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Novel targets for prostate cancer chemoprevention

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Abstract

Among many endocrine-related cancers, prostate cancer (PCa) is the most frequent male malignancy, and it is the second most common cause of cancer-related death in men in the United States. Therefore, this review focuses on summarizing the knowledge of molecular signaling pathways in PCa because, in order to better design new preventive strategies for the fight against PCa, documentation of the knowledge on the pathogenesis of PCa at the molecular level is very important. Cancer cells are known to have alterations in multiple cellular signaling pathways; indeed, the development and the progression of PCa are known to be caused by the deregulation of several selective signaling pathways such as the androgen receptor, Akt, nuclear factor- κ B, Wnt, Hedgehog, and Notch. Therefore, strategies targeting these important pathways and their upstream and downstream signaling could be promising for the prevention of PCa progression. In this review, we summarize the current knowledge regarding the alterations in cell signaling pathways during the development and progression of PCa, and document compelling evidence showing that these are the targets of several natural agents against PCa progression and its metastases.

Introduction

Despite significant effort made in the fight against cancers, prostate cancer (PCa) is still the most frequent non-cutaneous male malignancy, and it is the second most common cause of cancer death in the United States with an estimated 192 280 new cases and 27 360 deaths expected in 2009 (Jemal et al. 2009). Treatments for PCa include surgery, radiation, chemotherapy, or hormonal ablation therapy. Since PCa is an endocrine-related cancer driven by androgens, androgen deprivation can be used to shrink the cancer significantly even though androgen ablation therapy alone is not the optimal therapy to eradicate PCa; androgen ablation combined with other novel therapies may be more effective. Despite the initial efficacy of androgen-deprivation therapy, most patients with advanced PCa eventually develop resistance to this therapy and progress to castrate-resistant PCa (CRPC) for which there is no curative therapy (Bracarda et al. 2005). The emergence of CRPC and its subsequent metastases contributes to overall poor survival and high mortality (Donovan et al. 2010). Declining mortality trends for PCa have been observed since the early 1990s, suggesting that early detection using the prostate-specific antigen (PSA) test or digital rectal exam is beneficial. However, it is important to note that prevention should still be the fundamental strategy by which the mortality due to PCa should be reduced.

Recently, a significant proportion of cancers have been believed to be preventable. It is estimated that one-third of all cancers are preventable simply through modification of diet,

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Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

maintenance of optimum body weight, and regular physical activity (American Cancer Society 2009, Amin *et al.* 2009). For the prevention of PCa, chemoprevention could be an important avenue aiming to reduce both the incidence and the mortality through the use of active 'natural agents' to prevent, reverse, or delay the carcinogenic process. So far, some chemopreventive agents have been considered to reduce PCa risks. Several agents including 5- α -reductase inhibitors (finasteride and dutasteride), selenium, vitamins E and D, lycopene, soy isoflavones, green tea polyphenols, 3,3'-diindolylmethane (DIM) and curcumin have demonstrated their various activities in the inhibition of prostate carcinogenesis, with mixed results.

In recent years, significant efforts have been made to understand the biological and molecular mechanisms driving PCa development and progression. It is necessary to reveal the molecular determinants involved in the processes of cancer development and progression of PCa, in order to design or find novel chemopreventive agents that could be useful in targeted prevention and/ or treatment strategies against PCa. In this review, we summarize the current knowledge regarding the alterations in cell signaling pathways during the development and progression of PCa, and outline the evidence supporting the development of innovative strategies for targeting selective pathways by novel chemopreventive agents for the prevention of PCa progression.

Cell signaling pathways involved in the development and progression of PCa

Cancer cells are known to have alterations in multiple cellular signaling pathways. In PCa cells, the altered proteins produced as a result of mutations or defects of genes affect the way these cells communicate with each other. The cellular signaling pathways that are known to be important in PCa cells include the androgen receptor (AR), Akt, nuclear factor- κ B (NF- κ B), Wnt, Hedgehog (Hh) and Notch pathways, among many others (Fig. 1). The alterations in these pathways could occur at different stages of PCa from early to advanced disease.

AR signaling

AR is a ligand-activated transcription factor of the nuclear receptor superfamily that plays a critical role in male physiology and pathology. In prostate epithelial cells, ligand-free AR is sequestered in the cytoplasm and bound to heat shock proteins (HSPs). Binding of androgens to the AR induces a conformational change in the AR, which causes the dissociation of HSPs and phosphorylation of the AR. The conformational change in the AR also allows AR nuclear localization, increased AR phosphorylation in the nuclear compartment, AR homodimer formation, and its interaction with DNA (Heinlein & Chang 2002, 2004). The activated AR then initiates gene transcription by binding to specific androgen response elements in the promoter regions of target genes (Fig. 1), promoting prostate epithelial cell growth (Heinlein & Chang 2002, 2004). Androgens and the AR are involved in all stages of prostate carcinogenesis including initiation, progression, and treatment resistance (Montgomery et al. 2001); therefore, AR signaling has been believed to be a critical target for PCa prevention and/ or treatment. Moreover, one of the androgen-responsive genes, PSA, is a clinically important marker that is routinely used to monitor diagnosis, treatment response, prognosis, and progression in patients with suspected PCa (Kupelian et al. 1996, Sato et al. 1996). It is known that during the progression of PCa from an androgen-sensitive status to the androgenindependent stage classically known as CRPC, the majority of PCa cells still express AR, suggesting that AR signaling plays a critical role in the development and progression of PCa (Heinlein & Chang 2004). Moreover, phosphorylation of the AR by molecules in other cell signaling pathways could also influence AR transactivation (Lee & Chang 2003). Studies have shown that Akt can phosphorylate the AR at Ser210/213 and Ser790/791, and transactivate the activity of AR independent of androgen signaling (Wen et al. 2000). The phosphorylation by Akt also sensitizes the AR to low circulating levels of androgen, such as those present during maximum androgen blockade (Rochette-Egly 2003). This sensitization allows low levels of

androgens to induce phosphorylation at specific sites, which is required for the translocation of the AR to the nucleus. Therefore, Akt is an important activator of the AR, which is required for the androgen-independent survival and growth of PCa cells.

PI3K/Akt/mammalian target of rapamycin signaling

The phosphoinositide-3-kinase (PI3K)/Akt/mammalian target of rapamycin (mTOR) pathway plays critical roles in mammalian cell survival signaling and is activated in various cancers including PCa (Liu et al. 2009, Morgan et al. 2009, Sarker et al. 2009, de Souza et al. 2009). It has been reported that Akt is activated by phospholipid binding and phosphorylation at Thr308 by PDK1 or at Ser473 by PDK2 (Alessi et al. 1996). Activated Akt functions to promote cell survival by inhibiting apoptosis through inactivation of several pro-apoptotic factors including Bad, Forkhead transcription factors, and caspase-9 (Cardone et al. 1998, Brunet et al. 1999). In addition to promoting cell survival through the inhibition of apoptosis, the Akt pathway regulates cell growth, proliferation, and angiogenesis through the mTOR and PTEN signaling pathways, which facilitates translation of important signaling molecules such as c-Myc, cyclin D, and vascular endothelial growth factor (VEGF). Restoration of functional PTEN activity or inhibition of mTOR activity can block the growth of PTEN^{-/-} PCa xenografts in mice and restore sensitivity to chemotherapy (Neshat et al. 2001). It has been estimated that PI3K/Akt/mTOR signaling is up-regulated in 30-50% of PCa cases, often due to the loss of PTEN function (Morgan et al. 2009). The alteration of molecules in the PI3K/Akt/mTOR signaling pathway has been found in PCa when comparing malignant prostatic epithelium with normal epithelium. More importantly, the activation of PI3K/Akt/mTOR signaling is associated with increasing tumor stage, grade, and risk of biochemical recurrence; therefore, the Akt pathway is an attractive target for cancer prevention and/or treatment (Sarker et al. 2009). It is also important to note that insulin-like growth factor (IGF) is an upstream molecule of Akt signaling, and the up-regulation of IGF, which activates Akt, could promote the development of PCa in animal models (Adhami et al. 2004, 2009), suggesting the interrelationship of IGF and Akt signaling in PCa. Studies have also shown that Akt regulates NF- κ B signaling via the phosphorylation and activation of molecules in the NF- κ B pathway (Fig. 1; Ozes et al. 1999, Romashkova & Makarov 1999); therefore, NF-KB signaling is also a critical pathway for the development and progression of PCa, and this pathway is an appropriate target for the management of PCa development and progression.

