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Signaling in Adult Neurogenesis

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Abstract

Neural stem cells (NSCs) in the adult brain continuously supply new neurons to the hippocampal dentate gyrus (DG) and the olfactory bulb (OB). Recent studies indicate that the progression from neural precursor cells (NPCs) to mature neurons is tightly controlled by coordinate cell-intrinsic programs and external signals within the neurogenic niche. In this review, we summarize both classes of regulatory factors involved in distinct stages of adult neurogenesis, including proliferation and lineage differentiation of NSCs, migration of neuroblasts and integration of newborn neurons. A full understanding of the wide variety of signaling pathways will ultimately provide precise targets for therapeutic applications.

Introduction

Over the last two decades, it has become apparent that persistent neurogenesis throughout life occurs in two specific brain areas of adult mammals: the subventricular zone (SVZ) of the lateral ventricles and the subgranular zone (SGZ) of the DG (Figure 1a) [1–4]. The newborn neuronal cells originate from adult NSCs in the germinal zones, which are defined by their ability to self-replicate and differentiate into multiple neural lineages, including neurons, astrocytes, and oligodendrocytes [5]. Two types of NSCs have been identified based on their morphology, proliferative behavior and marker expression, although their origin and identity remain to be defined (Figure 1b and c) [6–8]. In the SVZ, slowly-dividing, radial glia-like progenitors (type B cells) that express GFAP and CD133 have been hypothesized to be the primary NSCs in vivo. They are hypothesized to generate rapidly dividing, transit-amplifying progenitors (type C cells) that typically have either no or a very short process and are characterized as positive for Dlx2, Mash1, and EGFR. The majority of these intermediate progenitors subsequently give rise to DCX⁺ PSA-NCAM⁺ neuroblasts (type A cells) that migrate into the OB through the rostral migratory stream (RMS) and differentiate into GABAand dopamine-producing interneurons. In parallel, a population of GFAP⁺ Sox2⁺ Nestin⁺ radial cells (type 1 cells) is found to act as quiescent NSCs in the SGZ. They may generate actively self-renewing nonradial progenitors (type 2 cells) expressing Sox2 and Nestin but not GFAP, and type 2 cells in turn give rise to DCX⁺ neuroblasts that predominantly differentiate into local glutamatergic dentate granule cells (DGCs). It was recently found that a subset of type 2 Sox2⁺ cells has the potential to self-renew and generate both neurons and astrocytes, indicating a possible reciprocal lineage relationship between type 1 and 2 cells in the SGZ [9].

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Although NSCs have been derived from a variety of adult brain areas, active neurogenesis seems to be restricted to SVZ and SGZ under physiological conditions in vivo. Committed NPCs from SVZ differentiate into glia when grafted outside their normal neurogenic environment [10], whereas glial progenitors derived from spinal cord generate neurons when transplanted into the DG [11]. Furthermore, adult hippocampal NPCs grafted into RMS differentiate appropriately into neurons of non-hippocampus phenotype, whereas those grafted into nonneurogenic sites showed no neuronal differentiation [12]. These transplantation experiments clearly demonstrate that the microenvironment or neurogenic niche plays a crucial role in determining where and how neurogenesis can occur. However, it is also important to recognize that, even within the germinal zones of the adult brain, only a subset of markerexpressing cells are capable of generating neurons. Besides, the populations of adult NSCs are inherently diverse in their nature, as evidenced by the fact that NSCs from different regions of SVZ produce different neuronal subtypes, even when heterotopically grafted or grown in culture [13]. Thus, the progression from NSCs to mature neurons is subject to a tightly coordinated control by a multitude of cell-intrinsic and extracellular factors. Here we review recent progress in our understanding of the molecular mechanisms regulating the developmental steps of neurogenesis in the adult hippocampus and forebrain, including proliferation and fate specification of NPCs, their subsequent migration, and functional maturation.

