

Brief Communication

Increased brain microvascular MMP-9 and incidence of haemorrhagic transformation in obese mice after experimental stroke

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Obesity is an independent risk factor for stroke and is associated with poorer outcome after stroke. We investigated whether this poorer outcome is related to brain microvascular disruption. Focal cerebral ischaemia was induced in lean or obese (*ob/ob*) mice by transient middle cerebral artery occlusion. The incidence of haemorrhagic transformation and the volume of ischaemic brain damage were significantly greater in obese mice. Blood–brain barrier permeability and brain microvascular MMP-9 expression were also markedly increased in obese mice. These effects were independent of leptin or glycaemic status, suggesting that obesity potentiates brain microvascular disruption after experimental stroke.

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Introduction

Obesity is an independent risk factor for stroke, and is associated with atherosclerosis, diabetes, and hypertension, conditions that predispose to stroke (Winter *et al*, 2008). The increased risk of stroke in obese individuals may also be accompanied by poorer prognosis after the ischaemic insult (Razinia *et al*, 2007). In support of this, recent studies have demonstrated greater brain damage in obese rodents after experimental stroke (Mayanagi *et al*, 2008; Terao *et al*, 2008). However, the mechanisms responsible for these detrimental effects of obesity on cerebrovascular injury are poorly delineated.

A crucial site of convergence for pathophysiological mechanisms involved in obesity and stroke is the brain microvascular endothelium. Obesity is a state of chronic systemic inflammation and is associated with vascular oxidative stress (Dandona *et al*, 2004). Inflammation and oxidative stress are important factors that contribute to disruption of the blood–brain barrier (BBB). Mechanisms include disruption of inter-endothelial tight-junction complexes (Schreibelt *et al*, 2007) and induction of proteases, in particular matrix metalloproteinases (MMPs), that

degrade constituents of the vascular basement membrane (Zhao *et al*, 2007). We have shown previously that systemic inflammation exacerbates ischaemic brain damage through MMP-9-dependent alterations in BBB integrity (McColl *et al*, 2008). Clinically, loss of BBB integrity after stroke is associated with serious complications such as brain oedema and haemorrhagic transformation (HT) that correlate with poor prognosis (Simard *et al*, 2007). Accordingly, it is feasible that obesity may predispose to poorer outcome after stroke through aggravation of brain microvascular disruption.

In the present study, we sought to determine if obesity exacerbates brain microvascular disruption after experimental stroke.

Materials and methods

Focal Cerebral Ischaemia

Experiments were performed on 8- to 12-week-old male obese *ob/ob* (C57BL/6OlaHsd-Lep^{ob}) or lean littermate mice (Harlan-Olac, Bicester, UK) according to the Animals (Scientific Procedures) Act (1986). Body weight and blood glucose concentration at the time of surgery were as follows: body weight, lean 29.0 ± 0.5 g versus *ob/ob* 47.3 ± 0.9 g ($P < 0.001$) and glucose, lean 9.3 ± 0.7 mmol/L versus *ob/ob* 10.1 ± 1.4 mmol/L ($P > 0.05$). Focal ischaemia was induced by transient (40 mins) middle cerebral artery occlusion (MCAo) as previously described (McColl *et al*, 2008). Briefly, under isoflurane anaesthesia, the carotid

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arteries were exposed and a 6-0 nylon monofilament (Dermalon) with a 2 mm tip (180 μ m diameter) coated in thermo-melting glue (Jet Melt) was introduced into the external carotid artery and advanced 9 mm along the internal carotid artery until occluding the origin of the MCA. After 40 mins, the filament was withdrawn to establish reperfusion and the wound sutured.

In a separate experiment, lean and obese *ob/ob* mice were administered an intraperitoneal injection of vehicle (0.9% sterile saline) or leptin (4 mg/kg in saline; Peprotech, London, UK) at the onset of MCAo. This dose of leptin has been shown previously to reduce ischaemic injury (Zhang *et al*, 2007).

Tissue Processing

Twenty-four hours after MCAo mice were perfused transcardially with 0.9% saline followed by 4% paraformaldehyde and brains post-fixed, cryoprotected, and frozen. Coronal brain sections (20 μ m) were prepared on a cryostat.

Measurement of HT and Ischaemic Damage

To assess HT brain sections were stained with haematoxylin and eosin. Areas of HT were delineated at coronal levels (400 μ m apart) and the total area of HT was calculated. For ischaemic damage, adjacent brain sections were stained with cresyl violet. Briefly, areas of damage were delineated at eight anatomically defined coronal levels and the total volume was calculated.