NF-kB signaling

It is now well accepted that the NF- κ B signaling pathway plays important roles in the control of cell growth, apoptosis, inflammation, stress response, and many other physiological processes (Yamamoto & Gaynor 2001, Karin et al. 2002, Li & Verma 2002, Lin & Karin 2003, Storz & Toker 2003). There are several important molecules such as NF-κB, inhibitor of κ light polypeptide gene enhancer in B-cells (I κ B), and I κ B-kinase (IKK) involved in the NF- κ B signaling pathway; however, NF- κ B is the key protein in the pathway, and has been described as a major culprit and therapeutic target in cancer (Biswas et al. 2001, Bharti & Aggarwal 2002, Haefner 2002, Orlowski & Baldwin 2002). The activation of NF-kB has been frequently observed in PCa. The constitutive activation of NF-KB observed in PCa cells is likely to be due to the involvement of other multiple signal transduction pathways including tyrosine kinase, NF-KB inducing kinase (NIK), and IKK activation (Suh et al. 2002). Moreover, nuclear translocation and activation of NF-KB has been reported to be significantly greater in PCa patients with lymph node metastasis compared with controls. Such up-regulation of NFκB activity was observed in the tumor cells as well as in the surrounding lymphocytes (Ismail et al. 2004), suggesting that NF-KB plays critical roles in the development and progression of PCa. Indeed, blockade of NF-κB activity in human PCa cells suppressed angiogenesis, invasion, and metastasis in PCa cells (Huang et al. 2001). Furthermore, constitutive activation of PI3K/Akt and NF-kB was also observed during PCa progression in an autochthonous

transgenic mouse model (Shukla *et al.* 2005), suggesting that both Akt and NF-κB are potential molecular targets for the prevention and/or therapeutic intervention in PCa.

Wnt signaling

Wnt signaling plays important roles in the embryonic developmental processes including cell proliferation, differentiation, and epithelial-mesenchymal interactions (Angers & Moon 2009). The aberrant activation of the canonical Wnt/ β -catenin signaling pathway is one of the most frequent signaling abnormalities known in human cancers. In human cancer, activated What signaling promotes β -catenin accumulation in the nucleus, resulting in the transcriptional activation of specific target genes and the development of cancer (Fig. 1; Behrens 2000, Peifer & Polakis 2000). The inappropriate expression of the Wnt ligand and Wnt-binding proteins and the inappropriate activation of the Wnt signaling pathway have been found in a variety of human cancers including PCa (Taipale & Beachy 2001, Reya & Clevers 2005, Verras & Sun 2006). However, the activation of Wnt signaling may occur in a different manner in PCa than in colorectal cancer or other human malignancies because mutations in adenomatous polyposis coli and other components of the β -catenin destruction complex are rare in PCa cells. Therefore, other regulatory mechanisms could play dominant roles in the activation of β -catenin in PCa. In PCa cells, Wnt-3a was found to stimulate proliferation selectively in AR-positive CWR22Rv1 and LNCaP cells; however, T-cell factor (TCF)-dependent reporter gene transcription was not induced in LNCaP cells, suggesting that the activation of Wnt signaling in AR-positive PCa cells may be through AR-dependent mechanisms rather than classical TCFdependent mechanisms (Cronauer *et al.* 2005). It was also found that β -catenin enhanced the function of AR and that nuclear translocation of β -catenin takes place in PCa tissue, indicating that Wnt signaling is required for disease progression (Chesire et al. 2002). In addition, loss or reduction of E-cadherin and abnormal expression of Wnt ligands, receptors, inhibitors, and other co-regulators could also contribute to the activation of the Wnt signaling pathway in PCa. Therefore, inhibition of aberrant Wnt activity in PCa cells could provide an opportunity for the prevention and/or treatment of PCa (Dihlmann & von Knebel 2005, Barker & Clevers 2006, Verras & Sun 2006).

Hh signaling

Another important signaling pathway involved in cell development and proliferation is the Hh signaling pathway, which is a major regulator of cell differentiation, tissue polarity, and cell proliferation. Hh ligands, Sonic Hh and Indian Hh, stimulate GLI transcription factors, which constitute the final effectors of the Hh signaling pathway (Fig. 1). It has been found that germline mutations that subtly affect Hh pathway activity are associated with developmental disorders (Varjosalo & Taipale 2008). More importantly, somatic mutations that activate the Hh pathway have been linked to a variety of human cancers (Varjosalo & Taipale 2008). Emerging evidence clearly suggests the activation of Hh signaling in various human cancers, including basal cell carcinomas, medulloblastomas, leukemia, gastrointestinal, lung, ovarian, breast, and PCa (Yang et al. 2010). Furthermore, because Hh plays a central role in the control of cell proliferation and differentiation of both embryonic stem cells and adult stem cells, the aberrant activation of Hh signaling could lead to the development of cancer and the generation of cancer stem cells (Medina et al. 2009). It has been known that epithelial expression of Hh ligand during prostate development exerts autocrine and paracrine signaling activities that regulate growth and differentiation. Increased Hh signaling has been associated with PCa progression and has also been shown to accelerate PCa growth (Vezina & Bushman 2007). Studies have revealed the critical role of Hh signaling in PCa, and demonstrated that autocrine Hh signaling by tumor cells is required for the proliferation, viability, and invasive behavior of PCa (Antón Aparicio et al. 2007). Therefore, the development of Hh inhibitors such as those that are currently coming through the drug pipeline hold great promise for the prevention and/ or treatment of PCa.

Notch signaling

Hh, Wnt, transforming growth factor- β (TGF- β)/BMP, and Notch signaling pathways all are involved in embryonic development, adult tissue homeostasis, and tumorigenesis. It is known that Notch signaling plays a critical role in the regulation and maintenance of stem cells; therefore, normal functioning of Notch signaling is required for development during early life. Emerging evidence suggests that deregulation of Notch signaling contributes to the development and progression of a number of cancers (Rizzo et al. 2008, Zardawi et al. 2009). Up-regulation of Notch receptors and their ligands has been observed in cervical, lung, colon, head and neck, renal and pancreatic cancers, and in Hodgkin and large-cell lymphomas (Miele et al. 2006). Controversial results have been reported regarding the role of Notch in PCa. Shou et al. (2001) reported that the expression of Notch ligands was low or undetectable in PCa cells, and that overexpression of a constitutively active form of Notch-1 inhibited the proliferation of various PCa cells, suggesting that Notch acts as a tumor suppressor. However, we and other investigators have found that Notch ligand Jagged-1 expression was associated with PCa metastasis and recurrence (Santagata et al. 2004), and that down-regulation of Notch-1 and Jagged-1 inhibited PCa cell growth, migration, and invasion, while inducing apoptosis via inactivation of Akt, mTOR, NF-KB, MMP-9, and uPA signaling pathways (Bin et al. 2009, Wang et al. 2010). Therefore, Notch signaling could be an important target for the prevention and/or treatment of PCa; however, more in-depth molecular investigations are needed to address this controversy because the consequences of aberrant Notch signaling could depend on cell context, dose, and timing (Maillard & Pear 2003).

Other signaling pathways

Other signaling pathways that are involved in PCa include epidermal growth factor receptor (EGFR) signaling, VEGF receptor (VEGFR) signaling, IGF receptor (IGFR) signaling, and mitogen-activated protein kinase (MAPK) signaling. The activation of these signaling pathways could stimulate the development of PCa through the activation of PI3K/Akt and NFκB signaling. It has been found that increased levels of circulating IGF1 and decreased levels of IGF binding protein 3 (IGFBP3) were associated with a higher risk of developing PCa (Renehan *et al.* 2004). Moreover, tumor angiogenesis is an important biological component of PCa metastasis; the contribution of increased angiogenic molecules such as VEGF was investigated in PCa, and found to correlate with advanced clinical stage in PCa (Duque *et al.* 1999, Shariat *et al.* 2004). Therefore, EGFR, VEGFR, IGFR, and MAPK signaling pathways also participate in the development and progression of PCa, suggesting that the inhibitors of these signaling pathways could be important for the prevention of PCa progression and/or treatment.

It is important to note that cellular signaling is a complex signal network with positive or negative feedback loops and is also regulated by compensatory mechanisms (Fig. 1). In PCa cells, deregulations of several signaling pathways often exist; therefore, targeting multiple signaling pathways is needed for the prevention and/or treatment of PCa in the future.

