

Caloric restriction or catalase inactivation extends yeast chronological lifespan by inducing H₂O₂ and superoxide dismutase activity

Ana Mesquita^a, Martin Weinberger^b, Alexandra Silva^a, Belém Sampaio-Marques^a, Bruno Almeida^a, Cecília Leão^a, Vítor Costa^{c,d}, Fernando Rodrigues^a, William C. Burhans^{b,1}, and Paula Ludovico^{a,1}

^aInstituto de Investigação em Ciências da Vida e Saúde (ICVS), Escola de Ciências da Saúde, Universidade do Minho, Campus de Gualtar, 4710-057 Braga, Portugal; ^bDepartment of Molecular and Cellular Biology, Roswell Park Cancer Institute, Buffalo, NY 14263; ^cInstituto de Biologia Molecular e Celular (IBMC), Universidade do Porto, 4150-180 Porto, Portugal; and ^dInstituto de Ciências Biomédicas Abel Salazar (ICBAS), Departamento de Biologia Molecular, Universidade do Porto, 4099-003 Porto, Portugal

Edited* by Fred Sherman, University of Rochester School of Medicine and Dentistry, Rochester, NY, and approved July 23, 2010 (received for review April 1, 2010)

The free radical theory of aging posits oxidative damage to macromolecules as a primary determinant of lifespan. Recent studies challenge this theory by demonstrating that in some cases, longevity is enhanced by inactivation of oxidative stress defenses or is correlated with increased, rather than decreased reactive oxygen species and oxidative damage. Here we show that, in *Saccharomyces cerevisiae*, caloric restriction or inactivation of catalases extends chronological lifespan by inducing elevated levels of the reactive oxygen species hydrogen peroxide, which activate superoxide dismutases that inhibit the accumulation of superoxide anions. Increased hydrogen peroxide in catalase-deficient cells extends chronological lifespan despite parallel increases in oxidative damage. These findings establish a role for hormesis effects of hydrogen peroxide in promoting longevity that have broad implications for understanding aging and age-related diseases.

aging | hydrogen peroxide | hormesis | antioxidant enzymes | oxidative damage

The longstanding free radical theory has guided investigations into the causes and consequences of aging for more than 50 y (1). However, the results of a number of recent studies have failed to provide support for the free radical theory or suggest that this theory is at best incomplete (2). Studies of naked mole rats, for example, demonstrated that this extremely long-lived rodent exhibits high levels of oxidative damage compared with mice or rats, whose lifespans are $\approx 1/10$ that of naked mole rats (3). In addition, caloric restriction (CR), which extends the lifespans of a variety of eukaryotic organisms, promotes longevity in *Caenorhabditis elegans* by a mechanism that involves increased oxidative stress (4). In fact, in contrast to the destructive effects of reactive oxygen species (ROS), recent evidence indicates that in mammals, hydrogen peroxide (H₂O₂) and other forms of ROS function as essential secondary messengers in the regulation of a variety of physiological processes (reviewed in ref. 5). For example, H₂O₂ activates pro-survival signaling pathways mediated by p53, NF- κ B, AP-1, and other molecules (6). Furthermore, increases in the intracellular steady-state production of H₂O₂ by *SOD2* overexpression can block the activation of cellular processes required for programmed cell death (7). However, a causal relationship between CR and effects on oxidative stress has been difficult to establish.

To better understand how CR impacts oxidative stress and longevity in the model organism *Saccharomyces cerevisiae*, in this study we examined intracellular levels of H₂O₂ and superoxide anions (O₂⁻), which are two forms of ROS implicated in aging in all eukaryotes, under CR and other conditions. Our findings indicate that CR or inactivation of catalases extends chronological lifespan (CLS) by inducing elevated levels of H₂O₂, which activate superoxide dismutases that inhibit the accumulation of O₂⁻. These findings establish a role for hormesis effects of H₂O₂ in promoting

longevity induced by CR conditions that are likely to be conserved in complex eukaryotes. In catalase-deficient cells, increased H₂O₂ extends CLS despite parallel increases in oxidative damage. This violates a fundamental tenet of the free radical theory that posits oxidative damage as a primary determinant of aging.

