

Review Article

Skin wound healing modulation by macrophages

Mathieu P. Rodero, Kiarash Khosrotehrani

University of Queensland Centre for Clinical Research, Experimental Dermatology Group, Brisbane, Australia

Received July 5, 2010; accepted July 23, 2010; available online July 25, 2010

Abstract: Skin wound healing is a multi stage phenomenon that requires the activation, recruitment or activity of numerous cell types as keratinocytes, endothelial cells, fibroblast and inflammatory cells. Among the latter, macrophages appear to be central to this process. They colonize the wound at its very early stage and in addition to their protective immune role seem to organize the activity of other cell types at the following stages of the healing. Their benefit to this process is however controversial, as macrophages are described to promote the speed of healing but may also favour the fibrosis resulting from it in scars. Moreover wound healing defects are associated with abnormalities in the inflammatory phase. In this review, we summarise our knowledge on what are the Wound Associated Macrophages, and how they interact with the other cell types to control the reepithelialisation, angiogenesis and the extracellular matrix remodelling. We believe this knowledge may open new avenues for therapeutic intervention on skin wounds.

Keywords: Macrophage, skin wound, healing, inflammation, cytokines, growth factors, M1, alternatively activated

Introduction

Macrophages are mature myeloid cells, mostly derived from the differentiation of circulating monocytes after tissue infiltration. Far to be a homogeneous population, macrophages display a wide range of phenotypes and physiological properties depending on the cytokines inducing their maturation [1]. Moreover, macrophages are very plastic cells, able to switch from one functional sub-population to another depending on the stimulus received [2]. Another aspect of macrophage heterogeneity is the tissue specialisation of resident macrophages, as microglial cells in the brain, Kupffer cells in the liver or alveolar macrophages in the lung. However, it is still not clear if these resident macrophages are derived in steady state condition from circulating monocytes and if they are terminally differentiated cells [1]. After birth, macrophages are known to affect different stages of skin wound healing, modulating the function of the different cell types involved in this process. The benefit of inflammation and inflammatory cells in the wound healing is a matter of debate as in normal conditions it promotes wound closure but

also the fibrosis associated with scar formation. Wound Associated Macrophages (WAM) have a central role in the control of wound inflammation. Here, we will present an overview of the main functions of WAM and their influence on the other major cell types present in the wound during the healing process and how modulating these processes might result in better wound healing.

Wound healing

In humans, and more widely in all mammalian species, the wound healing process can be subdivided in three consecutive and overlapping stages: inflammation, new tissue formation and remodelling [3]. The transition from one stage to another depends on the maturation and differentiation of the main cell populations involved, among which the keratinocytes, the fibroblasts and the macrophages.

Inflammation

The first event occurring after injury is the formation of a blood clot by activated platelets.

The blood plug will be composed of various cell types including platelet, red and white blood cells. The initial plug is stabilised by fibrin fibres and will be a scaffold for the various infiltrating cells. The first inflammatory cells recruited are the neutrophils [4]. They infiltrate massively the wound during the first 24h post injury [5] attracted by the numerous inflammatory cytokines produced by the activated platelets, endothelial cells, as well as by the degradation products from pathogens. Neutrophils enter apoptosis soon after infiltrating the wound and the release of cytokines during this apoptotic process is an important component in macrophage recruitment. Macrophages infiltrate the wound massively 2 days post injury and exacerbate at this stage an intense phagocytic activity [6].

New tissue formation

The reepithelialisation process begins few hours after the wound formation. Keratinocytes from the wound edges migrate over the wound bed at the interface between the wound dermis and the fibrin clot. This migration is facilitated by the production of specific proteases such as the collagenase by the epidermal cells to degrade the extracellular matrix [7]. Activated fibroblasts also migrate to the wound bed and form, with the macrophages, the granulation tissue. A massive angiogenesis allowing the supply of oxygen and nutrients necessary for the healing process also occurs within this tissue [8]. Later, some of the fibroblasts differentiate into myofibroblasts. These contractile cells will help bridge the gap between the wound edges [9]. During the same time, growth factors produced by the granulation tissue will favour proliferation and differentiation of epithelial cells restoring the epithelial barrier integrity.

Remodelling

The last stage of the wound healing process consists in a gradual involution of the granulation tissue and dermal regeneration. This step is associated with the apoptosis of myofibroblasts, endothelial cells and macrophages. The remaining tissue is therefore composed mostly of extracellular matrix proteins, essentially collagen type III that will be remodelled by the metalloproteinase produced by the epidermal cells, endothelial cells, fibroblasts and the macrophages remaining in the scar and be replaced by collagen type I [8].

Macrophages and wound healing

Wound macrophage phenotypes

Macrophages used to be divided in several sub-populations depending of the way they had been activated, their cell surface markers or their functionality. [1, 10,11]. Schematically, macrophages activated by microbial agents and cytokines like Interferon gamma (IFN γ) are classified as M1 macrophages. These macrophages produce an important level of Nitric Oxide (NO) and pro inflammatory cytokines such as Tumor Necrosis Factor alfa (TNF α), IL-1 β , IL-6, or IL-12, and overexpress MHC class II molecules. They bear microbicidal and antitumoral properties. M2 macrophages are a much more heterogeneous population composed of all macrophages that do not correspond to M1 characteristics [12]. M2 macrophages could be divided in 3 sub-populations. The alternatively activated macrophages or M2a, that promote a Th2 type of inflammation resulting in increased IgE as observed in allergy and parasite immunity, the M2b macrophages, that promote Th2 inflammation and bear some immunoregulation properties, and the deactivated macrophages or M2c, able to control the inflammation and implicated in tissue remodelling [12]. In another classification, M1 macrophages are considered as the classically activated macrophages as compared to the alternatively activated macrophages. The best described alternative activation consists in the stimulation of the IL-4R by IL-4 and IL-13 that induces pro Th2 macrophages [13]. Of course, all these sub populations should not be considered as distinct populations *in vivo* but more as different stages of a continuum of activation and differentiation of macrophage populations [14]. The phenotype of skin wound infiltrating macrophages is not yet fully characterised, but it already appears that it changes during the healing process suggesting that macrophages have different roles in the diverse phases of skin repair [15, 16].

