ ) than EF-Hand sites. The orders of ligand preference for Non-EF-Hand and EF-Hand sites, respectively, were  $\text{H}_2\text{O}$  (33.1%) > sidechain Asp (24.5%) > mainchain carbonyl (23.9%) > sidechain Glu (10.4%), and sidechain Asp (29.7%) > sidechain Glu (26.6%) > mainchain carbonyl (21.4%) >  $\text{H}_2\text{O}$  (13.3%).

The Non-EF-Hand proteins exhibit increased solvent interaction although the data for EF-Hand sites follow previously-reported trends (carboxylates > carbonyls > water > hydroxyl atoms) [14]. Reducing the cutoff to  $2.9 \text{ \AA}$  to compare distributions of water oxygen ligands (Table 1) revealed only a modest difference from previous studies for Non-EF-Hand sites [15,29], although a more significant difference was observed with the 13.3% reported for EF-Hand sites, where all water ligands fell within  $2.9 \text{ \AA}$  of the ion. Additionally, while the results obtained with respect to coordination number for Non-EF-Hand binding sites are consistent with those reported by Pidcock [26], they differed significantly from 6.7 and 7.0 reported in earlier studies which did not exclude EF-Hand sites [29,36].

For Non-EF-Hand sites, the higher than expected percentage of Asp residues and lower than expected percentage of Glu residues involved in calcium binding sites may be due to the more common occurrence of Asp in Asx turns, or a preference by  $\text{Ca}^{2+}$  for less sterically bulky sidechains [26]. However, the more equivalent distribution of Asp and Glu in EF-Hand sites is less readily understood. Canonical EF-Hand sites typically include 2–3 Asp and 1 Glu residues that bind  $\text{Ca}^{2+}$  with carboxyl oxygen atoms. Glu residues are of strategic importance in EF-Hand binding sites as bidentate, anchoring ligands for  $\text{Ca}^{2+}$  [29,37–40] and exhibit strong propensity towards helix formation which is consistent with its observed distribution in EF-Hand binding sites (Fig. (2)).

## b. Binding Site Geometry

In EF-Hand binding sites, the ligand oxygen atoms, usually six to seven, exhibit bipyramidal/monopyramidal pentagonal geometry (Fig. (1b)). Five oxygen atoms of the pentagon and  $\text{Ca}^{2+}$  lie nearly on the same plane, with variations [29, 37, 41]. The O-Ca-O angles in the pentagonal plane (Fig. (1b)) are approximately  $72^\circ$ . In monopyramidal geometry, the extraplanar peak oxygen atom is usually provided by an Asp/Asn sidechain, and the line between it and the calcium ion is nearly vertical with the pentagonal plane. In the bipyramidal geometry, the second extraplanar oxygen atom is typically from water. Other Non-EF-Hand  $\text{Ca}^{2+}$ -binding sites usually do not have similar, well-defined geometrical structures.

The mean Ca-O ligand binding distance has been reported, from different studies, as either  $2.4 \pm 0.1 \text{ \AA}$  or  $2.42 \text{ \AA}$  with a range of  $2.01 - 3.15 \text{ \AA}$  [2,14,24,42–45]. The mean Ca-O distance values reported for EF-Hand and Non-EF-Hand indicate very little difference between carbonyl and side-chain oxygens, and between the different classes for each ligand type. However, bidentate ligand distances are slightly longer for both EF-Hand ( $2.5 \pm 0.2 \text{ \AA}$ ) and Non-EF-Hand ( $2.6 \pm 0.3 \text{ \AA}$ ) than for the carbonyl and side-chain ligands in their respective classes [28]. A more pronounced change was observed for the bidentate mean Ca-C distances, which were  $0.5-0.6 \text{ \AA}$  shorter than the distances found for carbonyl and side-chain ligand oxygen atoms, resulting in overlap between the Ca-O and Ca-C shells [28].

The mean Ca-O-C angles (Fig. (1b)) are observed to be different among carbonyl, side-chain, and bidentate oxygen ligands. The Ca-O-C angles were largest for carbonyl ( $151.5 \pm 15.8^\circ$  and  $159.8 \pm 12.5^\circ$ ), followed by side chain ( $140.4 \pm 15.2^\circ$  and  $136.7 \pm 16.0^\circ$ ) and bidentate ( $93.6 \pm 11.3^\circ$  and  $92.9 \pm 6.8^\circ$ ) for Non-EF-Hand and EF-Hand, respectively. It was observed that a Gaussian distribution of Ca-O-C angle values is associated with Non-EF-Hand ligands, and the range values for both classes are nearly identical for carbonyl, side chain, and bidentate.

For the bidentate ligands, the dihedral angle is the angle between the plane formed by the side-chain carboxyl group (-COO), and the plane formed by the two carboxyl oxygen atoms and the  $\text{Ca}^{2+}$  ion (Fig. (1c)). For Non-EF-Hand and EF-Hand the mean and standard deviation values for dihedral angles were found to be  $168.1 \pm 9.7^\circ$  and  $170.6 \pm 7.1^\circ$ , indicating that they almost lie in the same plane.

This analysis presented a variety of flexible coordination schemes associated with  $\text{Ca}^{2+}$ -binding and tremendous structural diversity, consistent with earlier conclusions by Martin [46].

### c. Charge in the Binding Site Microenvironment

Formal charge (FC) by site is typically simplified to account only for negatively charged side-chain carboxyl groups (-1) from Glu and Asp [28,34]. Mean negative formal charge values of  $1 \pm 1$  and  $3 \pm 1$  have been reported for Non-EF-Hand and EF-Hand sites, respectively. However, only a small percentage of Non-EF-Hand sites exhibit a negative charge greater than 2. While binding affinity may be enhanced by an increase in the number of negatively-charged ligands, there appears to be an optimal level for this [47,48] which varies based on ion charge (e.g., trivalent cations are better charge acceptors than divalent cations) and the presence of bidentate ligands in the binding site [13–15]. However, recent work in our laboratory has demonstrated that increasing the number of negatively-charged ligands from three to five in an engineered  $\text{Ca}^{2+}$ -binding site increases the binding affinity for both  $\text{Ca}^{2+}$  and  $\text{La}^{3+}$  [49]. Additionally, removal of charges distant from the binding site may lead to significant decreases in binding affinity [50] due to reduction in the calcium kinetic on-rate [51].  $\text{Ca}^{2+}$ -binding sites with high negative formal charges are likely located in flexible loop regions of the protein [52]. Binding sites of zero formal charge were identified within EF-Hand sites, such as the protein calprotectin (1xk4.pdb) [53], and Non-EF-Hand proteins, although this was predictably more evident in Non-EF-Hand (20%) than EF-Hand (4%) sites. Detailed structural analyses of the protein environments of these charge-deficient sites reveal that charge-charge stabilization beyond the chelated metal ion can lead to the exclusion of available negatively charged side-chain residues and facilitate the binding of  $\text{Ca}^{2+}$  with carbonyl oxygen atoms [28].

Increasing the number of bidentate ligands is energetically favorable, and bidentate binding may allow for an increase in the number of charged ligands in the binding site. This subsequently leads to better protection for the cation from other intracellular anions. The carboxylate binding mode is highly-significant with respect to  $\text{Ca}^{2+}$ -binding, as the conserved

bidentate anchor in EF-Hand proteins is instrumental in conformational changes associated with protein function [54,55].

#### d. Calcium-Induced Conformational Change

The comparative analyses of static properties may be further extended to evaluate dynamic aspects of  $\text{Ca}^{2+}$ -binding. A significant body of research is available related to the mechanistic and functional aspects of protein conformational change associated with binding [7,55–60]. In general, these conformational analyses can be divided into two categories: the analyses of conformational changes of individual proteins and proteins on a database scale.

Vertical studies (i.e., between native and metal-complexed structures of the same protein) for individual proteins have compared changes in interatomic distances and angles, and hydrophobicity or  $\beta$ -factor changes associated with  $\text{Ca}^{2+}$ -binding. Zhang *et al.* reported the  $\text{Ca}^{2+}$ -induced conformational transition observed by comparing NMR structures of apo- and holo-calmodulin [55]. Chrysin *et al.* compared the X-ray structures of apo- and holo- $\alpha$ -lactalbumin with respect to  $\text{Ca}^{2+}$ -induced protein folding [59]. In addition, protein conformational changes have been investigated using simulations (e.g., Molecular Dynamics, Monte Carlo simulations) [61–63]. In a different approach, Dudev and Lim [15] used *ab initio* calculations, continuum dielectric methods (CDM) representing different dielectric media, and density functional theory (DFT) to compute thermodynamic values related to the exchange of metal-bound water molecules with small molecule analogs for Asp and Glu (formate); Asn, Gln and backbone peptide (forma-mide); and His (imidazole). For  $\text{Ca}^{2+}$  sites, which tend to have higher coordination numbers (CN) than  $\text{Mg}^{2+}$  or  $\text{Zn}^{2+}$ , the presence of more charged ligands leads to higher charge repulsion in the binding site, thus increasing the cavity size and contributing to the selectivity of  $\text{Ca}^{2+}$  over the more abundant  $\text{Mg}^{2+}$ .