Results

Caloric Restriction or Inactivation of Catalases Extends *S. cerevisiae* Chronological Lifespan by Increasing Intracellular Levels of H₂O₂. CLS of budding yeast is determined by measuring the survival of non-dividing stationary phase cells, which is impacted by conserved factors that affect aging of postmitotic cells of complex eukaryotes, including humans (8). As reported previously (9), CR by decreasing the concentration of glucose extended CLS of budding yeast (Fig. 1A). Surprisingly, however, the CLS-extending effects of CR were accompanied by an increase in the fraction of cells containing high levels of intracellular ROS, detected by staining cells with dihydrorhodamine 123 (DHR) (Fig. 1B and Fig. S1). CR-induced increases in ROS were also detected by staining cells with 2',7'-dichlorodihydrofluorescein diacetate (H₂DCF-DA) (Fig. S2). CR also induced the activity of the peroxisomal catalase Cta1p, as well as the cytosolic catalase Ctt1p (Fig. S3), which are the two main H₂O₂ scavenging enzymes in this organism. However, increased catalase activity did not contribute to the CLS-extending effects of CR, because CLS was also extended by mutational inactivation of *CTA1* and the longer CLS of Δ *cta1* cells was extended further by CR (Fig. 1C). Similar results were obtained upon mutational inactivation of *CTT1* or pharmacological inhibition of the synthesis of glutathione, which scavenges H₂O₂ (Fig. S4). These findings identify H₂O₂ as a form of intracellular ROS induced by CR in stationary phase budding yeast cells and establish that catalase activity or glutathione synthesis inhibits longevity in the chronological aging model.

The longer CLS of Δ *cta1* cells was accompanied by an increased fraction of cells containing high levels of intracellular H₂O₂ under non-CR conditions (Fig. 1C and D). Under CR conditions, the fraction of Δ *cta1* cells exhibiting high intracellular levels of H₂O₂ was similar to CR wild-type cells or non-CR Δ *cta1* cells at day 6 and lower at day 12 (Fig. 1D). Increased levels of H₂O₂ were also detected in Δ *ctt1* and in Δ *cta1\Delta**ctt1* compared with wild-type cells

Author contributions: V.C., W.C.B., and P.L. designed research; A.M., M.W., A.S., B.S.-M., W.C.B., and P.L. performed research; M.W., B.S.-M., B.A., C.L., V.C., F.R., W.C.B., and P.L. analyzed data; and F.R., W.C.B., and P.L. wrote the paper.

The authors declare no conflict of interest.

*This Direct Submission article had a prearranged editor.

¹To whom correspondence may be addressed. E-mail: wburhans@buffalo.edu or pludovico@ecsaude.uminho.pt.

This article contains supporting information online at www.pnas.org/lookup/suppl/doi:10.1073/pnas.1004432107/-DCSupplemental.