Several reviews refer to wound macrophages as the IL4 alternatively activated macrophages because of numerous markers of tissues remodelling, such as YM1, Resistin Like Molecule Alfa (RELM α), Insulin Growth Factor 1 (IGF1), factor XIII-A or arginase that are expressed after *in vitro* activation of macrophages with IL4. [13, 17-20]. Recent work on aseptic wounds using an implantable sponge model described wound

Table 1. Impact of macrophages depletion strategy on wound healing

Depletion stage	Depletion process	Wound closure	Gr T formation / Angiogenesis	Collagen / fibrosis/αSMA	Ref
Early	Macrophage anti-serum	↓	ND	↓	[6]
Early	CD11b DTR	↓	↓	↓	[22]
Early	lysM-Cre DTR	↓	↓	↓	[23]
Early	lysM-Cre DTR	↓	↓	↓	[16]
Middle	lysM-Cre DTR	↓	↓	ND	[16]
Late	lysM-Cre DTR	=	=	=	[16]

Gr T: Granulation tissue

macrophages as cells bearing both classical activation markers, like TNF α expression, and alternatively activated markers like the mannose receptor [15]. The authors also described the evolution of the macrophage phenotype. Day 1 macrophages produced more TNF α and IL6 and less Tumor Growth Factor beta (TGF β) compared to day 7 macrophages suggesting a transition from an inflammatory to an immuno-regulatory or tissue remodelling state. However, it is important to consider that neither IL4 nor IL13 were detectable in this model, perhaps because of the aseptic experimental condition. However, it is not clear to what extent the bacterial colonization of the wound would modify Wound Associated Macrophage phenotypes and healing functions, as it is clear that parasitic and bacterial component are major macrophage activators [13].

Macrophage importance in wound healing

In order to show the implication of macrophages in the control of wound healing, several studies have analysed skin wound healing upon macrophage depletion (**Table 1**). The first attempt consisted in corticosteroids associated with anti-macrophage sera to deplete macrophages in wounded guinea pigs. This treatment resulted in a delayed infiltration of the wound by fibroblasts and decreased fibrosis [6]. These results are in accordance with observations that fetal wounds heal without fibrosis and are not infiltrated by macrophages. In a model of PU.1 null mice where animals lacked macrophages, mast cells and functional neutrophils due to defective myelopoiesis, wounds performed on newborns additionally treated with antibiotic healed at the same speed as wild types, but without scar formation, suggesting that inflammatory cells are

not needed for wound closure [21]. However, several recent models of specific inducible macrophage depletion, based on genetically modified mice resulted in detrimental effect of pre-injury depletion of macrophages [16, 22, 23]. Mice depleted before injury typically show a defect in re-epithelialisation, granulation tissue formation, angiogenesis, wound cytokine production and myofibroblast associated wound contraction. Recently, the effect of a post-injury depletion of the WAM has also been investigated. Macrophage depletion during the granulation tissue formation, about 3 days after injury, is associated with vascularisation defect, delay in wound closure as well as in granulation tissue maturation [16]. Finally, in their study, Lucas et al did not find any morphological or biological differences between mice that were or were not depleted 9 days after injury suggesting no further implication of macrophages at later stages.

During the healing process numerous cytokines and growth factors are produced by the various cell types present in the wound or at the wound edge [24]. The level of production of the different cytokines depends on the regulation of the cross talk between the major cell populations in the wound: epithelial cells, endothelial cells, fibroblasts and inflammatory cells. However, the literature clearly lacks *in vivo* characterisation of the cytokine production kinetics by the various cell types during skin wound healing. Most of our current knowledge concerning the ability of a cell population to produce or to respond to a specific cytokine is based on *in vitro* studies.

Cross talk with keratinocyte

Injured and inflamed keratinocytes produce

several cytokines allowing the recruitment and activation of the WAM, including chemokines, interleukins and growth factors [25, 26]. On the other hand, the ability of immune cells to produce factors able to regulate keratinocyte growth in vitro has been described since 1988 [27]. Interestingly, conditioned medium from co-culture of macrophages and allogenic T cells were much more efficient in inducing keratinocyte proliferation compared to conditioned medium from macrophages alone. However, the direct implication of WAM in producing cytokines or growth factors to influence directly or indirectly keratinocyte migration and proliferation is not clear *in vivo*. Several cytokines and growth factors, that can be produced by macrophages, have been associated with reepithelialisation, mostly *in vitro*. Among the Epithelial growth Factor family (EGF), the main members involved in wound healing are EGF, TGF α and Heparin Bound EGF (EGF-HB) [28-32]. The activation of the EGF Receptor (EGFR) on keratinocytes promotes cell migration and proliferation. Although, only TGF α and EGF-HB have been described to be produced by macrophages *in vitro*, their production by WAM *in vivo* is more enigmatic [33, 34]. TGF α has been detected in macrophages collected from sub epidermal wound cylinders 6 days after implantation in mice [35]. However, no significant wound healing abnormalities were observed in TGF α -/-mice [36]. In addition, all EGFR ligands are synthesised as membrane-anchored forms, which can be released as a soluble form after proteolysis by MMP [37]. Therefore, another possible role for WAM might be the regulation of EGF family release by the production of MMPs. Other cytokines as IL-6, IL1 and TNF- α produced by macrophages are also associated with reepithelialisation. Even if the secretion of these cytokines by WAM has not been clearly demonstrated yet, the ability of macrophages activated by the inflammatory cytokine IFN γ and/or bacterial products via Toll Like Receptor 4 (TLR4) to produce IL-6, IL1 β and TNF α makes this hypothesis probable [13]. The implication of IL-1 and IL-6 in the reepithelialisation is indirect and is mediated by other cell types present in the wound, at least the fibroblast [38, 39]. On the contrary, TNF α directly stimulates transcription of genes associated with numerous cell functions including inflammation, mobility, cell division and survival in keratinocytes [25]. Of note, in a model of sponge implanted under aseptic conditions in the dermis, macrophages ex-

tracted on day 1 produced more TNF α compared to day 7 [15]. Finally, the other cytokine known to be massively produced by macrophages, and highly implicated in reepithelialisation, is TGF β , although its effect on keratinocyte proliferation and migration remains controversial [36, 40-43]. Cultured WAM extracted from implanted sponges start producing TGF β from the very first day of the wound [15].

