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T1 ρ MRI Quantification of Arthroscopically-Confirmed Cartilage Degeneration

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Abstract

9 asymptomatic subjects and 6 patients underwent T1 ρ MRI to determine whether Outerbridge grade 1 or 2 cartilage degeneration observed during arthroscopy could be detected noninvasively. MRI was performed 2–3 months post-arthroscopy using sagittal T1-weighted and axial and coronal T1 ρ MRI from which spatial T1 ρ relaxation maps were calculated from segmented T1-weighted images. Median T1 ρ relaxation times of patients with arthroscopically documented cartilage degeneration and asymptomatic subjects were significantly different ($p < 0.001$) and median T1 ρ exceeded asymptomatic articular cartilage median T1 ρ by 2.5 to 9.2 ms. In 8 observations of mild cartilage degeneration at arthroscopy (Outerbridge grades 1 and 2), mean compartment T1 ρ was elevated in 5, but in all observations, large foci of increased T1 ρ were observed. It was determined that T1 ρ could detect some, but not all, Outerbridge grade 1 and 2 cartilage degeneration but that a larger patient population is needed to determine the sensitivity to these changes.

Keywords

Cartilage degeneration; T1 ρ ; arthroscopy

Introduction

The need to noninvasively detect the earliest changes in the degeneration of articular cartilage in order to both use and validate disease modifying osteoarthritic drugs (DMOADs) has stimulated considerable interest in the development of techniques that can directly probe the macromolecular structure. Three MRI techniques that were developed to directly assess the loss of proteoglycan that occurs during the early phases of osteoarthritis (OA) are sodium imaging, delayed Gadolinium-enhanced magnetic resonance imaging of cartilage (dGEMRIC) and T1 ρ -weighted MRI. T2-mapping is also widely used to assess the macromolecular constitution of cartilage (1–4), but collagen concentration and orientation

alters the relaxation rate, thereby it is not a specific measure of proteoglycan. On the other hand, changes in collagen orientation may be among the first changes to occur and have been visualized by T2 mapping (5).

Only a limited amount of research, however, has been performed to investigate the accuracy of the techniques used to probe proteoglycan content with the use of arthroscopy as the gold standard. In 15 patients with arthroscopically confirmed early changes of OA, who were studied with the dGEMRIC technique, T1 was lower in the compartment with those changes than in a reference compartment, indicating the loss of proteoglycan (6). In a similar study employing the dGEMRIC technique, a decrease in T1 was observed to correlate with abnormal arthroscopic findings and average T1 relaxation times were significantly different between radiographic grade 0 and grade 1 changes (7). No research currently exists to correlate sodium concentration to arthroscopy. All three techniques are limited in some respects, dGEMRIC requires the administration of exogenous contrast agents, sodium MRI requires specialized hardware, has relatively lower concentration than protons, short relaxation times and lower magnetic polarizability (8) and, as will be discussed further, T1 ρ is sensitive to a number of molecular processes including the rate of proton magnetic dipole reorientation, chemical exchange and residual dipolar effects.

T1 ρ was used previously to explore proteoglycan loss in patients with confirmed cartilage loss. A study of two patients with arthroscopically confirmed posttraumatic cartilage injury found increased T1 ρ compared to the healthy compartments in both patients (9). T1 ρ was increased ($p < 0.002$) in six subjects with OA scored using a Western Ontario and McMaster University (WOMAC) scale (10). In a recent study of 10 patients with radiographic OA scored using the Kellgren-Lawrence (KL) scale (11), T1 ρ was significantly increased from 45.5 ± 3.3 ms (KL = 0) to 55.6 ± 0.4 ms (KL = 4) (12). The specificity and sensitivity of T1 ρ imaging of articular cartilage lesions at arthroscopy has not yet been determined. In this preliminary, blinded, retrospective study, we tested the hypothesis that T1 ρ MRI can reliably detect cartilage degeneration demonstrated at arthroscopy in 6 patients.

Materials and Methods

Recruitment

The study was compliant with the Institutional Review Board (IRB) of the University of Pennsylvania and the Health Insurance Portability and Accountability Act (HIPAA). Prior to study participation, all asymptomatic subjects and patients were screened through a telephone interview to determine whether the inclusion and exclusion criteria, as detailed below, were satisfied. All asymptomatic subjects and patients gave informed consent to MRI.