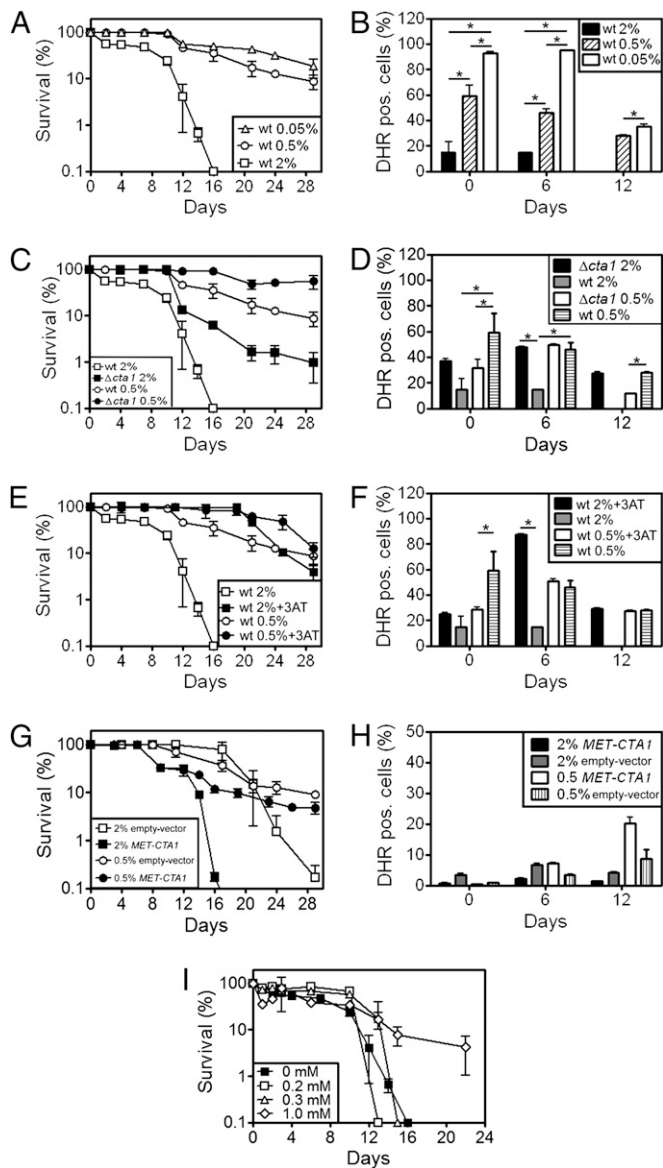


Fig. 1. Caloric restriction (CR) or inactivation of catalases extends *Saccharomyces cerevisiae* chronological lifespan by increasing intracellular levels of H₂O₂. Survival of (A) wild-type (BY4742) cells, (C) $\Delta cta1$ cells, (E) wild-type cells in the absence or presence of the catalase inhibitor 10 mM 3-amino-1,2,4-triazole (3AT), and (G) wild-type cells transformed with an empty vector or a plasmid that overexpresses *CTA1* ("MET-CTA1"). (I) Effects of ectopic exposure of wild-type cells to indicated concentrations of H₂O₂. Cell viability was measured at 2- to 3-d intervals beginning the day cultures achieved stationary phase (day 0) and is expressed as % survival compared with survival at day 0 (100%). Percentage of cells exhibiting high levels of intracellular ROS detected by FACS measurements of fluorescence of the probe dihydrorhodamine 123 (DHR) in (B) wild-type cells, (D) $\Delta cta1$ cells, (F) wild-type cells in the absence or presence of 3AT, and (H) wild-type cells transformed with an empty vector or a plasmid that overexpresses *CTA1* ("MET-CTA1"). Three to five biological replicates of each experiment were performed. Survival and DHR positive cell values are mean \pm SD or mean \pm SEM, respectively, in all experiments. Statistical significance ($*P < 0.05$) was determined by Student's *t*-test.

(Fig. S5). Pharmacological inactivation of catalases by 3-amino-1,2,4-triazole (3AT) also extended CLS, and CR extended the CLS of 3AT-treated cells even further (Fig. 1E). Treatment with 3AT also increased the fraction of cells containing high levels of intracellular H₂O₂ (Fig. 1F). However, CR did not increase the

fraction of 3AT-treated cells containing high intracellular levels of H₂O₂ further (Fig. 1F). Overexpression of *CTA1* had effects opposite to those associated with inactivation of catalases—it decreased CLS in non-CR and CR cells (Fig. 1G) and reduced the fraction of cells containing high levels of intracellular H₂O₂ (Fig. 1H). However, the fraction of cells containing high intracellular levels of H₂O₂ was also reduced (albeit to a lesser extent) in cells harboring an empty vector. This may reflect effects of the different medium required to maintain plasmids in these but not other experiments.

These observations suggest that H₂O₂ promotes longevity in the budding yeast CLS model. To address this possibility directly, we asked whether ectopic application of H₂O₂ over a range of concentrations from 0 to 1 mM would extend CLS of non-CR wild-type cells. The lowest H₂O₂ concentrations tested did not alter CLS compared with cells that were not exposed to H₂O₂ (Fig. 1I). In contrast, exposure to 1 mM H₂O₂ resulted in a significant increase in longevity (Fig. 1I). Overall these results indicate that H₂O₂ extends CLS of budding yeast and is an important component of the CLS-extending effects of CR.

CR is known to extend CLS by inhibiting the accumulation of acetic acid in culture medium (10). As reported earlier, buffering medium to eliminate acetic acid also extends CLS (10, 11). However, CR increased intracellular H₂O₂ in experiments performed in buffered medium (Fig. S6). This suggests that some of the CLS-extending effects of CR occur independently of reduced levels of acetic acid in medium. Furthermore, the CLS-extending effects of increased H₂O₂ detected in catalase mutants occurred in the absence of a change in medium pH compared with cultures of wild-type cells (Table S1). This establishes that the effects of increased H₂O₂ on CLS can occur independently of changes in levels of acetic acid in culture medium.

Elevated H₂O₂ Levels Induced by CR or Inactivation of Catalases Are Accompanied by a Reduction in Levels of Superoxide Anions.

