



Published in final edited form as:

Gastroenterol Clin North Am. 2010 September ; 39(3): 697–707. doi:10.1016/j.gtc.2010.08.014.

Peroxisome proliferator-activated receptors in chronic inflammation and colorectal cancer

Dingzhi Wang¹ and Raymond N. DuBois^{2,*}

¹Department of Cancer Biology, The University of Texas MD Anderson Cancer Center, Houston, TX 77030-4009

²Gastrointestinal Medical Oncology, The University of Texas MD Anderson Cancer Center, Houston, TX 77030-4009

Abstract

Peroxisome proliferator-activated receptors (PPARs) are members of the nuclear hormone receptor superfamily and have been implicated in a variety of physiological and pathological processes, such as nutrient metabolism, energy homeostasis, inflammation and cancer. In this review, we highlight breakthroughs in our understanding of the potential roles of PPARs in inflammatory bowel disease and colorectal cancer. These PPAR receptors might hold the key to some of the questions pertinent to the pathophysiology of inflammatory diseases and colorectal cancer and could possibly serve as drug targets for new anti-inflammatory therapeutic and anti-cancer agents.

Keywords

peroxisome proliferator-activated receptor; chronic inflammation; inflammatory bowel disease; colorectal cancer

INTRODUCTION

The recognition of chronic inflammation caused by infections or autoimmune diseases as the seventh trait of cancer has highlighted the contribution of inflamed stroma to tumor initiation, growth and metastasis. Epidemiologic studies indicate that chronic inflammation is clearly associated with increased cancer risk in a number of instances, including esophageal, gastric, hepatic, pancreatic and colorectal cancer. For example, it has been long known that patients with persistent hepatitis B infection, *Helicobacter pylori* infection, or an immune disorder such as inflammatory bowel disease (IBD), have a higher risk for the development of liver or gastrointestinal tract cancer. It has been estimated that chronic inflammation contributes to the development of approximately 15% of malignancies worldwide (1). The best evidence for the link between inflammation and tumor progression comes from recent epidemiologic studies

© 2010 Elsevier Inc. All rights reserved.

*Correspondence to: Raymond N. DuBois, M.D., Ph. D. The University of Texas MD Anderson Cancer Center Unit 118; 1515 Holcombe Blvd. Houston, TX 77030-4009 Phone: 713-745-4495 FAX: 713-745-1812 rdubois@mdanderson.org.

Dingzhi Wang, Ph. D. The University of Texas MD Anderson Cancer Center Unit 1014; S7.8316A, 6767 Bertner Ave. Houston, TX 77030 Phone: 713-729-6379 FAX: 713-729-6375 dwang11@mdanderson.org

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Financial disclosures and conflicts of interest: The authors have nothing to disclose.

and clinical trials showing that long-term use of non-steroidal anti-inflammatory drugs (NSAIDs) reduced the relative risk of developing colorectal cancer (CRC) by 40-50% (2).

The gastrointestinal mucosa forms a complex, semi-permeable barrier between the host and the largest source of foreign antigens. The mucosal barrier consists of epithelial cell junctions and the underlying stromal elements including immune cells. An abnormal mucosal immune response to bacteria, which make up the intestinal flora, is thought to result in chronic inflammation and the development of inflammatory bowel disease (IBD). IBD, with its two clinical manifestations of Crohn's Disease (CD) and Ulcerative Colitis (UC), is a chronic inflammatory disorder of the gastrointestinal tract. Chronic IBD (especially pan-colitis) significantly increases the risk of developing CRC (3). In support of this notion, the observation that 5-aminosalicylic acid (5-ASA), currently used in the treatment of UC, suppresses the development of colitis-associated cancer in an animal model (4).

A large body of evidence indicates that genetic mutations, epigenetic changes, chronic inflammation, diet and lifestyle are risk factors for cancer (5-7). Similar to other solid tumors, colorectal cancer (CRC) is a heterogeneous disease with at least three major forms: hereditary, sporadic, and colitis-associated CRC. Patients with familial adenomatous polyposis (FAP), due to a germ-line mutation in one allele of the tumor suppressor gene adenomatous polyposis coli (*APC*), have a near 100% risk of developing CRC by the age of 40, if untreated. Somatic loss of *APC* function occurs in about 85% of sporadic colorectal adenomas and carcinomas (8-10). Hereditary nonpolyposis colorectal cancer (HNPCC), which is due to inherited mutations in genes for DNA mismatch repair such as *MLH1*, *MSH2*, and *MSH6*, is responsible for approximately 2 to 7 percent of all diagnosed cases of CRC. The average age of patients with this syndrome develop cancer around 44 years old, as compared to 64 years old in the general population. Together with the hereditary syndromes of FAP and hereditary nonpolyposis CRC, IBD is among the top three high-risk conditions for CRC; therefore, patients with IBD face an increased lifetime risk for developing CRC. Compared with sporadic CRC, colitis-associated CRC affects individuals at a younger age than the general population.

