Biased agonism

Terry Kenakin

Address: Department of Biological Reagents and Assay Development, Molecular Discovery, GlaxoSmithKline Research and Development, 5 Moore Drive, Research Triangle Park, NC 27709, USA

Email: terry.p.kenakin@gsk.com

Abstract

Seven-transmembrane receptors are commonly coupled to multiple signaling pathways in cells. The simple model describing agonists for these receptors as producing a common active state to induce uniform activation of the pathways linked to the receptor has been shown to be untenable in light of a large body of data that suggest that some agonists produce activation of some but not all available pathways. These agonists are referred to as 'biased' in that they select which signaling pathways become activated upon binding to the receptor. The data to support this mechanism as well as ideas on the possible therapeutic application of this effect will be discussed.

Introduction and context

Seven-transmembrane receptors are a family of signaling proteins that bind hormones, autacoids, and neurotransmitters to mediate a myriad of cellular functions. Like all proteins, these exist in collections of tertiary conformations called protein ensembles; receptors sample these conformations according to changes in the thermal energy in the system. Some of these conformations can be designated as 'active states', denoting their ability to activate cytosolic signaling mechanisms. The current model of pharmacological seven-transmembrane receptor agonism describes the selective binding of agonists to these active states to enrich their presence within the ensemble through a process referred to as 'conformational selection' [1]. Thus, through Le Chatelier's principle, upon agonist binding, the make-up of the ensemble is directed toward the conformations possessing the highest affinity for the agonist and these are stabilized at the expense of other conformations. The product of this thermodynamic process is a collection of membrane proteins that activate cellular signaling processes and agonism ensues. Early discussions of this mechanism and agonist efficacy in general used the most simple assumptions that the receptor is the minimal unit of control for this process and that a receptor activated by any agonist triggers all cellular signaling processes mediated by that particular receptor type.

Two experimental findings required the modification of this idea. First, it was observed that seven-transmembrane receptors pleiotropically interact with a wide range of cytosolic coupling proteins. Second, new technology revealed multiple biochemical behaviors of receptors such as the propensity to be phosphorylated and to interact with multiple G-proteins and β-arrestin, internalization, and desensitization. Subsequent studies have indicated that not all agonists uniformly produce activation of these multiple receptor behaviors. For example, the peptides $PACAP_{1-27}$ and $PACAP_{1-38}$ activate PACAP (pituitary adenylate cyclase-activating polypeptide type 1) receptors to elevate cyclic AMP and increase production of IP_3. However, the relative potency of these agonists for these responses is reversed; the relative efficacy of $PACAP_{1-27}$ for cyclic AMP elevation is higher than that of $PACAP_{1-38}$ but lower for elevation of IP_3 [2]. This indicates that the receptor is not the minimal unit of control of agonism, it is the agonist-receptor complex that controls the ultimate signaling event; the data leading to this conclusion also clearly indicate that agonist activation of multiple signaling mechanisms is not uniform but rather is often 'biased' toward some but not all signaling pathways. This concept has been put into a formal model showing that agonist-selective states can produce biased agonism [3].

In terms of the molecular mechanism of biased agonism, two additional factors are relevant to the discussion, namely the nature of the receptor-active state and the interaction of the activated receptor with multiple cytosolic signaling proteins. The selective activation of cellular pathways with biased agonists is consistent with there being 'agonist-specific' ensembles of receptor conformations mediating these events; subsequent studies have given independent corroboration of this mechanism through separate lines of experimentation that directly show that ligands can stabilize unique receptor conformations [4-9]. Similarly, molecular dynamics predicts that when proteins such as receptors change conformation, different regions of the receptor change independently (i.e., the protein does not form uniform global conformations) [10-13]. Therefore, the fact that signaling proteins interact with different regions of the receptor (e.g. [14,15]) suggests that different conformations would not produce uniform coupling to all signaling proteins; that is, the unique receptor conformations stabilized by agonists most likely will result in differential (biased) activation of cell signaling pathways [16]. In fact, the activation of a receptor that interacts with multiple signaling components in a cell most likely will never produce equal activation of all pathways; therefore, from this standpoint, every agonist will have a bias in signaling. However, a useful point of reference is the natural agonist for the receptor; this will have a natural signaling bias and can be used as a standard with which other agonists can be compared. Within this scale, functionally selective agonists are defined as having a signaling bias different from that of the natural agonist.