To determine whether CR or other experimental manipulations also increase levels of superoxide anions, we measured O₂⁻ levels in stationary phase cells using dihydroethidium (DHE), which can detect O₂⁻ (12). In non-CR conditions, O₂⁻ levels increased in wild-type cells from day 0 to day 3 of stationary phase, whereas H₂O₂ levels remained unchanged (Fig. 2A). However, CR caused a significant reduction in O₂⁻ levels compared with levels in non-CR cells, despite a pronounced increase in H₂O₂ in the same cells (Fig. 2A). Similar to the effects of CR in wild-type cells, O₂⁻ levels were decreased and H₂O₂ levels were increased in $\Delta cta1$ compared with wild-type cells at day 0 and day 3 of stationary phase (Fig. 2A). O₂⁻ levels in stationary phase $\Delta cta1$ cells were also reduced by CR conditions, comparable to the effects of CR on stationary phase wild-type cells (Fig. 2A). The reduction in O₂⁻ levels in CR $\Delta cta1$ cells was accompanied by an increase in H₂O₂ at day 0 (Fig. 2A). Treatment of wild-type cells with 3AT or inactivation of both catalases ($\Delta cta1 ctt1$ cells) also reduced O₂⁻ levels at the same time that it increased intracellular H₂O₂ in the same cells (Fig. 2B and Fig. S5B). These findings suggest that the longevity-promoting effects of intracellular H₂O₂ in CR conditions or when catalases are inactivated are related to inhibition of the accumulation of O₂⁻. Although buffering medium to eliminate the effects of acetic acid also inhibits the accumulation of O₂⁻ in wild-type cells in stationary phase (11), CR reduced levels of O₂⁻ in buffered medium (Fig. S6). Therefore, CR inhibition of O₂⁻ accumulation is mediated by both acetic acid-dependent and -independent mechanisms.

Induction of Superoxide Dismutase Activity by Intracellular H₂O₂.

Ectopic application of sublethal concentrations of H₂O₂ in budding yeast induces the transcription of genes encoding both the cytosolic Cu/Zn-dependent and the mitochondrial Mn-dependent superoxide dismutases *SOD1* and *SOD2* (13, 14) as well as an increase in levels of the corresponding proteins (13). Ectopic H₂O₂