Serial studies (i.e., between datasets of different proteins) for comparison of protein conformational changes on a database scale have evaluated different calcium-binding pockets for common features. These analyses are dependent upon (1) the availability of empirical and experimental data deposited in the PDB, and (2) the methodologies and measurements used to analyze the dataset. A database-scaled study by Babor *et al.* analyzed 58 apo/holo pairs of calcium binding sites from the PDB to evaluate conformational change based on comparing changes in Root Mean Square Deviation (RMSD), distance, angle,  $\beta$ -factor and solvent accessibility as a result of  $\text{Ca}^{2+}$ -binding [64]. Babor *et al.* evaluated conformational changes between the apo/holo forms, where conformational change was defined based on side-chain dihedral angles that were required to differ by a minimum of  $40^\circ$  between structures to qualify as a conformational change. Results of this study suggested that approximately 20% of  $\text{Ca}^{2+}$ -binding sites undergo backbone rearrangements upon  $\text{Ca}^{2+}$  binding, while only a small percentage of binding sites (~5%) exhibits significant rotation in more than two side-chains if the backbone is not rearranged upon calcium-binding.

Conversely, Eyal *et al.*, analyzing conformational change associated with point mutations, considered a  $60^\circ$  change as a cutoff for defining conformational change [9]. Similarly, Zhao *et al.* evaluated multiple paired PDB protein structures that did not involve metal-protein complex formation and concluded that the residue angle differences in terms of one paired entry are intrinsic properties of the individual residues, and that inaccurate conclusions may be drawn based on arbitrary selection of a single cutoff value for all residues [65].

Future comparison of the holo- and apo- protein forms of NMR structures on a database scale will provide additional important information on dynamic conformational changes in solution.



### e. Methodologies to Identify Ca<sup>2+</sup>-Binding Sites

While statistical analyses identify structural, chemical and energetic characteristics necessary for the characterization of Ca<sup>2+</sup>-binding sites, computational methods are required to exploit these data to identify or predict the location of Ca<sup>2+</sup>-binding sites in the protein structure. These methodologies include: homology analyses against known Ca<sup>2+</sup>-binding sequences or structures; energy or pseudo-energy calculations using Ca<sup>2+</sup>-binding ligands (e.g., oxygen clusters) or rotamer libraries of Ca<sup>2+</sup>-binding residues; grid functions; Bayesian statistical methods or scoring functions based on Ca<sup>2+</sup>-binding characteristics (e.g., charge, distance); and graph theory applications identifying Ca<sup>2+</sup>-binding ligand clusters. Many current applications combine two or more of these methods with statistical parameters as filters. Details and examples of these methodologies are discussed in the next section.

## 3. ALGORITHMS: EVOLUTION, DESIGN AND APPLICATIONS

### a. Primary Sequence Analysis and Structure Prediction

Ca<sup>2+</sup>-binding sites may be predicted or identified based on primary sequence or three-dimensional tertiary structure. Software is also available to predict or identify secondary structure, including PDBSum (<http://www.ebi.ac.uk/thornton-srv/databases/pdbsum/>). Prediction is dependent on matrices that evaluate existing databases to provide predictions based on known sequence and structure data, which may include static structural information (e.g., ligand number, ligand type, bond angles, etc.) and dynamic parameters (e.g., binding affinity).

Significant achievements have been made in the prediction or identification of Ca<sup>2+</sup>-binding sites utilizing programs which align primary sequences with related conserved regions. Examples of these would include the MacVector package (Oxford Molecular Group), ClustalW (<http://www.ebi.ac.uk/clustalw/>), FASTA (<http://www.ebi.ac.uk/Tools/fasta/index.html>) and BLAST (<http://blast.ncbi.nlm.nih.gov/Blast.cgi>). Similarly, programs such as CALM [66] and CaPS [35] apply pattern matching to identify target motifs in a sequence based on derived flexible patterns. In canonical EF-Hand proteins first analyzed by Kretsinger *et al.* [67], Ca<sup>2+</sup>-binding sites were found to be highly conserved with respect to a helix-loop-helix motif and, to a certain degree, by the relative locations of certain residues and residue ligand preference within the motif regions. Variants of the EF-Hand, including the lesser conserved pseudo EF-Hand and EF-Hand like motifs, also demonstrate less pronounced but recognizable primary sequence patterns. Patterns describing these motifs are usually generated by first conducting multiple sequence alignments and identifying highly-conserved residues. Accordingly, the sequence pattern search method is a simple and efficient way to predict EF-Hand and EF-Hand like sites in proteins.

Using the sequence searching software CALM [66] which was designed to search for continuous calcium binding sites, Bertrand and coworkers [68] identified five calcium binding amino acid sequences which possessed significant sequence homology with EF-Hand III loop of calmodulin in the N-terminal domain of neuronal nicotinic receptors. All of these sequences were rich in hydrophilic and acidic amino acids and included a terminal glutamate. The site-directed mutation of all the Glu to Gln residues in the five sequences reduced the Ca<sup>2+</sup> binding affinity, thus providing confirmation of the predicted binding ligands.

Similarly, Egmond and coworkers [69] reported identification of a calcium binding site in *Staphylococcus hyicus* Lipase (SHL) by sequence alignment with other staphylococcal lipases. Sequence alignment was conducted by MacVector package (Oxford Molecular Group) [70]. Replacing predicted Ca<sup>2+</sup>-binding ligands generated a Ca<sup>2+</sup>-insensitive staphylococcus hyicus lipase.

Zhou *et al.* developed new PROSITE-like motifs for the prediction of EF-Hand and EF-Hand-Like  $\text{Ca}^{2+}$ -binding sites by first conducting extensive sequence analyses across various databases and then utilizing these results (Fig. (1a)) as the basis for sequential patterns [35]. These patterns were then applied to bacterial proteins on a database scale. A total of 467 canonical EF-Hand and zero pseudo EF-Hand sites were identified, which suggested a later phylogenetical evolution of pseudo EF-Hand compared with canonical EF-Hand motif. An online server for CaPS is available at <http://lithium.gsu.edu/faculty/Yang/Calciomics.htm>. Zhou *et al.* later applied CaPS to identify a putative EF-Hand calcium binding motif within the rubella virus [71]. Similar prediction results were obtained using the programs PSIPRED [72], JPRED [73], and PredictProtein [74] to analyze the helix-loop-helix structure of the predicted calcium binding sequence. Parallel evidence came from similar prediction results obtained using the GG algorithm [75] based on a homology model of the rubella virus nonstructural pro-tease generated by the program SWISS-MODEL [76]. Experimental validation was obtained when the predicted EF-Hand motif from rubella virus was grafted onto a scaffold protein CD2, the wild-type form of which exhibits no  $\text{Ca}^{2+}$ -binding capability [77]. This grafting approach allowed for analysis of the intrinsic  $\text{Ca}^{2+}$ -binding capability of an isolated  $\text{Ca}^{2+}$ -binding site. Site-directed mutagenesis of presumed  $\text{Ca}^{2+}$ -binding ligands was conducted, and subsequent affinity studies validated the predicted site, which is believed to be required for the stability of rubella virus nonstructural protease under physiological conditions [71].

#### b. Prediction Based on Structural Data: Algorithm Design, Parameter Definition

Currently, most efforts to predict  $\text{Ca}^{2+}$ -binding sites may more accurately be described as the identification of known  $\text{Ca}^{2+}$ -binding sites using some computational methods or combination of methods to accurately locate a documented  $\text{Ca}^{2+}$  ion when little or no significant conformational change is observed. Conversely, true prediction will be achieved when the location of a  $\text{Ca}^{2+}$  binding site in the holo (loaded) form of the protein can be determined *in silico* based on only the apo (free) protein structure or the sequence alone. Many prediction algorithms employ structural parameters as distinguishing factors between  $\text{Ca}^{2+}$ -binding and nonbindingsites. This includes parameters related to ligand number and type and binding geometry restrictions related to distance and angle values between  $\text{Ca}^{2+}$  and its ligands as well as between the ligands with each other. Frequently, multiple algorithms are combined, typically using some type of scoring function with a grid system or other method of spatial analysis (e.g., graph theory) to evaluate physicochemical characteristics systematically and identify either binding ligands or binding sites based on comparative scoring. It is relevant to note that these functions are dependent on *a priori* statistical analyses to define or constrain function parameters based on structural, chemical or thermodynamic data. Prediction algorithms discussed in this section are summarized in Table 2.

### HYDROPHOBICITY CONTRAST FUNCTION

One of the first programs to predict calcium binding sites in three-dimensional structures was based on a hydrophobicity contrast function, which identified centers of high hydrophobic contrast as the metal-binding sites [11]. To quantitatively evaluate the correlation between points of high hydrophobicity expressed by the function and metal-binding sites, this program examined the structures of 10 metalloproteins and 13 metal-host molecules (five with documented  $\text{Ca}^{2+}$ -binding sites) with metal ions deleted. By embedding the structure into 0.5 Å grids and then computing at each grid the value of the function, the program was able to verify whether grid points returning high values were indeed near the metal-binding sites.