Peroxisome proliferator-activated receptors (PPARs), which were initially identified as mediators of the peroxisome proliferators in the early 1990s (11), belong to the nuclear hormone receptor superfamily and are also ligand-dependent transcription factors. PPARs play a central role in regulating the storage and catabolism of dietary fats via complex metabolic pathways, including fatty acid oxidation and lipogenesis (12). To date, three mammalian PPARs have been identified and are referred to as PPAR α (NR1C1), PPAR δ/β (NR1C2) and PPAR γ (NR1C3). Each PPAR isotype displays a tissue-selective expression pattern. PPAR α and PPAR γ are predominantly present in the liver and adipose tissue, respectively, while PPAR δ is expressed in diverse tissues (13) and its expression in the gastrointestinal tract is very high compared with other tissues (14). As ligand-dependent transcription factors, transcriptional activation by PPARs depends on ligand binding and the interaction of co-regulators. PPAR ligands are chemically unrelated molecules including a variety of fatty acids, fatty acid derivatives, and steroids, as well as synthetic compounds. Polyunsaturated fatty acids activate all three PPAR isotypes with relative low affinity (15). The endogenous fatty-acid derivatives, which are mainly converted by cyclooxygenase and lipoxygenase enzymes, selectively bind and activate each PPAR isotype. For example, 15-deoxy- Δ^{12},Δ^{14} PGJ₂ (15dPGJ₂), a dehydration product of PGD₂, is a natural ligand for the PPAR γ (16,17), while PGI₂ can transactivate PPAR δ (18,19).

It is well established that modulation of PPAR activity maintains cellular and whole-body glucose and lipid homeostasis. Hence, great efforts have been made to develop drugs targeting these receptors. For example, PPAR γ synthetic agonists, troglitazone, rosiglitazone and pioglitazone, are clinically used for the therapy of non-insulin-dependent diabetes mellitus.

The anti-atherosclerotic and hypolipidemic agents including fenofibrate and gemfibrozil are PPAR α synthetic agonists that induce hepatic lipid uptake and catabolism. Genetic and pharmacological studies have also revealed that PPAR δ agonists are potential drugs for use in the treatment of dyslipidemias, obesity and insulin resistance (20-23). Therefore, the PPAR δ agonist (GW501516) is currently in phase III clinical trials to evaluate its use for treatment of patients with hyperlipidemias and obesity. In addition to modulation of lipid homeostasis and energy balance, PPARs have emerged as essential molecules in the pathogenesis of IBD and CRC.

PPARS AND IBD

The currently available therapies for IBD include 5-aminosalicylic acid (5-ASA), corticosteroids, antibiotics, immune modulators and immunosuppressive agents such as azathioprine, 6-mercaptopurine, and cyclosporine. Corticosteroids and immunosuppressive agents are associated with significant risks of unwanted side effects and not all patients respond to these medications. For 5-ASA agents, these medications are generally safe but only induce remission in approximately 50% of patients with UC (24). It is, therefore, essential to develop newer therapeutic interventions for patients with IBD. A growing body of evidence indicates that PPAR α and PPAR γ have an anti-inflammatory effect on IBD and its agonists might serve as a new class of effective therapy for IBD. The role of PPAR δ in IBD remains ambiguous. This deserves significant attention and future research must be directed to better understand the role of PPARs in regulating chronic inflammation in IBD.

PPAR α

PPAR α is highly expressed in mouse colonic epithelial cells facing the intestinal lumen (25) and its expression induced by glucocorticoids (GC) (26). Subsequent studies further demonstrated that PPAR α mediates anti-inflammatory effects of GC in a mouse model of chemically-induced colitis (27). In this study, treatment with dexamethasone, a potent synthetic member of the glucocorticoid class of steroid drugs, suppressed dinitrobenzene sulfuric acid (DNBS)-induced colitis formation in wild-type mice, but not in PPAR α knockout mice. Consistent with the above results, deletion of PPAR α promoted more severity of colitis in DNBS-treated mice, whereas activation of PPAR α by its agonist activity significantly reduced colonic inflammation in this mouse model (28). However, there is no report thus far, on the precise role of PPAR α in genetic models of IBD (transgenic and knock-out models).

PPAR γ

Although PPAR γ is predominantly present in the liver and adipose tissue, it is also expressed in the intestinal epithelium, immune cells and adipocytes. However, patients with UC, but not CD, show decreased PPAR γ levels in colon epithelial cells in comparison to normal controls (29). This observation raises the hypothesis that microbe-host interactions, chronic inflammation and/or genetic predisposition may lead to low PPAR γ levels in colonic epithelial cells, which in turn may result in unrestrained inflammation. Several lines of evidence support the notion that PPAR γ may serve as a new therapeutic target in IBD. In mouse models of chemically-induced colitis, 5-ASA treatment had a beneficial effect on colitis only in wild-type but not in heterozygous PPAR γ ^{+/-} mice, demonstrating that PPAR γ mediates the anti-inflammatory effect of 5-ASA (30). Furthermore, treatment of a PPAR γ ligand, thiazolidinedione, markedly reduced colonic inflammation in mouse models of chemically-induced colitis (31,32) and IL-10 deficient mice (a genetic model of colitis) (33), suggesting that activation of PPAR γ suppresses inflammation in IBD.

Since PPAR γ is expressed in intestinal epithelial cells, macrophage, and T and B lymphocytes, it is critical to understand the contribution of PPAR γ in each cell type to this protection. The

results from two studies showed that the disruption of PPAR γ in colonic epithelial cells worsened colonic inflammatory lesions in DSS-treated mice, indicating that PPAR γ expression in epithelial cells is required for the prevention of experimental IBD (34,35). Similarly, mice with deficiency of PPAR γ in CD4 T cells are more sensitive to trinitrobenzene sulfonic acid-induced colitis, because the deficiency of PPAR γ in Treg cells impaired their ability to prevent effector CD4 T cell-induced colitis (36). Moreover, mice with a targeted disruption of PPAR γ in macrophages displayed an increased susceptibility to DSS-induced colitis compared with wild-type littermates, demonstrating that PPAR γ is required for macrophage-mediated protection against colitis (37). Consistent with these results, an increase in PPAR γ expression by adenovirus-mediated gene transfer attenuated colonic inflammation induced by DSS in mice (38). In addition, a recent study showed that the anti-inflammatory effects of PPAR γ on IBD is via maintenance of innate antimicrobial immunity in the mouse colon (39). Importantly, the studies from one randomized placebo-controlled trial and one open-label trial showed that a PPAR γ agonist, rosiglitazone, has therapeutic efficacy in humans with UC (40,41). Collectively, all of these studies support a rationale to develop PPAR γ agonists as potential therapeutic and prophylactic agents against IBD.