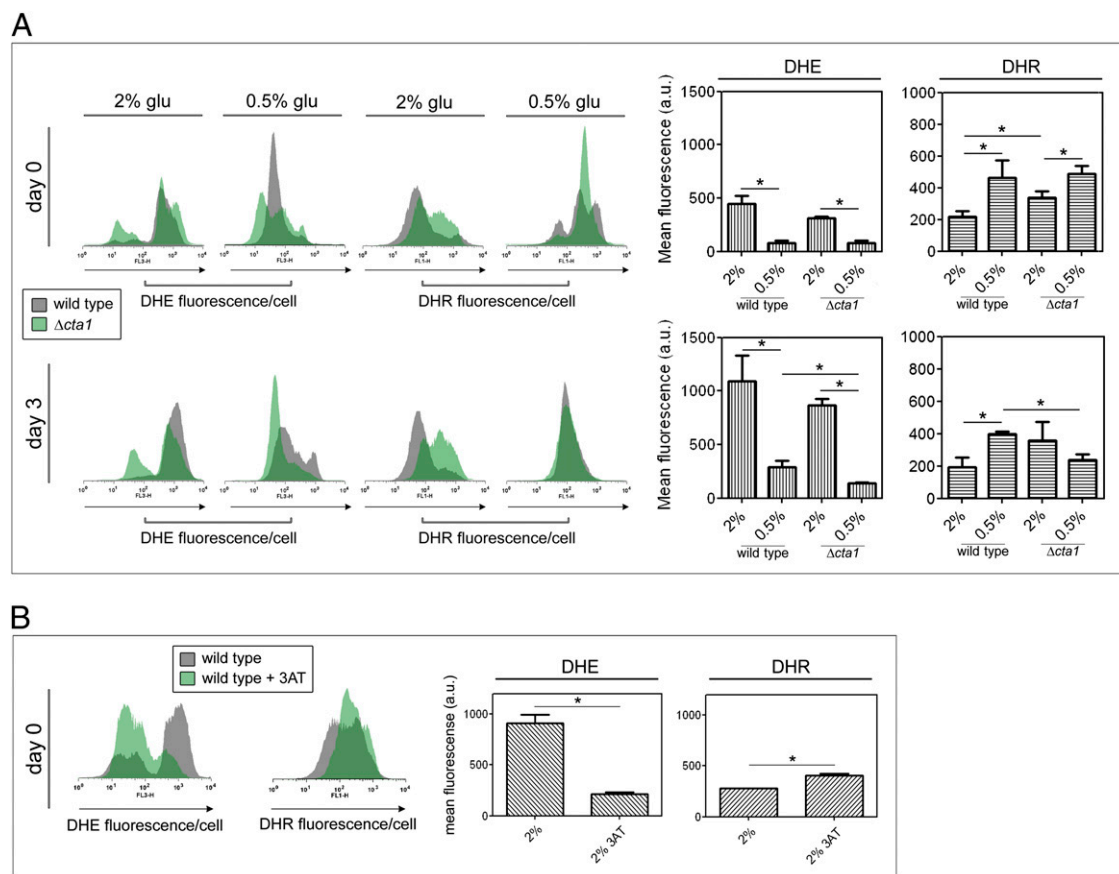


Fig. 2. The longevity-promoting effects of high intracellular H_2O_2 levels induced by CR or inactivation of catalases are accompanied by a reduction in the chronological age-dependent accumulation of superoxide anions. (A) FACS measurements of superoxide anions using the probe dihydroethidium (DHE) in parallel with measurements of H_2O_2 using dihydrorhodamine 123 (DHR) in wild-type (gray histograms) and $\Delta cta1$ (green histograms) cells at day 0 and day 3 of stationary phase. Bar graphs indicate mean \pm SD fluorescence/cell (arbitrary units) measured in 25,000 cells/sample in three independent experiments. (B) FACS measurements of superoxide anions (DHE) and H_2O_2 (DHR) in wild-type cells in the absence (gray histograms) or presence (green histograms) of 10 mM 3-amino-1,2,4-triazole (3AT) at day 0 of stationary phase. Bar graphs indicate mean \pm SD fluorescence/cell (arbitrary units) as described above. Statistical significance ($*P < 0.05$) was determined by Student's *t*-test.

also induces transcription of the superoxide dismutase *sodA* in *Escherichia coli* (15) and the transcription and activity of MnSOD, but not Cu/Zn-SOD in rat cells (16). To determine whether induction of Sod1p and/or Sod2p activity underlies the reduction in O_2^- levels that accompanies increases in intracellular H_2O_2 , we measured the activities of Sod1p and Sod2p under CR and non-CR conditions in wild-type and *CTA1* mutant cells. CR or *CTA1* deletion resulted in minor increases in Sod1p activity at day 0 that were not detected at day 3 of stationary phase (Fig. 3A and B). However, at day 6 both CR and deletion of *CTA1* increased the activity of Sod1p (Fig. 3A and B). CR or deletion of *CTA1* increased the activity of Sod2p at day 0 compared with wild-type cells in non-CR conditions (Fig. 3A and B). Larger increases in Sod2p activity were induced by CR conditions in wild-type cells at day 3 and day 6 (Fig. 3A and B). Deletion of *CTA1* also induced large increases in Sod2p activity at day 3 and day 6 under non-CR conditions and under CR conditions at day 3 (Fig. 3A and B). Similar observations were made in $\Delta ctt1$ cells at these same time points (Fig. S7). Exposure of wild-type cells to 1 mM H_2O_2 also resulted in an increase in Sod2p but not Sod1p activity (Fig. 3C). These findings indicate that intracellular H_2O_2 induced by CR or by inactivation of catalase activity or ectopic exposure to H_2O_2 induces superoxide dismutase activity in budding yeast, especially Sod2p activity. The more robust induction of Sod2p activity in these experiments is consistent with earlier reports that ectopic

exposure to H_2O_2 induces higher levels of Sod2p compared with Sod1p (13, 16, 17).

Effects of Increased H_2O_2 Induced by Caloric Restriction or by Inactivation of *CTA1* on Oxidative Damage to Macromolecules. According to the free radical theory, oxidative damage to macromolecules is a primary factor in aging. Our observations show that under different experimental conditions, H_2O_2 levels are increased, whereas O_2^- levels are decreased in the same cells (Fig. 2). To determine the overall impact these divergent changes in different types of ROS have on oxidative damage, we examined levels of protein carbonylation, which is a form of oxidative damage, under different experimental conditions. Protein carbonylation was increased in $\Delta cta1$ cells compared with wild-type cells at day 0 of stationary phase (Fig. 4A). A similar increase in protein carbonylation was observed in stationary phase $\Delta ctt1$ cells at this time point (Fig. S8). CR conditions also increased protein carbonylation in wild-type cells at day 0. Nevertheless, CR conditions decreased protein carbonylation in wild-type and $\Delta cta1$ cells at day 6 (Fig. 4A).

We also measured changes in cellular autofluorescence, which is produced by global oxidative damage to proteins and lipids (18, 19). Similar to the increased protein carbonylation detected in $\Delta cta1$ compared with wild-type cells, autofluorescence of $\Delta cta1$ cells was increased at day 0 and day 3 (Fig. 4). Comparable results were obtained in $\Delta ctt1$ cells (Fig. S8). In contrast, CR conditions