Signals directing proliferation and fate commitment of NSCs

The extracellular signaling mechanisms that are present in the microenvironments of the SVZ and SGZ provide them with the unique ability to support and promote neurogenesis. (See Table 1) Signaling molecules that are critical during embryonic development of the nervous system are conserved and continue to modulate NSC activity and adult neurogenesis. The Wnt signaling pathway influences NSC proliferation and differentiation during embryonic development. Recent studies have also identified the Wnt signaling pathway as a key regulator of adult hippocampal neurogenesis. NSCs in the adult hippocampus express several Wnts as well as corresponding receptors, and therefore receive Wnt signals produced not only from astrocytes but also from themselves [14,15]. The autocrine signaling of Wnt supports the proliferation and multipotency of NSCs through the canonical pathway involving GSK3 β/β -catenin [15]. Activation of the Wnt/ β -catenin pathway increases expression of NeuroD1, which is known to promote neuronal differentiation in NSCs. Additionally, in response to Wnt, NSCs increase expression of LINE-1, a retrotransposon important in NSC survival [16]. These findings demonstrate the involvement of the Wnt signaling pathway in all aspects of adult neurogenesis.

Sonic hedgehog (Shh) has been studied extensively for its role in the developing nervous system, and it is now known to also be important in regulating the activity of adult NSCs, specifically acting as a mitogen and increasing proliferation [17]. The effects of Shh are mediated by primary cilia found on the NSCs and thus dependent on Kif3a, an essential motor for assembling primary cilia. Mice with a conditional loss of primary cilia on NSCs exhibit significantly reduced levels of proliferation in the DG, which is accompanied by a loss of Shh signaling. Disruption of Shh signaling by a conditional loss of Smoothened (Smo), a downstream target of Shh, results in a similar phenotype with reduced NSC proliferation [18].

In addition to the extracellular cues within the microenvironment of SVZ and SGZ, cellintrinsic mechanisms comprise another major component of control over the process of adult neurogenesis. An orphan nuclear receptor, Tlx, plays an essential role in maintenance and selfrenewal of adult NSCs [19–21], presumably by complexing with histone deacetylases (HDACs) to repress cell cycle genes p21 and PTEN [22]. It was recently revealed that Tlx can

The Sox family of genes, which encode for transcription factors, plays a prominent role in NSC activity, as demonstrated by the expression of Sox2 in NSCs. Additional functions of Sox2 include the regulation of NSC maintenance or proliferation, possibly by repressing GFAP transcription [24]. Adult mice bearing a Sox2 regulatory mutant allele exhibit a substantial decrease in NPC proliferation and generation of new neurons in both SVZ and SGZ [25]. The absence of Sox2 in the NSCs leads to abnormal neuronal differentiation, with the development of shorter and fewer processes and reduced expression of mature neuronal genes. Interestingly, Sox2 is necessary for neuronal differentiation only in the early stages, as expression of Sox2 during this stage rescues the arborizations and maturation of the neurons [24].

Furthermore, a member of the Sox family has been recently implicated in regulating neurogenesis through its interaction with microRNAs (miRNAs). miRNAs are endogenous, non-coding RNAs that regulate gene expression [26]. The microRNA miR-124, found abundantly in the brain [27], and its target gene Sox9 regulate neurogenesis in the SVZ. Expression of miR-124 is found in low levels in the SVZ, but miR-124 is elevated strictly in neuroblasts isolated from this region and increases with neuronal differentiation. The generation of neuroblasts from type C cells and neuronal differentiation in the adult SVZ requires downregulation of the transcription factor Sox9 by miR-124 [28]. In contrast, miR-124 overexpression or knockdown in the chick neural tube model has no effect on NPC proliferation and neuronal differentiation [29], suggesting that the role of miR-124 may be different depending on the developing and adult nervous system. miRNAs represent an exciting new member of the intrinsic factors that modulate adult neurogenesis.