## COMBINATORY ALGORITHMS

An early FORTRAN algorithm for predicting  $\text{Ca}^{2+}$ -binding sites, which closely followed the approach reported by Yamashita [11], combined a fine grid system with a scoring function to describe points of valency based on electrostatic contribution of ligand atoms [27]. The scoring function parameters for the prediction of  $\text{Ca}^{2+}$ -binding were determined based on *a priori* analysis of the 62 available  $\text{Ca}^{2+}$ -binding proteins deposited in the PDB at that time which revealed that 371 of the 376 inner coordination shell atoms (i.e., within 2.7 Å) were oxygen. Based on these data, this algorithm required that  $\text{Ca}^{2+}$  be ligated by at least three protein oxygen atoms, where the metal-ligand distance must be smaller than 3.4 Å but larger than the van der Waals radius between the two. Each point in the grid system was then evaluated to determine if the oxygen coordination requirements were met and then a valence score was assigned to that point. An identification-removal procedure was used to iteratively eliminate all valence points within 3.5 Å of the highest valence point identified, leaving only the highest valence point in that region as the  $\text{Ca}^{2+}$ -binding site. Results demonstrated that 58/62  $\text{Ca}^{2+}$ -binding sites had valence greater than or equal to 1.4 and that 87% of the high valence points fell within 1.0 Å of the documented  $\text{Ca}^{2+}$  ion.

The Fold-X algorithm combines geometric pattern matching and energy calculations based on Fold-X empirical force field [78,79], where  $\text{Ca}^{2+}$ -binding sites are constrained to four to six ligand oxygen atoms. In the preprocess stage, the algorithm first identifies canonical positions [80] of  $\text{Ca}^{2+}$  with respect to each type of oxygen atom. The canonical ions are then superimposed onto a given protein where conflicting canonical ions are either removed or fused together depending on their relative proximities. Finally, the Fold-X force field [35,79] is used for energy calculation and consequently for the optimization of the position of the predicted calcium ion. This algorithm can also coarsely predict binding affinity based on energy-optimized placement of the  $\text{Ca}^{2+}$  ion to distinguish between low and high affinity sites, as well as high affinity  $\text{Ca}^{2+}$ - and  $\text{Mg}^{2+}$ -binding sites. Additionally, this approach was applicable to predicting  $\text{Mg}^{2+}$ ,  $\text{Ca}^{2+}$ ,  $\text{Zn}^{2+}$ ,  $\text{Mn}^{2+}$ , and  $\text{Cu}^{2+}$  and water molecules associated with the structure. Although these results indicate higher prediction rates than previous efforts, this method was less successful at identifying either multiple binding sites or sites with lower coordination numbers ( $\text{CN}<4$ ).

## GRAPH THEORY-BASED ALGORITHMS

The program GG (Graph theory and Geometry) presented a new approach to prediction based on a combination of graph theory and geometric parameters [75]. Based on the established parameters associated with  $\text{Ca}^{2+}$ -binding, the GG algorithm was designed to identify oxygen clusters as the basis for predicting  $\text{Ca}^{2+}$ -binding sites. In this method a graph  $G(V, E)$  was first constructed with oxygen atoms as its vertices and edges between two vertices if their spatial distance was no more than 6.0 Å. Oxygen clusters corresponding to cliques in  $G(V, E)$  consisting of exactly four vertices were identified, and the calcium center was determined at an equidistant center within each cluster if the distance ranged from 1.8 to 3.0 Å. If no such qualified center was located, that cluster was abandoned; otherwise, the center was the predicted calcium position and the atoms in the cluster were the predicted ligand atoms. This algorithm did not include a procedure to merge overlapping, predicted  $\text{Ca}^{2+}$  locations, resulting in multiple, adjacent predicted sites near the documented  $\text{Ca}^{2+}$ -binding site. Nonetheless, this algorithm produces rapid results, mainly due to the absence of a grid algorithm, and achieves approximately 90% site sensitivity and 80% site selectivity on testing datasets. These results are comparable to those reported by other algorithms. GG was implemented in C++. An online server is available at <http://chemistry.gsu.edu/faculty/Yang/Calciomics.htm>.



To fully utilize known geometric properties such as distance, angles and dihedral angles of  $\text{Ca}^{2+}$ -binding sites, which are essential in pinpointing  $\text{Ca}^{2+}$  position, Wang *et al.* developed the MUG (Multiple Geometries) program, which is able to predict  $\text{Ca}^{2+}$ -binding sites with different coordination numbers in proteins with atomic resolution. MUG requires that potential calcium-binding sites consist of at least four oxygen atoms [82] although MUG further allows oxygen atoms from either water or cofactors in addition to amino acid residues. Spatial analysis is achieved based on graph theory modeling as well as imposition of a grid system. After first identifying all possible oxygen clusters by finding maximal cliques, a calcium center (CC) for each cluster, corresponding to the potential  $\text{Ca}^{2+}$  position, is located to maximally regularize the structure of the (cluster, CC) pair. The structure is then inspected by geometric filters. An unqualified (cluster, CC) pair is further handled by recursively removing oxygen atoms and relocating the CC until its structure is either qualified or contains fewer than four ligand atoms. Ligand coordination is then determined for qualified structures. MUG also has the capability of predicting both low coordination sites and multiple  $\text{Ca}^{2+}$ -binding sites that share a common ligand. The MUG algorithm was implemented in Java and is available for use through an online server at <http://chemistry.gsu.edu/faculty/Yang/Calciomics.htm>.

## MACHINE LEARNING TECHNIQUES

Machine learning techniques, which are general approaches to any classification problem, are particularly effective at classifying ligands as binding residues and determining whether the target atom is close enough to the ion for binding [81,85,86]. In this method, a window including neighboring residues slides through the sequence so that each successive residue is viewed as a target residue in turn, to identify binding residues. The training procedures include applying the same learning machine to a training dataset consisting of sequences with known metal-binding residues. Classification features include location in the primary sequence, secondary structure, metal-binding propensity, and solvent accessibility. Additional information on machine learning methods for predicting functional classes can be found through the online server SVMProt [87] at <http://jing.cz3.nus.edu.sg/cgi-bin/svmprot.cgi>.

## TOWARDS APO STRUCTURE PREDICTION

Wei and Altman [25] reported the development of the FEATURE program (a machine learning system) that utilizes a Bayesian approach to classify potential  $\text{Ca}^{2+}$ -binding sites based on positive identification of predefined  $\text{Ca}^{2+}$ -binding characteristics. Following identification of the distributions of these features using FEATURE, the query protein structure was embedded in a 3D-grid of 2 Å grid cubes, and a score was computed for each grid (e.g., query site) based on a scoring function that uses Bayes' Rule to combine prior known distributions and observed frequencies (e.g., derived from a training dataset and the query site) to indicate the likelihood of  $\text{Ca}^{2+}$ -binding in the queried site. An online server WebFEATURE is available at <http://feature.stanford.edu/webfeature/>. Besides being applied to static X-ray crystal structures, FEATURE has recently been applied to multiple conformations of protein parvalbumin  $\beta$  generated from molecular dynamics (MD) simulation [88]. For MD generated conformations of both holo and apo forms, FEATURE correctly identifies the calcium-binding sites and non-sites with some interesting fluctuations in score. The combination of machine learning techniques such as FEATURE with MD simulations may provide new insights into  $\text{Ca}^{2+}$ -binding affinity, persistence and the stability of calcium-binding sites.

More recently, Babor *et al.* demonstrated related progress towards the prediction of metal (Zn, Fe, Cu, Co, Ni and Mn ions) binding sites for given protein apo structures [89] based on previous statistical analysis results, indicating that transition metal-binding residues were almost exclusively limited to Cys, His, Asp and Glu (CHED) residues (Table 2) [64]. An online server related to this method is available at <http://ligin.weizmann.ac.il/ched>.

Currently the MUG algorithm developed by Wang *et al.* [82] has been modified to accurately prediction protein calcium-binding sites including multiple-binding sites that undergo local conformational change or side chain rotations upon calcium-binding. Given an apo (unbound) protein structure, this new approach (MUG<sup>SR</sup>) conducts a side chain rotation procedure according to a rotamer library [90] to find binding sites with possible side-chain movements (local conformational changes) caused by calcium binding. Given an apo protein structure, MUG<sup>SR</sup> is able to capture calcium binding sites that may undergo local conformational changes, which is validated by comparison with the corresponding holo protein structure sharing more than 98% sequence similarity with the apo protein. For the test dataset consisting of 47 documented binding sites in the holo structures, MUG<sup>SR</sup> was able to predict 44/47 sites using the corresponding apo structures (manuscript in preparation).

#### a. Application of Algorithms for the Prediction or Design of Ca<sup>2+</sup>-Binding Proteins

Pre-experimental, computational design of Ca<sup>2+</sup>-binding proteins facilitates the transition from prediction and theory to empirical analysis and application. Various computational or prediction methods have been utilized for the rational design of proteins, the objective being to identify target sequences or structures with increased probability of obtaining some desired characteristics. This design phase may include sequential and statistical analysis, structural modeling, charge analysis, rotamer analysis and energy minimization as precursors to synthesis of novel proteins (Fig. (3)).

Previous work by Yang and coworkers [37] has shown that different classes of naturally evolved calcium binding sites can be re-identified based on local geometric properties using DEZYMER [83] which utilizes a two-step process to generate one or more potential ligand binding sites in proteins (apo or holo) of known structure.