Immunohistochemistry

Primary antibodies were used as follows: goat anti-MMP-9 (1:400; R&D Systems, Abington, UK), and rabbit anti-laminin (1:25; Sigma, Poole, UK). Endogenous peroxidase activity (except for double immunofluorescence) and nonspecific binding sites were blocked before incubating (4°C) sections in primary antibody solution. For peroxidase-based staining, sections were then incubated in biotinylated secondary antibody (1:200; Vector Laboratories, Peterborough, UK), immersed in avidin-biotin-peroxidase complex (ABC; Vector Laboratories) and colour-developed using a 0.05% diaminobenzidine solution (in 0.01% H₂O₂). For assessment of BBB disruption, primary antibody was omitted and a biotinylated anti-mouse IgG secondary antibody was used. For double labelling immunofluorescence, after primary antibody application sections were incubated in donkey anti-goat Alexa 488 and donkey anti-rabbit 594 (Molecular Probes, Paisley, UK), mounted, and coverslipped with ProLong Gold mounted medium (Invitrogen, Paisley, UK).

Statistical Analysis

For all analyses, data are represented as mean \pm s.d. Parametric data were analysed using Student's *t*-test for single comparisons, or one-way analysis of variance followed by a Tukey's test for multiple comparisons. The incidence of HT was compared using χ^2 -test.

Results

Increased Incidence of HT and Ischaemic Brain Damage in Obese Mice

Haemorrhagic transformation was observed in the ipsilateral ischaemic hemisphere of *ob/ob* mice. Multiple foci of bleeds were evident throughout the striatum and cortex, and in some cases in the corpus callosum and hippocampus (Figure 1A–1C). HT was not detected in the contralateral hemisphere of *ob/ob* mice, suggesting that HT was a consequence of the ischaemic challenge and did not occur spontaneously in the absence of ischaemia in *ob/ob* mice. The incidence of HT was significantly increased in *ob/ob* mice as compared with that in the lean controls (lean, $n = 1/6$ versus *ob/ob*, $n = 5/6$; $P < 0.05$, χ^2 -test of independence). The HT observed in the one lean mouse was restricted to a small striatal haemorrhage. The volume of ischaemic brain damage was significantly greater (120%) in *ob/ob* as compared with that in lean mice 24 h after MCAo ($P < 0.05$; Figure 1D and 1E). There were two mortalities in the *ob/ob* group, and post mortem examination of their brains revealed extensive brain damage and HT.

Increased HT and Ischaemic Brain Damage in Obese Mice are Independent of Leptin Deficiency

Neuroprotective actions of leptin have been demonstrated previously (Zhang *et al*, 2007); therefore, in a separate experiment, we assessed the effect of acute leptin replacement on ischaemic brain damage in lean and obese mice. Leptin administration did not affect the incidence of HT, as haemorrhages were present in the striatum and cortex of all *ob/ob* mice (*ob/ob* vehicle, $n = 4/4$ versus *ob/ob* leptin, $n = 4/4$) 24 h after MCAo. The total area of HT in the ischaemic tissue of *ob/ob* mice was also unaffected by leptin treatment (*ob/ob* vehicle, 0.35 ± 0.15 mm² versus *ob/ob* leptin, 0.56 ± 0.09 mm²; $P > 0.05$). There was no HT observed in the brains of lean mice treated with either vehicle or leptin (lean vehicle, $n = 0/6$ versus lean leptin, $n = 0/5$). Leptin administration did not affect the exacerbation of ischaemic damage observed in obese mice (Figure 1F). One mortality after MCAo was noted in both obese groups (*ob/ob* vehicle and *ob/ob* leptin) and a post-mortem examination of the brains of these mice revealed the presence of HT and extensive ischaemic damage.

To verify that the absence of an effect of leptin administration on brain pathology was not due to lack of biological activity, we assessed the effect of leptin on food intake and body weight. Leptin (4 mg/kg, intraperitoneal.) significantly reduced food intake (vehicle, 4.0 ± 0.2 g versus leptin, 3.2 ± 0.3 g, $P < 0.05$; $n = 5-6$) and caused body weight loss (Δ body weight; vehicle, 0.2 ± 0.1 g versus leptin, -0.4 ± 0.1 g, $P < 0.01$; $n = 5-6$) 24 h after injection in lean mice.