The fate commitment of adult NSCs is sequentially controlled by a family of proneural proteins called basic helix-loop-helix (bHLH) transcription factors, whose functions are largely conserved during embryonic and adult neurogenesis. In the SVZ, Ascl1 (Mash1) is expressed in type C cells destined to become GABAergic interneurons in the OB [30], whereas Neurog2 and Tbr2 are found in a subset of dorsal SVZ progenitors that produce glutamatergic juxtaglomerular neurons [31]. In agreement with these *in vivo* analyses, all progeny of *in vitro* expanded adult NSCs of SVZ acquire a glutamatergic identity when forced to express Neurog2, whereas only GABAergic neurons are generated upon expression of Mash1 [32]. Similarly, Ascl1 and Neurog2 are transiently expressed by adult neural progenitors in the hippocampal neurogenic niche that mature into glutamatergic DGCs [30,33]. Interestingly, retrovirus-mediated overexpression of Ascl1 in the proliferating adult hippocampal NSCs causes them to exclusively generate cells of oligodendrocytic lineage instead of neurons [34], suggesting that expression of a single gene may direct fate choice of adult NSCs in their *in vivo* niche.

There is also increasing evidence that epigenetic mechanisms, such as DNA methylation, chromatin remodeling, histone modification and non-coding RNA expression, are closely associated with multiple aspects of adult neurogenesis. Adult mice lacking MBD1, a member of the methylated DNA-binding protein family, have decreased neurogenesis in the DG [35], whereas those with MeCP2 deletion have normal production but deficient maturation of newborn neurons [36]. Knockout of Gadd45b, a protein required for activity-induced DNA demethylation, is found to compromise both activity-induced proliferation of neural progenitors and dendritic growth of newborn neurons in the adult hippocampus [37]. Similarly, a chromatin remodeling factor Mll1 is required for neuronal differentiation of adult NSCs in the SVZ [38]. Furthermore, inhibition of HDACs induces neuronal differentiation of adult hippocampal neural progenitors [39], and a deficiency in the MYST family of histone acetyltransferases reduces the ability of SVZ NSCs to self-renew and differentiate [40]. Taken

together, these findings highlight the complexity and specificity of the control over the process of adult neurogenesis.

Neuroblast migration from the SVZ to the OB

Neuroblasts generated in the SVZ migrate along the RMS by chain migration to the OB [41], where they differentiate into GABAergic neurons that integrate into the preexisting circuits of the granule cell layer and contribute to olfactory learning [42]. The proper migration of the neuroblast from the germinal zone to their target destination underlies the ability of the newborn neurons to populate the OB. Recent studies have identified Shh as playing a crucial role in neuroblast migration along the RMS. In conditional Smoothened-null mice, where Hedgehog signaling has been disrupted, neuroblasts fail to migrate to the OB, which is accompanied by a thin RMS and reduced population of these cells. Interestingly, when the neuroblasts isolated from the conditional Smo-null mice are transplanted into the SVZ of wildtype mice, the mutant neuroblasts migrate along the RMS to the OB, suggesting that the migratory defect of the mutant neuroblasts can be rescued when they are placed in a permissive environment with proper Hedgehog signaling [43]. Specifically, Shh activity modulates neuroblast migration both in vitro and in vivo by serving as a chemoattractant for neuroblasts. Injection of Shhproducing cells into the dorsal telencephalon is sufficient to redirect migration of neuroblasts from the RMS, demonstrating that these cells respond and migrate towards Shh [44]. Insulinlike growth factor-1 (IGF-1) is important in proliferation and differentiation of neural progenitors and their survival as neurons. Additionally, IGF-1 signaling is necessary for neuroblast migration from the SVZ, as is evident in IGF- $1^{-/-}$ mice that exhibit neuroblast accumulation in the SVZ and improper migration to the OB [45]. Once the neuroblasts have reached the OB, they migrate radially to their final destination toward the periphery, utilizing blood vessels as their migrating scaffold. In the OB, neuroblasts are found along the blood vessels and associated with astrocytic endfeet [46]. The interactions between the neuroblasts and astrocytes from the RMS and OB are essential in the process of migration, as astrocytes from other regions such as the cortex do not support neuroblast migration [47].