Asymptomatic Subjects—9 asymptomatic subjects (2 men and 7 women) participated in the study. Asymptomatic was defined as having no knee pain at the time of the MRI examination and no previous history of surgery or major trauma to the knee. All asymptomatic subjects were between 30 and 59 years of age. Radiographs of asymptomatic subjects were not obtained.

Patients—6 patients (3 men and 3 women) participated in the study and were found to have one or more regions of cartilage softening or surface fibrillation (Outerbridge grade I or II cartilage degeneration as defined in *Arthroscopy* below). Some of these subjects also had regions of more severe cartilage degeneration which was documented at arthroscopy. All patients were between ages 40 and 76 and were identified for surgery for tears to one or both menisci.

Arthroscopy

All arthroscopy was performed by an orthopedic surgeon (J.H.L.) who specializes in disorders of the knee and has more than 15 years of experience in clinical practice.

All arthroscopic procedures were performed with general anesthesia, followed by local injection with 0.5% bupivacaine with epinephrine. No corticosteroids were injected into the knees at the termination of surgery. Inferolateral and inferomedial parapatellar portals were utilized for surgery. Areas of the most severe degenerative changes were recorded for each compartment and photographs of those areas were taken.

During surgery, areas of cartilage degeneration were not treated, unless the edges of chondral defects were unstable, in which case a chondroplasty of those unstable chondral edges was gently performed. Cartilage grading was performed according to the classification of Outerbridge modified to include the femorotibial joint (13) and was as follows:

Grade I: Softening and swelling of the cartilage.

Grade II: Early superficial fibrillation, which does not reach the subchondral bone, and is less than 0.5 inch in diameter.

Grade III: Fissuring that reaches the subchondral bone, which is not exposed, and greater than 0.5 inch in diameter.

Grade IV: Exposed subchondral bone of any diameter.

MRI

MRI was conducted 2–3 months post-arthroscopy to avoid the potential confounding variables of acute and subacute postoperative changes. It was determined that patients in the first two months following surgery found that the knee coil caused a good deal of pain.

MRI was performed on a 1.5 T (Sonata Model, Siemens Medical Solutions USA, Inc., Malvern, PA) clinical imaging system equipped with 40 mT/m gradients. RF was delivered by a body transmit coil and received by an 8-channel (Invivo, Orlando, FL) knee extremity coil.

Sagittal T1-weighted images were obtained using a magnetization prepared rapid gradient echo (mp-rage) acquisition pulse sequence with water selective spectral RF excitation (matrix = 256×256 , FOV = 140 mm^2 , ST = 0.5 mm, TR = 2700 ms, TE = 1.86 ms, IS = 20 mm, BW = 130 Hz/pixel, $\alpha = 15^\circ$, inversion time (TI) = 300 ms, 208 slices).

Axial and coronal T1 ρ -weighted images were obtained using a T1 ρ and half-alpha prepared phase alternated, multishot 3D balanced gradient echo sequence (matrix = 256×128 , FOV = 140 mm^2 , ST = 3 mm, TR = 6000 ms, slice thickness = 3 mm, BW = 130 Hz/pixel, $\alpha = 30^\circ$, 20 slices) (14,15). The sequence was also prepared by inversion recovery (TI = 1700 ms) and fat saturation with a 3 s delay between readout and subsequent inversion recovery of the next acquisition shot. Ideally, the spin lock field amplitude is chosen to coincide with the proton chemical exchange rate on hydroxyl and amide functional groups from the proteoglycan ($\nu_1 \approx 1500 \text{ Hz}$) (16,17), however, the high specific absorption rate of radiation (SAR) limited the amplitude $\nu_1 = 500 \text{ Hz}$. Five images were obtained with varying T1 ρ contrast (TSL = 2–40 ms) chosen so the longest TSL would coincide with the approximate T1 ρ of healthy cartilage (T1 $\rho \approx 40 \text{ ms}$) (18).