The subsequent modification/engineering of Ca<sup>2+</sup>-binding proteins may be accomplished by either site-directed mutagenesis or through a grafting approach [77,91–93]. For the study of Ca<sup>2+</sup>-binding proteins, these well-established methods usually involve restructuring the charge environment in the binding site [94,95] which may alter the binding affinity. Both design and grafting have been applied to overcome the complexities encountered in cooperative, multi-site binding associated with natural Ca<sup>2+</sup>-binding proteins, where novel proteins have been engineered with a single Ca<sup>2+</sup>-binding site in order to dissect the key structural factors that control Ca<sup>2+</sup>-binding affinity, conformational change and cooperativity, as well as to function as novel reagents for research. Several *de novo* calcium-binding sites have been designed into a non-calcium binding protein CD2 with strong metal selectivity based on the local geometric properties and common chemical features of calcium binding sites in proteins [96]. The NMR solution structure of a novel protein (1t6w.pdb) revealed that Ca.CD2 binds Ca<sup>2+</sup> at the intended site with the designed arrangement, which validated a general strategy for designing *de novo* Ca<sup>2+</sup>-binding proteins [97].

Ye *et al.* reported the first estimation of the intrinsic Ca<sup>2+</sup> affinities of the four EF-Hand loops of calmodulin and a better estimation of the cooperativity of coupled EF-Hand proteins based on a designed grafting approach [77]. This approach has also been successfully applied to obtain site-specific calcium binding affinity of a predicted single EF-Hand motif in the non-structural protein of rubella virus [35] and Non-EF-Hand continuous calcium binding sites in GPCR, pumps and channels.

Shifman *et al.* observed a method of rational design to modify CaM for domain limited binding of only two Ca<sup>2+</sup> ions (either C-terminal or N-terminal) to circumvent cooperativity problems with CaM-Ca<sup>2+</sup> complexes and their interaction with protein kinase II [98]. The program ORBIT [84] was applied to minimize the energy of a given protein by optimizing the sequence, which was based on empirical atom-based force field calculation and a fast side-chain selection

algorithm. Far-UV circular dichroism was then applied to evaluate correct folding of the engineered CaMs. Their results indicated that CaM, bound with only two  $\text{Ca}^{2+}$  ions in the C-terminal could both bind to CaMKII and partially activate the kinase.

Recent studies based on rational design have shown that calcium and lanthanide binding affinities and protein stability can be systematically tuned by their arrangement in the coordination shell and its protein environment [49,99]. Jones *et al.* analyzed electrostatic interactions and engineered a novel  $\text{Ca}^{2+}$ -binding site on CD2, which was found to increase its CD48 binding affinity [99]. In addition, Li *et al.* further designed by site-directed mutagenesis a calcium-dependent CD2, trigger the conformation of which can be reversibly switched upon calcium binding without using coupled EF-Hand motifs [97,100]. The extensive *in silico* design of this protein variant based on pre-experimental analysis of structure, charge interactions and energy minimizations, provides an excellent example illustrating the progression from prediction and modeling to the creation of a desired product. This designed  $\text{Ca}^{2+}$  binding protein is very useful since it serves not only as a model system for understanding  $\text{Ca}^{2+}$  dependent cell adhesion but also as a basis for designing  $\text{Ca}^{2+}$ -dependent trigger proteins for the control of protein structures and conformations by metal binding.

Bunick *et al.* reported efforts to design a variant of calbindin D9k capable of responding to  $\text{Ca}^{2+}$ -binding with conformational changes similar to CaM in order to understand how differences in the amino acid sequence lead to differences in the  $\text{Ca}^{2+}$ -binding response [101]. Calbindin D9k was re-engineered with 15 mutations based on comparative analyses of sequence and structures combined with model building using MODELLER [102] to generate calbindo-modulin (CBM-1), which did not exhibit any non-native-like molten globule properties despite the large number, and, in some cases, the nonconservative nature, of the mutations.  $\text{Ca}^{2+}$ -induced changes in CD intensity and in the binding of the hydrophobic probe, ANS, implied that CBM-1 did undergo  $\text{Ca}^{2+}$  sensor-like conformational changes. The X-ray crystal structure of  $\text{Ca}^{2+}$ -CBM-1 determined at 1.44 Å resolution revealed the anticipated increase in hydrophobic surface area relative to the wild-type protein.

Huang *et al.* recently identified several calcium binding sites in the extracellular domain of calcium sensing receptors using algorithms based on the geometric description and surface electrostatic potentials [5].  $\text{Ca}^{2+}$ -sensing receptors (CaSRs) represent a class of receptors that respond to changes in the extracellular  $\text{Ca}^{2+}$  concentration ( $[\text{Ca}^{2+}]_o$ ) and activate multiple signaling pathways. A major barrier to advancing our understanding of the role of  $\text{Ca}^{2+}$  in regulating CaSRs is the lack of adequate information regarding the locations of their  $\text{Ca}^{2+}$ -binding sites due to the absence of a solved three-dimensional structure and rapid off rates related to low  $\text{Ca}^{2+}$ -binding affinities. In the absence of a solved CaSR structure, Huang *et al.* used SWISS-MODEL [76] and MODELLER to generate a putative CaSR structure model that was based on its homology with the previously-solved mGluR structure. Mutation of the predicted ligand residues in the full-length CaSR caused abnormal responses to  $[\text{Ca}^{2+}]_o$ , similar to those observed with naturally occurring activating or inactivating mutations of the CaSR, supporting the essential role of these predicted  $\text{Ca}^{2+}$ -binding sites in the sensing capability of the CaSR.

In a later related work, Huang *et al.* employed a similar computational strategy to probe the intrinsic  $\text{Ca}^{2+}$ -binding properties of predicted CaSR  $\text{Ca}^{2+}$ -binding sites [103]. For this study, two predicted continuous  $\text{Ca}^{2+}$ -binding sequences were individually engineered into a scaffold protein provided by a non- $\text{Ca}^{2+}$ -binding protein, CD2. Metal-binding affinities of these predicted sites in the CaSR were calculated by monitoring aromatic-sensitized  $\text{Tb}^{3+}$  fluorescence energy transfer. Removal of the predicted  $\text{Ca}^{2+}$ -binding ligands resulted in the loss or significant weakening of cation binding. The potential  $\text{Ca}^{2+}$ -binding residues were shown to be involved in  $\text{Ca}^{2+}/\text{Ln}^{3+}$  binding by high resolution NMR and site-directed

mutagenesis, further validating predictions of  $\text{Ca}^{2+}$ -binding sites within the extracellular domain of the CaSR. To further evaluate the potential calcium binding capabilities of predicted noncontinuous  $\text{Ca}^{2+}$ -binding sites in the ECD, the intact CaSR was dissected into three globular subdomains, each of which contained two to three predicted  $\text{Ca}^{2+}$ -binding sites. This approach allowed for analysis of the mechanisms underlying the binding of multiple metal ions to extended polypeptides derived from a location within the ECD of the CaSR, which would be anticipated to more closely mimic the structure of the native CaSR ECD. Studies on these subdomains suggested the existence of multiple metal-binding sites and metal-induced conformational changes that might be responsible for the switching on and off of the CaSR by the transition between its open inactive form and closed active form [103].

Application of these methods has also been used to develop novel, protein-based reagents. A rational grafting approach was used to develop calcium sensors capable of real-time quantitative  $\text{Ca}^{2+}$  concentration measurements in specific sub-cellular environments without using natural  $\text{Ca}^{2+}$  binding proteins such as calmodulin [104]. These engineered  $\text{Ca}^{2+}$  sensors have been shown to exhibit large ratiometric fluorescence and absorbance changes upon  $\text{Ca}^{2+}$  binding with affinities corresponding to the  $\text{Ca}^{2+}$  concentrations found in the ER ( $K_d$  values ranging from 0.4 to 2 mM). The developed  $\text{Ca}^{2+}$  sensor was successfully targeted to the ER of mammalian cell lines to monitor  $\text{Ca}^{2+}$  changes occurring in this organelle in response to stimulation with agonists. Similarly, computational design was utilized in the recent development of a new class of protein-based MRI contrast agents with significantly improved longitudinal and transverse relaxation [35,105].

## CONCLUSIONS AND PERSPECTIVES

The evolution of bioinformatics has led to an incredibly diverse set of algorithms, databases and tools that are rapidly becoming indispensable to all scientific inquiry in fields related to biology, chemistry and medicine. The availability of structured data collections, prediction algorithms and software for modeling structural and energetic characteristics has facilitated an important new phase in research, allowing for pre-experimental evaluation of hypotheses and the rational design of research methodologies. Due to the scope of the field, this review was restricted to a concentrated focus with respect to  $\text{Ca}^{2+}$ -binding as a central theme; however, it should be clear that the studies presented here can be extended beyond CaBPs as summarized in this work. The progression from prediction to development illustrates not only the increasing sophistication of bioinformatics tools, but their relative applicability to move beyond theoretical models and provide precise data for real-world design and development. Within the context of  $\text{Ca}^{2+}$ -binding research, our ability to make assumptions regarding the structure and function of CaBPs has advanced in step with the evolution of algorithms capable of recognizing or predicting binding sites, and the development of tools to predict binding sites in apo-structures may become routine in the near future. Using prediction as a basis for rational design, it is anticipated that the engineering of proteins with precise affinity and metal selectivity will significantly improve our ability to provide function-specific proteins for therapeutic, drug-delivery and diagnostic purposes.

