

# Correction of respiratory disorders in a mouse model of Rett syndrome

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**Rett syndrome (RTT) is an autism spectrum disorder caused by mutations in the X-linked gene that encodes the transcription factor methyl-CpG-binding protein 2 (MeCP2). A major debilitating phenotype in affected females is frequent apneas, and heterozygous *Mecp2*-deficient female mice mimic the human respiratory disorder. GABA defects have been demonstrated in the brainstem of *Mecp2*-deficient mice. Here, using an intact respiratory network, we show that apnea in RTT mice is characterized by excessive excitatory activity in expiratory cranial and spinal nerves. Augmenting GABA markedly improves the respiratory phenotype. In addition, a serotonin 1a receptor agonist that depresses expiratory neuron activity also reduces apnea, corrects the irregular breathing pattern, and prolongs survival in *MeCP2* null males. Combining a GABA reuptake blocker with a serotonin 1a agonist in heterozygous females completely corrects their respiratory defects. The results indicate that GABA and serotonin 1a receptor activity are candidates for treatment of the respiratory disorders in Rett syndrome.**

apnea | GABA | respiration | serotonin 1a receptor

**R**ett syndrome (RTT) is an autism spectrum disorder that is caused by mutations in the X-linked gene that encodes methyl-CpG-binding protein 2 (*MeCP2*) (1). The role of this transcription factor is incompletely understood. Until recently, on the basis of animal studies primarily in embryonic or early neonatal brain, *Mecp2* was considered a translational repressor (2, 3). Recently, however, Skene et al. (4) found in neuronal nuclei of mature (6–8 wk) WT littermates of *Mecp2* null males that *Mecp2* expression is  $\approx$ fivefold greater than that at birth. The amount of *Mecp2* bound to DNA was proportional to the methylation density of CpG sequences. Importantly, in adult mice, it has been shown that restoring *Mecp2* reverses the abnormalities in mobility, gait, hind limb clasp, tremor, breathing, and general condition that they developed during the period that the transcription factor was deficient (5). Thus, *Mecp2* deficiency does not cause neuronal degeneration, nor is it necessary for the correct development of neuronal networks. Strategies aimed at pharmacological corrections of symptoms are therefore essential in treating RTT.

Respiratory disorders are prominent and one of the most disturbing features of RTT (6, 7). These abnormalities are faithfully mimicked in mouse models of RTT (8). In *Mecp2* null male animals studied in situ phrenic (PN) apneas were characterized by prolonged postinspiratory (post-I) activity in the central vagus nerve (9). Post-I activity is linked to the control of laryngeal adductors that control expiratory airflow and protect the lower airways from aspiration during swallowing. Active breath-hold with laryngeal closure is a common feature of RTT (6, 7, 10).

This raises the possibility that the apneas are due to overactive brainstem expiratory neurons in *Mecp2* heterozygote mice, perhaps consequent to a lack of synaptic inhibitory control. In this regard, examination of GABA synaptic inhibition in the ventrolateral medulla of *Mecp2* null male mice revealed that it was markedly reduced compared with WT (11). These findings

led us to hypothesize that insufficient synaptic inhibition of expiratory neurons underlies the respiratory disturbances in RTT.

Using a combination of in situ studies in which PN, hypoglossal (HN), central vagus (cVN), and abdominal (AbN) nerves were recorded simultaneously from adult *Mecp2*-deficient female mice and separately monitored pleural pressure in awake, freely moving animals, we have performed a detailed characterization of respiratory motor pattern and examined the effects of blocking GABA reuptake and of allosteric modulation of its type A receptor. Because serotonin 1a receptor agonists have been shown to both inhibit expiratory neurons (12) and reinstate breathing after opioid-induced central apneas (13, 14), we tested whether 8-hydroxy-dipropyl-aminotetralin (8-OH-DPAT) (a serotonin 1a receptor agonist) could depress the respiratory apneas in *Mecp2* heterozygote mice. Our findings reveal that both treatments significantly reduced apneas and periodic breathing in *Mecp2*-deficient mice and restored regularity to their cycle intervals. Combining GABA reuptake block with 8-OH-DPAT resulted in a respiratory pattern similar to that in WT.