Influence on fibroblasts and myofibroblast

Fibroblasts begin to infiltrate the wound during the first steps of granulation tissue formation. Fibroblasts are involved in several processes of the wound healing. They contribute to the granulation tissue formation, produce the cytokines that favour keratinocyte proliferation and migration [44], and finally differentiate in myofibroblasts to promote wound closure. Several molecules, such as PDGF-bb, TNF α , IL-1 or IL-6, produced by activated macrophages are able to induce *in vitro* the production of pro-reepithelialisation molecules by fibroblasts such as KGF [45-48]. Interestingly, the effect of most of these molecules is dependent on the presence of serum in the fibroblast culture media, indicating that the stimulation resulting in their production is a cooperative process [49]. Mori et al demonstrated nicely that macrophage secretion of PDGF-bb isoform induced the production of osteopontin by fibroblasts [50]. The inhibition of osteopontin was associated with lower scar fibrosis [50]. Finally, the molecule produced by macrophages that has been the most extensively studied for its actions on wound fibroblasts is TGF β . It is a family of three cytokines, TGF β 1, 2 and 3, produced by several cell types during the healing process, including platelets and epidermal cells, but the main and sustained source of production are the WAM [16, 51]. All TGF β family members are secreted coupled to a "latent complex" which has the ability to bind the Extra Cellular Matrix (ECM). The latent forms have to be activated via various mechanisms to release the mature TGF β , including proteolysis by MMPs known to be extensively secreted by macrophages as reviewed by Annes et al [52]. While TGF β 1 and 2 promote the inflammation within the wound and are associated with scar formation, TGF β 3 is associated with scar free wound healing [53]. However, the expression of TGF β 3 in adult wound is rare while the two other forms are highly expressed [53]. TGF β 1 is chemotactic for fibro-

Table 2. Targeting of macrophages to control healing of pathological wound

Molecules / cells	Delivery	Stage	Pathological model	Wound model	Effect on MΦ	Effect on healing	Ref
MALP-2	local	Early to middle	Diabetic C57Bl6 db/db	Acute	↑ recruitment	↑	[89]
GM-CSF	local	NA	Human CVLU	Chronic	↑ VEGF production	↑	[88]
Anti TNFα	Systemic	Early to late	Diabetic ob/ob	Acute	↓ recruitment	↑	[90]
IL1β activated db/db MΦ	Local	Early	Diabetic C57Bl6 db/db	Acute	↑ VEGFC/VEGFR3 transcription	↑	[72]
GM-CSF	local	Early	Strpz induce diabetic mice	Acute	↑ Recruitment	↑	[86]

CVLU: Chronic Veinous Leg Ulcer. Strpz : streptozotocin

blasts [54, 55]. It induces the production of several growth factors by fibroblasts, including Connective Tissue Growth Factor (CTGF) [56], that results in their proliferation by an autocrine loop [57]. But the main action of TGF β on fibroblast is to promote their differentiation into myofibroblast and to favour collagen production [58]. The secretion of TGF β and MMP by the macrophages is critical in the control of the ECM composition. Accordingly, macrophage depletion is associated with a defect of Alpha Smooth Muscle Actin (α SMA) positive cell in the granulation tissue [23]. However, it is estimated that 30 to 50% of the granulation tissue α SMA+ cells do not derive from fibroblasts, but more likely from bone marrow derived fibrocytes or mesenchymal stem cells [59-61]. The implication of macrophage produced cytokines in the recruitment, activation and differentiation of these marrow derived fibrocytes in the wound bed is unknown and has to be further investigated.

Cross talk with endothelial cell

Neo-angiogenesis in the granulation tissue is an important process allowing the supply of nutrients necessary for the healing process. The main angiogenic factor in the wound is the Vascular Endothelial Growth Factor (VEGF) and its reduced expression results in a wound healing defect [62]. VEGF promotes wound vascularisation by multiple mechanisms targeting directly or indirectly endothelial cells as reviewed by

Eming et al [63]. VEGF promotes endothelial cell and precursor recruitment [64] but it also acts as a mitogen and survival factor on the endothelial cells [65-67]. The increase of VEGF production in the wound is described in migratory keratinocytes and in macrophages infiltrating the granulation tissue [62]. The keratinocyte production of VEGF is indirectly promoted by macrophages as well through the secretion of cytokines as TNF α or TGF β [68]. CTGF produced by fibroblasts [56], also favours endothelial cell proliferation and new vessel formation [69, 70]. VEGF family members, mostly VEGF-C and VEGF-D, have also been recognized for their ability to modulate lymphangiogenesis [71]. Decreased macrophage numbers and activation has been associated with reduced lymphatic vessel formation in diabetic mouse wounds [72]. It is important to note that double positive F4/80/Lyve-1 cells have been described to be integrated in lymphatic vessels in several models of neolymphangiogenesis suggesting that macrophages could be directly implicated in their formation [72, 73].

The other main pro-angiogenic molecule during the wound healing is the Placental Growth Factor (PIGF), a member of the VEGF family [67] [74]. PIGF expression is induced in vitro by TGF α and TGF β , and is produced during the angiogenic stage of the healing process [75]. The effects of PIGF are similar to those of VEGF, promoting monocytes and endothelial precursor cell migration [76] and favouring endothelial

Skin wound healing modulation by macrophages

cell survival [77]. PIGF stimulates also the secretion of VEGF by the monocyte/macrophages [78]. Interestingly, even if PIGF is described to have chemotactic properties by his own on VEGFR1+ expressing cells, the synergy between PIGF and VEGF is important in angiogenesis [79]: i) VEGFR1 homodimers may be a decoy receptor. Binding of PIGF homodimer to VEGFR1 homodimer make more VEGF available to bind to activated VEGFR2. ii) VEGF/PIGF heterodimers bind to VEGFR1/VEGFR2 heterodimer and induce more potent angiogenic signals iii) binding of PIGF homodimers to VEGFR1 homodimers favours VEGFR2 homodimer phosphorylation [80]. Overall, PIGF seems to potentiate the effect of VEGF in wound angiogenesis. These results altogether point to the important role of macrophages in coordinating the angiogenic signal during wound healing.