Image Processing and Analysis

Images reconstructed online from k-space data were exported in dicom format for offline processing in Matlab (v. 7.5.0, Natick, MA). Images were sorted according to contrast and aspect. The T1-weighted images were interpolated to 0.5 mm^3 and resliced along coronal and axial aspects and interpolated again along each aspect to $0.5 \text{ mm} \times 0.54 \text{ mm}^2$ to match the resolution of T1 ρ -weighted images. An initial manual alignment of the axial T1 ρ -weighted images to the resliced axial T1-weighted image was performed by in-plane rotation and translation after which a region of interest containing cartilage and bone was selected. Masked, axial T1 ρ -weighted images were exported to 3DVIEWNIX (MIPG, University of Pennsylvania, Philadelphia, PA) where T1 ρ -weighted images were registered automatically to T1-weighted images. The cartilage was semi-automatically segmented from the T1-weighted images using a LiveWire algorithm (19) and masks were applied to all T1 ρ -weighted images.

Masked, coregistered T1 ρ -weighted images were imported again into Matlab. The T1 ρ -weighted magnetization is modeled as a single exponential relaxation process $\ln(S) = -TSL/T1\rho + \ln(S_0)$ (20).

Arthroscopic reports, photos and T1 ρ maps were viewed by three of the authors (W.R.T.W., B.J.K., and J.H.L.), who categorized areas of the most severe degenerative changes as either encompassing a whole facet (diffuse) or localized to a region within the facet (focal). Diffuse lesions were quantified by the average compartment T1 ρ . Severe focal lesions (Outerbridge grade 3 or 4) were also quantified by the average compartment T1 ρ , which was necessary owing to the absence of cartilage at the site of exposed subchondral bone. Mild focal lesions (Outerbridge grade 1 or 2) were quantified by mean T1 ρ in an ROI drawn manually at the site of the lesion and extending across multiple slices.

Statistical Analysis

Statistical analysis was performed using the statistics toolbox in Matlab (v. 7.5.0, Natick, MA). To determine normality, a normal probability plot was constructed and normality was determined at the 5% significance level using a Lilliefors test.

Differences in median T1 ρ across all compartments between asymptomatic subjects and patients was assessed by means of a Wilcoxon signed rank test. Confidence intervals for differences in median T1 ρ were constructed by a 95% bootstrap confidence interval test performed by sampling with replacement (N = 1000).

Three-way ANOVA and multiple comparisons tests were performed to quantify the effects of population (asymptomatic subject or patient), side (medial or lateral) and compartment (patellar, femoral or tibial) on T1 ρ . Although the ANOVA model requires normally distributed data, the population T1 ρ distribution was only weakly non-normal. In addition, the population differences measured by the bootstrap confidence interval test were consistent with those measured by ANOVA.

Results

Group Results

T1 ρ relaxation times were quantified by side (medial or lateral) and facet (patellar, femoral or tibial) for both (post-arthroscopy) patients and asymptomatic subjects. Both patient and asymptomatic normal probability plots deviated from linearity indicating the T1 ρ was not normally distributed among populations. This was confirmed by a Lilliefors test (patient: $p < 0.03$; symptomatic: $p < 0.02$). T1 ρ was positively skewed, indicating a minimum, positive,

nonzero T1 ρ and high T1 ρ outliers. Median T1 ρ across all facets among patients and asymptomatic subjects was significantly different by a nonparametric Wilcoxon rank sum test ($p < 0.001$) and shown graphically in Figure 1. Patient exceeded asymptomatic articular cartilage median T1 ρ by 2.5 to 9.2 ms by a bootstrap confidence interval(CI)test (CI = 95%; N=1000).

A three-way ANOVA was performed to test the effects of population (patient or subject), side (medial or lateral) and facet (patellar, femoral or tibial) on T1 ρ while accounting for possible interactions between all three factors. Patient T1 ρ was significantly higher than asymptomatic T1 ρ by 1.6 to 9.6 ms (CI=95%; $p < 0.01$). This was in close agreement with the bootstrap CI test considered above. T1 ρ was significantly correlated with location ($p < 0.01$) and the patellar facet was 2.5 to 8.3 ms higher than the tibial facet T1 ρ (CI = 95%; $p < 0.01$), however the femoral facet was not significantly different from either the patellar or tibial facets, although median femoral T1 ρ was lower than patellar and higher than tibial cartilage facets. No interaction was observed between the three factors. It should be emphasized that the T1 ρ distribution was weakly non-normal as determined by the Lilliefors test above, however, the results of the multiple comparisons test for differences in populations were in close agreement with the bootstrap confidence interval test.

