

Mapping of the *supP* (*Su6*⁺) Amber Suppressor Gene in *Escherichia coli*

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The *supP* (*Su6*⁺) amber suppressor gene has been mapped on the clockwise side of the *valS* locus near min 95 on the *Escherichia coli* chromosome.

The *supP51* (formerly *Su6*⁺) amber suppressor was originally described by Chan and Garen (3) and shown to cause the amber triplet UAG to code for leucine. Fractionation of leucyl tRNA from *supP*⁺ (wild type) and *supP51* strains (5) revealed two fractions of UUG-binding leucyl-tRNA in the *supP*⁺ strain, whereas only one of these fractions was found in the *supP51* strain. In the *supP51* strain the other fraction had been replaced by UAG-binding leucyl tRNA (suppressor tRNA). Thus, the *supP51* mutation appears to have transformed one of two UUG-binding leucyl tRNA fractions normally present in *Escherichia coli* into amber suppressor tRNA.

The map position of the *supP* gene has not been known, but it has been reported to be located in the region covered by the F' factor F14 (9). Recently an amber suppressor gene, *supJ*, has been mapped in the corresponding region of the *Salmonella typhimurium* chromosome, closely linked to the *cya* locus (11).

Little is known about the distribution of leu-

cyl-tRNA genes on the *E. coli* chromosome. However, approximate mapping of tRNA genes in *E. coli* (6) has indicated the presence of leucyl tRNA genes in 3 widely separated chromosomal regions: min 66-69, 82-85, and 90-93.

After having obtained mapping results which indicated that the *supP* gene is not located in the F14 region, we carried out conjugation experiments to determine its approximate map position. These experiments showed that *supP* is located in the clockwise neighborhood of the *purA* gene, which is near min 93.5 on the *E. coli* map (1). Further mapping was done by P1-mediated transduction involving the *purA*, *pyrB*, *argI*, and *valS* genes. The last three genes are closely linked to each other near min 95 on the map (1). Their order has been determined in *E. coli* B and found to be as shown above (8). The recipients all carry the amber mutations *his-85* and *trpA9605*. Selection and scoring for *supP51* transductants was carried out on minimal media without histidine and tryptophan.

Our transduction mapping (Table 1 and Fig.

TABLE 1. Mapping of *supP51* (*Su6*⁺) by transduction with P1^a

Recipient strain	Selection	No. of transductants scored	Unselected marker	No. of cotransductants	% Cotransduction
GE554	<i>supP51</i>	295	<i>argI</i> ⁺	140	47.5
			<i>purA</i> ⁺	1	0.34
	<i>purA</i> ⁺	300	<i>argI</i> ⁺	6	2.0
GE556			<i>supP51</i>	0	0
	<i>supP51</i>	272	<i>pyrB</i> ⁺	100	36.8
			<i>purA</i> ⁺	1	0.37
	<i>purA</i> ⁺	282	<i>pyrB</i> ⁺	14	5.0
			<i>supP51</i>	1	0.35
GE557	<i>supP51</i>	250	<i>valS</i> ⁺	211	84.4
			<i>purA</i> ⁺	0	0
	<i>valS</i> ⁺	233	<i>supP51</i>	169	72.5
			<i>purA</i> ⁺	4	1.7

^a Strain CGSC4487 (strain S26, *Su6*⁺, of A. Garen; Hfr P02A *phoA4 relA1 tonA22 supP51* = *Su6*⁺) was used as donor in all experiments. All recipient strains are *supP*⁺ and carry the amber mutations *his-85* and *trpA9605*. Other characteristics of the recipient strains: GE554 F⁻ *purA54 argF58 argI61 thr-25 rpsL190 tsx-84*; GE556 F⁻ *purA54 pyrB9 argF58 rpsL190 tsx-84*; GE557 F⁻ *purA54 valS7 argF58 rpsL190 tsx-84*. Selection and scoring for *supP51* were carried out on minimal medium without tryptophan and histidine; *valS*⁺ transductants were selected on L-agar at 42°C. Selection for *pyrB*⁺ in strain GE556 resulted in preferential recovery of *purA*⁺ *supP*⁺ cotransductants (data not shown). Transduction procedures were as described by Thorbjarnardóttir et al. (10).

1) shows that the *supP* gene is cotransducible at high frequency with the *pyrB*, *argI*, and *valS* genes and at very low frequency (0.2 to 0.3%) with the *purA* gene. The order *purA*-(*pyrB*, *argI*, *valS*)-*supP* is established, placing the *supP* gene near 95 min on the map. This map position of the *supP* gene indicates that it may be very closely linked to the *ts-210* mutation which has recently been mapped by Isono and Kitakawa (7). One tRNA locus, *glyV*, has previously been mapped within the region shown in Fig. 1 (1, 4, 6). However, its exact position has not been reported. According to the approximate mapping referred to above (6), a leucyl tRNA locus, *leuV*, lies in the counterclockwise neighborhood of *purA*. The *leuV* locus is known to specify a CUG-binding leucyl-tRNA, tRNA^{Leu} (2, 6).

It can be concluded that the *supP* locus is neither identical to the *supJ* locus of *S. typhi-*

murium nor to any previously mapped leucyl tRNA locus in *E. coli*.

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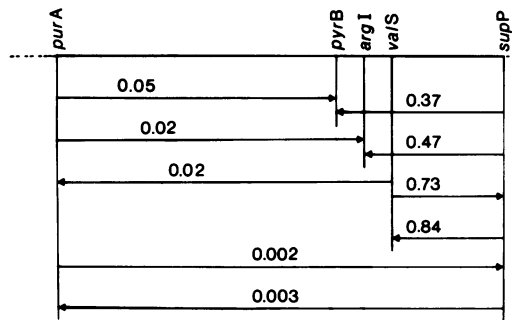


FIG. 1. Transduction map of the *purA*-*supP* region. Cotransduction frequencies are all derived from Table 1. Arrows point from selected marker to unselected marker.