Major recent advances

Studies using a wide variety of technologies now indicate that biased agonism is a common phenomenon. Agonists have been shown to differentially activate different G-proteins and β-arrestin and have differing susceptibility to phosphorylation, desensitization, and internalization in a wide variety of receptor systems (for reviews, see [17-23]). In particular, many studies now specifically show how differential activation of G-proteins versus β-arrestin results from biased stabilization of receptor conformation [24,25]. Observed most often with synthetic agonist ligands, bias can also be detected in natural systems such as the chemokine CCR7 receptor. Thus, CCL19 and CCL21, two natural agonists for the CCR7 chemokine receptor, differ in the type of pathway stimulation they elicit; although both agonists produce G-protein activation, only CCL19 (not CCL21) causes receptor agonist-dependent phosphorylation and recruitment of β-arrestin to terminate the G-protein stimulus [26]. In addition to G-proteins and β-arrestin,

seven-transmembrane receptors have been shown to couple to many other signaling proteins such as JAK/STATs (Janus kinase/signal transducers and activators of transcription), Src-family tyrosine kinases, G-protein receptor kinases (GRKs), and PDZ domain-containing proteins [27]. Finally, other receptor types also are involved in biased signaling. For example, different proteases have been shown to differentially activate protease-activated receptors through stabilization of distinct conformation (much like biased ligands [28]).

The discovery that some ligands can produce activation of some but not all receptor-linked stimulus mechanisms can introduce ambiguity in the classification of drugs [29]. For instance, the active internalization of receptors by some antagonists indicates that a label of 'agonist' should be placed on this one aspect of receptor behavior (internalization) [17,30,31]. Similarly, while propranolol is a well-known antagonist and inverse agonist of agonist activation of G_s-coupled effects of β-adrenoceptors [32], it has also been shown to be an active activator (agonist) of extracellular signal-regulated kinase (ERK) [33]. Biased agonism can be complex and need not necessarily involve excitatory signaling events; that is, some ligands can be positive agonists for one pathway and antagonists or inverse agonists for another [34]. Such divergences in efficacy have been used to propose that efficacy be considered 'pluridimensional'; that is, ligands may have a range of different efficacies to cause a range of receptor behaviors [35].

Future directions

The therapeutic relevance of biased agonism is still unknown. The emphasis has clearly shifted from 'does biased agonism occur?' to 'when it does occur, how can it be harnessed therapeutically?' This trend is consistent with the improving technology to detect whole-cell effects through label-free technology; these data provide cell-specific and detailed information regarding signaling patterns of ligands [36-40]. Thus, while assay technology is making it increasingly possible to detect and characterize agonist bias, focus is shifting to the mechanisms responsible for this effect.

Emphasis has also shifted from the cell surface to the cytosol in an effort to understand biased signaling. For example, detailed studies of biased ligand-induced β-arrestin-mediated signaling indicate significant differences in functionally distinct pools of β-arrestin accessed through conformational control of receptor sensitivity to phosphorylation by GRK isoforms [41]. These unique phosphorylation patterns result in effective 'bar coding' of ligand-bound receptors that lead to further downstream instruction of β-arrestin partners within the

cell [41]. Moreover, just as receptors adopt different conformations to achieve selective signaling, it has now been shown that β-arrestin itself forms different conformations and that these are associated with selective functions within the cell [42].