## Results

**Characterization of Apnea in Heterozygous *Mecp2*-Deficient Female Mice (*Mecp2*<sup>-/+</sup>) Studied in Situ.** Studies were performed in WT and heterozygous *Mecp2*-deficient mice between 4 and 14 mo of age. Apneas were significantly more common in *Mecp2*<sup>-/+</sup> ( $152 \pm 18 \cdot \text{h}^{-1}$ ,  $n = 17$ ) compared with *Mecp2*<sup>+/+</sup> ( $50 \pm 10 \cdot \text{h}^{-1}$ ,  $n = 16$ ) ( $P \leq 0.0001$ , unpaired *t* test) (Fig. 1C). The incidence of apnea was not affected by age (Fig. S1). The arrest of PN activity was characterized by both a prolonged post-I phase of cVN restricted to the initial portion of the apnea and tonic discharge of HN throughout the apnea. AbN activity was obtained in seven *Mecp2*<sup>-/+</sup> mice. Tonic AbN associated with PN apnea was seen in some of the apneas in all seven, and in four animals it exceeded 10% (range, 12.3–47.9%) (Fig. 1B). The temporal relationship for the onset of tonic AbN relative to that of HN was random: 29.9%  $\pm$  3.2% concurrent, 26.5%  $\pm$  6.6% preceded HN, and 43.5%  $\pm$  4.9% followed HN. In a number of eupneic respiratory cycles HN was biphasic with a distinct early expiratory component (Fig. 1B, arrow, and D). The association of tonic HN with that in AbN, an expiratory nerve (15), and with postinspiration in cVN suggests that it is the expiratory component of HN that tonically discharges in PN apnea. The duration of apnea in

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perfusate) (Fig. S34.1). The irregularity score, however, was not changed (Fig. S34.2).

To determine the long-term effects of a serotonin 1a agonist, *Mecp2* null males were implanted with osmotic pumps to provide a slow release of 8-OH-DPAT. On the basis of animal weight at the time the pumps were placed, the dose of 8-OH-DPAT ranged between 43 and 52  $\mu\text{g}$  per kg per h. The median survival of treated mice (77 d) exceeded that of untreated animals (63.5 d) ( $P = 0.035$ , Kaplan–Meier log–rank test) (Fig. 4A.4). This represents a lower estimate of the serotonin 1a agonist's effect on survival because the drug was only present for 28 d.

**Effect of Combined GABA Reuptake Block and Serotonin 1a Agonist on Apnea and Irregularity in *Mecp2*-Deficient Mice.** Because neither augmenting GABA synaptic inhibition nor serotonin stimulation at 1a receptors reduced apnea to the incidence seen in WT animals, and because their postulated modes of action are different, we investigated the effects of combining these treatments. NO-711 (5.2  $\mu\text{M}$  in perfusate) followed 10 min later by the addition of 8-OH-DPAT (0.1  $\mu\text{M}$ ) reduced the occurrence of apnea in *Mecp2*<sup>-/+</sup> animals studied *in situ* to that of *Mecp2*<sup>+/+</sup> mice. In four of six *Mecp2*<sup>-/+</sup> animals apnea was eliminated with the combined treatment (Fig. 5A). In one mouse apnea was reduced from 164/h to 36/h. The sixth animal showed periodic breathing, and the serotonin agonist reduced this pattern from 56.3% to 17.1%. The combined treatment restored breath cycle regularity. In *Mecp2* null males the combination of NO-711 (1.0 mg/kg) and 8-OH-DPAT (150  $\mu\text{g}/\text{kg}$ ) reduced the incidence of apnea (Fig. 5C). The magnitude of reduced apnea (82.0%  $\pm$  5.5%) was larger than that seen with 300  $\mu\text{g}/\text{kg}$  8-OH-DPAT alone (71.3%  $\pm$  7.5%).