Chemopreventive agents and their targets in PCa prevention

To prevent the development and progression of PCa, the strategy should target the cell signaling pathways that are deregulated in benign and malignant prostate tumors. Thus far, several chemopreventive agents including $5-\alpha$ -reductase inhibitors (finasteride and dutasteride), selenium, vitamins E and D, lycopene, soy isoflavones, green tea polyphenols and curcumin have shown their various activities in the inhibition of prostate carcinogenesis through the regulation of major cell signaling pathways such as the AR, Akt, NF- κ B, Wnt, Hh and Notch. However, the results to date have been mixed as summarized in the subsequent paragraphs.

5-α-reductase inhibitors

It has been well accepted that the activation of AR signaling plays critical roles in the development and progression of PCa. To activate AR signaling, androgens including testosterone and dihydrotestosterone (DHT), a metabolic product of testosterone, bind to the AR and stimulate the activation of the AR. However, DHT has a much higher affinity, leading to different kinetic processes. It is known that both type I and type II 5- α -reductases are responsible for synthesizing DHT from testosterone in prostatic tissue and in peripheral tissues. Therefore, 5- α -reductase inhibitors have been used to inhibit AR activation for the prevention of PCa. It has been found that the use of clinically available 5- α -reductase inhibitors leads to a reduction in prostatic volume of around 30%, and serum PSA levels are reduced by 50–60% in men with benign prostatic enlargement (Marberger 2006); however, the precise role of 5- α -reductase inhibitors for the prevention of PCa development and progression remains to be tested in a large population.

Finasteride is an agent that targets type II 5- α -reductase. This agent was used in the PCa Prevention Trial (PCPT), which tested a hypothesis that treatment with finasteride could lower DHT levels and thereby inhibit the activation of the AR, leading to the prevention of PCa. The PCPT was the first large-scale trial and included 18 882 men with a normal PSA level. Finasteride (5 mg/day) or placebo was given for 7 years. It was found that the prevalence of PCa was reduced by 24.8% in those randomized to finasteride compared with placebo (Thompson et al. 2003). However, the prevalence of tumors at Gleason scores 7-10 was higher in the finasteride group than in the placebo group, suggesting that the benefit of reducing the risk of PCa by finasteride must be weighed against the increased risk of development of highgrade PCa (Thompson et al. 2003). Recently, several studies have been conducted to re-analyze the data of the PCPT. It was found that sampling density bias alone could explain the excess of high-grade cancers among the finasteride-assigned participants in the PCPT (Cohen et al. 2007) and that after adjusting for biopsy sampling density, finasteride significantly reduced PCa risk relative to placebo across multiple Gleason scores in the PCPT, including the most frequently detected intermediate- and high-grade (Gleason scores 6 and 7) PCa (Kaplan et al. 2009).

Ongoing trials are addressing the unanswered questions from the PCPT. The second largescale trial of a 5- α -reductase inhibitor is the Reduction by Dutasteride of Prostate Cancer Events (REDUCE) trial (Andriole *et al.* 2004*a*, Crawford *et al.* 2010). The REDUCE clinical trial is an international, multi-center, double-blind, placebo-controlled chemoprevention trial. The study will examine the effects of the dual 5- α -reductase inhibitor dutasteride on the natural history of PCa in men at increased risk of this malignancy (Andriole *et al.* 2004*a*). The study will also examine biomarkers and genetic linkage for PCa. Dutasteride is used in the treatment of benign prostatic hyperplasia. It reduces serum PSA levels by ~50% at 6 months and total prostate volume by 25% after 2 years. Dutasteride differs from finasteride in that it inhibits both isoenzymes of 5- α -reductase, type I and type II. Preliminary data suggest a decrease in PCa incidence in dutasteride-treated patients (Andriole *et al.* 2004*b*, Musquera *et al.* 2008); however, we await the final outcome of this trial, which could be promising.

The major target of $5-\alpha$ -reductase inhibitors is AR signaling through the inhibition of $5-\alpha$ -reductase (Table 1). Finasteride significantly inhibits the activation of the AR; however, other cell signaling is also involved. It was found that finasteride significantly inhibited the proliferation of LNCaP PCa cells through regulation of the expression of AKR1B1, PTEN, NKX3.1, PMEPA1, PSA, and XRCC2 (Chen *et al.* 2005). In addition, finasteride could also induce apoptosis through the regulation of Akt, caspases, XIAP, and TGF- β signaling (Saez *et al.* 1998, Sawaya *et al.* 2002, Li & Kim 2009). Dutasteride has also been found to effectively inhibit both the viability and proliferation of LNCaP PCa cells, and to induce apoptosis. Dutasteride disrupted genes and cellular pathways that are involved in metabolic, cell cycle,

and apoptotic responses with alterations in TRADD, caspase-7, caspase-8, BIRC1, and other genes (Schmidt *et al.* 2004,2009,Biancolella *et al.* 2007). These mechanistic studies could be useful in explaining the positive or the negative outcome of the ongoing clinical trials using dutasteride.

Vitamin D

Two major forms of vitamin D that are important for humans are vitamin D_2 and D_3 . The active form of vitamin D in the body is 1,25-dihydroxyvitamin D, which can be made from either vitamin D_2 or vitamin D_3 . Some studies suggest that higher intakes of vitamin D from food and/or supplements and thereby higher levels of vitamin D in the blood are associated with reduced risk of cancer (Garland et al. 2006). Several epidemiological studies have suggested that vitamin D could be a preventive agent for PCa. It was found that reduced levels of active vitamin D resulted in a higher PCa incidence and mortality (Garland et al. 2006). Native Japanese men, whose diet is rich in vitamin D, have a low incidence of PCa, further supporting the protective role of vitamin D. A study also showed that dietary supplementation with >600IU of vitamin D reduced the risk of PCa (Ahn et al. 2007). However, the results of other studies were conflicting or negative (Whittemore et al. 1995, Lee et al. 1998), which suggests complexities in vitamin D signaling in PCa. The data from several studies also appeared to show a protective role of sunlight/UVB exposure, which induces vitamin D, against PCa (Gupta et al. 2009). Although the results of these studies are conflicting, one should not ignore the fact that vitamin D deficiency is associated with PCa risks. Further molecular understanding of the signaling pathways related to vitamin D is expected to resolve these controversies in the future. The precise mechanisms of vitamin D action on the prevention of PCa are not clear, although it is known that vitamin D regulates the expression of genes such as CYP24A1, osteopontin (Spp1), LRP5, TRPV6 and VDR (Meyer et al. 2010, Pike et al. 2010; Table 1); however, the significance of the regulation of these genes in relation to vitamin D and PCa remains unclear.

Selenium

Selenium is an essential trace element found in grains, fish, meat, poultry, or eggs. Selenium is currently available in over-the-counter supplements and multi-vitamins. It has been established that selenium is distributed in body tissues and has an antioxidant effect. Epidemiological studies also showed that selenium could be a protective agent against the development of PCa (Klein 2004). A study was conducted to test the level of selenium in serum and prostate of 52 men after selenium supplementation. It was found that selenium supplementation resulted in a significantly higher level of selenium in the prostatic tissue (Gianduzzo et al. 2003), suggesting the high bioavailability of selenium. To evaluate the effect of selenium on the prevention of PCa, a large clinical trial has been conducted. It was found that selenium supplementation significantly reduced the overall incidence of PCa with relative risk of 0.51 (95% confidence interval 0.29–0.87; Duffield-Lillico et al. 2003). The protective effect of selenium supplementation appeared to be confined to those with a baseline PSA level of <4 ng/ml and low serum levels of selenium (Duffield-Lillico et al. 2003). In an animal study, diet with selenium supplementation was given for 7 months and it was found that the extent of DNA damage in prostate cells and peripheral blood lymphocytes, as determined by the alkaline comet assay, was lower, while apoptotic cell death was higher in seleniumsupplemented dogs compared with the control dogs (Waters et al. 2003), suggesting that selenium is a potent antioxidant. In vitro studies have revealed that selenium could inhibit cellular proliferation, induce apoptosis, and modulate genes related to cell growth, apoptosis, and androgen regulation, leading to the suppression of prostatic tumorigenesis (Dong et al. 2003, Zhao et al. 2004).