PPAR δ

Relatively little is known about the role of PPAR δ in IBD and the results from two mouse models of IBD are controversial. Deletion of PPAR δ significantly exacerbated colitis, whereas treatment of a PPAR δ agonist didn't affect the clinical symptoms in the DSS-treated mouse model (42). This study implies that PPAR δ , like PPAR γ , exerts anti-inflammatory effects in IBD via a ligand independent mechanism. In contrast with this observation, administration of a PPAR δ agonist caused enhanced colitis in IL-10-deficient mice (a genetic model of colitis), suggesting that PPAR δ has a pro-inflammatory effect (43). Therefore, further studies are necessary to clarify the biological functions of PPAR δ in the modulation of IBD.

PPARS AND COLORECTAL CANCER

In addition to these metabolic and inflammatory properties, the roles of PPARs in CRC progression have been extensively investigated. PPARs can function as either tumor suppressors or accelerators, suggesting that these receptors are potential candidates as drug targets for cancer prevention and treatment.

PPAR α

Less is known about the role of PPAR α in human cancers although long-term administration of a PPAR α agonist induces the development of hepatocarcinomas in mice but not in PPAR α null animals, conclusively demonstrating that PPAR α mediates these effects in promoting liver cancer (44). In spite of the fact that activation of PPAR α by exogenous agonists generally causes inhibition of tumor cell growth in cell lines derived from CRC, melanoma, and glial brain tumors (45-47), the physiological significance of PPAR α in the regulation of CRC progression is also less well characterized than that of PPAR γ and PPAR δ ,

PPAR γ

Due to elevated expression of PPAR γ in CRC (48) and its involvement in regulating cellular differentiation, PPAR γ has become a point of interest in CRC studies. However, studies of PPAR γ mutation in human colon tumor samples and CRC cell lines have produced controversial results. One study showed that 8% of primary human colorectal cancers had a loss of function mutation in one allele of the PPAR γ gene (49). Recent data revealed that a Pro12Ala (P12A) polymorphism in the PPAR γ gene is associated with an increased risk of CRC (50,51). These results suggest a putative role for this receptor as a tumor suppressor. In

contrast, another study showed that the mutant PPAR γ gene was not detected in human colon carcinoma samples or CRC cell lines, suggesting that PPAR γ mutations in human CRC may be a rare event (52).

It is well established that activation of PPAR γ results in growth arrest of colon carcinoma cells through induction of cell-cycle arrest or/and apoptosis in numerous *in vitro* studies. However, the effect of PPAR γ on CRC progression *in vivo* is controversial due to conflicting results from different mouse models of colon cancer. Although PPAR γ agonists inhibit colorectal carcinogenesis in xenograft models and in the azoxymethane (AOM)-induced colon cancer model (53,54), these drugs are reported to have either tumor-promoting or tumor-inhibiting effects in a mouse model of FAP, the *Apc*^{Min/+} mouse. Multiple studies showed that administration of PPAR γ agonists significantly increased the number of colon adenomas in the *Apc*^{Min/+} mice (55-57) and even in wild-type C57BL/6 mice (58). However, other studies showed that treatment of 2 different *Apc* mutant models (*Apc*^{Min/+} and *Apc*^{Δ1309}) with a PPAR γ agonist pioglitazone resulted in a reduction of polyp number in both small and large intestines in a dose-dependent manner (59,60). These divergent effects of PPAR γ might be related to drug doses and bioavailability and/or the animal models employed. These paradoxical observations appear to have been resolved by genetic studies, showing that the heterozygous disruption of PPAR γ is sufficient to increase tumor number(s) in AOM-treated mice and that intestinal-specific PPAR γ knockout promotes tumor growth in *Apc*^{Min/+} mice (61,62). Thus, genetic evidence supports the hypothesis that PPAR γ serves as a tumor suppressor in CRC. In addition, a combined treatment of mice with a selective COX-2 inhibitor and a PPAR γ agonist significantly inhibited both the incidence and multiplicity of inflammation-associated colonic adenocarcinoma induced by AOM/DSS (63). Interestingly, a retrospective cohort study revealed that treatment of diabetic patients with a PPAR γ agonist (thiazolidinediones) exhibited a mild trend toward a risk reduction of CRC, although this difference did not reach statistical significance (64). Collectively, these findings further support a rationale to develop PPAR γ agonists as anti-tumor agents.

PPAR δ

The role of PPAR δ in colorectal carcinogenesis is more controversial than that of PPAR γ . The first evidence linking PPAR δ to carcinogenesis actually emerged from studies on gastrointestinal cancer. PPAR δ was identified as a direct transcriptional target of APC/b-catenin/Tcf pathway and as a repression target of NSAIDs (65,66). A large case-control study showed that the protective effect of NSAIDs against colorectal adenomas was reported to be modulated by a polymorphism in the *PPAR*TM gene (67). Moreover, COX-2-derived PGI₂ directly transactivates PPAR δ (68) and COX-2-derived PGE₂ indirectly induces PPAR δ activation in CRC, hepatocellular carcinoma, and cholangiocarcinoma cells (69-71). In addition, PPAR δ expression and activity are also induced by oncogenic K-Ras (72). These studies indicate that PPAR δ is a focal point of cross-talk between oncogenic signaling pathways.

