A Cautionary Note as to the Use of pBi-L and Related Luciferase/Transgenic Vectors in the Study of Thyroid Endocrinology

Dear Editor:

The pBi-L plasmid (Clontech, Incorporated, Mountain View, CA) allows dual expression of a gene of interest together with a luciferase marker, and both genes in this vector can be regulated in response to doxycycline or tetracycline. As a result, pBi-L and related vectors are of considerable utility for the generation and study of transgenic animals, and have become increasingly popular for this purpose. With the goal of generating transgenic mice expressing defined thyroid hormone receptor alleles (TRs), we generated a pBi-L vector containing the wild-type TRa1 isoform. Our initial characterizations of this recombinant vector in cultured Hepa 1-6 mouse hepatoma cells confirmed, as anticipated, that both the luciferase gene and the TRa1 gene were expressed in the presence of a tTA transactivator, and that both genes were negatively regulated by doxycycline under these conditions (compare lane 7 with lane 5 in Fig. 1A and lane 3 with lane 1 in Fig. 1B). Unexpectedly expression of both $TR\alpha 1$ and luciferase was also strongly negatively regulated by thyroid hormone (T_3) (compare lane 6 with lane 5 in Fig. 1A) and lane 2 with lane 1 in Fig. 1B). The negative regulation by T_3 could be observed whether the TRa1 was introduced in *cis* in the pBi-L vector itself (Fig. 1A, lanes 5-8) or was instead expressed in trans from another vector construct (Fig. 1A, lanes 9-12), but was absent in cells lacking TRs (Fig. 1A, lanes 1-4). Similar results were observed when using the TR β 1 isoform (Fig. 1A, lanes 13–20) and in additional cell lines (e.g. human HepG2, Fig. 1B). Notably a fragment of the pBi-L vector created by restriction enzyme digestion and limited to the bidirectional O-Tet-CMV promoter and the luciferase coding region (tetO-Luc) also displayed negative T₃ regulation (Fig. 1C; data not shown). In contrast, pUHD10-3, a tet-regulated, but unidirectional expression vector lacking a luciferase marker, did not display this negative T₃ regulation (lanes 5-8 in Fig. 1B). Unlike positive T₃ response elements, there is no single agreed consensus sequence for a negative T₃ response element (e.g. Saatcioglu et al. [2]) and we have not yet mapped the negative response element to a specific sequence within the luciferase gene; we also note that although our data maps one negative T₃ response element (nTRE) in pBi-L to the luciferase gene, we cannot exclude the possibility that additional nTREs exist elsewhere in this vector.

The powerful T₃-mediated down-regulation of pBi-L transcription is likely to introduce significant distortions in the expression patterns of these constructs in different tissues and under different physiological states. We therefore recommend caution in the use of pBi-L and related vectors in studies of thyroid hormone endocrinology. A negative T₃ response element has also been identified in vectors possessing a thymidine-kinase promoter-driven luciferase construct, indicating that this may be a general concern when using luciferase as a reporter (3). However, luciferase-based vectors are widely used in cell transfections and many do not appear to display the negative T₃ response reported here. Presumably the specific sequence and topology of each vector construct can influence the negative T₃ response, indicating that a given vector must be individually evaluated experimentally for its suitability in a particular experiment.

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FIG. 1. Regulation of pBi-L by doxycycline and thyroid hormone (T₃). (**A**) From left to right, the pBi-L plasmid itself (pBi-L), the same vector with a TRα1 insert, the pBi-L plasmid together with a pSG5-TRα1 expression vector, the pBi-L vector with a TRβ1 insert, or the pBi-L plasmid together with a pSG5-TRβ1 expression vector were transiently transfected into Hepa 1-6 cells using the lipofection procedure previously described (1). Transfections also included a pCH110 β-galactosidase construct as an internal normalization control (1). Cells were maintained at 37°C in Dulbecco-modified Eagles medium containing 10% hormone-depleted fetal bovine serum. After 24 hours the transfection medium was replaced with medium containing doxycycline, T₃, or both, as indicated below each panel. After an additional 24 hours the cells were harvested and relative luciferase activity was calculated (1) and is presented. (**B**) The overall transfection protocol in panel A was repeated using the luciferase gene (pUHD10-3), and either Hepa 1-6 cells or HepG2 cells as indicated. After harvesting the cells, mRNA was isolated and quantified for levels of TRα1, luciferase, and GAPDH using a reverse transcription PCR protocol (4). (**C**) Alternatively, the pBi-L vector, a linearized version of the pBi-L vector (lin), or a fragment of the same construct limited to the luciferase assay as described in panel A.