Regulation of neuronal integration in the adult brain

A large portion of newborn neurons dies within 4 weeks after birth. Their survival is subject to regulation by diverse mechanisms, and so is their morphological/physiological development before integration into the existing neural circuitry. (See Fig. 2) Neuroblasts born in the postnatal SVZ express NMDA receptors (NMDARs) during migration to the OB. The NMDAR activity is regulated by glutamate released from astrocyte-like cells that ensheathe the neuroblasts. Single-cell NMDAR knockout leads to neuroblast apoptosis along the migratory path and therefore reduced neurogenesis, suggesting that astrocytic glutamate signaling through NMDARs plays a critical role in controlling the number of adult-born neurons that integrate into the synaptic network [48]. Interestingly, lack of electrical input to the OB does not affect the generation, migration, and early differentiation of olfactory granule cells, but significantly reduces the number of young granule cells in the OB soon after they develop spines and presumably become synaptically connected, which is between 15 and 45 days after their birth [49]. In parallel, knockout of the NR1 subunit of NMDARs in hippocampal progenitors reduces neuronal survival 2-3 weeks after their birth, and this reduction can be partially rescued by global inhibition of neuronal activity [50], suggesting a glutamatedependent critical period for survival of newborn DGCs. Owing to expression of NR2Bcontaining NMDARs, immature DGCs between 4 and 6 week old have enhanced capacity for synaptic plasticity, as shown by reduced induction threshold and increased magnitude of longterm potentiation (LTP), indicating a second glutamate-dependent critical period [51]. Although the role of NMDARs in synaptic plasticity in the OB remains to be elucidated, LTP expression is exclusively found in 2-8 week old olfactory granule cells but not in those mature

ones [52]. These results indicate that newborn neurons may provide a unique substrate for certain types of learning and memory. GABA, a major inhibitory neurotransmitter in the mature CNS, initially depolarizes newborn DGCs and tangentially migrating SVZ progenitors due to their high intracellular concentration of chloride ions [53,54]. Ambient GABA-induced depolarization and Ca²⁺-influx is required for dendritic initiation and elongation in postnatally generated olfactory interneurons [55]. Genetically switching GABAergic depolarization to hyperpolarization by knocking down the Cl⁻ importer NKCC1 in hippocampal progenitors reduces dendritic arborization and delays synapse formation [56]. In addition to neurotransmitters, the survival and synaptic integration of newly born cells are subject to regulation by neurotrophic factors. Knock-in of a variant form of BDNF (Val66Met) that results in reduced activity-dependent BDNF secretion impairs survival of SVZ neuroblasts and spontaneous olfactory discrimination [57]. Likewise, deletion of TrkB receptors in hippocampal progenitors reduces the growth of dendrites and spines in adult-born DGCs and compromises cell survival. It also leads to impaired LTP and remarkably increased anxietylike behavior [58]. In conditional knockout mice that lack expression of FGFR1, a major receptor for FGF-2, decreased neurogenesis is accompanied by a severe impairment of LTP at medial perforant path-DGC synapses and deficits in memory consolidation but not spatial learning [59]. In contrast, conditional knockout of NT-3 gene results in decreased LTP at lateral perforant path-DGC synapses and impaired performance on spatial memory tasks [60]. Furthermore, some molecules playing a cell-intrinsic role become identified, too. In both adult OB and DG, CREB activity positively modulates dendritic development and survival of nascent neurons [61,62]. Cdk5 is required for survival and dendritic growth of adult-generated hippocampal neurons [63,64]. The cell-intrinsic proteins, DISC1 and Klf-9, act as the negative and the positive regulator of synaptic integration in the adult DG, respectively [65,66]. NeuroD1 plays an essential role in the survival and maturation of newborn neurons in both hippocampus and OB [67]. It was recently found that expression of NeuroD1 in the periventricular region in vivo leads to the appearance of mature, neuron-like cells in the SVZ and RMS, where normally do not show addition of new neurons [68]. Taken together, various mechanisms coordinate to link adult neurogenesis and network activity, thereby allowing for experience-dependent construction of neural circuits, which in turn contributes to learning and memory. The evidence in support of the reciprocal relationship between learning and adult neurogenesis has been extensively reviewed elsewhere [8,69,70]. Interestingly, two recent findings respectively suggest that newborn neurons may be required for pattern separation function in the DG [71] and disruption of old memories encoded in the hippocampus [72].