Macrophages and defective wound healing

Despite their heterogeneous aetiology, most chronic wounds have in common a defect in the progression from the inflammatory to the tissue formation stage. Loots et al reported increased infiltration of chronic and diabetic wounds by WAM compared to control acute wounds. In addition they observed higher amounts of ECM in the wound edge [81]. Similarly, in diabetic db/db mice, wounds are characterising by a prolonged expression of inflammatory cytokines and larger infiltration and persistence of the WAM [82]. However, these cells seem to have altered sensitivity towards exogenous signals such as VEGF or IGF1 [83, 84] as well as altered ability to release cytokine [85]. Several studies have targeted macrophages, by different strategies in order to improve defective wound healing (**Table 2**). In the obese diabetic

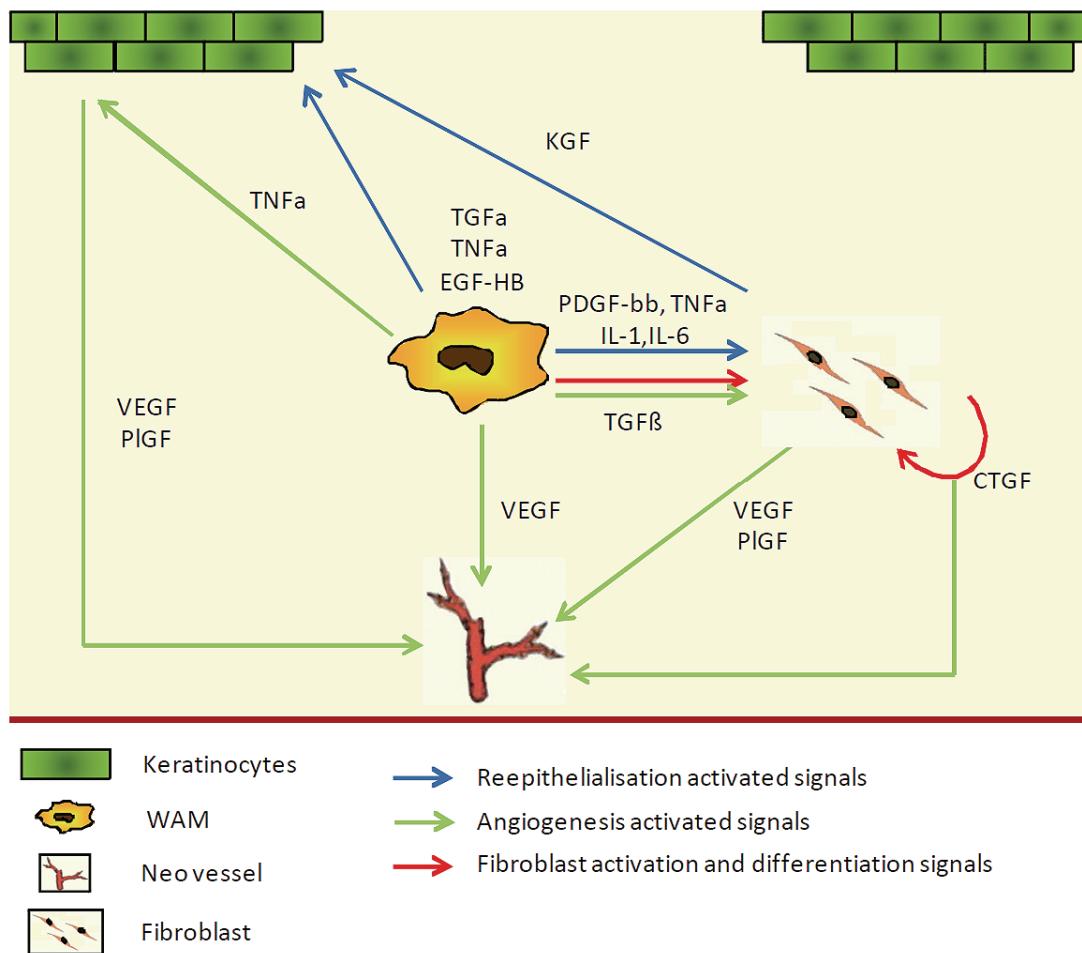


Figure 1. Direct and indirect control of reepithelialisation, angiogenesis and fibroblast activation by macrophages.

db/db mouse model, the injection of peritoneal macrophages activated by IL-1 β at the wound site was associated with an increased production of prolymphangiogenic molecules such as VEGF-C, resulting in improved lymphangiogenesis and wound healing [72]. Local application of GM-CSF is associated with better healing both in human and mouse pathological wounds in a clinical setting. In Streptozotocin induced diabetic mice, GM-CSF is associated with a stronger infiltration of the wound by macrophages, increased angiogenesis and a better healing, while no effect was observed in normal wound healing [86] [87]. Similarly, in a pilot study in humans, GM-CSF treatment improves chronic vascular ulcers, probably by promoting the secretion of VEGF by macrophages [88]. With a similar strategy, Deiters et al injected Macrophage-Activating Lipopeptide-2 (MALP-2) in wounds on obese diabetic db/db mice. MALP-2 induced the transcription of several genes associated with wound healing, including GM-CSF and IL-1 [89] and resulted in increased infiltration by WAM and more rapid wound closure. In opposition to these activating approaches, Goren et al used a depleting strategy based on the systemic injection of a neutralizing anti-TNF α in ob/ob mice [90]. The treatment resulted in a systemic and local depletion of macrophages that was associated with a faster healing. Taken together, these results sustain the hypothesis that in chronic or diabetic wounds different strategies that might affect macrophage phenotype might modify the healing.