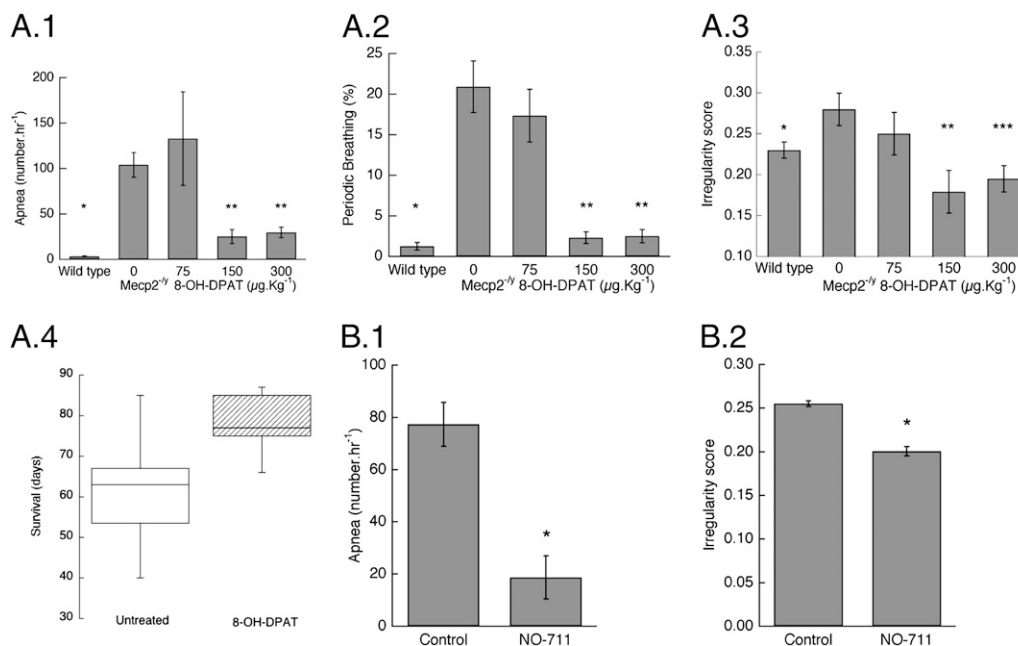
Locomotor activity was greater in heterozygous *Mecp2*-deficient females aged 6.4–7.8 mo after treatment with NO-711 (1.0 mg/kg) combined with 8-OH-DPAT (50  $\mu\text{g}/\text{kg}$ ); 3.4  $\pm$  0.4

$\text{min} \cdot 5 \text{ min}^{-1}$  than after vehicle injection, 1.9  $\pm$  0.4  $\text{min} \cdot 5 \text{ min}^{-1}$  ( $P = 0.002$ ) (two-way repeated-measures ANOVA with strain and treatment as the two factors). In addition, whereas in the control condition *Mecp2*<sup>-/+</sup> mice were less active than *Mecp2*<sup>+/+</sup>, after combined treatment there was no difference: *Mecp2*<sup>-/+</sup> 3.3  $\pm$  0.4, *Mecp2*<sup>+/+</sup> 3.5  $\pm$  0.3 ( $P = 0.68$ ) (Fig. S64). Vertical rearing movements were not significantly less in *Mecp2*<sup>-/+</sup> animals compared with *Mecp2*<sup>+/+</sup> ( $P$  values between 0.06 and 0.55) (Fig. S6B).

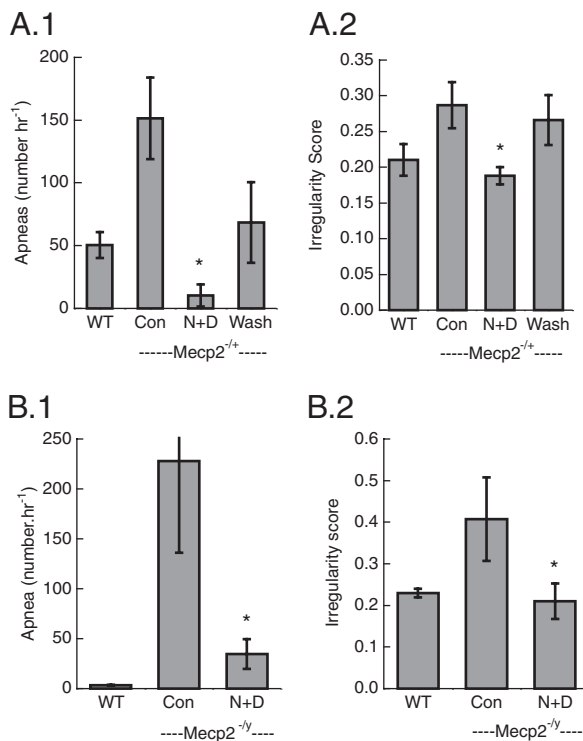
As with *Mecp2*-deficient females, locomotor activity was decreased in *Mecp2*<sup>-/-</sup> mice compared with WT. Combined treatment did not affect locomotor activity in either strain (Fig. S7A). It reduced vertical rearing activity in WT; however, a similar effect was seen with vehicle injections. Vertical rearing was not significantly affected in *Mecp2*<sup>-/-</sup> animals (Fig. S7B).