The targets of selenium include the AR, estrogen receptor (ER), NF- κ B, antioxidant genes and pro-inflammatory molecules (Table 1). It was found that selenium could exert its anticancer property through increasing the expression of a humoral defense gene (*A2M*) and the tumor suppressor-related genes (*IGFBP3* and *HHIP*) while decreasing pro-inflammatory gene expression (*CXC L9* and *HSPB2*) (Zeng & Botnen 2007, Vunta *et al.* 2008). High selenium intake reduced the expression of AR, 24-dehydro-cholesterol reductase (Dhcr24), and ATPbinding cassette sub-family C member 4 (ABCC4; Legg *et al.* 2008, Schmidt *et al.* 2009). Selenium could also inhibit ER α signaling (Shah *et al.* 2005). In addition, treatment with selenium virtually eliminated the binding of NF- κ B to target DNA and reduced transcription of NF- κ B-regulated genes (Christensen *et al.* 2007); these findings are consistent with the antioxidant effects of selenium. The role of selenium in the prevention of PCa is further discussed below.

Vitamin E

Vitamin E is a lipid-soluble antioxidant found in green leafy vegetables, nuts, seeds, sunflower, and plant oils. Several forms of vitamin E have been identified; however, α -tocopherol is the most active, abundant, and predominant form of vitamin E in human tissues. In addition to the antioxidant effect, as it contains a chromanol moiety, vitamin E could also have antiandrogen activity (Thompson & Wilding 2003). In the 1990s, NIH conducted the Alpha-Tocopherol, Beta-Carotene (ATBC) Cancer Prevention Study, which was a placebo-controlled, randomized intervention trial to test the hypothesis of whether β -carotene and α -tocopherol (vitamin E) supplements could prevent lung and other cancers. It was found that the vitamin E group had reduced incidence of PCa compared with the group not receiving vitamin E (number of cases 99 compared with 151) (Albanes *et al.* 1995). Several years later, a prospective nested case–control study to determine the association between serum carotenoids, retinoids, and tocopherols (vitamin E) on both lung and PCa incidence was reported. For PCa, low serum levels of α -tocopherol (vitamin E) were associated with a higher risk of PCa (Goodman *et al.* 2003), suggesting a protective effect of vitamin E against PCa although the results on lung cancer were disappointing.

The mechanism involved in the prevention of PCa by vitamin E is not very clear. It was found that vitamin E mainly modulated two major signal transduction pathways including PI3K/Akt and protein kinase C (Azzi *et al.* 2004), leading to a change in cell proliferation. Several other genes were also regulated by vitamin E partly because of the effects of vitamin E on kinases. These genes include *P450*, glutathione *S*-transferase, *MMP-1*, *MMP-19*, *IL-2*, *IL-4*, cyclin D1, cyclin E, *Bcl-2*, *p27*, *CD95* (APO-1/Fas ligand), $5 - \alpha$ -reductase type I, NF- κB and activator protein 1 (*AP1*), (Azzi *et al.* 2004; Table 1). The antioxidant-responsive element and the TGF- β -responsive element were also regulated by vitamin E (Azzi *et al.* 2004). Recent animal studies have also shown that vitamin E could induce p21 signaling and significantly increase the median lifespan of C57BL/6 mice by 15%, an effect which appeared to be independent of any antioxidant effect of vitamin E (Banks *et al.* 2010); however, the role of vitamin E as a single agent for the prevention of PCa remains to be established.

Since epidemiological and biological studies showed that selenium and vitamin E may prevent PCa, a phase III trial is currently assessing the value of selenium and vitamin E in the prevention of PCa. This trial, known as the Selenium and Vitamin E Cancer Prevention Trial (SELECT), is a randomized, prospective, double-blind study designed to determine whether selenium and vitamin E alone or in combination can reduce the risk of PCa among healthy men. SELECT is the second large-scale study of chemoprevention for PCa, and enrollment began in 2001 with final results anticipated in 2013 (Klein *et al.* 2003). However, the results reported so far are disappointing. As of October of 2008, median overall followup was 5.46 years. A statistically non-significant increased risk of PCa was found in the vitamin E group (P=0.06)

but not in the selenium plus vitamin E group. Selenium or vitamin E alone or in combination at the doses and formulations used did not prevent PCa in this population of relatively healthy men (Lippman *et al.* 2009). A recent animal study also does not support the hypothesis that selenium and vitamin E are potent cancer chemopreventive agents against PCa (McCormick *et al.* 2010), suggesting that more detailed epidemiological and biological studies are needed to investigate the effects of selenium and vitamin E prior to conducting expensive large-scale intervention trials for reducing the risk of PCa.

Soy isoflavone

Isoflavones are a subclass of the more ubiquitous flavonoids and are much more narrowly distributed in soybeans. Genistein, daidzein, and glycitein are three isoflavones found in soybeans and most soy protein products. Several epidemiological studies have shown that soy could have protective effects against prostate and other cancers (Adlercreutz *et al.* 1991, 1993, Hebert *et al.* 1998, Jacobsen *et al.* 1998). A prospective study of 12 395 California Seventh-Day Adventist men who often drank soy milk showed that frequent consumption (more than once a day) of soy milk was associated with a 70% reduction in the risk of PCa (Jacobsen *et al.* 1998), suggesting the possible association between a high intake of soy isoflavones and a reduced risk of PCa. Experimental studies have also revealed that isoflavones, particularly genistein, exert antioxidant effects on human cells. It has been known that genistein protects cells against reactive oxygen species (ROS) by scavenging free radicals and reducing the expression of stress–response-related genes.

Our laboratory has investigated the effects of the isoflavone genistein on several signaling pathways. We have found that isoflavones significantly inhibited the activation of AR, Akt, NF-κB, and Notch signaling (Li & Sarkar 2002*a*,*b*, Wang *et al.* 2006*a*, Li *et al.* 2008, Sarkar et al. 2008; Table 1). The isoflavone genistein has also been found to inhibit the molecules in the MAPK pathway. It was reported that genistein blocked the activation of p38 MAPK by TGF-β; p38 MAPK is necessary for TGF-β-mediated induction of MMP-2 and cell invasion in PCa (Huang et al. 2005). Therefore, genistein could inhibit cancer cell invasion and metastasis by blocking the activation of p38 MAPK. We have also found that genistein downregulated the expression of MMP-9, protease M, uPAR, VEGF, neuropilin, TSP, BPGF, LPA, TGF- β 2, TSP-1, and PAR-2, which are involved in angiogenesis, tumor cell invasion, and metastasis of PCa cells (Li & Sarkar 2002b). Mechanistic studies revealed that isoflavones upregulate the expression of GSK-3 β , enhance GSK-3 β binding to β -catenin, and increase the phosphorylation of β -catenin, suggesting that isoflavones could inactivate Wnt signaling to inhibit PCa cell growth (Li et al. 2008). Other investigators have also reported that genistein diminished basal and Wnt-1-induced cell proliferation, attenuated Wnt-1 targets such as c-Myc and cyclin D1 expression (Su & Simmen 2009), and that isoflavones inhibit the expression of Wnt-5a (Su et al. 2007). Moreover, genistein could reduce Gli1 mRNA concentrations and down-regulate Gli reporter activity (Slusarz et al. 2010). These results suggest an inhibitory effect of isoflavones on Wnt and Hh signaling. In addition, genistein could also inhibit the growth of PCa cells through the induction of neuroendocrine differentiation (Pinski et al. 2006), suggesting genistein has effects on multiple signaling pathways.

Lycopene

Tomatoes are rich in lycopene, which is the pigment principally responsible for the deep-red color of tomato and its products. Tomato products including ketchup, tomato juice, and pizza sauce are the richest sources of lycopene in the US diet. The consumption of tomatoes and tomato products containing lycopene is associated with a decreased risk of chronic diseases including cardiovascular diseases and cancers. Lycopene is a potent antioxidant, and it has been established that lycopene is a biologically occurring carotenoid, which exhibits a high physical quenching rate constant with singlet oxygen, suggesting its high activity as an

antioxidant. Giovannucci *et al.* (2002) reported that frequent consumption of tomato products is associated with a lower risk of PCa. Inverse associations between plasma lycopene and PCa have also been reported (Gann *et al.* 1999, Lu *et al.* 2001). Experimental studies have also shown that lycopene inhibits cell growth in breast, prostate, and endometrial cancer cells by regulation of cell cycle-related genes (Nahum *et al.* 2001, Kim *et al.* 2002, Bureyko *et al.* 2009). An in vivo animal study showed that lycopene had anti-tumor effects that could be potentiated by vitamin E, an antioxidant that is also present in tomatoes (Limpens *et al.* 2004), which confirmed the anticancer activity of lycopene. A phase II clinical trial from our group has shown that lycopene supplements reduced tumor size and PSA level in localized PCa (Kucuk *et al.* 2001, 2002), suggesting a promising effect in PCa prevention and/or treatment. Another clinical trial with lycopene intake also showed a small but statistically significant reduction in serum PSA as shown below. In addition, compared with pre-intervention levels, the oxidative DNA damage in both leukocyte and prostate was significantly reduced after intervention (Chen *et al.* 2001), further suggesting an antioxidant effect for lycopene.