The significance of biased signaling is increasingly appreciated as signaling pathways become linked to normal physiology and pathophysiology. For example, the reported ERK activation through β-arrestin by some antagonists (i.e. propranolol [33]) is interesting in light of recent data that suggest that angiotensin receptor-mediated β-arrestin stimulation may be beneficial in the treatment of heart failure [43] or, alternatively, may be linked to the progression of heart failure through aldosterone pathways [44]. Biased signaling has been implicated in a unique profile of antagonism and agonism in breast cancer. Specifically, substance-P analogues SP-D and SP-G have been shown to produce biased signaling at vasopressin V_{1A} receptors and receptors for gastrin-releasing peptide to yield a profile of blocked G_q-protein-mediated calcium release and concomitant activation of ERK. This is postulated to produce a unique antiproliferative profile of activity [45].

In terms of specific examples in which bias may be a practical aspect of drug activity, there are interesting divergences in activity for opioid agonists to produce analgesia with reduced propensity to produce desensitization and δ-opioid receptor internalization [46-50] and intriguing functionally selective dopamine [51] and serotonin [52,53] agonists for the treatment of psychiatric disorders. Similarly, there are data that suggest that functionally selective thyrotropin agonists may be able to differentiate thyroid growth and thyroid hormone synthesis [54] and orexin receptor functionally selective agonists may differentiate effects on catecholamine release and adrenal steroid production [55,56]. The consequences of biased agonism currently are an intense subject of study. While initial work in the field considered acute effects such as desensitization, receptor internalization, and differential activation of G-proteins, more recent studies center on long-term effects of biased signaling such as prolonged signaling through β-arrestin activation and effects such as G-protein regulation [57].

In general, it is not yet clear to what extent these biased stimulations of cellular signaling pathways will yield favorable therapeutic phenotypic agonism. What is clear is that seven-transmembrane receptors cannot be considered on/off switches and synthetic agonists cannot be considered surrogates of natural agonists. Stabilization of receptor-active states by different molecules has the potential to traffic receptor stimulus in unique ways to cause complex patterns of cellular activation. The challenge now is to harness this powerful effect.

Abbreviations
CCR7, chemokine (C-C motif) receptor 7; CCL, chemokine (C-C motif) ligand; ERK, extracellular signal-regulated kinase; GRK, G-protein receptor kinase; PACAP, pituitary adenylate cyclase-activating polypeptide type 1.

Competing interests
The author declares that he has no competing interests.

References
1. Burgen AS: **Conformational changes and drug action.** *Fed Proc* 1966, **40**:2723-8.

2. Spengler D, Waeber C, Pantoloni C, Holsboer F, Bockaert J, Seeburg PH, Journot L: **Differential signal transduction by five splice variants of the PACAP receptor.** *Nature* 1993, **365**:170-5.

3. Kenakin TP: **Agonist-receptor efficacy II: agonist trafficking of receptor signals.** *Trends Pharmacol Sci* 1995, **16**:232-8.

4. Gether U, Lin S, Kobilka BK: **Fluorescent labeling of purified β₂-adrenergic receptor: evidence for ligand specific conformational changes.** *J Biol Chem* 1995, **270**:28268-75.

5. Ghanouni P, Gryczynski Z, Steenhuis JJ, Lee TW, Farrens DL, Lakowicz JR, Kobilka BK: **Functionally different agonists produce distinct conformations in G-protein coupling domains of the β₂-adrenergic receptor.** *J Biol Chem* 2001, **276**:24433-6.

6. Hruby VJ, Tollin G: **Plasmon-waveguide resonance (PWR) spectroscopy for directly viewing rates of GPCR/G-protein interactions and quantifying affinities.** *Curr Opin Pharmacol* 2007, **7**:507-14.

7. Okada T, Palczewski K: **Crystal structure of rhodopsin: implications for vision and beyond.** *Curr Opin Struc Biol* 2001, **11**:420-6.

8. Palanche T, Ilien B, Zoffmann S, Reck M-P, Bucher B, Edelstein SJ, Galzi J-L: **The neurokinin A receptor activates calcium and cAMP responses through distinct conformational states.** *J Biol Chem* 2001, **276**:34853-61.