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## ABBREVIATIONS

**CaBP**      Calcium-Binding Protein

<b>CN</b>	Coordination Number
<b>FC</b>	Formal Charge
<b>PDB</b>	Protein Data Bank

## References

1. Holmes KC, Popp D, Gebhard W, Kabsch W. Atomic model of the actin filament. *Nature* 1990;347:44–9. [PubMed: 2395461]
2. Herzberg O, Moulton J, James MN. A model for the Ca<sup>2+</sup>-induced conformational transition of troponin C: a trigger for muscle contraction. *J Biol Chem* 1986;261:2638–44. [PubMed: 3949740]
3. Mann KG, Nesheim ME, Church WR, Haley P, Krishnaswamy S. Surface-dependent reactions of the vitamin K-dependent enzyme complexes. *Blood* 1990;76:1–16. [PubMed: 2194585]
4. Nelson MR, Chazin WJ. An interaction-based analysis of calcium-induced conformational changes in Ca<sup>2+</sup> sensor proteins. *Protein Sci* 1998;7:270–82. [PubMed: 9521102]
5. Huang Y, Zhou Y, Yang W, et al. Identification and dissection of Ca(2+)-binding sites in the extracellular domain of Ca(2+)-sensing receptor. *J Biol Chem* 2007;282:19000–10. [PubMed: 17478419]
6. Hofer AM, Curci S, Doble MA, Brown EM, Soybel DI. Intercellular communication mediated by the extracellular calcium-sensing receptor. *Nat Cell Biol* 2000;2:392–8. [PubMed: 10878803]
7. Bower MJ, Cohen FE, Dunbrack RL Jr. Prediction of protein side-chain rotamers from a backbone-dependent rotamer library: a new homology modeling tool. *J Mol Biol* 1997;267:1268–82. [PubMed: 9150411]
8. Gunasekaran K, Nussinov R. How different are structurally flexible and rigid binding sites? Sequence and structural features discriminating proteins that do and do not undergo conformational change upon ligand binding. *J Mol Biol* 2007;365:257–273. [PubMed: 17059826]
9. Eyal E, Najmanovich R, Edelman M, Sobolev V. Protein side-chain rearrangement in regions of point mutations. *Proteins* 2003;50:272–82. [PubMed: 12486721]
10. Najmanovich R, Kuttner J, Sobolev V, Edelman M. Side-chain flexibility in proteins upon ligand binding. *Proteins* 2000;39:261–8. [PubMed: 10737948]
11. Yamashita MM, Wesson L, Eisenman G, Eisenberg D. Where metal ions bind in proteins. *Proc Natl Acad Sci USA* 1990;87:5648–52. [PubMed: 2377604]
12. Bagley SC, Altman RB. Characterizing the microenvironment surrounding protein sites. *Protein Sci* 1995;4:622–35. [PubMed: 7613462]
13. Dudev T, Lim C. Effect of carboxylate-binding mode on metal binding/selectivity and function in proteins. *Acc Chem Res* 2007;40:85–93. [PubMed: 17226948]
14. Dudev T, Lim C. Principles governing Mg, Ca, and Zn binding and selectivity in proteins. *Chem Rev* 2003;103:773–87. [PubMed: 12630852]
15. Dudev T, Lin YL, Dudev M, Lim C. First-second shell interactions in metal binding sites in proteins: a PDB survey and DFT/CDM calculations. *J Am Chem Soc* 2003;125:3168–80. [PubMed: 12617685]
16. Babu CS, Dudev T, Casareno R, Cowan JA, Lim C. A combined experimental and theoretical study of divalent metal ion selectivity and function in proteins: application to *E. coli* ribonuclease H1. *J Am Chem Soc* 2003;125:9318–28. [PubMed: 12889961]
17. Chakrabarti P. Geometry of interaction of metal ions with histidine residues in protein structures. *Protein Eng* 1990;4:57–63. [PubMed: 2290835]
18. Chakrabarti P. Geometry of interaction of metal ions with sulfur-containing ligands in protein structures. *Biochemistry* 1989;28:6081–5. [PubMed: 2775752]
19. Chakrabarti P. Interaction of metal ions with carboxylic and carboxamide groups in protein structures. *Protein Eng* 1990;4:49–56. [PubMed: 2290834]
20. Glusker, JP.; Lewis, M.; Rossi, M. *Crystal structure analysis for chemists and biologists*. New York: VCH; 1994.