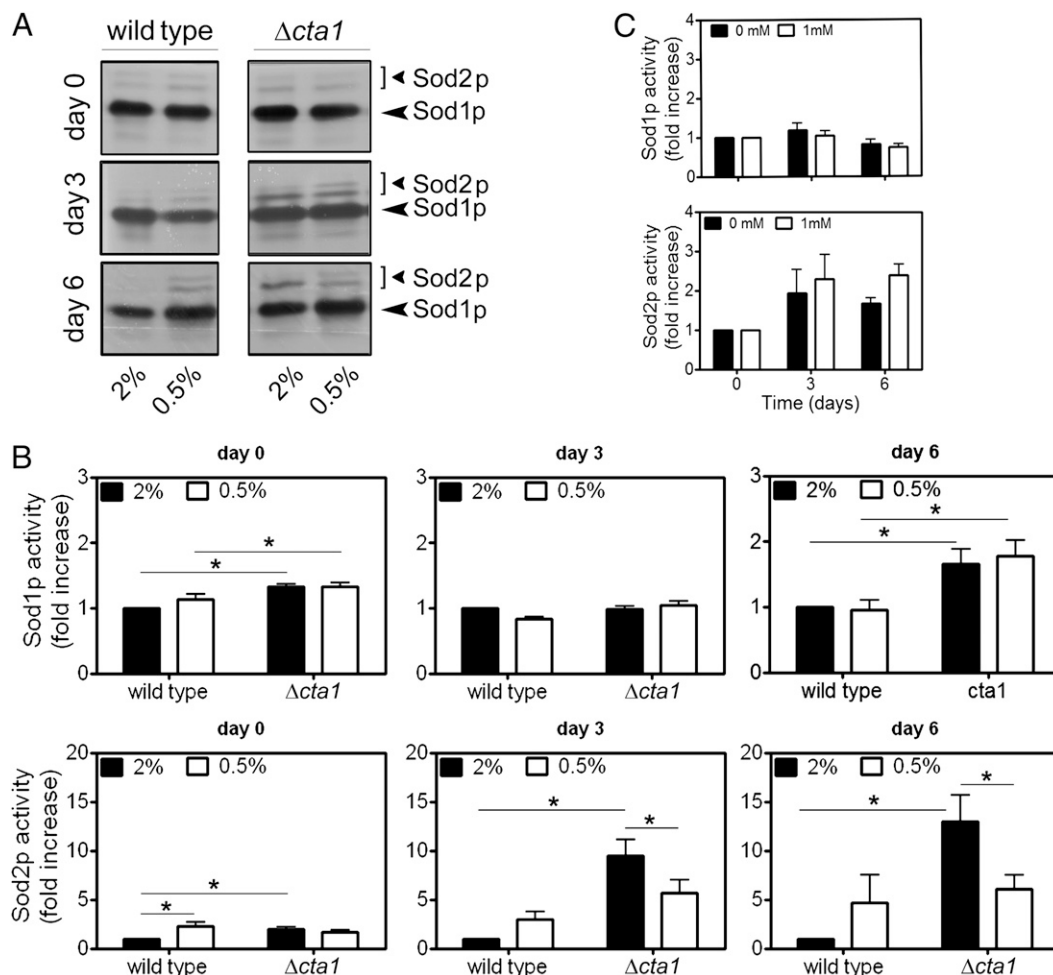


Fig. 3. Induction of superoxide dismutase activity by intracellular H_2O_2 . (A) In situ determination of superoxide dismutase activity in stationary phase wild-type and $\Delta cta1$ cells measured as previously described. MnSOD (Sod2p) activity was assessed in the presence of 2 mM potassium cyanide, which inhibits Sod1p, but not Sod2p activity (Fig. S7B). (B) Quantification of fold increases in Sod1p and Sod2p activity under indicated conditions in wild-type and $\Delta cta1$ cells. Sod1p and Sod2p activity at each time point was normalized to activity in wild-type cells under non-CR conditions (2% glucose). (C) Quantification of fold increases in Sod1p and Sod2p activity in wild-type cells after ectopic exposure to 1 mM H_2O_2 . Sod1p and Sod2p activity at each time point was normalized to activity in wild-type cells under non-CR conditions (2% glucose). Values indicate mean \pm SEM from three independent experiments. Statistical significance (* P < 0.05) was determined by Student's t -test.

led to a reduction in autofluorescence of wild-type cells at these time points, similar to the effects of CR on protein carbonylation in wild-type cells at day 6 (Fig. 4). CR conditions also reduced the autofluorescence of $\Delta cta1$ (Fig. 4B) and $\Delta ctt1$ cells (Fig. S8). Together, these findings establish that CR extends CLS in parallel with a reduction in oxidative damage to macromolecules in stationary phase budding yeast cells despite the induction of higher levels of H_2O_2 . In contrast, the CLS-extending effects of inactivating catalases are accompanied by parallel increases in levels of both H_2O_2 and oxidative damage, especially under non-CR conditions.