Similar to PPAR γ , investigation of PPAR δ expression in human and mouse colon tumor samples and CRC cell lines generated controversial results. Some reports showed that PPAR δ is elevated in most human colorectal cancers and in tumors arising in the *Apc*^{Min/+} mice and AOM-treated rats (65,68), in agreement with the observations that activation of the b-catenin/Tcf pathway by *APC* mutation or K-Ras upregulates PPAR δ expression. Importantly, the PPAR δ proteins are accumulated only in human CRC cells with highly malignant morphology (73). Downregulation of PPAR δ is correlated with anti-tumor effects of dietary fish oil/pectin in rats treated with radiation and AOM (74). However, other reports showed that PPAR δ expression is lower in human cancer tissues and adenomas from the *Apc*^{Min/+} mice than normal control tissues (75,76).

In a murine xenograft cancer model, the disruption of both PPAR δ alleles by deletion of its exons 4-6 in human HCT-116 colon carcinoma cells decreased tumorigenicity, suggesting that activation of PPAR δ promotes tumor growth (77). To further determine whether PPAR δ attenuates or promotes intestinal tumor growth, three mouse models of CRC were employed, including AOM-treated mice, *Apc*^{Min/+} mice and *Mlh*-null mice. The *Mlh* is a DNA mismatch repair gene that is involved in hereditary non-polyposis CRC. Conflicting data was obtained from studies in AOM-treated and *Apc*^{Min/+} mice. For example, activation of PPAR δ by a selective synthetic PPAR δ agonist (GW501516) or a PPAR δ endogenous activator (PGE₂) accelerated intestinal adenoma growth in *Apc*^{Min/+} mice by promoting tumor cell survival (69,78). In contrast, another PPAR δ ligand (GW0742) inhibited colon carcinogenesis in AOM-treated mice, but promoted small intestinal polyp growth in *Apc*^{Min/+} mice (79). It is not clear whether PPAR δ mediates the effects of GW0742 in this model. A genetic study showed that loss of PPAR δ by deletion of its exons 4-5 attenuated both small and large intestinal adenoma growth and demonstrated that PPAR δ mediated the tumor-promoting effects of the PPAR δ ligand (GW501516) and PGE₂ in *Apc*^{Min/+} mice (69,80). A recent study with a tissue-specific deletion of PPAR δ exon 4, inhibited colonic carcinogenesis in AOM-treated mice (81), further confirmed the notion that PPAR δ serves as a tumor accelerator. On the other hand, several other studies have shown different results when using PPAR δ mutant mice generated by germline deletion of PPAR δ exon 8. Deletion of PPAR δ exon 8 enhances polyp growth in *Apc*^{Min/+} and AOM-treated mice, in the absence of exogenous PPAR δ stimulation (76,82). In *Mlh*-null mice, no significant differences are evident in the number and size of intestinal adenomas between wild-type and PPAR δ mutant mice (deletion of PPAR δ exon 8) (76). The conflicting results regarding the effect of PPAR δ on intestinal tumorigenesis in *Apc*^{Min/+} and AOM-treated mice may have been attributed to differences in the specific targeting strategy employed to delete PPAR δ . Deletion of PPAR δ exon 4 and/or 5, which encode an essential portion of the DNA binding domain, is thought to disrupt PPAR δ function as a nuclear transcriptional factor and to inhibit tumorigenesis. The deletion of exon 8, the last PPAR δ exon, is postulated to generate a hypomorphic allele, which retains some aporeceptor function. Indeed, the observation that the high rates of embryonic mortality, subsequent to abnormal trophoblastic giant cell differentiation and abnormal placental development occurred in deletion of PPAR δ exon 4-5, but not in deletion of PPAR δ exon 8 mice supports this hypothesis (83,84). Taken together, not enough evidence is available to establish whether PPAR δ has pro- or anti-tumorigenic effects on CRC progression and the role of PPAR δ in cancer biology remains unclear.

SUMMARY

Emerging evidence indicates that PPAR γ suppresses inflammation in IBD and tumor growth in CRC. In contrast to PPAR γ , conflicting results have emerged regarding the role of PPAR δ in IBD and colon carcinogenesis. Therefore, further investigation is warranted prior to considering modulation of PPAR δ as an effective therapy for chemoprevention and treatment of IBD and CRC.

Acknowledgments

This work is supported, in part, from the National Institutes of Health Grants RO1DK 62112, P01-CA-77839, R37-DK47297, and P30 CA068485 (RND). RND (R37-DK47297) is the recipient of a NIH MERIT award. We also thank the National Colorectal Cancer Research Alliance (NCCRA) for their generous support (RND).