Individual Results

In 6 patients, 14 diffuse or focal areas of the most severe cartilage degeneration were made at arthroscopy. In 5 observations of grade 1 arthroscopic chondromalacia, T1 ρ was observed to be 1 standard deviation higher than asymptomatic controls in 3, normal in 1 and decreased in 1. In addition, elevated focal and heterogeneous T1 ρ was observed in 4. In 3 observations of grade 2 arthroscopic chondromalacia, T1 ρ was elevated in 2 and normal in a third (patient 6). In 6 observations of grade 3 and 4 focal and diffuse chondromalacia, T1 ρ was always elevated in the remaining, surrounding articular cartilage.

Representative arthroscopic photos and T1 ρ relaxation maps are shown for 1 asymptomatic subject and 3 patients in Figures 3–5 and documented changes of cartilage degeneration at arthroscopy and on T1 ρ MRI are briefly summarized in what follows. The asymptomatic subject (Figure 2) was observed to have homogeneously, smoothly varying patellar cartilage together with a characteristic increase in T1 ρ from the deep cartilage adjacent to the subchondral bone to the superficial cartilage adjacent to the synovium. At an ROI drawn at the cartilage surface, T1 ρ = 41.7 ms.

Patient 1 was observed at arthroscopy to have diffuse grade 1 degeneration throughout the entire knee joint (Figure 3). No focal defects or thinning was observed in either patellar or femorotibial compartments on MRI, however, a heterogeneous T1 ρ distribution was observed, as well as local elevated T1 ρ and cartilage thinning was observed in the lateral patellar superficial compartment and elevated T1 ρ = 47.2–56.3 across six slices. Diffuse, low femoral condyle compartment T1 ρ was observed (T1 ρ = 38.2 ms).

Patient 3 was observed at arthroscopy to have grade 2 patellar (Figure 4). Focal elevated T1 ρ on the medial patellar facet was observed by MRI with a patellar ROI for which T1 ρ = 49.2–62.7 ms, simultaneously with cartilage thinning.

Patient 6 was observed at arthroscopy to have grade 1 degeneration of the lateral patellar facet and grade 2 degeneration of the medial patellar facet (Figure 5). At MRI, both the medial and lateral patellar facets had focal T1 ρ lesions (T1 ρ = 49.6–56.5 and 53.4–62.3 ms, respectively). Mean T1 ρ in each femorotibial facet was 41.9 ms lateral and 44.7 ms medial indicating that mean T1 ρ was too conservative and misrepresented the local nature of the

disease. Grade 2 degeneration was observed in the medial femorotibial facet at arthroscopy and elevated $T1\rho = 43.8\text{--}45.3$ ms was observed laterally.

Discussion

A previous study of individuals with a history of knee pain found that $T1\rho$ was 25–30% higher in these subjects than in subjects without knee pain, even in cases where there was no observed cartilage loss on the $T1\rho$ -weighted image (10). The differences in $T1\rho$ relaxation times measured here were lower (6–20% difference corresponding with the bootstrap confidence interval of 2.5–9.2 ms), reflecting quite possibly severe, and certainly unknown, cartilage degeneration in patients recruited for that study. For asymptomatic subjects, $T1\rho$ was between 45–55 ms, nearly 5–10 ms higher than values reported here. This discrepancy might be explained by a partial volume containing differentially suppressed joint space fluid owing to the use of a short TR acquisition in that study and inversion recovery preparation in this study. Significant non-rigid body rotation of the knee coupled with a long scan duration (25 minutes to acquire all weighted images) or, alternatively, the estimation of $T1\rho$ by a single, manual region-of-interest drawn in a single slice in that study might explain this discrepancy.

More recently, a study quantifying both $T1\rho$ and $T2$ among patients classified according to KL score found significantly increased $T1\rho$ between osteoarthritic patients and controls and a greater percentage increase and effect size compared to $T2$ (12) and employed a multislice sequence of similar scan duration, having additionally performed registration of weighted images. In that study, mean $T1\rho$ among asymptomatic subjects were approximately 5 ms higher, possibly owing to the use of a multislice $T1\rho$ acquisition and post-processing removal of voxels containing a mixture of both cartilage and joint space fluid. As previously mentioned, it was desirable here to use fluid suppression to improve evaluation of the cartilage surface, which was often intact.