Conclusions

Our current knowledge points to macrophages as key players in skin wound healing after birth. Healing is a complex and evolving process, and because of their plasticity, WAM progress with the wound, adapting their cytokine expression profile [15]. Because of this plasticity and of their central position, WAM seem to be the ideal target for clinical interventions in chronic wounds, to accelerate closure or to attenuate fibrosis (**Figure 1**). For this purpose, there is still a need to further characterise this population. It seems obvious that, similar to other inflammation models, WAM are essentially composed of infiltrating blood monocytes that change from an inflammatory phenotype to a repair one [15, 91, 92]. It is not clear if the multiple functions of WAM are exclusive to a unique macrophage

population or fulfilled by specialised sub populations at different time points? Should the future therapeutic strategies consist in the control of the WAM polarisation or in the specific recruitment or depletion of sub populations? These strategies will also have to define the appropriate timing for their effectiveness as WAM bear different functions during the healing time course [16].

Acknowledgement

This work was supported by the Ramaciotti foundation.

Please address correspondence to: Kiarash Khosrotehrani, MD, PhD, University of Queensland Centre for Clinical Research, Building 71/918, Royal Brisbane & Women's Hospital Campus, Herston, QLD, 4029, Tel: +6173346 6077, Fax : +61733465598, E-mail: k.khosrotehrani@uq.edu.au

References

- [1] Gordon S and Taylor PR. Monocyte and macrophage heterogeneity. *Nat Rev Immunol* 2005; 5: 953-964.
- [2] Stout RD and Suttles J. Functional plasticity of macrophages: reversible adaptation to changing microenvironments. *J Leukoc Biol* 2004; 76: 509-513.
- [3] Gurtner GC, Werner S, Barrandon Y and Longaker MT. Wound repair and regeneration. *Nature* 2008; 453: 314-321.
- [4] Ross R and Odland G. Human wound repair. II. Inflammatory cells, epithelial-mesenchymal interrelations, and fibrogenesis. *J Cell Biol* 1968; 39: 152-168.
- [5] Kim MH, Liu W, Borjesson DL, Curry FR, Miller LS, Cheung AL, Liu FT, Isseroff RR and Simon SI. Dynamics of neutrophil infiltration during cutaneous wound healing and infection using fluorescence imaging. *J Invest Dermatol* 2008; 128: 1812-1820.
- [6] Leibovich SJ and Ross R. The role of the macrophage in wound repair. A study with hydrocortisone and antimacrophage serum. *Am J Pathol* 1975; 78: 71-100.
- [7] Pilcher BK, Dumin JA, Sudbeck BD, Krane SM, Welgus HG and Parks WC. The activity of collagenase-1 is required for keratinocyte migration on a type I collagen matrix. *J Cell Biol* 1997; 137: 1445-1457.
- [8] Singer AJ and Clark RA. Cutaneous wound healing. *N Engl J Med* 1999; 341: 738-746.
- [9] Gabbiani G. The myofibroblast in wound healing and fibrocontractive diseases. *J Pathol* 2003; 200: 500-503.
- [10] Gordon S. Alternative activation of macrophages. *Nat Rev Immunol* 2003; 3: 23-35.

Skin wound healing modulation by macrophages

- [11] Mantovani A, Sozzani S, Locati M, Allavena P and Sica A. Macrophage polarization: tumor-associated macrophages as a paradigm for polarized M2 mononuclear phagocytes. *Trends Immunol* 2002; 23: 549-555.
- [12] Mantovani A, Sica A, Sozzani S, Allavena P, Vecchi A and Locati M. The chemokine system in diverse forms of macrophage activation and polarization. *Trends Immunol* 2004; 25: 677-686.
- [13] Martinez FO, Helming L and Gordon S. Alternative activation of macrophages: an immunologic functional perspective. *Annu Rev Immunol* 2009; 27: 451-483.
- [14] Stout RD, Jiang C, Matta B, Tietzel I, Watkins SK and Suttles J. Macrophages sequentially change their functional phenotype in response to changes in microenvironmental influences. *J Immunol* 2005; 175: 342-349.
- [15] Daley JM, Brancato SK, Thomay AA, Reichner JS and Albina JE. The phenotype of murine wound macrophages. *J Leukoc Biol* 2010; 87: 59-67.
- [16] Lucas T, Waisman A, Ranjan R, Roes J, Krieg T, Muller W, Roers A and Eming SA. Differential Roles of Macrophages in Diverse Phases of Skin Repair. *J Immunol* 2009;