## Discussion

The present study confirms, in heterozygous female mice, previous observations in *Mecp2* null males that showed prolonged discharge in the post-I portion of cVN at the onset of PN apnea (9). Our characterization of respiratory disorders extended this earlier study by showing (*i*) all PN apneas are associated with large-amplitude tonic activity in HN that persists throughout the PN silence; (*ii*) a number of apneas are accompanied with tonic activity in AbN; (*iii*) augmenting GABA<sub>A</sub> inhibition significantly decreases the incidence of apnea and periodic breathing and corrects cycle irregularity; (*iv*) the serotonin 1a agonist 8-OH-DPAT reduces apnea and periodic breathing in a dose-dependent manner, induces regular breathing in *Mecp2* null males, and when given chronically extends their survival; and (*v*) the combination of blocking GABA reuptake and 8-OH-DPAT reduces apnea to the level seen in WT.



**Fig. 4.** Effect of blocking GABA reuptake and of the serotonin 1a receptor agonist 8-OH-DPAT on respiratory disorders in *Mecp2* null male mice. (A) Dose-dependent effect of 8-OH-DPAT on apnea, periodic breathing, and irregularity score. (A.1) Apnea.  $*P < 0.001$  vs. *Mecp2*<sup>-/-</sup> 0 mg/kg;  $**P = 0.046$  and  $0.02$  vs. *Mecp2*<sup>-/-</sup> 0 mg/kg ( $n = 9$  for WT, 10 for *Mecp2*<sup>-/-</sup> 0 and 300  $\mu\text{g}/\text{kg}$ , and 4 for *Mecp2*<sup>-/-</sup> 75 and 150  $\mu\text{g}/\text{kg}$ ) (two-way repeated-measures ANOVA with strain and treatment as the two factors). (A.2) Periodic breathing.  $*P \leq 0.001$  vs. *Mecp2*<sup>-/-</sup> 0 mg/kg;  $**P = 0.003$  and  $<0.001$  vs. *Mecp2*<sup>-/-</sup> 0 mg/kg. (A.3) Irregularity score.  $*P = 0.001$  vs. *Mecp2*<sup>-/-</sup> 0 mg/kg;  $**P = 0.048$  vs. *Mecp2*<sup>-/-</sup> 0 mg/kg;  $***P = 0.009$  vs. *Mecp2*<sup>-/-</sup> 0 mg/kg. (A.4) Box plot for untreated ( $n = 22$ ) and *Mecp2* null males treated with 8-OH-DPAT osmotic pump.  $P = 0.035$  ( $n = 9$ ) (Kaplan–Meier log–rank test). (B) Effect of NO-711 (5 mg/kg *i.p.*) on apnea and irregularity score. (B.1) Apnea.  $*P = 0.001$  ( $n = 4$ ) (paired *t* test). (B.2) Irregularity score.  $*P = 0.004$ .



**Fig. 5.** Effect of combined GABA reuptake block and serotonin 1a agonist on apnea and irregularity score in *Mecp2*<sup>-/-</sup> studied in situ and conscious *Mecp2*<sup>-/-</sup> mice. (A.1) Apnea in *Mecp2*<sup>-/-</sup> female mice treated with combined NO-711 (5.2  $\mu$ M) and 8-OH-DPAT (0.1  $\mu$ M). \* $P = 0.01$  ( $n = 6$ ) (ANOVA). (A.2) Irregularity score. \* $P = 0.001$ . (B.1) Apnea in *Mecp2*<sup>-/-</sup> males treated with combined NO-711 (1.0 mg/kg) and 8-OH-DPAT (150  $\mu$ g/kg). \* $P = 0.016$  ( $n = 5$ ) (paired  $t$  test). (B.2) Irregularity score. \* $P = 0.05$ .