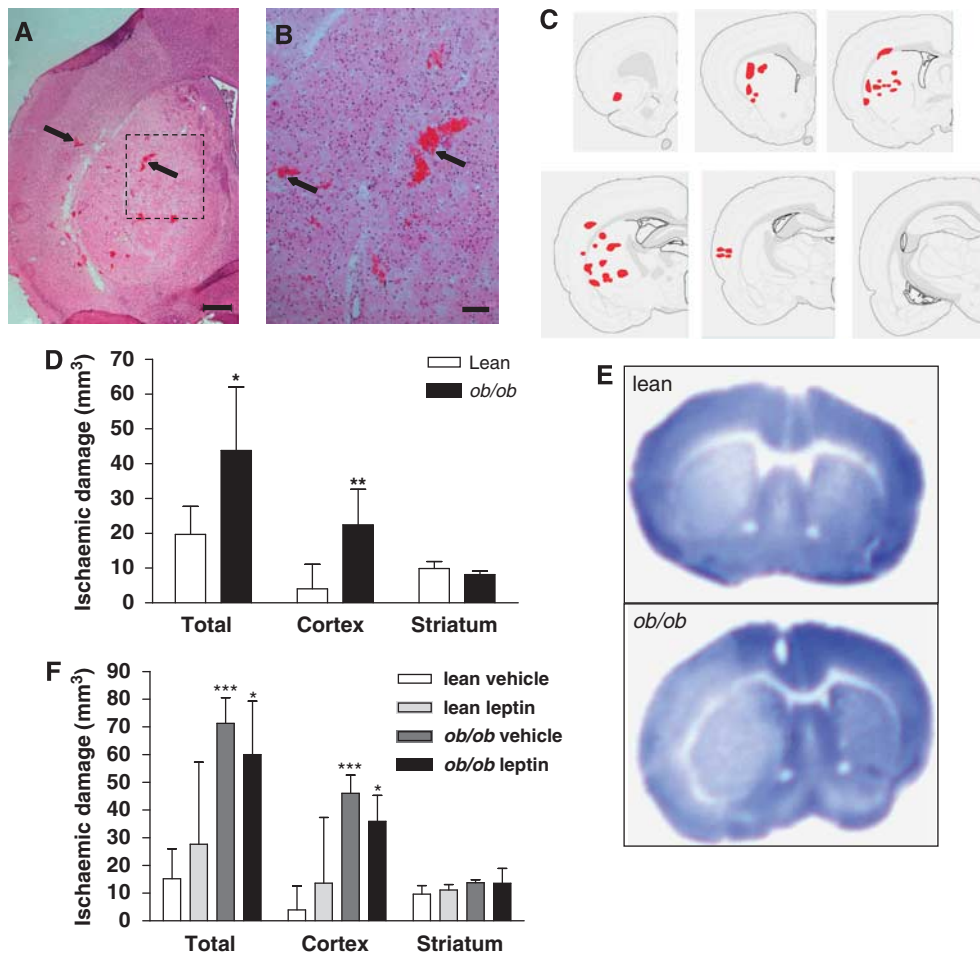


Figure 1 HT and ischaemic damage are increased in obese mice 24 h after MCAo. **(A)** Coronal brain sections stained with haematoxylin and eosin illustrate discrete foci of haemorrhages (arrows) in the ipsilateral striatum and cortex of *ob/ob* mice. **(B)** A higher magnification view of the area outlined by the dashed box in panel **A**. **(C)** The typical distribution of HT at six coronal brain sections. **(D)** An increase in the volume of ischaemic brain damage was observed in obese *ob/ob* mice 24 h after MCAo. **(E)** Representative cresyl violet-stained brain sections illustrate the exacerbation of brain damage in *ob/ob* mice. **(F)** Increased ischaemic damage in the brains of obese mice is independent of leptin. Leptin (4 mg/kg, intraperitoneal) or vehicle was administered at the onset of MCAo to lean and obese *ob/ob* mice and volume of ischaemic injury was assessed at 24 h. Data are mean \pm s.d. for $n = 4-6$ mice per group. * $P < 0.05$, ** $P < 0.01$ and *** $P < 0.001$ versus lean mice. Scale bars, 200 μ m **(A)** and 25 μ m **(B)**.