The molecular targets of lycopene include NF-κB, AR, Akt, Wnt and MAPK (Table 1). It has been found that lycopene significantly inhibited the DNA-binding activity of NF- κ B and the expression of the NF-KB target gene MMP-9, leading to inhibition of the invasion of cancer cells (Huang et al. 2007). The inhibition of NF-kB DNA-binding activity by lycopene was mediated through the down-regulation of IκB phosphorylation, NF-κB expression, and NFκB p65 subunit translocation from the cytosol to the nucleus (Huang et al. 2007). Another study showed that pretreatment with lycopene markedly inhibited the lipopolysaccharide (LPS)-induced up-regulation of p-ERK, p-p38, p-JNK, and NF-kB (Kim et al. 2004), suggesting an inhibitory effect by lycopene on MAPK and NF-KB signaling pathways. Lycopene also showed an inhibitory effect on Akt signaling and cell proliferation (Tang et al. 2008). Lycopene treatment suppressed Akt activation, suppressed non-phosphorylated activated β -catenin, increased the phosphorylated form of β -catenin proteins and increased the expression of p27Kip1 (Tang et al. 2008). Lycopene also induced apoptosis through downregulation of pAkt, cyclin D1, and pBad (Palozza et al. 2005), suggesting an inhibitory effect on Akt signaling. It has been reported that lycopene inhibits IGF1-mediated Akt and AR signaling in rat PCa (Liu *et al.* 2008). Lycopene reduced AR and β -catenin nuclear localization, and inhibited IGF1-stimulated PCa growth, perhaps by attenuating the effects of IGF1 on phosphorylation of Akt and GSK38. These results suggest the broad effects of lycopene on multiple signaling pathways.

Green tea

Consumption of green tea has been associated with human health including the prevention of cancer and heart disease. Epidemiological observations showed lower incidence of PCa among Asian men with a high dietary intake of green tea, suggesting that green tea might be a preventive agent against PCa (Jian *et al.* 2007). The result from Japan's Public Health Centerbased Prospective Study showed that the consumption of green tea was associated with a dosedependent decrease in the risk of advanced PCa (Kurahashi *et al.* 2008). A clinical trial with oral administration of green tea catechins (GTC) showed that GTC decreased PSA levels and that the effect of PCa prevention by GTC was long lasting (Bettuzzi *et al.* 2006, Brausi *et al.* 2008). Green tea and its constituents have been studied both *in vitro* and *in vivo*. Green tea contains several catechins including epicatechin (EC), epigallocatechin (EGC), epicatechin-3-gallate (ECG), and epigallocatechin-3-gallate (EGCG). EGCG is believed to be the most potent among these catechins for the inhibition of oncogenesis and reduction of oxidative stress (Syed *et al.* 2007, 2008, Khan & Mukhtar 2008, Khan *et al.* 2009, Johnson *et al.* 2010). These results clearly suggest that further clinical trials are needed to appreciate the value of green tea and its active components in PCa prevention and/or treatment.

The inhibitory effects of EGCG on AR signaling have been reported in PCa in vitro and in vivo (Table 1). EGCG inhibited LNCaP cell growth and the expression of AR at both mRNA and protein levels (Ren et al. 2000). Moreover, EGCG showed a significant inhibitory effect on androgen-induced PSA promoter-mediated expression of PSA. It has been reported that EGCG treatment resulted in a significant dose- and time-dependent inhibition of the activation and translocation of NF- κ B to the nucleus by suppressing the degradation of I κ B α in the cytoplasm (Ahmad et al. 2000, Khan & Mukhtar 2008). EGCG has also been shown to inhibit the activation of IKK and the phosphorylation of IkBa (Yang et al. 2001, Chen et al. 2002). EGCG has been found to inhibit PI3K/Akt activation which, in turn, resulted in the modulation of the Bcl-2 family of proteins, leading to enhanced apoptosis of bladder cancer cells (Qin et al. 2007). EGCG also inhibited VEGF-induced angiogenesis in vitro through suppression of VE-cadherin phosphorylation and inactivation of Akt, suggesting an inhibitory effect of EGCG on the Akt signaling pathway (Tang et al. 2003). The Wnt and Hh signaling pathways have also been found to be inhibited by EGCG in a dose-dependent manner in cancer cells (Kim et al. 2006, Slusarz et al. 2010). EGCG treatment induced transcription of HBP1, which is a suppressor of Wnt signaling. It has been found that EGCG treatment resulted in a dosedependent increase of p53 in LNCaP cells which carry the wild-type p53 gene, but not in DU145 cells carrying a mutant p53 (Gupta et al. 2000). EGCG also induced stabilization of p53 and caused an up-regulation of its transcriptional activity, thereby causing activation of its downstream targets such as p21^{WAF1} and Bax, resulting in the induction of apoptosis.

Other agents

DIM is the dimeric product of indole-3-carbinol (I3C), which is produced from naturally occurring glucosinolates contained in a wide variety of plants, including members of the family Cruciferae. Under the acidic conditions of the stomach, I3C undergoes extensive and rapid self-condensation reactions to form several derivatives. DIM is believed to be the major derivative and condensation product of I3C. Epidemiological studies indicate that human exposure to indoles through cruciferous vegetable consumption could decrease cancer risk (Higdon et al. 2007). DIM has been shown to reduce oxidative stress and stimulate antioxidant response element-driven gene expression, suggesting that indole compounds have an antioxidant function (Benabadji et al. 2004, Nho & Jeffery 2004). An animal study showed that DIM is not toxic and has an *in vivo* preventive effect against the development of PCa in a mouse model (Fares et al. 2009). Furthermore, several experimental studies have shown that DIM inhibited oncogenesis and cancer cell growth, and induced apoptosis in PCa cells in vitro and in vivo, suggesting that DIM could serve as a potent agent for the prevention and/or treatment of cancers (Nachshon-Kedmi et al. 2004, Garikapaty et al. 2006). We and other investigators have also investigated the molecular targets of DIM. It has been found that DIM is a potent inhibitor of the AR (Le et al. 2003, Bhuiyan et al. 2006), suggesting it has inhibitory effects on PCa cell growth. Moreover, DIM also regulated Akt, Wnt, NF-KB, VEGF, and uPA (Le et al. 2003, Garikapaty et al. 2006, Kong et al. 2007, 2008, Ahmad et al. 2009), suggesting multiple molecular targets (Table 1).

Curcumin is another bioactive compound found in *Curcuma longa* (turmeric). Turmeric extract from the rhizomes, commonly called curcuminoids, is mainly composed of curcumin. Curcumin-related research has received considerable attention due to its pronounced antiinflammatory, anti-oxidative, immunomodulating, anti-atherogenic, and anti-carcinogenic activities (Miquel *et al.* 2002, Banerjee *et al.* 2003). It has been reported that curcumin inhibits IKK, suppresses both constitutive and inducible NF- κ B activation, and potentiates TNFinduced apoptosis (Bharti *et al.* 2003). Curcumin showed strong antioxidant and anticancer properties through regulating the expression of genes that require the activation of AP1 and NF- κ B (Duvoix *et al.* 2003). We and other investigators have found that curcumin inhibited Notch, Akt, Hh, and Wnt signaling (Wang *et al.* 2006*b*, Ryu *et al.* 2008, Yu *et al.* 2008, Slusarz

et al. 2010). In addition, a number of curcumin analogs have been identified as potential AR antagonists in the presence of the AR and the AR co-activator, ARA70 (Ohtsu *et al.* 2002). Considering the effects of curcumin on NF- κ B, Akt, Notch, Wnt, Hh, and AR signaling, curcumin could, in the future, be a non-toxic alternative for PCa prevention and/or treatment; indeed, making improvements in the bioavailability of curcumin and its analogues is an active area of research.