REFERENCES

1. Kuper H, Adami HO, Trichopoulos D. Infections as a major preventable cause of human cancer. *J Intern Med*;2000 248(3):171–83. [PubMed: 10971784]

2. Smalley WE, DuBois RN. Colorectal cancer and nonsteroidal anti-inflammatory drugs. *Adv Pharmacol* 1997;39:1–20. [PubMed: 9160111]
3. Lewis JD, Deren JJ, Lichtenstein GR. Cancer risk in patients with inflammatory bowel disease. *Gastroenterol Clin North Am Jun*;1999 28(2):459–77. x. [PubMed: 10372277]
4. Ikeda I, Tomimoto A, Wada K, et al. 5-aminosalicylic acid given in the remission stage of colitis suppresses colitis-associated cancer in a mouse colitis model. *Clin Cancer Res Nov 1*;2007 13(21):6527–31. [PubMed: 17975166]
5. Vogelstein B, Kinzler KW. Cancer genes and the pathways they control. *Nat Med Aug*;2004 10(8):789–99. [PubMed: 15286780]
6. Ting AH, McGarvey KM, Baylin SB. The cancer epigenome--components and functional correlates. *Genes Dev Dec 1*;2006 20(23):3215–31. [PubMed: 17158741]
7. Woutersen RA, Appel MJ, van Garderen-Hoetmer A, Wijnands MV. Dietary fat and carcinogenesis. *Mutat Res Jul 15*;1999 443(1-2):111–27. [PubMed: 10415435]
8. Powell SM, Zilz N, Beazer-Barclay Y, et al. APC mutations occur early during colorectal tumorigenesis. *Nature Sep 17*;1992 359(6392):235–7. [PubMed: 1528264]
9. Jen J, Powell SM, Papadopoulos N, et al. Molecular determinants of dysplasia in colorectal lesions. *Cancer Res Nov 1*;1994 54(21):5523–6. [PubMed: 7923189]
10. Smith AJ, Stern HS, Penner M, et al. Somatic APC and K-ras codon 12 mutations in aberrant crypt foci from human colons. *Cancer Res Nov 1*;1994 54(21):5527–30. [PubMed: 7923190]
11. Issemann I, Green S. Activation of a member of the steroid hormone receptor superfamily by peroxisome proliferators. *Nature 1990*;347(6294):645–50. [PubMed: 2129546]
12. Berger JP, Akiyama TE, Meinke PT. PPARs: therapeutic targets for metabolic disease. *Trends Pharmacol Sci May*;2005 26(5):244–51. [PubMed: 15860371]
13. Michalik L, Desvergne B, Wahli W. Peroxisome-proliferator-activated receptors and cancers: complex stories. *Nat Rev Cancer Jan*;2004 4(1):61–70. [PubMed: 14708026]
14. Higashiyama H, Billin AN, Okamoto Y, Kinoshita M, Asano S. Expression profiling of peroxisome proliferator-activated receptor-delta (PPAR-delta) in mouse tissues using tissue microarray. *Histochem Cell Biol May*;2007 127(5):485–94. [PubMed: 17333240]
15. Kersten S, Wahli W. Peroxisome proliferator activated receptor agonists. *Exs 2000*;89:141–51. [PubMed: 10997287]
16. Forman BM, Tontonoz P, Chen J, Brun RP, Spiegelman BM, Evans RM. 15-Deoxy-delta 12, 14-prostaglandin J2 is a ligand for the adipocyte determination factor PPAR gamma. *Cell 1995*;83(5):803–12. [PubMed: 8521497]
17. Kliewer SA, Lenhard JM, Willson TM, Patel I, Morris DC, Lehmann JM. A prostaglandin J2 metabolite binds peroxisome proliferator-activated receptor gamma and promotes adipocyte differentiation. *Cell 1995*;83(5):813–9. [PubMed: 8521498]
18. Gupta RA, Tan J, Krause WF, et al. Prostacyclin-mediated activation of peroxisome proliferator-activated receptor delta in colorectal cancer. *Proc Natl Acad Sci U S A 2000*;97(24):13275–80. [PubMed: 11087869]
19. Forman BM, Chen J, Evans RM. Hypolipidemic drugs, polyunsaturated fatty acids, and eicosanoids are ligands for peroxisome proliferator-activated receptors alpha and delta. *Proc Natl Acad Sci U S A 1997*;94(9):4312–7. [PubMed: 9113986]
20. Wang YX, Lee CH, Tiep S, et al. Peroxisome-proliferator-activated receptor delta activates fat metabolism to prevent obesity. *Cell Apr 18*;2003 113(2):159–70. [PubMed: 12705865]
21. Tanaka T, Yamamoto J, Iwasaki S, et al. Activation of peroxisome proliferator-activated receptor delta induces fatty acid beta-oxidation in skeletal muscle and attenuates metabolic syndrome. *Proc Natl Acad Sci U S A Dec 23*;2003 100(26):15924–9. [PubMed: 14676330]
22. Oliver WR Jr, Shenk JL, Snaith MR, et al. A selective peroxisome proliferator-activated receptor delta agonist promotes reverse cholesterol transport. *Proc Natl Acad Sci U S A Apr 24*;2001 98(9):5306–11. [PubMed: 11309497]
23. Evans RM, Barish GD, Wang YX. PPARs and the complex journey to obesity. *Nat Med Apr*;2004 10(4):355–61. [PubMed: 15057233]