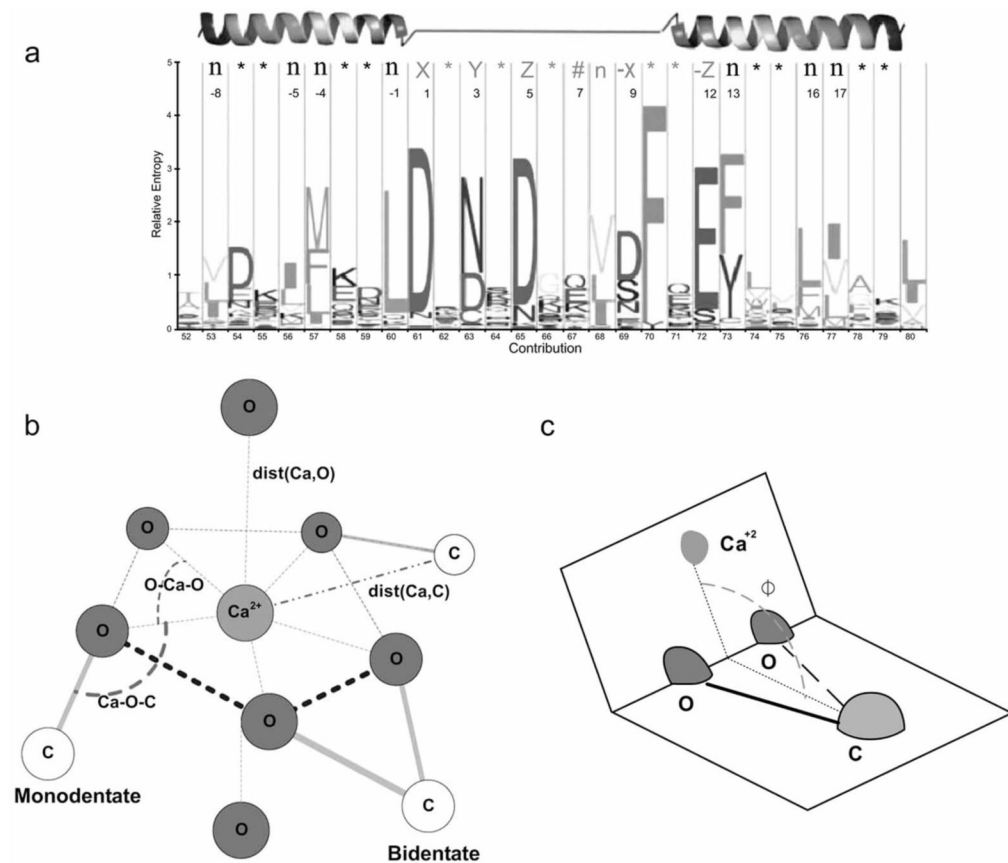


21. Harding MM. Geometry of metal-ligand interactions in proteins. *Acta Crystallogr D Biol Crystallogr* 2001;57:401–11. [PubMed: 11223517]
22. Harding MM. The geometry of metal-ligand interactions relevant to proteins. *Acta Crystallogr D Biol Crystallogr* 1999;55:1432–43. [PubMed: 10417412]
23. Harding MM. The geometry of metal-ligand interactions relevant to proteins. II. Angles at the metal atom, additional weak metal-donor interactions. *Acta Crystallogr D Biol Crystallogr* 2000;56:857–67. [PubMed: 10930832]
24. Glusker JP. Structural aspects of metal liganding to functional groups in proteins. *Adv Protein Chem* 1991;42:1–76. [PubMed: 1793004]
25. Wei, L.; Altman, RB. *Pac Symp Biocomput World Scientific*. 1998. Recognizing protein binding sites using statistical descriptions of their 3D environments.
26. Pidcock E, Moore GR. Structural characteristics of protein binding sites for calcium and lanthanide ions. *J Biol Inorg Chem* 2001;6:479–89. [PubMed: 11472012]
27. Nayal M, Di Cera E. Predicting Ca(2+)-binding sites in proteins. *Proc Natl Acad Sci USA* 1994;91:817–21. [PubMed: 8290605]
28. Kirberger M, Wang X, Deng H, Yang W, Chen G, Yang JJ. Statistical analysis of structural characteristics of protein Ca(2+)-binding sites. *J Biol Inorg Chem* 2008;13:1169–81. [PubMed: 18594878]
29. McPhalen CA, Strynadka NC, James MN. Calcium-binding sites in proteins: a structural perspective. *Adv Protein Chem* 1991;42:77–144. [PubMed: 1793008]
30. Hill E, Broadbent ID, Chothia C, Pettitt J. Cadherin superfamily proteins in *Caenorhabditis elegans* and *Drosophila melanogaster*. *J Mol Biol* 2001;305:1011–24. [PubMed: 11162110]
31. Truong K, Ikura M. The cadherin superfamily database. *J Struct Funct Genomics* 2002;2:135–43. [PubMed: 12836704]
32. Yang, JJ.; Yang, W. *Encyclopedia of inorganic chemistry*. New York: Wiley; 2005.
33. Rigden DJ, Galperin MY. The Dx Dx DG motif for calcium binding: multiple structural contexts and implications for evolution. *J Mol Biol* 2004;343:971–84. [PubMed: 15476814]
34. Marsden BJ, Shaw GS, Sykes BD. Calcium binding proteins. Elucidating the contributions to calcium affinity from an analysis of species variants and peptide fragments. *Biochem Cell Biol* 1990;68:587–601. [PubMed: 2198059]
35. Zhou Y, Yang W, Kirberger M, Lee HW, Ayalasomayajula G, Yang JJ. Prediction of EF-hand calcium-binding proteins and analysis of bacterial EF-hand proteins. *Proteins* 2006;65:643–55. [PubMed: 16981205]
36. Katz AK, Glusker JP, Beebe SA, Bock CW. Calcium ion coordination: a comparison with that of beryllium, magnesium, and zinc. *J Am Chem Soc* 1996;118:5752–63.
37. Yang W, Lee HW, Hellinga H, Yang JJ. Structural analysis, identification, and design of calcium-binding sites in proteins. *Proteins* 2002;47:344–56. [PubMed: 11948788]
38. Kawasaki H, Kretsinger RH. Calcium-binding proteins 1: EF-hands. *Protein Profile* 1995;2:297–490. [PubMed: 7553064]
39. Falke JJ, Drake SK, Hazard AL, Peersen OB. Molecular tuning of ion binding to calcium signaling proteins. *Q Rev Biophys* 1994;27:219–90. [PubMed: 7899550]
40. Marsden BJ, Hodges RS, Sykes BD. 1H-NMR studies of synthetic peptide analogues of calcium-binding site III of rabbit skeletal troponin C: effect on the lanthanum affinity of the interchange of aspartic acid and asparagine residues at the metal ion coordinating positions. *Biochemistry* 1988;27:4198–206. [PubMed: 3415981]
41. Strynadka NC, James MN. Crystal structures of the helix-loop-helix calcium-binding proteins. *Annu Rev Biochem* 1989;58:951–98. [PubMed: 2673026]
42. Einspahr, H.; Bugg, C. Calcium and its role in biology. In: Sigel, H., editor. *Crystal Structure Studies of Calcium Complexes and Implications for Biological Systems*. New York: M. Dekker; 1984. p. 52-97.
43. Harding MM. Small revisions to predicted distances around metal sites in proteins. *Acta Crystallogr D Biol Crystallogr* 2006;62:678–82. [PubMed: 16699196]

44. Swain AL, Kretsinger RH, Amma EL. Restrained least squares refinement of native (calcium) and cadmium-substituted carp parvalbumin using X-ray crystallographic data at 1.6-Å resolution. *J Biol Chem* 1989;264:16620–8. [PubMed: 2777802]
45. Vyas MN, Jacobson BL, Quioco FA. The calcium-binding site in the galactose chemoreceptor protein. Crystallographic and metal-binding studies. *J Biol Chem* 1989;264:20817–21. [PubMed: 2684986]
46. Martin, R. *Bioinorganic chemistry of calcium*. New York: Marcel Dekker; 1984.
47. Drake SK, Lee KL, Falke JJ. Tuning the equilibrium ion affinity and selectivity of the EF-hand calcium binding motif: substitutions at the gateway position. *Biochemistry* 1996;35:6697–705. [PubMed: 8639620]
48. Drake SK, Zimmer MA, Miller CL, Falke JJ. Optimizing the metal binding parameters of an EF-hand-like calcium chelation loop: coordinating side chains play a more important tuning role than chelation loop flexibility. *Biochemistry* 1997;36:9917–26. [PubMed: 9245425]
49. Maniccia AW, Yang W, Li SY, Johnson JA, Yang JJ. Using protein design to dissect the effect of charged residues on metal binding and protein stability. *Biochemistry* 2006;45:5848–56. [PubMed: 16669627]
50. Linse S, Helmersson A, Forsen S. Calcium binding to calmodulin and its globular domains. *J Biol Chem* 1991;266:8050–4. [PubMed: 1902469]
51. Martin SR, Linse S, Johansson C, Bayley PM, Forsen S. Protein surface charges and Ca<sup>2+</sup> binding to individual sites in calbindin D9k: stopped-flow studies. *Biochemistry* 1990;29:4188–93. [PubMed: 2193686]
52. Cheng Y, Sequeira SM, Malinina L, Tereshko V, Sollner TH, Patel DJ. Crystallographic identification of Ca<sup>2+</sup> and Sr<sup>2+</sup> coordination sites in synaptotagmin I C2B domain. *Protein Sci* 2004;13:2665–72. [PubMed: 15340165]
53. Korndorfer IP, Brueckner F, Skerra A. The crystal structure of the human (S100A8/S100A9)2 heterotetramer, calprotectin, illustrates how conformational changes of interacting alpha-helices can determine specific association of two EF-hand proteins. *J Mol Biol* 2007;370:887–98. [PubMed: 17553524]
54. Kuboniwa H, Tjandra N, Grzesiek S, Ren H, Klee CB, Bax A. Solution structure of calcium-free calmodulin. *Nat Struct Biol* 1995;2:768–76. [PubMed: 7552748]
55. Zhang M, Tanaka T, Ikura M. Calcium-induced conformational transition revealed by the solution structure of apo calmodulin. *Nat Struct Biol* 1995;2:758–67. [PubMed: 7552747]
56. Davis IW, Arendall WB 3rd, Richardson DC, Richardson JS. The backbone motion: how protein backbone shrugs when a sidechain dances. *Structure* 2006;14:265–74. [PubMed: 16472746]
57. Esposito L, De Simone A, Zagari A, Vitagliano L. Correlation between omega and psi dihedral angles in protein structures. *J Mol Biol* 2005;347:483–7. [PubMed: 15755444]
58. Dunbrack RL Jr, Karplus M. Conformational analysis of the backbone-dependent rotamer preferences of protein sidechains. *Nat Struct Biol* 1994;1:334–40. [PubMed: 7664040]
59. Chrysin ED, Brew K, Acharya KR. Crystal structures of apo- and holo-bovine alpha-lactalbumin at 2.2-Å resolution reveal an effect of calcium on inter-lobe interactions. *J Biol Chem* 2000;275:37021–9. [PubMed: 10896943]
60. Nelson MR, Chazin WJ. Structures of EF-hand Ca(2+)-binding proteins: diversity in the organization, packing and response to Ca<sup>2+</sup> binding. *Biomaterials* 1998;11:297–318. [PubMed: 10191495]
61. Iyer LK, Qasba PK. Molecular dynamics simulation of alpha-lactalbumin and calcium binding c-type lysozyme. *Protein Eng* 1999;12:129–39. [PubMed: 10195284]
62. Shesham RD, Bartolotti LJ, Li Y. Molecular dynamics simulation studies on Ca<sup>2+</sup>-induced conformational changes of annexin I. *Protein Eng Des Sel* 2008;21:115–20. [PubMed: 18283055]
63. Richardson RC, King NM, Harrington DJ, Sun H, Royer WE, Nelson DJ. X-Ray crystal structure and molecular dynamics simulations of silver hake parvalbumin (Isoform B). *Protein Sci* 2000;9:73–82. [PubMed: 10739249]
64. Babor M, Greenblatt HM, Edelman M, Sobolev V. Flexibility of metal binding sites in proteins on a database scale. *Proteins* 2005;59:221–30. [PubMed: 15726624]