9. Swaminath G, Xiang Y, Lee TW, Steenhuis J, Parnot C, Kobilka BK: **Sequential binding of agonists to the β₂ adrenoceptor: kinetic evidence for intermediate conformational states.** *J Biol Chem* 2004, **279**:686-91.

10. Frauenfelder H, Parak F, Young RD: **Conformational substrates in proteins.** *Annu Rev Biophys Biophys Chem* 1988, **17**:451-79.

11. Frauenfelder H, Sligar SG, Wolynes PG: **The energy landscapes and motions of proteins.** *Science* 1991, **254**:1598-603.

12. Hilser J, Freire E: **Predicting the equilibrium protein folding pathway: structure-based analysis of staphylococcal nuclease.** *Protein Struct Funct Bioinform* 1997, **27**:171-83.

13. Woodward C: **Is the slow-exchange core the protein folding core?** *Trends Biochem Sci* 1993, **18**:359-60.

14. Ikezu T, Okamoto T, Ogata E, Nishimoto I: **Amino acids 356-372 constitute a Gi-activator sequence of the alpha 2-adrenergic receptor and have a Phe substitute in the G protein-activator sequence motif.** *FEBS Lett* 1992, **311**:29-32.

15. Jones BW, Hinkle PM: **Arrestin binds to different phosphorylated regions of the thyrotropin-releasing hormone receptor with distinct functional consequences.** *Mol Pharmacol* 2008, **74**:195-202.

16. Kenakin TP: **Efficacy at G protein coupled receptors.** *Nat Rev Drug Discov* 2002, **1**:103-9.

17. Kenakin TP: **Collateral efficacy as a pharmacological problem applied to new drug discovery.** *Expert Opin Drug Discov* 2006, **1**:635-52.

18. Kenakin TP: **Collateral efficacy in drug discovery: taking advantage of the good (allosteric) nature of 7TM receptors.** *Trends Pharmacol Sci* 2007, **28**:407-15.

19. Kenakin TP: **Efficacy at G protein coupled receptors.** *Annu Rev Phamacol Toxicol* 2002, **42**:349-79.

20. Hermans E: **Biochemical and pharmacological control of the multiplicity of coupling at G-protein receptors.** *Pharmacol Ther* 2003, **99**:25-44.

21. Perez DM, Karnick SS: **Multiple signaling states of G-protein coupled receptors.** *Pharmacol Rev* 2005, **57**:147-61.

22. Kukkonen JP: **Regulation of receptor-coupling to (multiple) G proteins: a challenge for basic research and drug discovery.** *Receptors Channels* 2004, **10**:167-83.

23. Kenakin TP (Ed): **Special Issue on Allosterism and Collateral Efficacy.** *Trends Pharmacol Sci* 2007, **28**:359-446.

24. Drake MT, Violin JD, Whalen EJ, Wisler JW, Shenoy SK, Lefkowitz RJ: **β-arrestin-biased agonism at the β2-adrenergic receptor.** *J Biol Chem* 2008, **283**:5669-76.

   F1000 Factor 4.8 *Must Read*
   Evaluated by Soeren Sheikh 25 Feb 2008, Susan Steinberg 30 Jan 2009

25. Galandrin S, Oligny-Longpre G, Bonin H, Ogawa K, Gales C, Bouvier M: **Conformational rearrangements and signaling cascades involved in ligand-biased mitogen-activated protein kinase signaling through the β1-adrenergic receptor.** *Mol Pharmacol* 2008, **74**:162-72.

   F1000 Factor 4.8 *Must Read*
   Evaluated by Ralf Jockers 25 Jun 2008, Andrew Tobin 13 Aug 2008

26. Kohout TA, Nicholas SL, Perry SJ, Reinhart G, Junger S, Struthers RS: **Differential desensitization, receptor phosphorylation, β-arrestin recruitment, and ERK1/2 activation by the two endogenous ligands for the CC chemokine receptor 7.** *J Biol Chem* 2004, **279**:23214-22.