Discussion

Our study leads to the surprising conclusions that despite the induction of catalase activity by CR conditions that promote longevity, catalases are proaging factors in the budding yeast chronological aging model and both CR and catalase inactivation promote longevity in this model by inducing oxidative stress in the form of H_2O_2 . Although previous studies showed that glucose restriction increases ROS that promote longevity both in budding yeast (20) and in *C. elegans* (4), specific types of ROS were not identified in these studies. In our study, the effects of genetic or

pharmacological inactivation of catalases, catalase overexpression, and ectopic exposure to H_2O_2 combined with the use of probes that can distinguish between H_2O_2 and O_2^- definitively establish H_2O_2 as a longevity-promoting form of ROS induced by CR. The absence of toxic effects of H_2O_2 in these experiments reflects the well-documented dual roles of ROS in both survival and cell death pathways reported in other systems. Although H_2O_2 can induce programmed cell death (21), the intracellular concentrations that induce the beneficial hormesis-like effects in our study are likely to be far less than the concentrations that induce toxic effects. Furthermore, stationary phase cells are more resistant to H_2O_2 and other stresses compared with cells in exponential phase growth.

Our study also shows that the mechanism by which H_2O_2 exerts its prolongevity effects in budding yeast involves H_2O_2 induction of SOD activity. Hormesis theories posit that mild oxidative and other stresses induce adaptive responses that protect against further increases in stress (22). Our data identify SODs as targets of a H_2O_2 -induced stress response triggered by CR that confers hormesis-like protective effects against aging by reducing levels of O_2^- . This is consistent with previous reports that overexpression of SODs extends budding yeast CLS (23, 24). Although

promotes longevity despite increased oxidative damage to macromolecules by inducing superoxide dismutases that reduce levels of O_2^- . This mechanism likely underlies recent findings that challenge the validity of the free radical theory and provides a different paradigm for understanding how oxidative stress impacts aging and health.

Materials and Methods

Strains, Media, Chronological Lifespan, and Treatments. *S. cerevisiae* strain BY4742 cells (MAT α his3 Δ 1 leu2 Δ 0 lys2 Δ 0 ura3 Δ 0) and the respective knockouts in the *CTT1* and *CTA1* genes (European *Saccharomyces cerevisiae* Archive for Functional Analysis, EUROSCARF) were used for all experiments. Cell stocks were maintained in YEPD agar medium containing 0.5% yeast extract, 1% peptone, 2% agar, and 2% glucose. All experiments were performed in synthetic complete (SC) medium containing glucose as a carbon source and 0.67% yeast nitrogen base without amino acids (Difco Laboratories) supplemented with excess amino acids and bases for which the strains

were auxotrophic (50 μ g/mL histidine, 50 μ g/mL lysine, 300 μ g/mL leucine, and 100 μ g/mL uracil) with the exception that uracil was absent from medium in experiments that used the *CTA1* overexpressing strain, the control strain transformed with an empty vector or the double mutant *CTA1CTT1*. CLS was measured in different conditions as described in Supplementary Information. Construction of the *CTA1* overexpressing strain or the double mutant *CTA1CTT1* strain, treatment with pharmacological inhibitors of catalases or glutathione synthesis, measurements of intracellular reactive oxygen species, oxidative damage and SOD activity were performed using standard techniques: details are provided in Supplementary Information. Statistical analysis was performed according to the description in *SI Materials and Methods*.

ACKNOWLEDGMENTS. This work was supported by Fundação para a Ciência e Tecnologia, Portugal Grant PTDC/AGR-ALI/71460/2006, and National Cancer Institute Cancer Center Support Grant (P30 CA016056) to Roswell Park Cancer Institute. A.M., A.S., and B.S.-M. have fellowships from Fundação para a Ciência e Tecnologia (SFRH/BD/32464/2006, SFRH/BD/33125/2007, and SFRH/BD/41674/2007, respectively).

