

## Complete Genome Sequence of *Methanothermobacter marburgensis*, a Methanoarchaeon Model Organism<sup>∇</sup>

Heiko Liesegang,<sup>1</sup> Anne-Kristin Kaster,<sup>2</sup> Arnim Wiezer,<sup>1</sup> Meike Goenrich,<sup>2</sup> Antje Wollherr,<sup>1</sup> Henning Seedorf,<sup>2</sup> Gerhard Gottschalk,<sup>1</sup> and Rudolf K. Thauer<sup>2\*</sup>

Göttingen Genomics Laboratory, Institute of Microbiology and Genetics, Georg August University, 37077 Göttingen, Germany,<sup>1</sup> and Max Planck Institute for Terrestrial Microbiology, 35043 Marburg, Germany<sup>2</sup>

Received 19 July 2010/Accepted 13 August 2010

**The circular genome sequence of the chemolithoautotrophic euryarchaeon *Methanothermobacter marburgensis*, with 1,639,135 bp, was determined and compared with that of *Methanothermobacter thermautotrophicus*. The genomes of the two model methanogens differ substantially in protein coding sequences, in insertion sequence (IS)-like elements, and in clustered regularly interspaced short palindromic repeats (CRISPR) loci.**

*Methanothermobacter marburgensis* (DSM 2133) (formerly *Methanobacterium thermoautotrophicum* strain Marburg), a member of the *Methanobacteriales* (2), was isolated in 1978 from anaerobic sewage sludge in Marburg, Germany (5). The hydrogenotrophic methanogen grows even faster (2 h versus 3 h doubling time) and to higher cell concentrations (3 g versus 1.5 g dry mass per liter) than *Methanothermobacter thermautotrophicus* (DSM 1053) (formerly *Methanobacterium thermoautotrophicum* strain ΔH) (20) (for other differences, see references 3 and 19). Both methanogens were used in the last 35 years for the elucidation of the enzymes and coenzymes involved in CO<sub>2</sub> reduction to methane with H<sub>2</sub> (4, 16–18). The genome sequence of *M. thermautotrophicus* was reported in 1997 (15); that of *M. marburgensis* is announced here.

The genome size of *M. marburgensis* is 1,639,135 bp (that of *M. thermautotrophicus* is 1,751,377 bp), the genome G+C content is 48.64% (49.54% for *M. thermautotrophicus*), and the part coding is 90.94% (91.02% for *M. thermautotrophicus*). Comparison of the sequences (13) revealed that the two genomes have 1,607 protein coding sequences (CDS) in common and 411 CDS not in common (145 CDS are found only in *M. marburgensis* and 266 CDS only in *M. thermautotrophicus*) and show a high degree of synteny. The CDS not in common could be traced back to gene splitting (15%), gene deletion (30%), gene duplication (30%), and lateral gene transfer (24%) events (percentages given are for *M. marburgensis*). Of the 1,607 CDS in common, approximately 40% show BLAST search expectation values of  $>10^{-100}$  at the protein level, reflecting large differences in sequence divergence. Almost 470 CDS encode conserved hypothetical proteins.

The genome of *M. marburgensis* harbors 15 insertion sequence (IS)-like elements, whereas there is no evidence for a classically organized IS-like element in *M. thermautotrophicus*. Consistently, a CDS for a transposase is found only in *M. marburgensis*.

In the genome of *M. marburgensis* there is only one clustered regularly interspaced short palindromic repeat (CRISPR) locus with 36 repeats and only one CRISPR-associated (*cas*) gene (*csa3*), indicating that the organism is not protected from invasion by phage and plasmid DNA (7, 8, 10, 12). By comparison, in the genome of *M. thermautotrophicus* there are three CRISPR loci with 124, 4, and 47 repeats and 18 *cas* genes that encode proteins involved in adaptation and interference (<http://genoweb1.irisa.fr/Serveur-GPO/outils/repeatsAnalysis/CRISPR/>). The spacer sequences from locus 2 match DNA sequences found in phage ΨM1 of *M. marburgensis* (6, 11) and ΨM100 of *M. wolfei* (9), which supports the observation that *M. thermautotrophicus* is not lysed by those two phages. Unfortunately, there is no DNA sequence available for phage ΦF1, which is able to lyse *M. thermautotrophicus* (14), to compare it with the spacer sequences of the CRISPR regions. In the plasmid pM2001 (= pMTBMA4) (4,439-bp circular multicopy plasmid found only in *M. marburgensis*) (1, 19), no sequence identities for CRISPR spacer sequences of *M. thermautotrophicus* were found (14).

Approximately 200 CDS were identified that are required for the synthesis of the enzymes, coenzymes, and prosthetic groups involved in CO<sub>2</sub> reduction to methane and in the coupling of this process with energy conservation. Some of the genes have been found only recently; others, such as those for coenzyme F<sub>430</sub> biosynthesis, still remain to be discovered.

**Nucleotide sequence accession number.** The complete genome sequence of *M. marburgensis* was deposited in GenBank under accession numbers CP001710 (chromosome) and CP001711 (pMTBMA4).

This work was supported by the Max Planck Society, by the Fonds der Chemischen Industrie, and by a grant from the Niedersächsische Ministerium für Wissenschaft und Kultur.