Many other botanical compounds have been proposed for the prevention of cancer. It was found that apigenin, baicalein, quercetin, and resveratrol with IC₅₀ values ranging from <1 to 25 μ M could inhibit *Gli1* mRNA concentrations by up to 95%, suggesting an inhibitory effect on Hh signaling (Slusarz *et al.* 2010). These compounds also reduced or delayed PCa *in vivo* in TRAMP mice, consistent with *in vitro* data showing that these compounds inhibited the growth of human and mouse PCa cell lines, suggesting a potential role in the prevention of PCa (Slusarz *et al.* 2010); further in-depth investigation in pre-clinical models and human clinical trials is warranted.

Conclusions and perspectives

In conclusion, PCa cells are known to have alterations in multiple cellular signaling pathways, among which the AR, Akt, NF- κ B, Wnt, Hh, and Notch pathways appear more important than others. The alterations in these pathways could occur at different stages of PCa. Therefore, novel strategies targeting these important pathways could be promising for the prevention of PCa and its metastases. So far, the preventive effects of 5- α -reductase inhibitors show more promising results for the prevention of PCa; however, it is important to note that natural agents such as soy isoflavones, lycopene, EGCG, DIM, and curcumin which target multiple pathways could be useful for the prevention of tumor progression and/or treatment, which further suggests that agents harvested from the bounties of nature could be useful either alone or in combination with targeted or non-targeted conventional preventive or therapeutic agents for the prevention and/or treatment of PCa. Further in-depth mechanistic *in vitro* studies and animal model *in vivo* studies together with intelligently designed clinical trials are needed to fully appreciate the value of natural products for the prevention and/or treatment of human PCa.

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References

- Adhami VM, Siddiqui IA, Ahmad N, Gupta S, Mukhtar H. Oral consumption of green tea polyphenols inhibits insulin-like growth factor-I-induced signaling in an autochthonous mouse model of prostate cancer. Cancer Research 2004;64:8715–8722. [PubMed: 15574782]
- Adhami VM, Siddiqui IA, Sarfaraz S, Khwaja SI, Hafeez BB, Ahmad N, Mukhtar H. Effective prostate cancer chemopreventive intervention with green tea polyphenols in the TRAMP model depends on the stage of the disease. Clinical Cancer Research 2009;15:1947–1953. [PubMed: 19276266]
- Adlercreutz H, Honjo H, Higashi A, Fotsis T, Hamalainen E, Hasegawa T, Okada H. Urinary excretion of lignans and isoflavonoid phytoestrogens in Japanese men and women consuming a traditional Japanese diet. American Journal of Clinical Nutrition 1991;54:1093–1100. [PubMed: 1659780]

- Adlercreutz H, Markkanen H, Watanabe S. Plasma concentrations of phyto-oestrogens in Japanese men. Lancet 1993;342:1209–1210. [PubMed: 7901532]
- Ahmad N, Gupta S, Mukhtar H. Green tea polyphenol epigallocatechin-3-gallate differentially modulates nuclear factor kappaB in cancer cells versus normal cells. Archives of Biochemistry and Biophysics 2000;376:338–346. [PubMed: 10775421]
- Ahmad A, Kong D, Sarkar SH, Wang Z, Banerjee S, Sarkar FH. Inactivation of uPA and its receptor uPAR by 3,3'-diindolylmethane (DIM) leads to the inhibition of prostate cancer cell growth and migration. Journal of Cellular Biochemistry 2009;107:516–527. [PubMed: 19330806]
- Ahn J, Albanes D, Peters U, Schatzkin A, Lim U, Freedman M, Chatterjee N, Andriole GL, Leitzmann MF, Hayes RB. Dairy products, calcium intake, and risk of prostate cancer in the prostate, lung, colorectal, and ovarian cancer screening trial. Cancer Epidemiology, Biomarkers & Prevention 2007;16:2623–2630.
- Albanes D, Heinonen OP, Huttunen JK, Taylor PR, Virtamo J, Edwards BK, Haapakoski J, Rautalahti M, Hartman AM, Palmgren J, et al. Effects of alpha-tocopherol and beta-carotene supplements on cancer incidence in the Alpha-Tocopherol Beta-Carotene Cancer Prevention Study. American Journal of Clinical Nutrition 1995;62 1427S–1430S.
- Alessi DR, Andjelkovic M, Caudwell B, Cron P, Morrice N, Cohen P, Hemmings BA. Mechanism of activation of protein kinase B by insulin and IGF-1. EMBO Journal 1996;15:6541–6551. [PubMed: 8978681]
- American Cancer Society. Cancer Facts & Figures 2009. Atlanta: American Cancer Society Inc; 2009.
- Amin AR, Kucuk O, Khuri FR, Shin DM. Perspectives for cancer prevention with natural compounds. Journal of Clinical Oncology 2009;27:2712–2725. [PubMed: 19414669]
- Andriole G, Bostwick D, Brawley O, Gomella L, Marberger M, Tindall D, Breed S, Somerville M, Rittmaster R. Chemoprevention of prostate cancer in men at high risk: rationale and design of the Reduction by Dutasteride of Prostate Cancer Events (REDUCE) trial. Journal of Urology 2004a; 172:1314–1317. [PubMed: 15371831]
- Andriole GL, Roehrborn C, Schulman C, Slawin KM, Somerville M, Rittmaster RS. Effect of dutasteride on the detection of prostate cancer in men with benign prostatic hyperplasia. Urology 2004b;64:537– 541. [PubMed: 15351586]
- Angers S, Moon RT. Proximal events in Wnt signal transduction. *Nature Reviews*. Molecular Cell Biology 2009;10:468–477. [PubMed: 19536106]
- Antón Aparicio LM, Garcia CR, Cassinello EJ, Valladares AM, Reboredo LM, Diaz PS, Aparicio GG. Prostate cancer and Hedgehog signalling pathway. Clinical & Translational Oncology 2007;9:420– 428. [PubMed: 17652055]
- Azzi A, Gysin R, Kempna P, Munteanu A, Negis Y, Villacorta L, Visarius T, Zingg JM. Vitamin E mediates cell signaling and regulation of gene expression. Annals of the New York Academy of Sciences 2004;1031:86–95. [PubMed: 15753136]
- Banerjee M, Tripathi LM, Srivastava VM, Puri A, Shukla R. Modulation of inflammatory mediators by ibuprofen and curcumin treatment during chronic inflammation in rat. Immunopharmacology and Immunotoxicology 2003;25:213–224. [PubMed: 12784914]
- Banks R, Speakman JR, Selman C. Vitamin E supplementation and mammalian lifespan. Molecular Nutrition & Food Research 2010;54:719–725. [PubMed: 20205192]
- Barker N, Clevers H. Mining the Wnt pathway for cancer therapeutics *Nature Reviews*. Drug Discovery 2006;5:997–1014. [PubMed: 17139285]
- Behrens J. Control of beta-catenin signaling in tumor development. Annals of the New York Academy of Sciences 2000;910:21–33. [PubMed: 10911903]
- Benabadji SH, Wen R, Zheng JB, Dong XC, Yuan SG. Anticarcinogenic and antioxidant activity of diindolylmethane derivatives. Acta Pharmacologica Sinica 2004;25:666–671. [PubMed: 15132835]
- Bettuzzi S, Brausi M, Rizzi F, Castagnetti G, Peracchia G, Corti A. Chemoprevention of human prostate cancer by oral administration of green tea catechins in volunteers with high-grade prostate intraepithelial neoplasia: a preliminary report from a one-year proof-of-principle study. Cancer Research 2006;66:1234–1240. [PubMed: 16424063]
- Bharti AC, Aggarwal BB. Nuclear factor-kappa B and cancer: its role in prevention and therapy. Biochemical Pharmacology 2002;64:883–888. [PubMed: 12213582]