24. Sutherland LR, May GR, Shaffer EA. Sulfasalazine revisited: a meta-analysis of 5-aminosalicylic acid in the treatment of ulcerative colitis. *Ann Intern Med* Apr 1;1993 118(7):540–9. [PubMed: 8095128]
25. Mansen A, Guardiola-Diaz H, Rafter J, Branting C, Gustafsson JA. Expression of the peroxisome proliferator-activated receptor (PPAR) in the mouse colonic mucosa. *Biochem Biophys Res Commun* May 24;1996 222(3):844–51. [PubMed: 8651933]
26. Bernal-Mizrachi C, Xiaozhong L, Yin L, et al. An afferent vagal nerve pathway links hepatic PPARalpha activation to glucocorticoid-induced insulin resistance and hypertension. *Cell Metab* Feb;2007 5(2):91–102. [PubMed: 17276352]
27. Riccardi L, Mazzon E, Bruscoli S, et al. Peroxisome proliferator-activated receptor-alpha modulates the anti-inflammatory effect of glucocorticoids in a model of inflammatory bowel disease in mice. *Shock* Mar;2009 31(3):308–16. [PubMed: 18665053]
28. Cuzzocrea S, Di Paola R, Mazzon E, et al. Role of endogenous and exogenous ligands for the peroxisome proliferator-activated receptors alpha (PPAR-alpha) in the development of inflammatory bowel disease in mice. *Lab Invest* Dec;2004 84(12):1643–54. [PubMed: 15492755]
29. Dubuquoy L, Jansson EA, Deeb S, et al. Impaired expression of peroxisome proliferator-activated receptor gamma in ulcerative colitis. *Gastroenterology* May;2003 124(5):1265–76. [PubMed: 12730867]
30. Rousseaux C, Lefebvre B, Dubuquoy L, et al. Intestinal antiinflammatory effect of 5-aminosalicylic acid is dependent on peroxisome proliferator-activated receptor-gamma. *J Exp Med* Apr 18;2005 201(8):1205–15. [PubMed: 15824083]
31. Su CG, Wen X, Bailey ST, et al. A novel therapy for colitis utilizing PPAR-gamma ligands to inhibit the epithelial inflammatory response. *J Clin Invest* Aug;1999 104(4):383–9. [PubMed: 10449430]
32. Dubuquoy L, Bourdon C, Peuchmaur M, et al. [Peroxisome proliferator-activated receptor (PPAR) gamma: a new target for the treatment of inflammatory bowel disease]. *Gastroenterol Clin Biol* Aug-Sep;2000 24(8-9):719–24. [PubMed: 11011247]
33. Lytle C, Tod TJ, Vo KT, Lee JW, Atkinson RD, Straus DS. The peroxisome proliferator-activated receptor gamma ligand rosiglitazone delays the onset of inflammatory bowel disease in mice with interleukin 10 deficiency. *Inflamm Bowel Dis* Mar;2005 11(3):231–43. [PubMed: 15735429]
34. Adachi M, Kurotani R, Morimura K, et al. Peroxisome proliferator activated receptor gamma in colonic epithelial cells protects against experimental inflammatory bowel disease. *Gut* Aug;2006 55(8):1104–13. [PubMed: 16547072]
35. Mohapatra SK, Guri AJ, Climent M, et al. Immunoregulatory actions of epithelial cell PPAR gamma at the colonic mucosa of mice with experimental inflammatory bowel disease. *PLoS One* 2010;5(4):e10215. [PubMed: 20422041]
36. Hontecillas R, Bassaganya-Riera J. Peroxisome proliferator-activated receptor gamma is required for regulatory CD4+ T cell-mediated protection against colitis. *J Immunol* Mar 1;2007 178(5):2940–9. [PubMed: 17312139]
37. Shah YM, Morimura K, Gonzalez FJ. Expression of peroxisome proliferator-activated receptor-gamma in macrophage suppresses experimentally induced colitis. *Am J Physiol Gastrointest Liver Physiol* Feb;2007 292(2):G657–66. [PubMed: 17095756]
38. Katayama K, Wada K, Nakajima A, et al. A novel PPAR gamma gene therapy to control inflammation associated with inflammatory bowel disease in a murine model. *Gastroenterology* May;2003 124(5):1315–24. [PubMed: 12730872]
39. Peyrin-Biroulet L, Beisner J, Wang G, et al. Peroxisome proliferator-activated receptor gamma activation is required for maintenance of innate antimicrobial immunity in the colon. *Proc Natl Acad Sci U S A*. Apr 26;2010
40. Lewis JD, Lichtenstein GR, Deren JJ, et al. Rosiglitazone for active ulcerative colitis: a randomized placebo-controlled trial. *Gastroenterology* Mar;2008 134(3):688–95. [PubMed: 18325386]
41. Lewis JD, Lichtenstein GR, Stein RB, et al. An open-label trial of the PPAR-gamma ligand rosiglitazone for active ulcerative colitis. *Am J Gastroenterol* Dec;2001 96(12):3323–8. [PubMed: 11774944]