BBB Permeability and Brain Microvascular MMP-9 Expression are Increased after Focal Cerebral Ischaemia in Obese Mice

The marked increase in susceptibility to ischaemia-induced HT observed in *ob/ob* mice suggests severe loss of BBB integrity and an increase in cerebrovascular permeability. BBB permeability was determined by assessing the accumulation of endogenous IgG (which is excluded by an intact BBB) in the brain by immunohistochemistry. There was minimal IgG immunoreactivity in the brain tissue of lean mice after MCAo (Figure 2A). Marked increases in distribution and intensity of IgG immunoreactivity were observed in the ipsilateral hemisphere of brain sections from *ob/ob* mice 24 h after MCAo. Intense IgG staining was present in both

the striatum and cortex, corresponding to areas of ischaemic damage.

Experimental and clinical evidence implicates proteolytic disruption of BBB substrates by proteases such as MMP-9 in cerebrovascular disruption. We, therefore, assessed the expression of MMP-9 in the brain of obese and lean mice after MCAo. Extensive MMP-9 immunoreactivity was detected in the ipsilateral ischaemic striatum and cortex of *ob/ob* mice (Figure 2B), but not in the contralateral hemisphere. No MMP-9-positive cells were seen in the ipsilateral striatum or cortex of lean mice. The majority of MMP-9 in *ob/ob* mice was localised to blood vessels, which was confirmed by strong colocalisation between MMP-9 and laminin, a major basement membrane component of blood vessels (Figure 2C).