- [17] Mosser DM and Edwards JP. Exploring the full spectrum of macrophage activation. *Nat Rev Immunol* 2008; 8: 958-969.
- [18] Raes G, Noel W, Beschin A, Brys L, de Baetselier P and Hassanzadeh GH. FIZZ1 and Ym as tools to discriminate between differentially activated macrophages. *Dev Immunol* 2002; 9: 151-159.
- [19] Wynes MW and Riches DW. Induction of macrophage insulin-like growth factor-I expression by the Th2 cytokines IL-4 and IL-13. *J Immunol* 2003; 171: 3550-3559.
- [20] Torocsik D, Bardos H, Nagy L and Adany R. Identification of factor XIIIa as a marker of alternative macrophage activation. *Cell Mol Life Sci* 2005; 62: 2132-2139.
- [21] Martin P, D'Souza D, Martin J, Grose R, Cooper L, Maki R and McKercher SR. Wound healing in the PU.1 null mouse-tissue repair is not dependent on inflammatory cells. *Curr Biol* 2003; 13: 1122-1128.
- [22] Mirza R, DiPietro LA and Koh TJ. Selective and specific macrophage ablation is detrimental to wound healing in mice. *Am J Pathol* 2009; 175: 2454-2462.
- [23] Goren I, Allmann N, Yoge N, Schurmann C, Linke A, Holdener M, Waisman A, Pfeilschifter J and Frank S. A transgenic mouse model of inducible macrophage depletion: effects of diphtheria toxin-driven lysozyme M-specific cell lineage ablation on wound inflammatory, angiogenic, and contractive processes. *Am J Pathol* 2009; 175: 132-147.
- [24] Chen L, Tredget EE, Wu PY and Wu Y. Paracrine factors of mesenchymal stem cells recruit macrophages and endothelial lineage cells and enhance wound healing. *PLoS One* 2008; 3: e1886.
- [25] Banno T, Gazel A and Blumenberg M. Effects of tumor necrosis factor-alpha (TNF alpha) in epidermal keratinocytes revealed using global transcriptional profiling. *J Biol Chem* 2004; 279: 32633-32642.
- [26] McKay IA and Leigh IM. Epidermal cytokines and their roles in cutaneous wound healing. *Br J Dermatol* 1991; 124: 513-518.
- [27] Hancock GE, Kaplan G and Cohn ZA. Keratinocyte growth regulation by the products of immune cells. *J Exp Med* 1988; 168: 1395-1402.
- [28] Rheinwald JG and Green H. Epidermal growth factor and the multiplication of cultured human epidermal keratinocytes. *Nature* 1977; 265: 421-424.
- [29] Li Y, Fan J, Chen M, Li W and Woodley DT. Transforming growth factor-alpha: a major human serum factor that promotes human keratinocyte migration. *J Invest Dermatol* 2006; 126: 2096-2105.
- [30] Schultz G, Rotatori DS and Clark W. EGF and TGF-alpha in wound healing and repair. *J Cell Biochem* 1991; 45: 346-352.
- [31] Marikovsky M, Vogt P, Eriksson E, Rubin JS, Taylor WG, Joachim S and Klagsbrun M. Wound fluid-derived heparin-binding EGF-like growth factor (HB-EGF) is synergistic with insulin-like growth factor-I for Balb/MK keratinocyte proliferation. *J Invest Dermatol* 1996; 106: 616-621.
- [32] Shirakata Y, Kimura R, Nanba D, Iwamoto R, Tokumaru S, Morimoto C, Yokota K, Nakamura M, Sayama K, Mekada E, Higashiyama S and Hashimoto K. Heparin-binding EGF-like growth factor accelerates keratinocyte migration and skin wound healing. *J Cell Sci* 2005; 118: 2363-2370.
- [33] Edwards JP, Zhang X and Mosser DM. The expression of heparin-binding epidermal growth factor-like growth factor by regulatory macrophages. *J Immunol* 2009; 182: 1929-1939.
- [34] Hallbeck AL, Walz TM and Wasteson A. Interleukin-6 enhances transforming growth factor-alpha mRNA expression in macrophage-like human monocytoid (U-937-1) cells. *Biosci Rep* 2001; 21: 325-339.
- [35] Rappolee DA, Mark D, Banda MJ and Werb Z. Wound macrophages express TGF-alpha and other growth factors in vivo: analysis by mRNA phenotyping. *Science* 1988; 241: 708-712.
- [36] Grose R and Werner S. Wound-healing studies in transgenic and knockout mice. *Mol Biotechnol* 2004; 28: 147-166.
- [37] Massague J and Pandiella A. Membrane-anchored growth factors. *Annu Rev Biochem* 1993; 62: 515-541.
- [38] Gallucci RM, Sloan DK, Heck JM, Murray AR and O'Dell SJ. Interleukin 6 indirectly induces keratinocyte migration. *J Invest Dermatol* 2004; 122: 764-772.
- [39] Chedid M, Rubin JS, Csaky KG and Aaronson SA. Regulation of keratinocyte growth factor gene