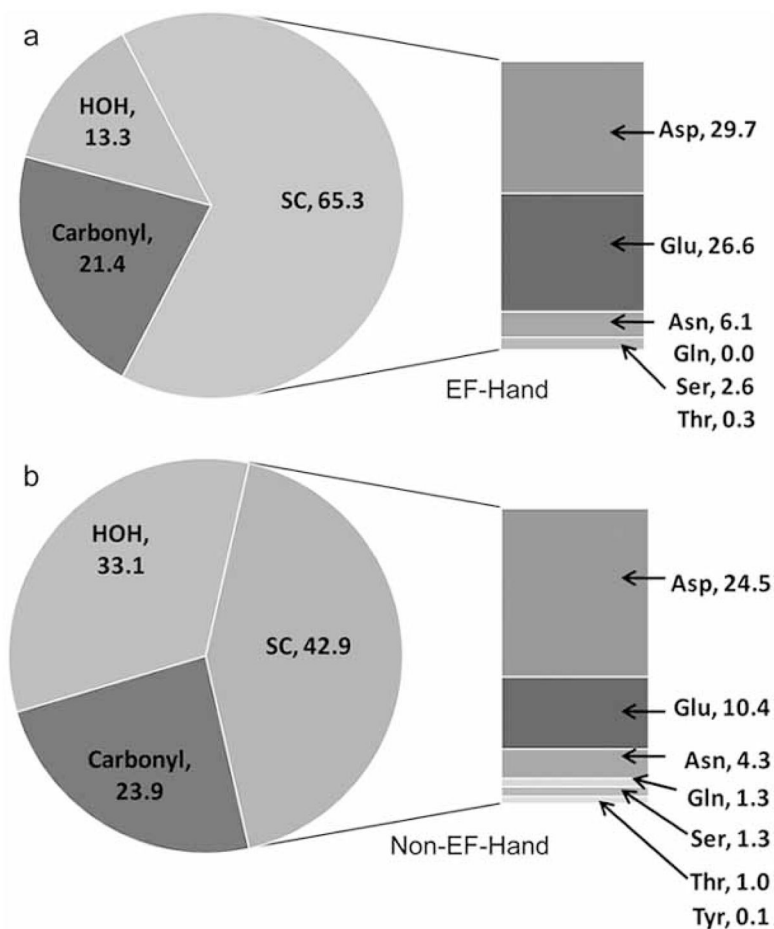
65. Zhao S, Goodsell DS, Olson AJ. Analysis of a data set of paired uncomplexed protein structures: new metrics for side-chain flexibility and model evaluation. *Proteins* 2001;43:271–9. [PubMed: 11288177]
66. Haiech J, Sallantin J. Computer search of calcium binding sites in a gene data bank: use of learning techniques to build an expert system. *Biochimie* 1985;67:555–60. [PubMed: 3839696]
67. Kretsinger RH, Nockolds CE. Carp muscle calcium-binding protein. II. Structure determination and general description. *J Biol Chem* 1973;248:3313–26. [PubMed: 4700463]
68. Galzi JL, Bertrand S, Corringer PJ, Changeux JP, Bertrand D. Identification of calcium binding sites that regulate potentiation of a neuronal nicotinic acetylcholine receptor. *EMBO J* 1996;15:5824–32. [PubMed: 8918460]
69. Simons JW, van Kampen MD, Ubarretxena-Belandia I, et al. Identification of a calcium binding site in *Staphylococcus hyicus* lipase: generation of calcium-independent variants. *Biochemistry* 1999;38:2–10. [PubMed: 9890877]
70. Rastogi, PA. MacVector: Integrated Sequence Analysis for the Macintosh, in *Bioinformatics Methods and Protocols*. Misener, S.; Krawetz, SA., editors. Humana Press; Totowa, N.J.: 2000.
71. Zhou Y, Tzeng WP, Yang W, et al. Identification of a Ca<sup>2+</sup>-binding domain in the rubella virus nonstructural protease. *J Virol* 2007;81:7517–28. [PubMed: 17475644]
72. McGuffin LJ, Bryson K, Jones DT. The PSIPRED protein structure prediction server. *Bioinformatics* 2000;16:404–5. [PubMed: 10869041]
73. Cuff JA, Clamp ME, Siddiqui AS, Finlay M, Barton GJ. JPred: a consensus secondary structure prediction server. *Bioinformatics* 1998;14:892–3. [PubMed: 9927721]
74. Rost B, Yachdav G, Liu J. The PredictProtein server. *Nucleic Acids Res* 2004;32:W321–6. [PubMed: 15215403]
75. Deng H, Chen G, Yang W, Yang JJ. Predicting calcium-binding sites in proteins - a graph theory and geometry approach. *Proteins* 2006;64:34–42. [PubMed: 16617426]
76. Schwede T, Kopp J, Guex N, Peitsch MC. SWISS-MODEL: an automated protein homology-modeling server. *Nucleic Acids Res* 2003;31:3381–5. [PubMed: 12824332]
77. Ye Y, Lee HW, Yang W, Shealy S, Yang JJ. Probing site-specific calmodulin calcium and lanthanide affinity by grafting. *J Am Chem Soc* 2005;127:3743–50. [PubMed: 15771508]
78. Schymkowitz JW, Rousseau F, Martins IC, Ferkinghoff-Borg J, Stricher F, Serrano L. Prediction of water and metal binding sites and their affinities by using the Fold-X force field. *Proc Natl Acad Sci USA* 2005;102:10147–52. [PubMed: 16006526]
79. Guerois R, Serrano L. The SH3-fold family: experimental evidence and prediction of variations in the folding pathways. *J Mol Biol* 2000;304:967–82. [PubMed: 11124040]
80. Pitt WR, Goodfellow JM. Modelling of solvent positions around polar groups in proteins. *Protein Eng* 1991;4:531–7. [PubMed: 1891460]
81. Lin HH, Han LY, Zhang HL, et al. Prediction of the functional class of metal-binding proteins from sequence derived physico-chemical properties by support vector machine approach. *BMC Bioinformatics* 2006;7 (Suppl 5):S13. [PubMed: 17254297]
82. Wang X, Kirberger M, Qiu F, Chen G, Yang JJ. Towards predicting Ca(2+)-binding sites with different coordination numbers in proteins with atomic resolution. *Proteins* 2008;75:787–98. [PubMed: 19003991]
83. Hellinga HW, Richards FM. Construction of new ligand binding sites in proteins of known structure. I. Computer-aided modeling of sites with pre-defined geometry. *J Mol Biol* 1991;222:763–85. [PubMed: 1749000]
84. Dahiyat BI, Mayo SL. *De novo* protein design: fully automated sequence selection. *Science* 1997;278:82–7. [PubMed: 9311930]
85. Sodhi JS, Bryson K, McGuffin LJ, Ward JJ, Wernisch L, Jones DT. Predicting metal-binding site residues in low-resolution structural models. *J Mol Biol* 2004;342:307–20. [PubMed: 15313626]
86. Lin CT, Lin KL, Yang CH, Chung IF, Huang CD, Yang YS. Protein metal binding residue prediction based on neural networks. *Int J Neural Syst* 2005;15:71–84. [PubMed: 15912584]

87. Cai CZ, Han LY, Ji ZL, Chen X, Chen YZ. SVM-Prot: web-based support vector machine software for functional classification of a protein from its primary sequence. *Nucleic Acids Res* 2003;31:3692–7. [PubMed: 12824396]
88. Glazer DS, Radmer RJ, Altman RB. Combining molecular dynamics and machine learning to improve protein function recognition. *Pac Symp Biocomput* 2008;13:332–343. [PubMed: 18229697]
89. Babor M, Gerzon S, Raveh B, Sobolev V, Edelman M. Prediction of transition metal-binding sites from apo protein structures. *Proteins* 2008;70:208–17. [PubMed: 17657805]
90. Dunbrack RL Jr, Cohen FE. Bayesian statistical analysis of protein side-chain rotamer preferences. *Protein Sci* 1997;6:1661–81. [PubMed: 9260279]
91. Ye Y, Lee HW, Yang W, et al. Metal binding affinity and structural properties of an isolated EF-loop in a scaffold protein. *Protein Eng* 2001;14:1001–13. [PubMed: 11809931]
92. Toma S, Campagnoli S, Margarit I, et al. Grafting of a calcium-binding loop of thermolysin to *Bacillus subtilis* neutral protease. *Biochemistry* 1991;30:97–106. [PubMed: 1899021]
93. Ye Y, Lee HW, Yang W, Yang JJ. Calcium and lanthanide affinity of the EF-loops from the C-terminal domain of calmodulin. *J Inorg Biochem* 2005;99:1376–83. [PubMed: 15917089]
94. Wilkins AL, Ye Y, Yang W, Lee HW, Liu ZR, Yang JJ. Metal-binding studies for a de novo designed calcium-binding protein. *Protein Eng* 2002;15:571–4. [PubMed: 12200539]
95. Falke JJ, Snyder EE, Thatcher KC, Voertler CS. Quantitating and engineering the ion specificity of an EF-hand-like Ca<sup>2+</sup> binding. *Biochemistry* 1991;30:8690–7. [PubMed: 1653605]
96. Yang W, Jones LM, Isley L, et al. Rational design of a calcium-binding protein. *J Am Chem Soc* 2003;125:6165–71. [PubMed: 12785848]
97. Yang W, Wilkins AL, Ye Y, et al. Design of a calcium-binding protein with desired structure in a cell adhesion molecule. *J Am Chem Soc* 2005;127:2085–93. [PubMed: 15713084]
98. Shifman JM, Choi MH, Mihalas S, Mayo SL, Kennedy MB. Ca<sup>2+</sup>/calmodulin-dependent protein kinase II (CaMKII) is activated by calmodulin with two bound calciums. *Proc Natl Acad Sci USA* 2006;103:13968–73. [PubMed: 16966599]
99. Jones LM, Yang W, Maniccia AW, Harrison A, van der Merwe PA, Yang JJ. Rational design of a novel calcium-binding site adjacent to the ligand-binding site on CD2 increases its CD48 affinity. *Protein Sci* 2008;17:439–49. [PubMed: 18287277]
100. Li S, Yang W, Maniccia AW, et al. Rational design of a conformation-switchable Ca<sup>2+</sup>- and Tb<sup>3+</sup>-binding protein without the use of multiple coupled metal-binding sites. *FEBS J* 2008;275:5048–61. [PubMed: 18785925]
101. Bunick CG, Nelson MR, Mangahas S, et al. Designing sequence to control protein function in an EF-hand protein. *J Am Chem Soc* 2004;126:5990–8. [PubMed: 15137763]
102. Marti-Renom MA, Stuart AC, Fiser A, Sanchez R, Melo F, Sali A. Comparative protein structure modeling of genes and genomes. *Annu Rev Biophys Biomol Struct* 2000;29:291–325. [PubMed: 10940251]
103. Huang Y, Zhou Y, Castiblanco A, Yang W, Brown EM, Yang JJ. Multiple Ca(2+)-binding sites in the extracellular domain of the Ca(2+)-sensing receptor corresponding to cooperative Ca(2+) response. *Biochemistry* 2009;48(2):388–98. [PubMed: 19102677]
104. Zou J, Hofer AM, Lurtz MM, et al. Developing sensors for real-time measurement of high Ca<sup>2+</sup> concentrations. *Biochemistry* 2007;46:12275–88. [PubMed: 17924653]
105. Yang JJ, Yang J, Wei L, et al. Rational design of protein-based MRI contrast agents. *J Am Chem Soc* 2008;130:9260–7. [PubMed: 18576649]