27. Sun Y, McGarrigle D, Huang XY: **When a G protein-coupled receptor does not couple to a G protein.** *Mol Biosyst* 2007, **3**:849-54.

28. Russo A, Soh UJK, Trjo J: **Proteases display biased agonism at protease-activated receptors: location matters.** *Mol Intervent* 2009, **9**:87-96.

29. Kenakin TP: **Pharmacological onomastics: what's in a name?** *Br J Pharmacol* 2008, **153**:432-8.

30. Gray JA, Roth BL: **Paradoxical trafficking and regulation of 5-HT$_{2A}$ receptors by agonists and antagonists.** *Brain Res Bulletin* 2001, **56**:441-51.

31. Roettger BF, Ghanekar D, Rao R, Toledo C, Yingling J, Pinon D, Miller LJ: **Antagonist-stimulated internalization of the G protein-coupled cholecystokinin receptor.** *Mol Pharmacol* 1997, **51**:357-62.

32. Baker JG, Hall IP, Hill SJ: **Agonist and inverse agonist actions of β-blockers at the human β2-adrenoceptor provide evidence for agonist-directed signaling.** *Mol Pharmacol* 2003, **64**:1357-69.

33. Azzi M, Charest PG, Anger S, Rouseau G, Kohout T, Bouvier M, Pineyro G: **β-arrestin-mediated activation of MAPK by inverse agonists reveals distinct active conformations for G-protein-coupled receptors.** *Proc Natl Acad Sci U S A* 2003, **100**:11406-11.

   F1000 Factor 3.0 *Recommended*
   Evaluated by Rafael Franco 27 Sep 2006

34. Galandrin S, Oligny-Longpre G, Bouvier M: **The evasive nature of drug efficacy: implications for drug discovery.** *Trends Pharmacol Sci* 2007, **28**:423-30.

35. Galandrin S, Bouvier M: **Distinct signaling profiles of β1 and β2 adrenergic receptor ligands toward adenylyl cyclase and mitogen-activated protein kinase reveals the pluridimensionality of efficacy.** *Mol Pharmacol* 2006, **70**:1575-84.

   F1000 Factor 3.0 *Recommended*
   Evaluated by Paul Simpson 30 Oct 2006

36. Peters MF, Scott CW: **Evaluating cellular impedance assays for detection of GPCR pleiotropic signaling and functional selectivity.** *J Biomol Screen* 2009, **14**:246-55.

37. Kenakin TP: **Cellular assays as portals to seven transmembrane receptor-based new drug discovery.** *Nat Rev Drug Discov* 2009, **8**:617-26.

38. van der Lee MMC, Blomenrohr M, van der Doelen AA, Wat JWY, Smits N, Hanson BJ, van Koppen CJ, Zaman GJR: **Pharmacological characterization of receptor redistribution and β-arrestin recruitment assays for cannabinoid receptor 1.** *J Biomol Screen* 2009, **14**:811-23.

39. Fang Y, Ferrie AM: **Label-free optical biosensor for ligand-directed functional selectivity acting on β$_2$-adrenoceptor in living cells.** *FEBS Lett* 2008, **582**:558-64.

40. Hanson BJ, Wetter J, Bercher MR, Kopp L, Fuerstenau-Sharp M, Vedvik KL, Zielinski T, Doucette C, Whitney PJ, Revankar C: **A homogeneous fluorescent live-cell assay for measuring 7-transmembrane receptor activity and agonist functional selectivity through beta-arrestin recruitment.** *J Biomol Screen* 2009, **14**:798-810.

41. Zidar DA, Violin JD, Whalen EJ, Lefkowitz RJ: **Selective engagement of G protein coupled receptor kinases (GRKs) encodes distinct functions of biased ligands.** *Proc Natl Acad Sci U S A* 2009, **106**:9649-54.

42. Shukla AK, Violin JD, Whalen EJ, Gesty-Palmer D, Shenoy S, Lefkowitz RJ: **Distinct conformational changes in β-arrestin report biased agonism at seven transmembrane receptors.** *Proc Natl Acad Sci U S A* 2008, **105**:9988-93.