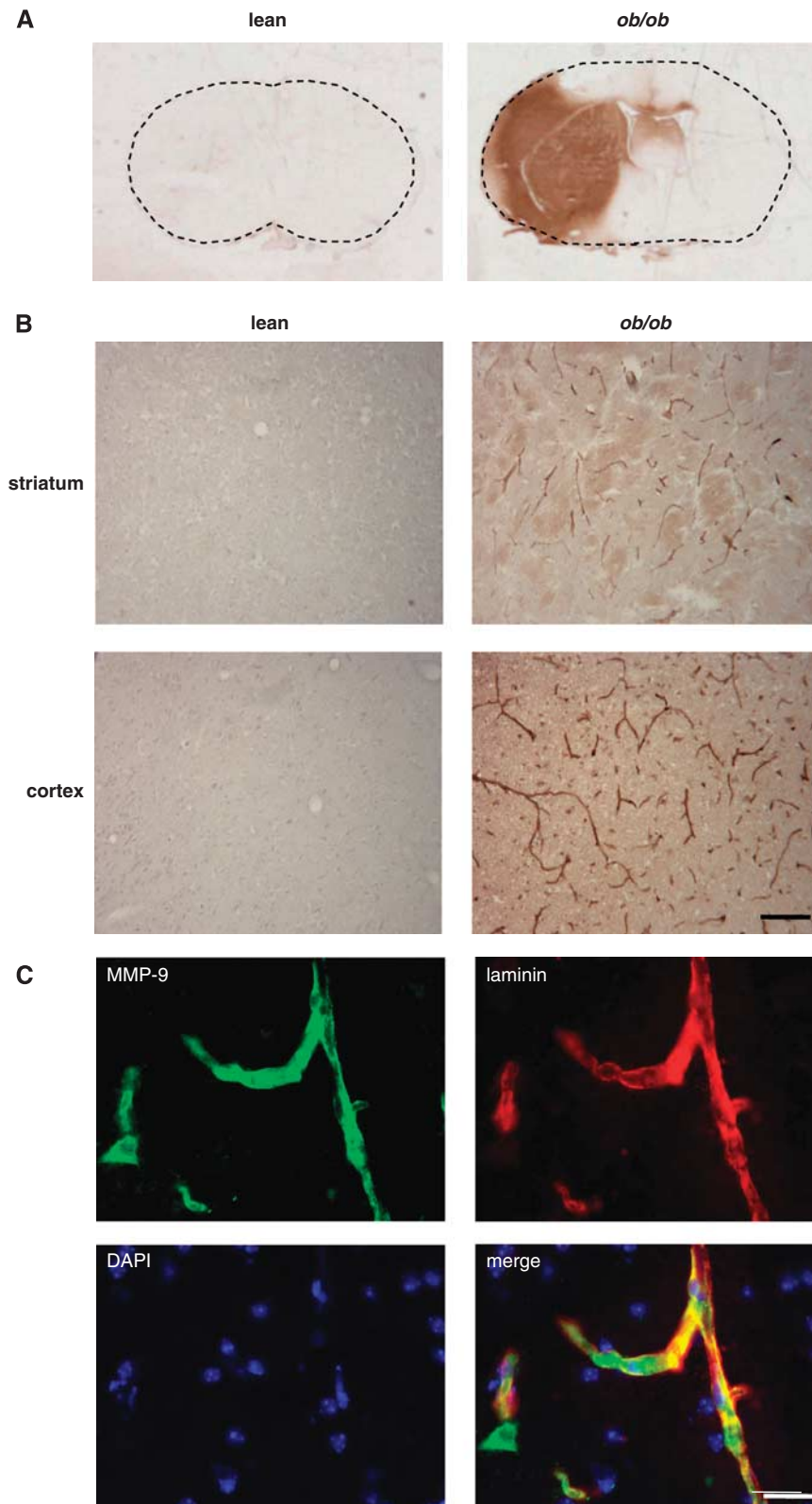


Figure 2 BBB permeability and MMP-9 expression are increased in obese mice after MCAo. BBB permeability was assessed by immunostaining for IgG. **(A)** Representative sections from lean and obese *ob/ob* mice 24 h after MCAo illustrate marked increased in the intensity and distribution of IgG immunoreactivity in *ob/ob* mice in the cortex and striatum indicating greater BBB disruption in obese mice. **(B)** Representative photomicrographs of MMP-9 expression in the brain of lean and obese *ob/ob* mice 24 h after MCAo. Increase in MMP-9 immunoreactivity was observed in the striatum and cortex of *ob/ob* mice. **(C)** Multi-labelling immunofluorescence shows colocalisation of laminin-positive blood vessels (red) and MMP-9 expression (green) in *ob/ob* mice indicating that the majority of MMP-9 is localised to cerebral blood vessels. Scale bars, 100 μm **(B)** and 25 μm **(C)**.

Discussion

In the present study, we show elevated microvascular MMP-9 expression and increased incidence of HT in obese, leptin-deficient *ob/ob* mice after experimental stroke. BBB permeability and infarct volume were also exacerbated in obese mice. Our data suggest that these effects are not due to hyperglycaemia or leptin deficiency, and, therefore, implicate obesity as an independent factor that promotes severe brain microvascular disruption after ischaemic brain injury.

In addition to clinical data that have indicated increased risk of stroke in obese individuals (Winter *et al*, 2008), recent studies have shown that the severity of brain damage is increased after experimental stroke in obese rodents (Mayanagi *et al*, 2008; Terao *et al*, 2008). Our data expand on these previous findings to show that obese mice are highly susceptible to HT, which is associated with elevated microvascular MMP-9 immunoreactivity. HT is a serious complication in ischaemic stroke patients that correlates with poor prognosis (Paciaroni *et al*, 2008). Conventional risk factors for HT include hyperglycaemia and hypertension (Paciaroni *et al*, 2008), both of which are common in obese subjects. However, these factors are unlikely to account for the increased frequency of HT in the present study, because pre-ischaemic blood glucose levels were similar in lean and obese mice, and previous studies have not found significant differences in blood pressure (Vachharajani *et al*, 2005). Thrombolytic treatment also predisposes to HT (Zhao *et al*, 2007). Our data suggest that obese patients may be at further risk of HT if treated with thrombolytic agents.

The extravasation of all blood constituents, including erythrocytes, that occurs during HT is indicative of a catastrophic failure of microvascular integrity. This failure also underlies the increased permeability of the BBB and brain oedema that accompanies HT. In the present study, we found marked increase in BBB permeability to IgG in obese mice, which is consistent with their increased susceptibility to HT and confirms that obesity promotes severe disruption to the BBB. We also observed localisation of extensive MMP-9 immunoreactivity to the microvasculature in obese mice, suggesting that MMP-9 may be an important mediator underlying increased BBB disruption and HT in obese mice. Quantitative assays of MMP-9 activity will be required to verify changes on a functional level. Previous studies have shown that MMP-9 can disrupt multiple components of the BBB, including the tight-junction proteins claudin-5 and occludin, and the basement membrane protein collagen-IV (Candelario-Jalil *et al*, 2009). Furthermore, inhibition of MMPs attenuates BBB disruption and reduces thrombolysis-induced HT after experimental stroke (Candelario-Jalil *et al*, 2009).

A chronically elevated inflammatory profile is a feature of obesity and inflammation is implicated in

microvascular dysfunction associated with obesity (Singer and Granger, 2007). Increased leukocyte–endothelium interactions and intracellular adhesion molecule-1 (ICAM-1) expression after experimental stroke in obese mice have recently been reported (Mayanagi *et al*, 2008; Terao *et al*, 2008). Leukocytes, in particular neutrophils, contain abundant proteases, including MMP-9; therefore, increased leukocyte adhesion could be a mechanism underlying the increased microvascular MMP-9 expression in the present study. In support of this, brain neutrophil accumulation was significantly greater after MCAo in obese mice (data not shown). More generally, growing evidence suggests that systemic inflammatory conditions, such as obesity, are important modifiers of stroke outcome (McColl *et al*, 2009).