Mean $T1\rho$ was elevated for each facet where there was documented grade 3 or 4 arthroscopic cartilage degeneration. However, for grade 1 or 2 cartilage degeneration, it was more difficult to discern documented degeneration when averaging $T1\rho$ over an entire facet. It must be understood, however, that those facets labeled as grade 1 or 2 changes, demonstrated considerable heterogeneity at arthroscopy with normal and abnormal cartilage intermixed and that the arthroscopic grade was assigned on the basis of the highest grade of abnormality present and not on the basis of some type of average. Thus, a facet average would underestimate the arthroscopic grade and the use of at least reasonably large foci of elevated $T1\rho$ to compare to the arthroscopic grade is justified in that it better corresponds to how the arthroscopic grade is assigned. For example, the significantly elevated $T1\rho$ ($T1\rho = 49.2\text{--}62.7$ ms) in patient 3 at the site of a focal lesion is clearly abnormal, despite the mean compartment $T1\rho$ for the medial patellar facet which was reported to be moderately lower ($T1\rho = 49.1$ ms). In addition, the small subject number limits the assessment of how reliably $T1\rho$ correlates with early degeneration.

One potentially confounding variable is the presence of clinically occult osteoarthritic changes in asymptomatic subjects. This would artifactually elevate the mean $T1\rho$ in the asymptomatic subjects, making it more difficult to discriminate between normal and abnormal values, especially for the more mild observations of OA. To help compensate for the problem of asymptomatic degenerative changes in subjects, we are prospectively measuring $T1\rho$ values in a large number of asymptomatic subjects over a wide age range. Because the prevalence of degenerative changes likely increases with age, it may be possible to account for these changes for a given age, although individual variation may prove greater than aggregate changes resulting from age as was the case here.

Although extensive *ex vivo* validation and *in vivo* validation in a porcine animal model has been performed (21–24), the specificity of T1 ρ relaxation times to changes in proteoglycan concentration *in vivo* in humans is not clear. T1 ρ relaxation occurs when energy is exchanged with the lattice in the rotating frame of reference and thereby is sensitive to low frequency molecular dynamics of water protons (25). In this experiment, T1 ρ is sensitive to spin energy exchange with the lattice on the 2 ms time scale ($\nu_1 = 500$ Hz), however, several physical processes occur to protons in cartilage on this scale, both static and dynamic. External B₀ and RF B₁ static field heterogeneity causes image banding artifacts, changes in the apparent relaxation time and changes in the effective field strength, although these effects are mitigated using rotary echoes and moderate spin lock amplitudes ($\nu_1 \gg \Delta\nu_0$) (26–28). The residual dipolar interaction from motionally restricted water was found to contribute to the T1 ρ relaxation in cartilage, although the effect was reduced at higher spin lock amplitudes ($\nu_1 \gg \nu_D$) (29). Water proton exchange with exchanging amide and hydroxyl protons in glycosaminoglycans was shown to change T1 ρ relaxation times in controlled studies (16,17). T1 ρ decreases with decreasing rotational correlation time (30) and will depend on total water content and concentration in cartilage.

In conclusion, these results demonstrate that T1 ρ relaxation mapping correlates with chondral lesions identified by arthroscopy. T1 ρ MRI and arthroscopically documented cartilage degeneration was observed for Outerbridge grades 3 and 4 damage, but agreement was only modest in the case of grade 1 or 2 damage when averaging across compartments. On the other hand, a granular, region-of-interest approach demonstrated a one-to-one correspondence between T1 ρ MRI and cartilage degeneration in all grades and is the suggested approach for further studies.

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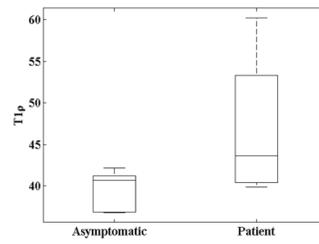


Figure 1. Boxplots of T1 ρ among asymptomatic subjects and patients. Median T1 ρ was significantly different by a nonparametric Wilcoxon rank sum test ($p < 0.001$).