42. Hollingshead HE, Morimura K, Adachi M, et al. PPARbeta/delta protects against experimental colitis through a ligand-independent mechanism. *Dig Dis Sci Nov*;2007 52(11):2912–9. [PubMed: 17404849]
43. Lee JW, Bajwa PJ, Carson MJ, et al. Fenofibrate represses interleukin-17 and interferon-gamma expression and improves colitis in interleukin-10-deficient mice. *Gastroenterology Jul*;2007 133(1): 108–23. [PubMed: 17631136]
44. Peters JM, Cattley RC, Gonzalez FJ. Role of PPAR alpha in the mechanism of action of the nongenotoxic carcinogen and peroxisome proliferator Wy-14,643. *Carcinogenesis Nov*;1997 18(11): 2029–33. [PubMed: 9395198]
45. Grabacka M, Plonka PM, Urbanska K, Reiss K. Peroxisome proliferator-activated receptor alpha activation decreases metastatic potential of melanoma cells in vitro via downregulation of Akt. *Clin Cancer Res May 15*;2006 12(10):3028–36. [PubMed: 16707598]
46. Strakova N, Ehrmann J, Bartos J, Malikova J, Dolezel J, Kolar Z. Peroxisome proliferator-activated receptors (PPAR) agonists affect cell viability, apoptosis and expression of cell cycle related proteins in cell lines of glial brain tumors. *Neoplasma 2005*;52(2):126–36. [PubMed: 15800711]
47. Grau R, Punzon C, Fresno M, Iniguez MA. Peroxisome-proliferator-activated receptor alpha agonists inhibit cyclo-oxygenase 2 and vascular endothelial growth factor transcriptional activation in human colorectal carcinoma cells via inhibition of activator protein-1. *Biochem J Apr 1*;2006 395(1):81–8. [PubMed: 16343055]
48. DuBois RN, Gupta R, Brockman J, Reddy BS, Krakow SL, Lazar MA. The nuclear eicosanoid receptor, PPARgamma, is aberrantly expressed in colonic cancers. *Carcinogenesis Jan*;1998 19(1): 49–53. [PubMed: 9472692]
49. Sarraf P, Mueller E, Smith WM, et al. Loss-of-function mutations in PPAR gamma associated with human colon cancer. *Mol Cell Jun*;1999 3(6):799–804. [PubMed: 10394368]
50. Slattery ML, Curtin K, Wolff R, et al. PPARgamma and colon and rectal cancer: associations with specific tumor mutations, aspirin, ibuprofen and insulin-related genes (United States). *Cancer Causes Control Apr*;2006 17(3):239–49. [PubMed: 16489531]
51. Landi S, Moreno V, Gioia-Patricola L, et al. Association of common polymorphisms in inflammatory genes interleukin (IL)6, IL8, tumor necrosis factor alpha, NFkB1, and peroxisome proliferator-activated receptor gamma with colorectal cancer. *Cancer Res Jul 1*;2003 63(13):3560–6. [PubMed: 12839942]
52. Ikezoe T, Miller CW, Kawano S, et al. Mutational analysis of the peroxisome proliferator-activated receptor gamma gene in human malignancies. *Cancer Res Jul 1*;2001 61(13):5307–10. [PubMed: 11431375]
53. Sarraf P, Mueller E, Jones D, et al. Differentiation and reversal of malignant changes in colon cancer through PPARgamma. *Nat Med Sep*;1998 4(9):1046–52. [PubMed: 9734398]
54. Osawa E, Nakajima A, Wada K, et al. Peroxisome proliferator-activated receptor gamma ligands suppress colon carcinogenesis induced by azoxymethane in mice. *Gastroenterology Feb*;2003 124 (2):361–7. [PubMed: 12557142]
55. Lefebvre AM, Chen I, Desreumaux P, et al. Activation of the peroxisome proliferator-activated receptor gamma promotes the development of colon tumors in C57BL/6J-APCMin/+ mice. *Nat Med Sep*;1998 4(9):1053–7. [PubMed: 9734399]
56. Saez E, Tontonoz P, Nelson MC, et al. Activators of the nuclear receptor PPARgamma enhance colon polyp formation. *Nat Med Sep*;1998 4(9):1058–61. [PubMed: 9734400]
57. Pino MV, Kelley MF, Jayyosi Z. Promotion of colon tumors in C57BL/6J-APC(min)/+ mice by thiazolidinedione PPARgamma agonists and a structurally unrelated PPARgamma agonist. *Toxicol Pathol Jan-Feb*;2004 32(1):58–63. [PubMed: 14713549]
58. Yang K, Fan KH, Lamprecht SA, et al. Peroxisome proliferator-activated receptor gamma agonist troglitazone induces colon tumors in normal C57BL/6J mice and enhances colonic carcinogenesis in Apc(1638 N/+) Mlh1(+/-) double mutant mice. *Int J Cancer Apr 7*;2005 116(4):495–9. [PubMed: 15818612]
59. Niho N, Takahashi M, Kitamura T, et al. Concomitant suppression of hyperlipidemia and intestinal polyp formation in Apc-deficient mice by peroxisome proliferator-activated receptor ligands. *Cancer Res Sep 15*;2003 63(18):6090–5. [PubMed: 14522940]