- Bharti AC, Donato N, Singh S, Aggarwal BB. Curcumin (diferuloylmethane) down-regulates the constitutive activation of nuclear factor-kappa B and IkappaBalpha kinase in human multiple myeloma cells, leading to suppression of proliferation and induction of apoptosis. Blood 2003;101:1053–1062. [PubMed: 12393461]
- Bhuiyan MM, Li Y, Banerjee S, Ahmed F, Wang Z, Ali S, Sarkar FH. Down-regulation of androgen receptor by 3,3'-diindolylmethane contributes to inhibition of cell proliferation and induction of apoptosis in both hormone-sensitive LNCaP and insensitive C4-2B prostate cancer cells. Cancer Research 2006;66:10064–10072. [PubMed: 17047070]
- Biancolella M, Valentini A, Minella D, Vecchione L, D'Amico F, Chillemi G, Gravina P, Bueno S, Prosperini G, Desideri A, et al. Effects of dutasteride on the expression of genes related to androgen metabolism and related pathway in human prostate cancer cell lines. Investigational New Drugs 2007;25:491–497. [PubMed: 17636412]
- Bin HB, Adhami VM, Asim M, Siddiqui IA, Bhat KM, Zhong W, Saleem M, Din M, Setaluri V, Mukhtar H. Targeted knockdown of Notch1 inhibits invasion of human prostate cancer cells concomitant with inhibition of matrix metalloproteinase-9 and urokinase plasminogen activator. Clinical Cancer Research 2009;15:452–459. [PubMed: 19147749]
- Biswas DK, Dai SC, Cruz A, Weiser B, Graner E, Pardee AB. The nuclear factor kappa B (NF-kappa B): a potential therapeutic target for estrogen receptor negative breast cancers. PNAS 2001;98:10386–10391. [PubMed: 11517301]
- Bracarda S, de CO, Greco C, Prayer-Galetti T, Valdagni R, Gatta G, de BF, Bartsch G. Cancer of the prostate. Critical Reviews in Oncology/Hematology 2005;56:379–396. [PubMed: 16310371]
- Brausi M, Rizzi F, Bettuzzi S. Chemoprevention of human prostate cancer by green tea catechins: two years later. A follow-up update. European Urology 2008;54:472–473. [PubMed: 18406041]
- Brunet A, Bonni A, Zigmond MJ, Lin MZ, Juo P, Hu LS, Anderson MJ, Arden KC, Blenis J, Greenberg ME. Akt promotes cell survival by phosphorylating and inhibiting a Forkhead transcription factor. Cell 1999;96:857–868. [PubMed: 10102273]
- Bureyko T, Hurdle H, Metcalfe JB, Clandinin MT, Mazurak VC. Reduced growth and integrin expression of prostate cells cultured with lycopene, vitamin E and fish oil *in vitro*. British Journal of Nutrition 2009;101:990–997. [PubMed: 18718045]
- Cardone MH, Roy N, Stennicke HR, Salvesen GS, Franke TF, Stanbridge E, Frisch S, Reed JC. Regulation of cell death protease caspase-9 by phosphorylation. Science 1998;282:1318–1321. [PubMed: 9812896]
- Chen L, Stacewicz-Sapuntzakis M, Duncan C, Sharifi R, Ghosh L, van BR, Ashton D, Bowen PE. Oxidative DNA damage in prostate cancer patients consuming tomato sauce-based entrees as a whole-food intervention. Journal of the National Cancer Institute 2001;93:1872–1879. [PubMed: 11752012]
- Chen PC, Wheeler DS, Malhotra V, Odoms K, Denenberg AG, Wong HR. A green tea-derived polyphenol, epigallocatechin-3-gallate, inhibits IkappaB kinase activation and IL-8 gene expression in respiratory epithelium. Inflammation 2002;26:233–241. [PubMed: 12238566]
- Chen G, Geng J, Zhang YF. Mechanism of inhibiting the proliferation of prostate cancer by finasteride: a study using cDNA microarray. Zhonghua Yi Xue Za Zhi 2005;85:1489–1492. [PubMed: 16061029]
- Chesire DR, Ewing CM, Gage WR, Isaacs WB. *In vitro* evidence for complex modes of nuclear betacatenin signaling during prostate growth and tumorigenesis. Oncogene 2002;21:2679–2694. [PubMed: 11965541]
- Christensen MJ, Nartey ET, Hada AL, Legg RL, Barzee BR. High selenium reduces NF-kappaBregulated gene expression in uninduced human prostate cancer cells. Nutrition and Cancer 2007;58:197–204. [PubMed: 17640166]
- Cohen YC, Liu KS, Heyden NL, Carides AD, Anderson KM, Daifotis AG, Gann PH. Detection bias due to the effect of finasteride on prostate volume: a modeling approach for analysis of the Prostate Cancer Prevention Trial. Journal of the National Cancer Institute 2007;99:1366–1374. [PubMed: 17848668]
- Crawford ED, Andriole GL, Marberger M, Rittmaster RS. Reduction in the risk of prostate cancer: future directions after the Prostate Cancer Prevention Trial. Urology 2010;75:502–509. [PubMed: 20035983]

- Cronauer MV, Schulz WA, Ackermann R, Burchardt M. Effects of WNT/beta-catenin pathway activation on signaling through T-cell factor and androgen receptor in prostate cancer cell lines. International Journal of Oncology 2005;26:1033–1040. [PubMed: 15753999]
- Dihlmann S, von Knebel DM. Wnt/beta-catenin-pathway as a molecular target for future anti-cancer therapeutics. International Journal of Cancer 2005;113:515–524.
- Dong Y, Zhang H, Hawthorn L, Ganther HE, Ip C. Delineation of the molecular basis for seleniuminduced growth arrest in human prostate cancer cells by oligonucleotide array. Cancer Research 2003;63:52–59. [PubMed: 12517777]
- Donovan MJ, Osman I, Khan FM, Vengrenyuk Y, Capodieci P, Koscuiszka M, Anand A, Cordon-Cardo C, Costa J, Scher HI. Androgen receptor expression is associated with prostate cancer-specific survival in castrate patients with metastatic disease. BJU International 2010;105:462–467. [PubMed: 19624594]
- Duffield-Lillico AJ, Dalkin BL, Reid ME, Turnbull BW, Slate EH, Jacobs ET, Marshall JR, Clark LC. Selenium supplementation, baseline plasma selenium status and incidence of prostate cancer: an analysis of the complete treatment period of the Nutritional Prevention of Cancer Trial. BJU International 2003;91:608–612. [PubMed: 12699469]
- Duque JL, Loughlin KR, Adam RM, Kantoff PW, Zurakowski D, Freeman MR. Plasma levels of vascular endothelial growth factor are increased in patients with metastatic prostate cancer. Urology 1999;54:523–527. [PubMed: 10475365]
- Duvoix A, Morceau F, Delhalle S, Schmitz M, Schneken-burger M, Galteau MM, Dicato M, Diederich M. Induction of apoptosis by curcumin: mediation by glutathione S-transferase P1-1 inhibition. Biochemical Pharmacology 2003;66:1475–1483. [PubMed: 14555224]
- Fares F, Azzam N, Appel B, Fares B, Stein A. The potential efficacy of 3,3'-diindolylmethane in prevention of prostate cancer development. European Journal of Cancer Prevention 2009;19:199– 203. [PubMed: 20010430]
- Gann PH, Ma J, Giovannucci E, Willett W, Sacks FM, Hennekens CH, Stampfer MJ. Lower prostate cancer risk in men with elevated plasma lycopene levels: results of a prospective analysis. Cancer Research 1999;59:1225–1230. [PubMed: 10096552]
- Garikapaty VP, Ashok BT, Tadi K, Mittelman A, Tiwari RK. 3,3'-Diindolylmethane downregulates prosurvival pathway in hormone independent prostate cancer. Biochemical and Biophysical Research Communications 2006;340:718–725. [PubMed: 16380095]
- Garland CF, Garland FC, Gorham ED, Lipkin M, Newmark H, Mohr SB, Holick MF. The role of vitamin D in cancer prevention. American Journal of Public Health 2006;96:252–261. [PubMed: 16380576]
- Gianduzzo TR, Holmes EG, Tinggi U, Shahin M, Mactaggart P, Nicol D. Prostatic and peripheral blood selenium levels after oral supplementation. Journal of Urology 2003;170:870–873. [PubMed: 12913719]
- Giovannucci E, Rimm EB, Liu Y, Stampfer MJ, Willett WC. A prospective study of tomato products, lycopene, and prostate cancer risk. Journal of the National Cancer Institute 2002;94:391–398. [PubMed: 11880478]
- Goodman GE, Schaffer S, Omenn GS, Chen C, King I. The association between lung and prostate cancer risk, and serum micronutrients: results and lessons learned from beta-carotene and retinol efficacy trial. Cancer Epidemiology, Biomarkers & Prevention 2003;12:518–526.
- Gupta S, Ahmad N, Nieminen AL, Mukhtar H. Growth inhibition, cell-cycle dysregulation, and induction of apoptosis by green tea constituent (—)-epigallocate-chin-3-gallate in androgen-sensitive and androgen-insen-sitive human prostate carcinoma cells. Toxicology and Applied Pharmacology 2000;164:82–90. [PubMed: 10739747]
- Gupta D, Lammersfeld CA, Trukova K, Lis CG. Vitamin D and prostate cancer risk: a review of the epidemiological literature. Prostate Cancer and Prostatic Diseases 2009;12:215–226. [PubMed: 19350051]
- Haefner B. NF-kappa B: arresting a major culprit in cancer. Drug Discovery Today 2002;7:653–663. [PubMed: 12110242]
- Hebert JR, Hurley TG, Olendzki BC, Teas J, Ma Y, Hampl JS. Nutritional and socioeconomic factors in relation to prostate cancer mortality: a cross-national study. Journal of the National Cancer Institute 1998;90:1637–1647. [PubMed: 9811313]