#### Tonic Activity in Expiratory Neurons Causes Apnea in *Mecp2*-Deficient Mice.

The increased amplitude and extended duration of cVN post-I bursts coupled with tonic activity in AbN strongly suggest that arrest of PN activity is associated with a high excitability state of expiratory neurons. This may trigger enhanced levels of inhibitory synaptic influences from population of expiratory neurons into the respiratory pattern generator. The tonic activity seen in HN is also likely to be expiratory in origin. Although HN is often described as having solely inspiratory activity, particularly from in vitro preparations, there is evidence that the output of this cranial nerve also contains expiratory motor activity especially in vivo (19–21). Thus, the in situ characterization of PN apnea in *Mecp2*<sup>-/-</sup> female mice reveals that three expiratory outputs—post-I in cVN, AbN, and expiratory component of HN—are tonic.

#### Insufficient GABA<sub>A</sub> Inhibition Underlies Tonic Activity in Expiratory Neurons.

The present experiments do not determine whether phasic or tonic GABA<sub>A</sub> receptor-mediated signaling account for the effects of NO-711. In the hippocampus this reuptake blocker induced a tonic conductance in pyramidal cells (22). The L-838,417 protocol does not answer this question because the compound potentiates activity in receptors with  $\alpha 2$  and  $\alpha 3$  (synaptic) and  $\alpha 5$  (extrasynaptic) subunits (18). L-838,417, however, established an important clinically relevant finding, namely that modulation of the  $\alpha 1$  subunit is not necessary for achieving beneficial effects on respiration. The  $\alpha 1$  subunit mediates the sedative (18) and addictive (17) properties of benzodiazepines. Blocking GABA reuptake in the present studies could act through GABA<sub>A</sub> and/or GABA<sub>B</sub> receptors. GABA<sub>B</sub> agonists have been shown to depress respiratory rate (23, 24). Thus the reduction in frequency

observed in *Mecp2*<sup>-/+</sup> mice treated with NO-711 may have involved GABA<sub>B</sub> receptors. This effect, however, was not seen in null males. Potentiation of GABA<sub>A</sub> receptor activation through allosteric benzodiazepines was equally effective in correcting respiratory disorders in *Mecp2*<sup>-/+</sup> animals. This suggests that the correction of respiratory disorders involves primarily GABA<sub>A</sub> receptors. The significant amelioration of apnea and periodic breathing, both of which were associated with tonic activity in HN, by augmenting GABA indicates that insufficient inhibition is the underlying mechanism.

#### Possible Expiratory Neuronal Groups That Are Tonic During Apnea.

Two observations in the present study suggest that expiratory neurons reflected by the expiratory portion of HN and by AbN, which are tonic during PN apnea, do not arise from a single population: (i) the temporal relationship between onset of tonic discharge in AbN relative to that in HN, when both were present, was equally distributed between concurrent, earlier, and later; and (ii) the effect of blocking GABA reuptake with NO-711 on these two nerves was markedly different. AbN tonic activity was virtually abolished in the apneas that remained after reuptake block, whereas it generally persisted in HN with a decrease in burst amplitude.

The finding of increased and prolonged discharge in post-I activity of cVN raises the question of where in the brainstem this activity originates. Recent evidence supports a role for the pons: microtransection studies in rat that eliminated the pons abolished post-I recorded from cVN (25, 26). Moreover, glutamate stimulation of Kölliker-Fuse (KF) neurons caused a prolongation of post-I recorded from the recurrent laryngeal nerve of juvenile rats in situ, whereas GABA receptor activation at the identical injection sites completely suppressed post-I activity (27). In addition, there is evidence that medullary sites also play a role. Bilateral microinjections of the GABA<sub>A</sub> agonist isoguvacine into retrotrapezoid nucleus (RTN) reduced post-I in cVN (15). This indicates that RTN either through afferents to KF or as a relay from KF to post-I neurons in the Böttinger complex plays an important role in the generation of post-I expression.

It is also likely that stage 2 or late expiratory (augmenting neurons) are driven during the respiratory apneas of the *Mecp2*-deficient mice. Recently, expiratory activity in the AbN has been well characterized in neonatal and juvenile rats examined with the in situ preparation (15). Under baseline conditions (5% CO<sub>2</sub>) only post-I discharge was present. Hypercapnia (7% or 10% CO<sub>2</sub>), however, induced late expiratory (late-E) bursts that were characterized by an incrementing shape and abrupt termination with the onset of PN activity. The tonic AbN seen in *Mecp2*<sup>-/+</sup> mice during PN apnea has a similar, albeit prolonged, pattern. Either pontine transection or bilateral isoguvacine injections into RTN abolished late-E AbN activity (15), and on the basis of this we propose that these regions could generate the pathological discharge in the abdominal motor outflow in *Mecp2*<sup>-/+</sup> mice.