Conclusions

These recent studies highlight the broad range of signaling mechanisms involved in the regulation of adult neurogenesis. A number of the signaling pathways, such as Wnt and Shh, are conserved and function prominently in both the developing nervous system and the germinal zones of the adult brain, supporting the neurogenic niche. Additionally, intrinsic factors such as miRNAs and transcription factors are increasingly demonstrating the cell-autonomous characteristics that provide the NSCs and NPCs with the potential to proliferate, differentiate, and survive as newborn neurons.

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- •• of outstanding interest

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Figure 1.

Neurogenesis in the adult brain. (a) Adult NSCs are primarily located in two germinal zones of the brain: the SVZ of the lateral ventricles and the SGZ of hippocampal DG. (b) A subset of relatively quiescent GFAP+ radial cells (type B cells) in the SVZ has the potential to serve as adult NSCs and generate rapidly dividing, transit-amplifying nonradial NSCs (type C cells), which in turn give rise to neuroblasts (type A cells) that migrate through the RMS toward the OB. (c) In the adult SGZ, a population of GFAP+ Sox2+ radial cells corresponds to quiescent NSCs (type 1 cells). They coexist with actively proliferating, GFAP- Sox2+ nonradial NSCs (type 2 cells) that generate both astrocytes and neuroblasts. Neuroblasts then migrate into the granule cell layer and mature into neurons.



Figure 2.

Factors regulating the functional integration of newborn neurons. (a) Critical factors in the OB modulate survival, dendritic arborization, and maturation of newborn neurons. Migrating neuroblasts express NMDAR and receive glutamate signaling from astrocytes, which is crucial to neuroblast survival. (b) Critical factors in the SGZ influence the survival, dendritic arborization, and synaptic plasticity and integration of newborn neurons.

Table 1

Summary of signaling in adult neurogenesis		References
Proliferation		
miR-124	decreases proliferation	28
Shh	increases proliferation	17_18
Sox2	increases proliferation	24_25
Tlx	increases proliferation	19_23
Wnt	increases proliferation	14_15
Differentiation		
Transcription factors		
Ascl1	overexpression generates oligodendrocytes	34
	expressed in NSCs to produce GABAergic interneurons in OB	30
	expressed in NSCs to produce glutamatergic neurons in hippocampus	30
Neurog2	expressed in NSCs to produce glutamatergic neurons in hippocampus	33
	expressed in NSCs to produce glutamatergic neurons in svz	31
Tbr2	expressed in NSCs to produce glutamatergic neurons in svz	31
Epigenetic mechanisms		
Gadd45b	necessary for dendritic arborization	37
MBD1	necessary for neuronal differentiation	35
MeCP2	necessary for neuronal maturation	37
M111	necessary for neuronal differentiation in svz	38
Migration		
IGF-1	necessary for neuroblast migration	45
Shh	necessary for neuroblast migration	43
Integration		
Extrinsic factors		
BDNF	increases neuronal survival and dendritic arborization	57_58
FGF-2	necessary for synaptic plasticity	59
GABA	necessary for dendritic arborization and synapse formation	55_56
Glutamate	necessary for neuronal survival and synaptic plasticity	48,50,_51
NT-3	necessary for synaptic plasticity	60
Intrinsic factors		
Cdk	necessary for neuronal survival and dendritic arborization	63_64
CREB	increases neuronal survival and dendritic arborization	61_62
DISC1	decreases synaptic integration	65
Klf-9	increases synaptic integration	66
NeuroDI	necessary for neuronal survival and maturation	67