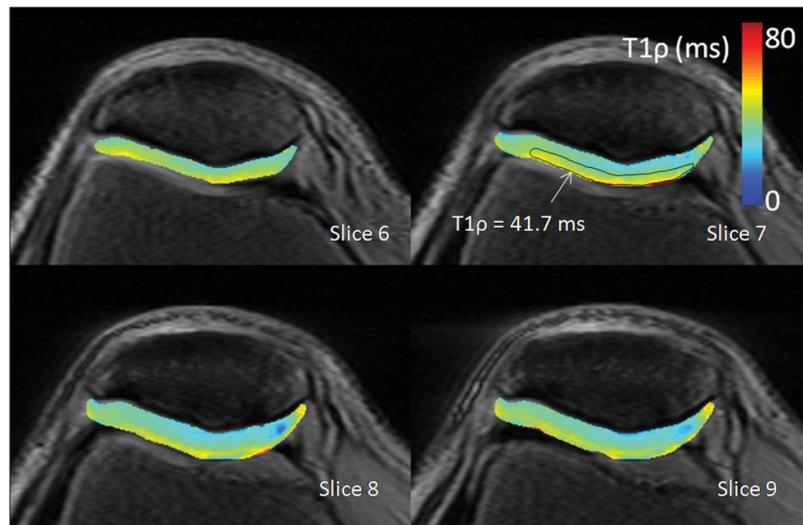


Figure 2. T1 ρ relaxation maps from a 30 year old male with no previous history of knee injury and no knee pain. Patellar cartilage is homogenously, smoothly varying and has a characteristic increase in relaxation time from the deep cartilage adjacent to the subchondral bone to the superficial cartilage adjacent to the synovium. At an ROI drawn at the cartilage surface, T1 ρ = 41.7 ms.

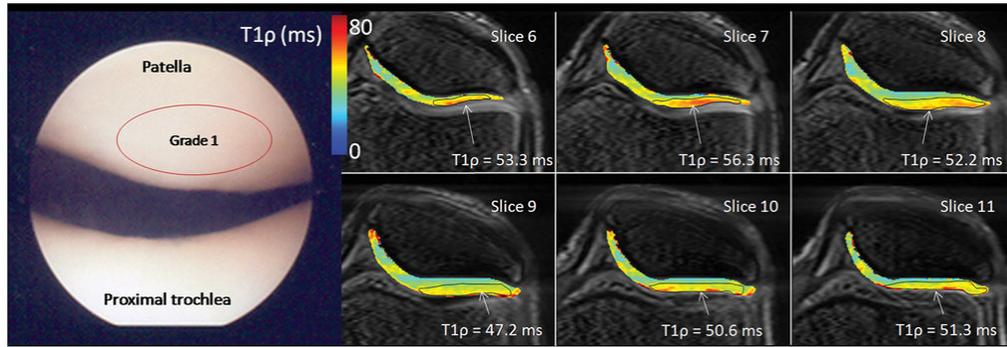


Figure 3.

Arthroscopic photographs and $T1\rho$ relaxation maps from a 40 year old male (patient 1). The patient was observed at arthroscopy to have diffuse grade 1 chondromalacia throughout the entire knee joint. No focal defects or thinning was observed in either patellar or femorotibial compartments on MRI, however, a heterogeneous $T1\rho$ distribution was observed, as well as local elevated $T1\rho$ and cartilage thinning was observed in the lateral patellar superficial compartment and elevated $T1\rho = 47.2\text{--}56.3$ across six slices. Diffuse, low femoral condyle compartment $T1\rho$ was observed ($T1\rho = 38.2$ ms).