To date, the location and characterization of GABA interneurons whose insufficiency may underlie the respiratory disorders in *Mecp2*-deficient mice are not known. However, Medrihan et al. (11) examined GABA synapses in the ventrolateral medulla of *Mecp2* null males at postnatal day 7. Neither the discharge pattern relative to PN nor markers to more clearly define the area under study were used. Neurokinin-1 receptor [a marker of pre-Böttinger neurons (28)] immunoreactive cells receive both GABAergic and glycinergic boutons onto their soma and dendrites (29). More recently, glutamic acid decarboxylase-green fluorescence protein (GAD67-GFP) knockin mice have been used to examine inhibition in respiratory related areas. Recordings from GFP-positive neurons in medullary slices that contained the pre-Böttinger complex showed that they fired concurrently with the integrated rootlet of HN (30). The spatial projections of these interneurons were not reported.

**Mechanism of Serotonin 1a Agonist Induced Inhibition of Expiratory Activity.** Serotonin 1a receptors are located presynaptically, where they act as heteroreceptors and modulate neurotransmitter release (reviewed in ref. 13). At this site they attenuate GABA release. Because insufficient GABA underlies the respiratory disorders in Mecp2-deficient mice, it is unlikely that the effects observed with 8-OH-DPAT are due to presynaptic serotonin 1a receptor activation. Postsynaptic 1a receptors by coupling to G $\alpha$ i subunits activate G protein-dependent inward rectifying K<sup>+</sup> channels with resultant hyperpolarization. This inhibitory effect has been demonstrated in late-E neurons of anesthetized cats (12) and most likely contributes to the effects seen with 8-OH-DPAT. More recently it has been shown that the glycinergic inhibitor strychnine abolishes the effects of 8-OH-DPAT in rats studied in situ (14). Thus glycine-mediated inhibition may also add to the correction of respiratory disturbances seen with the serotonin 1a agonist.

8-OH-DPAT also exerts effects through serotonin 7 receptors, albeit at concentrations 100-fold higher than that required at 1a receptors (31). Systemic administration of a serotonin 7 agonist in rats studied in situ, however, reduced respiratory frequency (32). In contrast, we observe no change in frequency with 8-OH-DPAT, suggesting that its primary site of action is mediated by serotonin 1a receptors.

In summary, we have demonstrated that either alone or especially in combination, augmenting GABA<sub>A</sub> inhibition or a serotonin 1a agonist markedly reduces the incidence of apnea and periodic breathing and restores breath cycle regularity in Mecp2-deficient mice. The treatments did not adversely affect animal behavior, and in Mecp2<sup>-/-</sup> mice the combination increased

activity over that seen with vehicle, to a level the same as in WT. Taken together with the demonstration that activation of Mecp2 in mature symptomatic mice reverses their phenotype (5), the results indicate that the respiratory network is intact in Mecp2-deficient mice. Inasmuch as drugs that act in this manner are currently approved to treat human disorders, although not specifically for respiratory disorders in RTT, the results have significant clinical relevance.

## Materials and Methods

Both heterozygous female and null male mice and their WT littermates were of the B6.129P2(C)-Mecp2<sup>tm1.1Bird</sup> strain. Mice were genotyped using a published method (33). In situ arterially perfused brainstem-spinal cord studies were performed as previously described (34). Telemetry recording of pleural pressure was obtained as previously described (35, 36). Respiratory pattern determined with body plethysmography was performed as previously described (37, 38). For chronic pharmacological treatment, respiratory pattern was examined at weekly intervals in Mecp2<sup>-y</sup> males until they developed an abnormal pattern (approximately postnatal day 40). Under general anesthesia (1.5% isoflurane in oxygen) a 100- $\mu$ L volume osmotic pump (Alzet model 1004) was placed in the peritoneal cavity through a midline abdominal incision. The pump contained 8-OH-DPAT (20 mM or 6.25  $\mu$ g/ $\mu$ L) and delivered 0.11  $\mu$ L/h for 28 d. A full description of methods is provided in *SI Materials and Methods*.

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