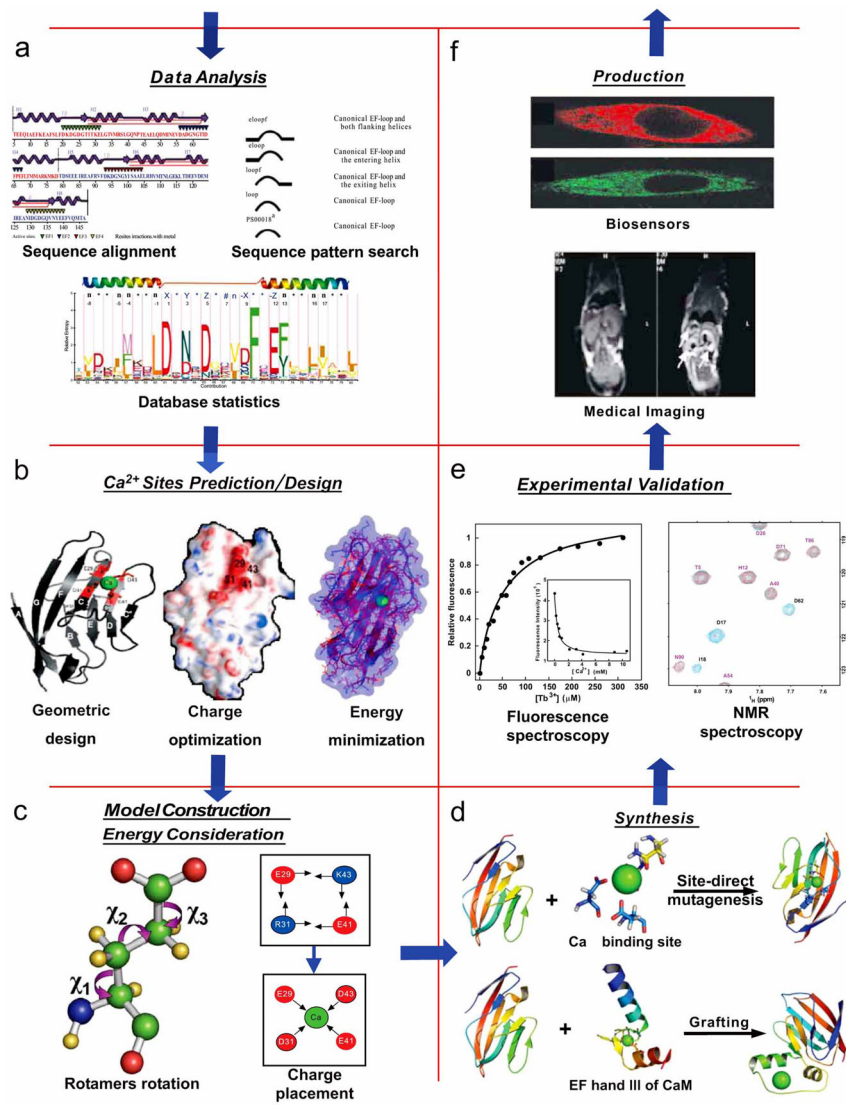


**Fig. 1.** (a) Canonical EF-Loop binding site. (b) Illustration of key EF-Hand structural characteristics. The physical relationships between the  $\text{Ca}^{2+}$  ion (Ca), the ligand oxygen (O), and the ligand oxygen atoms covalently-bound carbon (C) are defined by the angle  $\text{Ca}-\text{O}-\text{C}$  and distances  $\text{dist}(\text{Ca},\text{C})$  and  $\text{dist}(\text{Ca},\text{O})$ . (c) Dihedral angle of bidentate ligands between two planes formed by O-Ca-O and O-C-O.





**Fig. 2.** Distribution of ligand residues between water, carbonyl and sidechain (SC) residue oxygen ligands for (a) EF-Hand and (b) Non-EF-Hand.



**Fig. 3.** Schematic illustrating rational design of a  $Ca^{2+}$ -binding protein for biotechnological applications. (a) Calcium pattern search results and definition of  $Ca^{2+}$ -binding motifs, reproduced from Zhou *et al.* [35]. (b) Geometric design and charge optimization, and (c) charge placement for energy optimization, reproduced from Li *et al.* [100]. (d) General strategy for construction of  $Ca^{2+}$ -binding site on protein scaffold either by mutating existing residues (site-directed mutagenesis) or by inserting new residues (grafting). (e) Fluorescence spectrum (left) for identification of  $Ca^{2+}$ -binding site in Rubella virus, reproduced from Zhou [71] and NMR spectrum for designed CaBP, reproduced from Yang *et al.* [97]. (f) Application of designed proteins in medical imaging and as biosensors, reproduced from Yang *et al.* [105] and Zou *et al.* [104], respectively.

**Table 1**

Comparison of Ligand Distributions as Percentages of Total Ligand Numbers

Source	Water	SC Asp/Glu	SC Asn/Gln	BB Gly/Other	SC Hydroxyl
<i>a</i> Pidcock	27	25/10	6/<1	7/24	1
<i>b</i> Harding	40	34		22	4
<i>c</i> Kirberger EF	13 ( <sup>†</sup> 13)	30/27	6/0	21	3
<i>c</i> Kirberger NEF	33 ( <sup>†</sup> 25)	25/10	4/1	24	2

*a* [26]*b* [21]*c* [28] EF= EF-Hand, NEF=Non-EF-Hand.<sup>†</sup> If cutoff reduced to 2.9 Å.

SC=SideChain, BB=Backbone.

Table 2

## Summary of Prediction Algorithms

	Name	Input	Output	Method	Applicability
<i>a</i> Zhou	CaPS	Primary sequence	Binding pattern & region	Sequence pattern search	Identify EF-Hand and EF-like Ca <sup>2+</sup> -binding sites in proteins
<i>b</i> Yamashita	---	PDB file	Ca <sup>2+</sup> coordinates	Hydrophobicity contrast function	Predict metal (including Ca <sup>2+</sup> ) binding sites of insignificant conformational changes
<i>c</i> Nayal	---	PDB file	Ca <sup>2+</sup> coordinates	Valence function	Predict Ca <sup>2+</sup> -binding sites of insignificant conformational changes
<i>d</i> Wei	Feature	PDB file	Ca <sup>2+</sup> coordinates & physiochemical properties	Probability scoring function	Predict Ca <sup>2+</sup> -binding sites of insignificant conformational changes
<i>e</i> Lin	SVMProt	Primary sequence	Functional families of the input sequence	Machine learning	Assign functional families (including Ca <sup>2+</sup> and metal binding) to the input sequence
<i>f</i> Schymkowitz	---	PDB file	Ca <sup>2+</sup> coordinates & estimated binding affinity	Fold-X force field	Predict metal (include Ca <sup>2+</sup> ) binding sites of insignificant conformational changes
<i>g</i> Deng	GG	PDB file	Ca <sup>2+</sup> coordinates & ligand oxygen atoms	Graph theory	Predict Ca <sup>2+</sup> -binding sites with minor conformational changes
<i>h</i> Wang	MUG	PDB file	Ca <sup>2+</sup> coordinates & ligand oxygen atoms	Graph theory	Predict Ca <sup>2+</sup> -binding sites with minor conformational changes
<i>i</i> Babor	CHED	PDB file	Binding residues	Rotamer	Predict transition metal-binding sites with possible side-chain movement
<i>j</i> Hellinga	DEZYMER	PDB file	Binding residues & possible mutations	Rotamer	Suggest mutations for Ca <sup>2+</sup> -binding based on a PDB file
<i>k</i> Dahiyat	ORBIT	Backbone coordinates	Sequence appropriate for the backbone fold	Rotamer library & optimization	De novo design of protein (including CaBP) sequence based on a desired backbone fold

*a* [35].*b* [11].*c* [27].*d* [25].*e* [81].*f* [78].*g* [75].*h* [82].

$i$  [64],  
 $j$  [83],  
 $k$  [84].