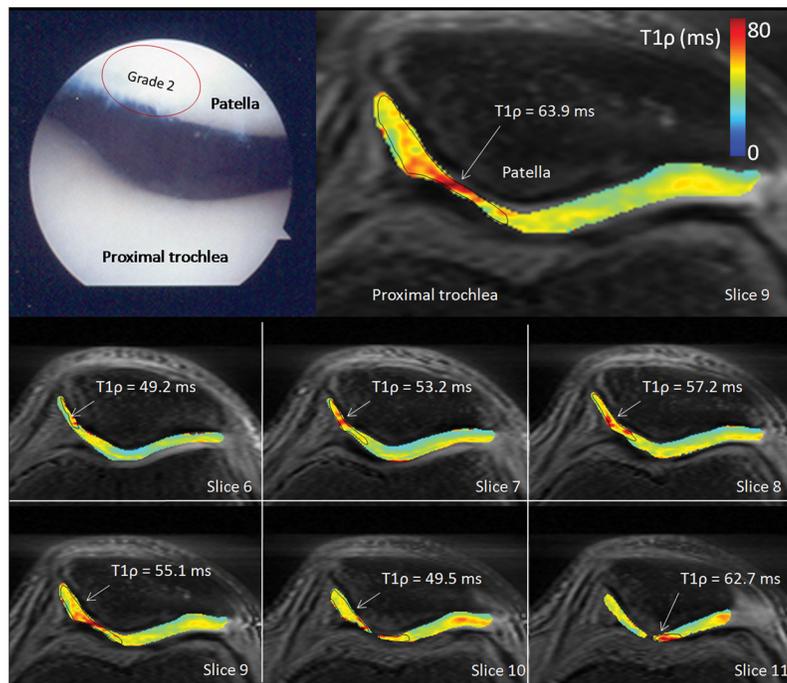


Figure 4. Arthroscopic photographs and T1 ρ relaxation maps from a 48 year old male (patient 3). This patient was observed at arthroscopy to have grade 2 patellar chondromalacia and a torn left medial meniscus for which a partial medial meniscectomy was performed. Focal elevated medial patellar T1 ρ was observed by MRI with a patellar ROI T1 ρ = 49.2–62.7 ms, simultaneously with cartilage thinning.

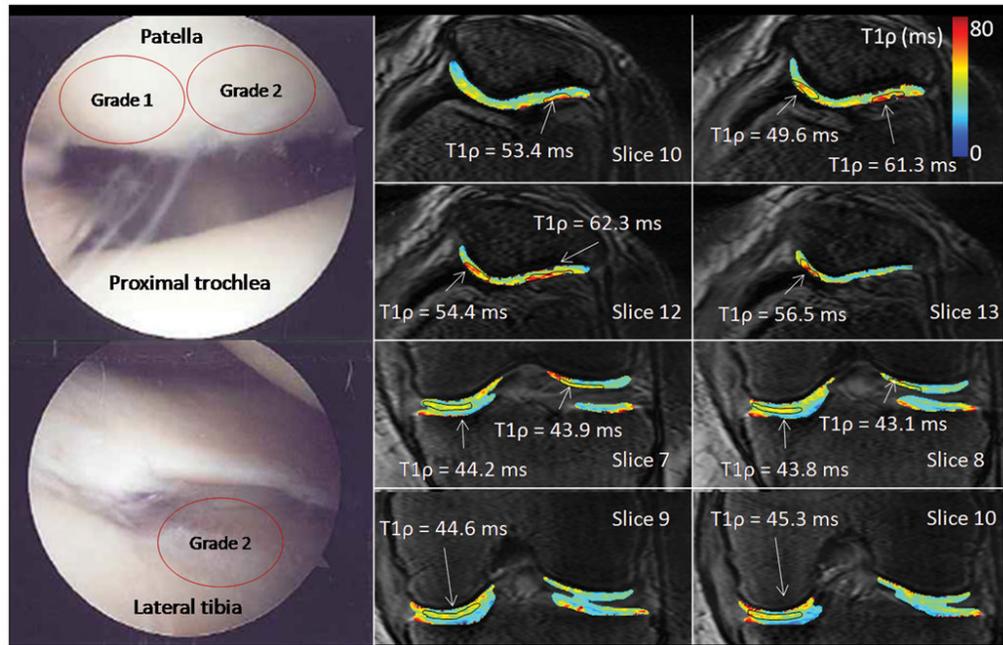


Figure 5. Arthroscopic photographs and T1 ρ relaxation maps from a 76 year old female (patient 6). This patient was observed at arthroscopy to have grade 1 chondromalacia of the lateral patellar facet and grade 2 chondromalacia of the medial patellar facet. At MRI, both the medial and lateral patellar facets had focal T1 ρ lesions (T1 ρ = 49.6–56.5 and 53.4–62.3 ms, respectively). Grade 2 chondromalacia was observed in the medial femorotibial compartment at arthroscopy and elevated T1 ρ = 43.8–45.3 ms was observed laterally.