- Harman D (1956) Aging: A theory based on free radical and radiation chemistry. *J Gerontol* 11:298–300.
- Blagosklonny MV (2008) Aging: ROS or TOR. *Cell Cycle* 7:3344–3354.
- Andziak B, et al. (2006) High oxidative damage levels in the longest-living rodent, the naked mole-rat. *Aging Cell* 5:463–471.
- Schulz TJ, et al. (2007) Glucose restriction extends *Caenorhabditis elegans* life span by inducing mitochondrial respiration and increasing oxidative stress. *Cell Metab* 6:280–293.
- Linnane AW, Kios M, Vitetta L (2007) Healthy aging: Regulation of the metabolome by cellular redox modulation and prooxidant signaling systems: the essential roles of superoxide anion and hydrogen peroxide. *Biogerontology* 8:445–467.
- Groeger G, Quiney C, Cotter TG (2009) Hydrogen peroxide as a cell-survival signaling molecule. *Antioxid Redox Signal* 11:2655–2671.
- Dasgupta J, et al. (2006) Manganese superoxide dismutase protects from TNF- α -induced apoptosis by increasing the steady-state production of H₂O₂. *Antioxid Redox Signal* 8:1295–1305.
- Madia F, Gattazzo C, Fabrizio P, Longo VD (2007) A simple model system for age-dependent DNA damage and cancer. *Mech Ageing Dev* 128:45–49.
- Weinberger M, et al. (2007) DNA replication stress is a determinant of chronological lifespan in budding yeast. *PLoS ONE* 2:e748.
- Burtner CR, Murakami CJ, Kennedy BK, Kaeberlein M (2009) A molecular mechanism of chronological aging in yeast. *Cell Cycle* 8:1256–1270.
- Burhans WC, Weinberger M (2009) Acetic acid effects on aging in budding yeast: Are they relevant to aging in higher eukaryotes? *Cell Cycle* 8:2300–2302.
- Benov L, Szejnberg L, Fridovich I (1998) Critical evaluation of the use of hydroethidine as a measure of superoxide anion radical. *Free Radic Biol Med* 25:826–831.
- Godon C, et al. (1998) The H₂O₂ stimulon in *Saccharomyces cerevisiae*. *J Biol Chem* 273:22480–22489.
- Gasch AP, et al. (2000) Genomic expression programs in the response of yeast cells to environmental changes. *Mol Biol Cell* 11:4241–4257.
- Semchyshyn H (2009) Hydrogen peroxide-induced response in *E. coli* and *S. cerevisiae*: Different stages of the flow of the genetic information. *Cent Eur J Biol* 4:142–153.
- Yoshioka T, et al. (1994) Oxidants induce transcriptional activation of manganese superoxide dismutase in glomerular cells. *Kidney Int* 46:405–413.
- Röhrdanz E, Schmuck G, Ohler S, Kahl R (2001) The influence of oxidative stress on catalase and MnSOD gene transcription in astrocytes. *Brain Res* 900:128–136.
- Delori FC, Dorey CK (1998) In vivo technique for autofluorescent lipopigments. *Methods Mol Biol* 108:229–243.
- Terman A, Dalen H, Eaton JW, Neuzil J, Brunk UT (2004) Aging of cardiac myocytes in culture: Oxidative stress, lipofuscin accumulation, and mitochondrial turnover. *Ann N Y Acad Sci* 1019:70–77.
- Agarwal S, Sharma S, Agrawal V, Roy N (2005) Caloric restriction augments ROS defense in *S. cerevisiae*, by a Sir2p independent mechanism. *Free Radic Res* 39:55–62.
- Madeo F, et al. (1999) Oxygen stress: A regulator of apoptosis in yeast. *J Cell Biol* 145:757–767.
- Gems D, Partridge L (2008) Stress-response hormesis and aging: "That which does not kill us makes us stronger." *Cell Metab* 7:200–203.
- Harris N, et al. (2005) Overexpressed Sod1p acts either to reduce or to increase the lifespans and stress resistance of yeast, depending on whether it is Cu(2+)-deficient or an active Cu,Zn-superoxide dismutase. *Aging Cell* 4:41–52.
- Fabrizio P, Pletcher SD, Minois N, Vaupel JW, Longo VD (2004) Chronological aging-independent replicative life span regulation by Msn2/Msn4 and Sod2 in *Saccharomyces cerevisiae*. *FEBS Lett* 557:136–142.
- Yokoo S, Furumoto K, Hiyama E, Miwa N (2004) Slow-down of age-dependent telomere shortening is executed in human skin keratinocytes by hormesis-like-effects of trace hydrogen peroxide or by anti-oxidative effects of pro-vitamin C in common concurrently with reduction of intracellular oxidative stress. *J Cell Biochem* 93:588–597.
- Passos JF, et al. (2007) Mitochondrial dysfunction accounts for the stochastic heterogeneity in telomere-dependent senescence. *PLoS Biol* 5:e110.
- Serra V, von Zglinicki T, Lorenz M, Saretzki G (2003) Extracellular superoxide dismutase is a major antioxidant in human fibroblasts and slows telomere shortening. *J Biol Chem* 278:6824–6830.
- Sarsour EH, Venkataraman S, Kalen AL, Oberley LW, Goswami PC (2008) Manganese superoxide dismutase activity regulates transitions between quiescent and proliferative growth. *Aging Cell* 7:405–417.
- Zmijewski JW, et al. (2009) Antiinflammatory effects of hydrogen peroxide in neutrophil activation and acute lung injury. *Am J Respir Crit Care Med* 179:694–704.
- Mitra S, Abraham E (2006) Participation of superoxide in neutrophil activation and cytokine production. *Biochim Biophys Acta* 1762:732–741.