One important caveat of the present study is that we cannot exclude that the effects of obesity are mediated through alterations in cerebral perfusion (e.g., through collateral vessels) and, indeed, given the pro-coagulatory state that is associated with obesity, it will be important to consider this potential mechanism in future studies. In summary, we have shown that obese *ob/ob* mice are highly susceptible to HT after experimental stroke, an effect that is associated with increased microvascular MMP-9 expression and BBB opening. Further work is ongoing to explore the mechanisms in more detail.

Acknowledgements

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Conflict of interest

The authors declare no conflict of interest.

References

- Candelario-Jalil E, Yang Y, Rosenberg GA (2009) Diverse roles of matrix metalloproteinases and tissue inhibitors of metalloproteinases in neuroinflammation and cerebral ischemia. *Neuroscience* 158:983–94
- Dandona P, Aljada A, Bandyopadhyay A (2004) Inflammation: the link between insulin resistance, obesity and diabetes. *Trends Immunol* 25:4–7
- Mayanagi K, Katakam PV, Gaspar T, Domoki F, Busija DW (2008) Acute treatment with rosuvastatin protects insulin resistant (C57BL/6J *ob/ob*) mice against transient cerebral ischemia. *J Cereb Blood Flow Metab* 28:1927–35
- McColl BW, Allan SM, Rothwell NJ (2009) Systemic infection, inflammation and acute ischemic stroke. *Neuroscience* 158:1049–61
- McColl BW, Rothwell NJ, Allan SM (2008) Systemic inflammation alters the kinetics of cerebrovascular tight junction disruption after experimental stroke in mice. *J Neurosci* 28:9451–62

- Paciaroni M, Agnelli G, Corea F, Ageno W, Alberti A, Lanari A, Caso V, Micheli S, Bertolani L, Venti M, Palmerini F, Biagini S, Comi G, Previdi P, Silvestrelli G (2008) Early hemorrhagic transformation of brain infarction: rate, predictive factors, and influence on clinical outcome: results of a prospective multicenter study. *Stroke* 39:2249–56
- Razinia T, Saver JL, Liebeskind DS, Ali LK, Buck B, Ovbiagele B (2007) Body mass index and hospital discharge outcomes after ischemic stroke. *Arch Neurol* 64:388–91
- Schreibelt G, Kooij G, Reijkerker A, van Doorn R, Gringhuis SI, van der Pol S, Weksler BB, Romero IA, Couraud PO, Piontek J, Blasig IE, Dijkstra CD, Ronken E, de Vries HE (2007) Reactive oxygen species alter brain endothelial tight junction dynamics via RhoA, PI3 kinase, and PKB signaling. *FASEB J* 21:3666–76
- Simard JM, Kent TA, Chen M, Tarasov KV, Gerzanich V (2007) Brain oedema in focal ischaemia: molecular pathophysiology and theoretical implications. *Lancet Neurol* 6:258–68
- Singer G, Granger DN (2007) Inflammatory responses underlying the microvascular dysfunction associated with obesity and insulin resistance. *Microcirculation* 14:375–87
- Terao S, Yilmaz G, Stokes KY, Ishikawa M, Kawase T, Granger DN (2008) Inflammatory and injury responses to ischemic stroke in obese mice. *Stroke* 39:943–50
- Vachharajani V, Russell JM, Scott KL, Conrad S, Stokes KY, Tallam L, Hall J, Granger DN (2005) Obesity exacerbates sepsis-induced inflammation and microvascular dysfunction in mouse brain. *Microcirculation* 12:183–94
- Winter Y, Rohrmann S, Linseisen J, Lanczik O, Ringleb PA, Hebebrand J, Back T (2008) Contribution of obesity and abdominal fat mass to risk of stroke and transient ischemic attacks. *Stroke* 39:3145–51
- Zhang F, Wang S, Signore AP, Chen J (2007) Neuroprotective effects of leptin against ischemic injury induced by oxygen-glucose deprivation and transient cerebral ischemia. *Stroke* 38:2329–36
- Zhao BQ, Tejima E, Lo EH (2007) Neurovascular proteases in brain injury, hemorrhage and remodeling after stroke. *Stroke* 38:748–52