### REFERENCES

1. Bokranz, M., A. Klein, and L. Meile. 1990. Complete nucleotide sequence of plasmid pME2001 of *Methanobacterium thermoautotrophicum* (Marburg). *Nucleic Acids Res.* **18**:363.
2. Boone, D. R., W. B. Whitman, and P. Rouvière. 1993. Diversity and taxonomy of methanogens, p. 35–80. *In* J. G. Ferry (ed.), *Methanogenesis*. Chapman & Hall, New York, NY.
3. Brandis, A., R. K. Thauer, and K. O. Stetter. 1981. Relatedness of strains DH and Marburg of *Methanobacterium thermoautotrophicum*. *Zentralbl. Bakt. Hyg. I Abt. Orig. C* **2**:311–317.

\* Corresponding author. Mailing address: Max Planck Institute for Terrestrial Microbiology, Karl-von-Frisch-Strasse 10, 35043 Marburg, Germany. Phone: 49 6421 178 101. Fax: 49 6421 178 109. E-mail: thauer@mpi-marburg.mpg.de.

† H. L. and A.-K. K. contributed equally to this work.

∇ Published ahead of print on 27 August 2010.

4. Dimarco, A. A., T. A. Bobik, and R. S. Wolfe. 1990. Unusual coenzymes of methanogenesis. *Annu. Rev. Biochem.* **59**:355–394.
5. Fuchs, G., E. Stupperich, and R. K. Thauer. 1978. Acetate assimilation and the synthesis of alanine, aspartate and glutamate in *Methanobacterium thermoautotrophicum*. *Arch. Microbiol.* **117**:61–66.
6. Jordan, M., L. Meile, and T. Leisinger. 1989. Organization of *Methanobacterium thermoautotrophicum* bacteriophage psi M1 DNA. *Mol. Gen. Genet.* **220**:161–164.
7. Karginov, F. V., and G. J. Hannon. 2010. The CRISPR system: small RNA-guided defense in bacteria and archaea. *Mol. Cell* **37**:7–19.
8. Lillestol, R. K., P. Redder, R. A. Garrett, and K. Brugger. 2006. A putative viral defence mechanism in archaeal cells. *Archaea* **2**:59–72.
9. Luo, Y., P. Pfister, T. Leisinger, and A. Wasserfallen. 2001. The genome of archaeal prophage psiM100 encodes the lytic enzyme responsible for autolysis of *Methanothermobacter wolfeii*. *J. Bacteriol.* **183**:5788–5792.
10. Marraffini, L. A., and E. J. Sontheimer. 2010. Self versus non-self discrimination during CRISPR RNA-directed immunity. *Nature* **463**:568–571.
11. Meile, L., U. Jenal, D. Studer, M. Jordan, and T. Leisinger. 1989. Characterization of psiM1, a virulent phage of *Methanobacterium thermoautotrophicum* Marburg. *Arch. Microbiol.* **152**:105–110.
12. Mojica, F. J., C. Diez-Villasenor, J. Garcia-Martinez, and E. Soria. 2005. Intervening sequences of regularly spaced prokaryotic repeats derive from foreign genetic elements. *J. Mol. Evol.* **60**:174–182.
13. Needleman, S. B., and C. D. Wunsch. 1970. A general method applicable to the search for similarities in the amino acid sequence of two proteins. *J. Mol. Biol.* **48**:443–453.
14. Nölling, J., A. Groffen, and W. M. Devos. 1993. phiF1 and phiF3, 2 novel virulent, archaeal phages infecting different thermophilic strains of the genus *Methanobacterium*. *J. Gen. Microbiol.* **139**:2511–2516.
15. Smith, D. R., L. A. Doucette-Stamm, C. Deloughery, H. Lee, J. Dubois, T. Aldredge, R. Bashirzadeh, D. Blakely, R. Cook, K. Gilbert, D. Harrison, L. Hoang, P. Keagle, W. Lumm, B. Pothier, D. Qiu, R. Spadafora, R. Vicaire, Y. Wang, J. Wierzbowski, R. Gibson, N. Jiwani, A. Caruso, D. Bush, H. Safer, D. Patwell, S. Prabhakar, S. McDougall, G. Shimer, A. Goyal, S. Pietrovski, G. M. Church, C. J. Daniels, J. Mao, P. Rice, J. Nölling, and J. N. Reeve. 1997. Complete genome sequence of *Methanobacterium thermoautotrophicum* ΔH: functional analysis and comparative genomics. *J. Bacteriol.* **179**:7135–7155.
16. Thauer, R. K. 1998. Biochemistry of methanogenesis: a tribute to Marjory Stephenson. *Microbiology* **144**:2377–2406.
17. Thauer, R. K., A. K. Kaster, M. Goenrich, M. Schick, T. Hiromoto, and S. Shima. 2010. Hydrogenases from methanogenic archaea, nickel, a novel cofactor, and H<sub>2</sub> storage. *Annu. Rev. Biochem.* **79**:507–536.
18. Thauer, R. K., A. K. Kaster, H. Seedorf, W. Buckel, and R. Hedderich. 2008. Methanogenic archaea: ecologically relevant differences in energy conservation. *Nat. Rev. Microbiol.* **6**:579–591.
19. Wasserfallen, A., J. Nölling, P. Pfister, J. Reeve, and E. Conway de Macario. 2000. Phylogenetic analysis of 18 thermophilic *Methanobacterium* isolates supports the proposals to create a new genus, *Methanothermobacter* gen. nov., and to reclassify several isolates in three species, *Methanothermobacter thermoautotrophicus* comb. nov., *Methanothermobacter wolfeii* comb. nov., and *Methanothermobacter marburgensis* sp. nov. *Int. J. Syst. Evol. Microbiol.* **1**:43–53.
20. Zeikus, J. G., and R. S. Wolfe. 1972. *Methanobacterium thermoautotrophicus* sp. n., an anaerobic, autotrophic, extreme thermophile. *J. Bacteriol.* **109**:707–713.