43. Aplin M, Bonde MM, Hansen JL: **Molecular determinant of angiotensin II type 1 receptor functional selectivity.** *J Mol Cell Cardiol* 2009, **46**:15-24.

44. Lymperopoulos A, Rengo G, Zincarelli C, Kim J, Soltys S: **An adrenal β-arrestin 1-mediated signaling pathway underlies angiotensin II-induced aldosterone production in vitro and in vivo.** *Proc Natl Acad Sci U S A* 2009, **106**:5825-30.

45. MacKinnon AC, Tufail-Hanif U, Lucas CD, Jodrell D, Haslett C, Sethi T: **Expression of V1A and GRP receptors leads to cellular transformation and increased sensitivity to substance-P analogue-induced growth inhibition.** *Br J Cancer* 2005, **92**:522-31.

46. Whistler JL, Van Zastrow M: **Morphine-activated opioid receptors elude desensitization by β-arrestin.** *Proc Natl Acad Sci USA* 1998, **95**:9914-9.

47. Zhang J, Ferguson SSG, Brak LS, Bodduluri SR, Laporte SA, Law P-Y, Caron MG: **Role for G protein-coupled receptor kinase in agonist-specific regulation of μ-opioid receptor responsiveness.** *Proc Natl Acad Sci U S A* 1998, **95**:7157-62.

48. Bailey CP, Couch D, Johnson E, Griffiths K, Kelly E, Henderson G: **μ-Opioid receptor desensitization in mature rat neurons: lack of interaction between DAMGO and morphine.** *J Neurosci* 2003, **23**:10515-20.

49. Bohn LM, Dykstra LA, Lefkowitz RJ, Caron MG, Barak LS: **Relative opioid efficacy is determined by the complements of the G protein coupled receptor desensitization machinery.** *Mol Pharmacol* 2004, **66**:106-12.

50. Koch T, Widera A, Bartzsch K, Schulz S, Brandenburg L-O, Wundrack N, Beyer A, Grecksch G, Höllt V: **Receptor endocytosis counteracts the development of opioid tolerance.** *Mol Pharmacol* 2005, **67**:280-7.

51. Mailman RB: **GPCR functional selectivity has therapeutic impact.** *Trends Pharmacol Sci* 2007, **28**:390-7.

52. Schmid CL, Raehal KM, Bohn LM: **Agonist-directed signaling of the serotonin 2A receptor depends on β-arrestin-2 interactions in vivo.** *Proc Natl Acad Sci USA* 2008, **105**:1079-84.

53. Meyer JH, McMain S, Kennedy SH, Korman L, Brown GM, DaSilva JN, Wilson AA, Blak T, Eynan-Harvey R, Goulding VS, Houle S, Links P: **Dysfunctional attitudes and 5-HT2 receptors during depression and self harm.** *Am J Psychiat* 2003, **160**:90-9.

54. Vassart G, Dumont D: **The thyrotropin receptor and the regulation of thyrocyte function and growth.** *Endocr Rev* 1992, **13**:596-611.

55. Mazzocchi G, Malendowicz LK, Aragona F, Nussdorfer GG: **Human pheochromocytomas express orexin receptor type 2 gene and display an in vitro secretory response to orexins A and B.** *J Clin Endocrinol Metab* 2001, **86**:4818-21.

56. Mazzocchi G, Malendowicz LK, Gottardo AF, Nussdorfer GG: **Orexin A stimulates cortisol secretion from human adreno-cortical cells through activation of the adenylate cyclase-dependent signaling cascade.** *J Clin Endocrinol Metab* 2001, **86**:778-82.

57. Xu H, Wang X, Partilla JS, Bishop-Mathis K, Benaderet TS, Dersch CM, Simpson DS, Prisinzano TE, Rothman RB: **Differential effects of opioid agonists on G protein expression in CHO cells expressing cloned human opioid receptors.** *Brain Res Bull* 2008, **77**:49-54.