Skin wound healing modulation by macrophages

- expression by interleukin 1. *J Biol Chem* 1994; 269: 10753-10757.
- [40] Ashcroft GS, Yang X, Glick AB, Weinstein M, Letterio JL, Mizel DE, Anzano M, Greenwell-Wild T, Wahl SM, Deng C and Roberts AB. Mice lacking Smad3 show accelerated wound healing and an impaired local inflammatory response. *Nat Cell Biol* 1999; 1: 260-266.
- [41] Li AG, Wang D, Feng XH and Wang XJ. Latent TGFbeta1 overexpression in keratinocytes results in a severe psoriasis-like skin disorder. *EMBO J* 2004; 23: 1770-1781.
- [42] Sellheyer K, Bickenbach JR, Rothnagel JA, Bundman D, Longley MA, Krieg T, Roche NS, Roberts AB and Roop DR. Inhibition of skin development by overexpression of transforming growth factor beta 1 in the epidermis of transgenic mice. *Proc Natl Acad Sci U S A* 1993; 90: 5237-5241.
- [43] Werner S and Grose R. Regulation of wound healing by growth factors and cytokines. *Physiol Rev* 2003; 83: 835-870.
- [44] Werner S, Krieg T and Smola H. Keratinocyte-fibroblast interactions in wound healing. *J Invest Dermatol* 2007; 127: 998-1008.
- [45] Hubner G, Brauchle M, Smola H, Madlener M, Fassler R and Werner S. Differential regulation of pro-inflammatory cytokines during wound healing in normal and glucocorticoid-treated mice. *Cytokine* 1996; 8: 548-556.
- [46] Brauchle M, Angermeyer K, Hubner G and Werner S. Large induction of keratinocyte growth factor expression by serum growth factors and pro-inflammatory cytokines in cultured fibroblasts. *Oncogene* 1994; 9: 3199-3204.
- [47] Takehara K. Growth regulation of skin fibroblasts. *J Dermatol Sci* 2000; 24 Suppl 1: S70-77.
- [48] Beer HD, Florence C, Dammeier J, McGuire L, Werner S and Duan DR. Mouse fibroblast growth factor 10: cDNA cloning, protein characterization, and regulation of mRNA expression. *Oncogene* 1997; 15: 2211-2218.
- [49] Wang X, Waldeck H and Kao WJ. The effects of TGF-alpha, IL-1beta and PDGF on fibroblast adhesion to ECM-derived matrix and KGF gene expression. *Biomaterials* 2001; 22: 2542-2548.
- [50] Mori R, Shaw TJ and Martin P. Molecular mechanisms linking wound inflammation and fibrosis: knockdown of osteopontin leads to rapid repair and reduced scarring. *J Exp Med* 2008; 205: 43-51.
- [51] Stramer BM, Mori R and Martin P. The inflammation-fibrosis link? A Jekyll and Hyde role for blood cells during wound repair. *J Invest Dermatol* 2007; 127: 1009-1017.
- [52] Annes JP, Munger JS and Rifkin DB. Making sense of latent TGFbeta activation. *J Cell Sci* 2003; 116: 217-224.
- [53] Occleston NL, Fairlamb D, Hutchison J, O'Kane S and Ferguson MW. Avotermin for the improvement of scar appearance: a new pharmaceutical in a new therapeutic area. *Expert Opin Investig Drugs* 2009; 18: 1231-1239.
- [54] Postlethwaite AE, Keski-Oja J, Moses HL and Kang AH. Stimulation of the chemotactic migration of human fibroblasts by transforming growth factor beta. *J Exp Med* 1987; 165: 251-256.
- [55] Border WA and Noble NA. Transforming growth factor beta in tissue fibrosis. *N Engl J Med* 1994; 331: 1286-1292.
- [56] Grotendorst GR, Okochi H and Hayashi N. A novel transforming growth factor beta response element controls the expression of the connective tissue growth factor gene. *Cell Growth Differ* 1996; 7: 469-480.
- [57] Igarashi A, Okochi H, Bradham DM and Grotendorst GR. Regulation of connective tissue growth factor gene expression in human skin fibroblasts and during wound repair. *Mol Biol Cell* 1993; 4: 637-645.
- [58] Desmouliere A, Geinoz A, Gabbiani F and Gabbiani G. Transforming growth factor-beta 1 induces alpha-smooth muscle actin expression in granulation tissue myofibroblasts and in quiescent and growing cultured fibroblasts. *J Cell Biol* 1993; 122: 103-111.
- [59] Direkze NC, Forbes SJ, Brittan M, Hunt T, Jeffery R, Preston SL, Poulsom R, Hodivala-Dilke K, Alison MR and Wright NA. Multiple organ engraftment by bone-marrow-derived myofibroblasts and fibroblasts in bone-marrow-transplanted mice. *Stem Cells* 2003; 21: 514-520.
- [60] Ishii G, Sangai T, Sugiyama K, Ito T, Hasebe T, Endoh Y, Magae J and Ochiai A. In vivo characterization of bone marrow-derived fibroblasts recruited into fibrotic lesions. *Stem Cells* 2005; 23: 699-706.
- [61] Herdrich BJ, Lind RC and Liechty KW. Multipotent adult progenitor cells: their role in wound healing and the treatment of dermal wounds. *Cyotherapy* 2008; 10: 543-550.
- [62] Brown LF, Yeo KT, Berse B, Yeo TK, Senger DR, Dvorak HF and van de Water L. Expression of vascular permeability factor (vascular endothelial growth factor) by epidermal keratinocytes during wound healing. *J Exp Med* 1992; 176: 1375-1379.
- [63] Eming SA, Brachvogel B, Odorosio T and Koch M. Regulation of angiogenesis: wound healing as a model. *Prog Histochem Cytochem* 2007; 42: 115-170.
- [64] Li B, Sharpe EE, Maupin AB, Teleron AA, Pyle AL, Carmeliet P and Young PP. VEGF and PIGF promote adult vasculogenesis by enhancing EPC recruitment and vessel formation at the site of tumor neovascularization. *FASEB J* 2006; 20: 1495-1497.
- [65] Dvorak HF, Brown LF, Detmar M and Dvorak AM. Vascular permeability factor/vascular endothelial growth factor, microvascular hyperpermeability, and angiogenesis. *Am J Pathol* 1995; 146: 1029-1039.
- [66] Gerber HP, Dixit V and Ferrara N. Vascular endothelial growth factor induces expression of the