60. Niho N, Takahashi M, Shoji Y, et al. Dose-dependent suppression of hyperlipidemia and intestinal polyp formation in Min mice by pioglitazone, a PPAR gamma ligand. *Cancer Sci Nov*;2003 94(11): 960–4. [PubMed: 14611672]
61. Girmun GD, Smith WM, Drori S, et al. APC-dependent suppression of colon carcinogenesis by PPARgamma. *Proc Natl Acad Sci U S A Oct 15*;2002 99(21):13771–6. [PubMed: 12370429]
62. McAlpine CA, Barak Y, Matisse I, Cormier RT. Intestinal-specific PPARgamma deficiency enhances tumorigenesis in ApcMin/+ mice. *Int J Cancer Nov 15*;2006 119(10):2339–46. [PubMed: 16858678]
63. Kohno H, Suzuki R, Sugie S, Tanaka T. Suppression of colitis-related mouse colon carcinogenesis by a COX-2 inhibitor and PPAR ligands. *BMC Cancer 2005*;5:46. [PubMed: 15892897]
64. Govindarajan R, Ratnasingham L, Simmons DL, et al. Thiazolidinediones and the risk of lung, prostate, and colon cancer in patients with diabetes. *J Clin Oncol Apr 20*;2007 25(12):1476–81. [PubMed: 17442990]
65. He TC, Chan TA, Vogelstein B, Kinzler KW. PPARdelta is an APC-regulated target of nonsteroidal anti-inflammatory drugs. *Cell 1999*;99(3):335–45. [PubMed: 10555149]
66. Ouyang N, Williams JL, Rigas B. NO-donating aspirin isomers downregulate peroxisome proliferator-activated receptor (PPAR)delta expression in APC(min/+) mice proportionally to their tumor inhibitory effect: Implications for the role of PPARdelta in carcinogenesis. *Carcinogenesis Feb*;2006 27(2):232–9. [PubMed: 16141240]
67. Siezen CL, Tjihuis MJ, Kram NR, et al. Protective effect of nonsteroidal anti-inflammatory drugs on colorectal adenomas is modified by a polymorphism in peroxisome proliferator-activated receptor delta. *Pharmacogenet Genomics Jan*;2006 16(1):43–50. [PubMed: 16344721]
68. Gupta RA, Tan J, Krause WF, et al. Prostacyclin-mediated activation of peroxisome proliferator-activated receptor delta in colorectal cancer. *Proc Natl Acad Sci U S A Nov 21*;2000 97(24):13275–80. [PubMed: 11087869]
69. Wang D, Wang H, Shi Q, et al. Prostaglandin E(2) promotes colorectal adenoma growth via transactivation of the nuclear peroxisome proliferator-activated receptor delta. *Cancer Cell Sep*;2004 6(3):285–95. [PubMed: 15380519]
70. Xu L, Han C, Wu T. A novel positive feedback loop between peroxisome proliferator-activated receptor-delta and prostaglandin E2 signaling pathways for human cholangiocarcinoma cell growth. *J Biol Chem Nov 10*;2006 281(45):33982–96. [PubMed: 16966336]
71. Xu L, Han C, Lim K, Wu T. Cross-talk between peroxisome proliferator-activated receptor delta and cytosolic phospholipase A(2)alpha/cyclooxygenase-2/prostaglandin E(2) signaling pathways in human hepatocellular carcinoma cells. *Cancer Res Dec 15*;2006 66(24):11859–68. [PubMed: 17178883]
72. Shao J, Sheng H, DuBois RN. Peroxisome proliferator-activated receptors modulate K-Ras-mediated transformation of intestinal epithelial cells. *Cancer Res Jun 1*;2002 62(11):3282–8. [PubMed: 12036946]
73. Takayama O, Yamamoto H, Damdinsuren B, et al. Expression of PPARdelta in multistage carcinogenesis of the colorectum: implications of malignant cancer morphology. *Br J Cancer Oct 9*;2006 95(7):889–95. [PubMed: 16969348]
74. Vanamala J, Glagolenko A, Yang P, et al. Dietary fish oil and pectin enhance colonocyte apoptosis in part through suppression of PPAR{delta}/PGE2 and elevation of PGE3. *Carcinogenesis. Nov 16*;2007
75. Chen LC, Hao CY, Chiu YS, et al. Alteration of gene expression in normal-appearing colon mucosa of APC(min) mice and human cancer patients. *Cancer Res May 15*;2004 64(10):3694–700. [PubMed: 15150130]
76. Reed KR, Sansom OJ, Hayes AJ, et al. PPARdelta status and Apc-mediated tumourigenesis in the mouse intestine. *Oncogene Nov 25*;2004 23(55):8992–6. [PubMed: 15480419]
77. Park BH, Vogelstein B, Kinzler KW. Genetic disruption of PPARdelta decreases the tumorigenicity of human colon cancer cells. *Proc Natl Acad Sci U S A Feb 27*;2001 98(5):2598–603. [PubMed: 11226285]
78. Gupta RA, Wang D, Katkuri S, Wang H, Dey SK, DuBois RN. Activation of nuclear hormone receptor peroxisome proliferator-activated receptor-delta accelerates intestinal adenoma growth. *Nat Med Mar*;2004 10(3):245–7. [PubMed: 14758356]

79. Marin HE, Peraza MA, Billin AN, et al. Ligand activation of peroxisome proliferator-activated receptor beta inhibits colon carcinogenesis. *Cancer Res* Apr 15;2006 66(8):4394–401. [PubMed: 16618765]
80. Wang D, Wang H, Guo Y, et al. Crosstalk between peroxisome proliferator-activated receptor delta and VEGF stimulates cancer progression. *Proc Natl Acad Sci U S A* Dec 12;2006 103(50):19069–74. [PubMed: 17148604]
81. Zuo X, Peng Z, Moussalli MJ, et al. Targeted genetic disruption of peroxisome proliferator-activated receptor-delta and colonic tumorigenesis. *J Natl Cancer Inst* May 20;2009 101(10):762–7. [PubMed: 19436036]
82. Harman FS, Nicol CJ, Marin HE, Ward JM, Gonzalez FJ, Peters JM. Peroxisome proliferator-activated receptor-delta attenuates colon carcinogenesis. *Nat Med.* Mar 28;2004
83. Peters JM, Lee SS, Li W, et al. Growth, adipose, brain, and skin alterations resulting from targeted disruption of the mouse peroxisome proliferator-activated receptor beta(delta). *Mol Cell Biol* Jul; 2000 20(14):5119–28. [PubMed: 10866668]
84. Nadra K, Anghel SI, Joye E, et al. Differentiation of trophoblast giant cells and their metabolic functions are dependent on peroxisome proliferator-activated receptor beta/delta. *Mol Cell Biol* Apr; 2006 26(8):3266–81. [PubMed: 16581799]