Skin wound healing modulation by macrophages

- antiapoptotic proteins Bcl-2 and A1 in vascular endothelial cells. *J Biol Chem* 1998; 273: 13313-13316.
- [67] Odorisio T, Cianfarani F, Failla CM and Zambruno G. The placenta growth factor in skin angiogenesis. *J Dermatol Sci* 2006; 41: 11-19.
- [68] Frank S, Hubner G, Breier G, Longaker MT, Greenhalgh DG and Werner S. Regulation of vascular endothelial growth factor expression in cultured keratinocytes. Implications for normal and impaired wound healing. *J Biol Chem* 1995; 270: 12607-12613.
- [69] Babic AM, Chen CC and Lau LF. Fisp12/mouse connective tissue growth factor mediates endothelial cell adhesion and migration through integrin alphavbeta3, promotes endothelial cell survival, and induces angiogenesis in vivo. *Mol Cell Biol* 1999; 19: 2958-2966.
- [70] Shimo T, Nakanishi T, Nishida T, Asano M, Kanayama M, Kuboki T, Tamatani T, Tezuka K, Takeamura M, Matsumura T and Takigawa M. Connective tissue growth factor induces the proliferation, migration, and tube formation of vascular endothelial cells in vitro, and angiogenesis in vivo. *J Biochem* 1999; 126: 137-145.
- [71] Nagy JA, Vasile E, Feng D, Sundberg C, Brown LF, Detmar MJ, Lawitts JA, Benjamin L, Tan X, Manseau EJ, Dvorak AM and Dvorak HF. Vascular permeability factor/vascular endothelial growth factor induces lymphangiogenesis as well as angiogenesis. *J Exp Med* 2002; 196: 1497-1506.
- [72] Maruyama K, Asai J, Ii M, Thorne T, Losordo DW and D'Amore PA. Decreased macrophage number and activation lead to reduced lymphatic vessel formation and contribute to impaired diabetic wound healing. *Am J Pathol* 2007; 170: 1178-1191.
- [73] Schledzewski K, Falkowski M, Moldenhauer G, Metharom P, Kzhyshkowska J, Ganss R, Demory A, Falkowska-Hansen B, Kurzen H, Ugurel S, Geginat G, Arnold B and Goerdt S. Lymphatic endothelium-specific hyaluronan receptor LYVE-1 is expressed by stabilin-1+, F4/80+, CD11b+ macrophages in malignant tumours and wound healing tissue in vivo and in bone marrow cultures in vitro: implications for the assessment of lymphangiogenesis. *J Pathol* 2006; 209: 67-77.
- [74] Ribatti D. The discovery of the placental growth factor and its role in angiogenesis: a historical review. *Angiogenesis* 2008; 11: 215-221.
- [75] Failla CM, Odorisio T, Cianfarani F, Schirotzma C, Puddu P and Zambruno G. Placenta growth factor is induced in human keratinocytes during wound healing. *J Invest Dermatol* 2000; 115: 388-395.
- [76] Hattori K, Heissig B, Wu Y, Dias S, Tejada R, Ferris B, Hicklin DJ, Zhu Z, Bohlen P, Witte L, Hendrikx J, Hackett NR, Crystal RG, Moore MA, Werb Z, Lyden D and Rafii S. Placental growth factor reconstitutes hematopoiesis by recruiting VEGFR1(+) stem cells from bone-marrow microenvironment. *Nat Med* 2002; 8: 841-849.
- [77] Adini A, Kornaga T, Firoozbakht F and Benjamin LE. Placental growth factor is a survival factor for tumor endothelial cells and macrophages. *Cancer Res* 2002; 62: 2749-2752.
- [78] Bottomley MJ, Webb NJ, Watson CJ, Holt L, Buhhari M, Denton J, Freemont AJ and Brenchley PE. Placenta growth factor (PIGF) induces vascular endothelial growth factor (VEGF) secretion from mononuclear cells and is co-expressed with VEGF in synovial fluid. *Clin Exp Immunol* 2000; 119: 182-188.
- [79] Carmeliet P, Moons L, Luttun A, Vincenti V, Compernolle V, De Mol M, Wu Y, Bono F, Devy L, Beck H, Scholz D, Acker T, DiPalma T, Dewerchin M, Noel A, Stalmans I, Barra A, Blacher S, Vandendriessche T, Ponten A, Eriksson U, Plate KH, Foidart JM, Schaper W, Charnock-Jones DS, Hicklin DJ, Herbert JM, Collen D and Persico MG. Synergism between vascular endothelial growth factor and placental growth factor contributes to angiogenesis and plasma extravasation in pathological conditions. *Nat Med* 2001; 7: 575-583.
- [80] Autiero M, Waltenberger J, Communi D, Kranz A, Moons L, Lambrechts D, Kroll J, Plaisance S, De Mol M, Bono F, Kliche S, Fellbrich G, Ballmer-Hofer K, Maglione D, Mayr-Beyrle U, Dewerchin M, Dombrowski S, Stanimirovic D, Van Hummelen P, Dehio C, Hicklin DJ, Persico G, Herbert JM, Shibuya M, Collen D, Conway EM and Carmeliet P. Role of PIGF in the intra- and intermolecular cross talk between the VEGF receptors Flt1 and Flk1. *Nat Med* 2003; 9: 936-943.
- [81] Loots MA, Lamme EN, Zeegelaar J, Mekkes JR, Bos JD and Middelkoop E. Differences in cellular infiltrate and extracellular matrix of chronic diabetic and venous ulcers versus acute wounds. *J Invest Dermatol* 1998; 111: 850-857.
- [82] Wetzler C, Kampfer H, Stallmeyer B, Pfeilschifter J and Frank S. Large and sustained induction of chemokines during impaired wound healing in the genetically diabetic mouse: prolonged persistence of neutrophils and macrophages during the late phase of repair. *J Invest Dermatol* 2000; 115: 245-253.
- [83] Tchaikovski V, Olieslagers S, Bohmer FD and Waltenberger J. Diabetes mellitus activates signal transduction pathways resulting in vascular endothelial growth factor resistance of human monocytes. *Circulation* 2009; 120: 150-159.
- [84] Goren I, Muller E, Pfeilschifter J and Frank S. Severely impaired insulin signaling in chronic wounds of diabetic ob/ob mice: a potential role of tumor necrosis factor-alpha. *Am J Pathol* 2006; 168: 765-777.
- [85] Zykova SN, Jenssen TG, Berdal M, Olsen R, Myklebust R and Seljelid R. Altered cytokine and nitric oxide secretion in vitro by macrophages from diabetic type II-like db/db mice. *Diabetes* 2000; 49: 1451-1458.
- [86] Fang Y, Shen J, Yao M, Beagley KW, Hambley BD

Skin wound healing modulation by macrophages

- and Bao S. Granulocyte-macrophage colony-stimulating factor enhances wound healing in diabetes via upregulation of proinflammatory cytokines. *Br J Dermatol* 2009;
- [87] Ure I, Partsch B, Wolff K and Petzelbauer P. Granulocyte/macrophage colony-stimulating factor increases wound-fluid interleukin 8 in normal subjects but does not accelerate wound healing. *Br J Dermatol* 1998; 138: 277-282.
- [88] Cianfarani F, Tommasi R, Failla CM, Viviano MT, Annessi G, Papi M, Zambruno G and Odorisio T. Granulocyte/macrophage colony-stimulating factor treatment of human chronic ulcers promotes angiogenesis associated with de novo vascular endothelial growth factor transcription in the ulcer bed. *Br J Dermatol* 2006; 154: 34-41.
- [89] Deiters U, Barsig J, Tawil B and Muhradt PF. The macrophage-activating lipopeptide-2 accelerates wound healing in diabetic mice. *Exp Dermatol* 2004; 13: 731-739.
- [90] Goren I, Muller E, Schiefelbein D, Christen U, Pfeilschifter J, Muhr H and Frank S. Systemic anti-TNFalpha treatment restores diabetes-impaired skin repair in ob/ob mice by inactivation of macrophages. *J Invest Dermatol* 2007; 127: 2259-2267.
- [91] Nahrendorf M, Swirski FK, Aikawa E, Stangenberg L, Wurdinger T, Figueiredo JL, Libby P, Weissleder R and Pittet MJ. The healing myocardium sequentially mobilizes two monocyte subsets with divergent and complementary functions. *J Exp Med* 2007; 204: 3037-3047.
- [92] Cochain C, Rodero MP, Vilar J, Recalde A, Richart A, Loinard C, Zouggari Y, Guerin C, Duriez M, Combadiere B, Poupel L, Levy BI, Mallat Z, Combadiere C and Silvestre JS. Regulation of monocyte subsets systemic levels by distinct chemokine receptors controls postischemic neovascularization. *Cardiovasc Res* 2010;in press.