- Heinlein CA, Chang C. Androgen receptor (AR) coregulators: an overview. Endocrine Reviews 2002;23:175–200. [PubMed: 11943742]
- Heinlein CA, Chang C. Androgen receptor in prostate cancer. Endocrine Reviews 2004;25:276–308. [PubMed: 15082523]
- Higdon JV, Delage B, Williams DE, Dashwood RH. Cruciferous vegetables and human cancer risk: epidemiologic evidence and mechanistic basis. Pharmacological Research 2007;55:224–236. [PubMed: 17317210]
- Huang S, Pettaway CA, Uehara H, Bucana CD, Fidler IJ. Blockade of NF-kappaB activity in human prostate cancer cells is associated with suppression of angiogenesis, invasion, and metastasis. Oncogene 2001;20:4188–4197. [PubMed: 11464285]
- Huang X, Chen S, Xu L, Liu Y, Deb DK, Platanias LC, Bergan RC. Genistein inhibits p38 map kinase activation, matrix metalloproteinase type 2, and cell invasion in human prostate epithelial cells. Cancer Research 2005;65:3470–3478. [PubMed: 15833883]
- Huang CS, Fan YE, Lin CY, Hu ML. Lycopene inhibits matrix metalloproteinase-9 expression and downregulates the binding activity of nuclear factor-kappa B and stimulatory protein-1. Journal of Nutritional Biochemistry 2007;18:449–456. [PubMed: 17049831]
- Ismail HA, Lessard L, Mes-Masson AM, Saad F. Expression of NF-kappaB in prostate cancer lymph node metastases. Prostate 2004;58:308–313. [PubMed: 14743471]
- Jacobsen BK, Knutsen SF, Fraser GE. Does high soy milk intake reduce prostate cancer incidence? The Adventist Health Study (United States). Cancer Causes & Control 1998;9:553–557. [PubMed: 10189040]
- Jemal A, Siegel R, Ward E, Hao Y, Xu J, Thun MJ. Cancer statistics, 2009. CA: A Cancer Journal for Clinicians 2009;59:225–249. [PubMed: 19474385]
- Jian L, Lee AH, Binns CW. Tea and lycopene protect against prostate cancer. Asia Pacific Journal of Clinical Nutrition 2007;16 Supplement 1:453–457. [PubMed: 17392149]
- Johnson JJ, Bailey HH, Mukhtar H. Green tea polyphenols for prostate cancer chemoprevention: a translational perspective. Phytomedicine 2010;17:3–13. [PubMed: 19959000]
- Kaplan SA, Roehrborn CG, Meehan AG, Liu KS, Carides AD, Binkowitz BS, Heyden NL, Vaughan ED Jr. PCPT: evidence that finasteride reduces risk of most frequently detected intermediate- and highgrade (Gleason score 6 and 7) cancer. Urology 2009;73:935–939. [PubMed: 19328538]
- Karin M, Cao Y, Greten FR, Li ZW. NF-kappaB in cancer: from innocent bystander to major culprit. *Nature Reviews*. Cancer 2002;2:301–310. [PubMed: 12001991]
- Khan N, Mukhtar H. Multitargeted therapy of cancer by green tea polyphenols. Cancer Letters 2008;269:269–280. [PubMed: 18501505]
- Khan N, Adhami VM, Mukhtar H. Review: green tea polyphenols in chemoprevention of prostate cancer: preclinical and clinical studies. Nutrition and Cancer 2009;61:836–841. [PubMed: 20155624]
- Kim L, Rao AV, Rao LG. Effect of lycopene on prostate LNCaP cancer cells in culture. Journal of Medicinal Food 2002;5:181–187. [PubMed: 12639392]
- Kim GY, Kim JH, Ahn SC, Lee HJ, Moon DO, Lee CM, Park YM. Lycopene suppresses the lipopolysaccharide-induced phenotypic and functional maturation of murine dendritic cells through inhibition of mitogen-activated protein kinases and nuclear factor-kappaB. Immunology 2004;113:203–211. [PubMed: 15379981]
- Kim J, Zhang X, Rieger-Christ KM, Summerhayes IC, Wazer DE, Paulson KE, Yee AS. Suppression of Wnt signaling by the green tea compound (—)-epigallocatechin 3-gallate (EGCG) in invasive breast cancer cells. Requirement of the transcriptional repressor HBP1. Journal of Biological Chemistry 2006;281:10865–10875. [PubMed: 16495219]
- Klein EA. Selenium: epidemiology and basic science. Journal of Urology 2004;171:S50–S53. [PubMed: 14713754]
- Klein EA, Lippman SM, Thompson IM, Goodman PJ, Albanes D, Taylor PR, Coltman C. The selenium and vitamin E cancer prevention trial. World Journal of Urology 2003;21:21–27. [PubMed: 12756490]
- Kong D, Li Y, Wang Z, Banerjee S, Sarkar FH. Inhibition of angiogenesis and invasion by 3,3'-diindolylmethane is mediated by the nuclear factor-kappaB downstream target genes MMP-9 and uPA that

regulated bioavailability of vascular endothelial growth factor in prostate cancer. Cancer Research 2007;67:3310–3319. [PubMed: 17409440]

- Kong D, Banerjee S, Huang W, Li Y, Wang Z, Kim HR, Sarkar FH. Mammalian target of rapamycin repression by 3,3' -diindolylmethane inhibits invasion and angiogenesis in platelet-derived growth factor-D-overexpressing PC3 cells. Cancer Research 2008;68:1927–1934. [PubMed: 18339874]
- Kucuk O, Sarkar FH, Sakr W, Djuric Z, Pollak MN, Khachik F, Li YW, Banerjee M, Grignon D, Bertram JS, et al. Phase II randomized clinical trial of lycopene supplementation before radical prostatectomy. Cancer Epidemiology, Biomarkers & Prevention 2001;10:861–868.
- Kucuk O, Sarkar FH, Djuric Z, Sakr W, Pollak MN, Khachik F, Banerjee M, Bertram JS, Wood DP Jr. Effects of lycopene supplementation in patients with localized prostate cancer. Experimental Biology and Medicine 2002;227:881–885. [PubMed: 12424329]
- Kupelian P, Katcher J, Levin H, Zippe C, Klein E. Correlation of clinical and pathologic factors with rising prostate-specific antigen profiles after radical prostatectomy alone for clinically localized prostate cancer. Urology 1996;48:249–260. [PubMed: 8753737]
- Kurahashi N, Sasazuki S, Iwasaki M, Inoue M, Tsugane S. Green tea consumption and prostate cancer risk in Japanese men: a prospective study. American Journal of Epidemiology 2008;167:71–77. [PubMed: 17906295]
- Le HT, Schaldach CM, Firestone GL, Bjeldanes LF. Plant-derived 3,3'-diindolylmethane is a strong androgen antagonist in human prostate cancer cells. Journal of Biological Chemistry 2003;278:21136–21145. [PubMed: 12665522]
- Lee HJ, Chang C. Recent advances in androgen receptor action. Cellular and Molecular Life Sciences 2003;60:1613–1622. [PubMed: 14504652]
- Lee MM, Wang RT, Hsing AW, Gu FL, Wang T, Spitz M. Case-control study of diet and prostate cancer in China. Cancer Causes & Control 1998;9:545–552. [PubMed: 10189039]
- Legg RL, Tolman JR, Lovinger CT, Lephart ED, Setchell KD, Christensen MJ. Diets high in selenium and isoflavones decrease androgen-regulated gene expression in healthy rat dorsolateral prostate. Reproductive Biology and Endocrinology 2008;6:57. [PubMed: 19025659]
- Li J, Kim J. Molecular profiles of finasteride effects on prostate carcinogenesis. Cancer Prevention Research 2009;2:518–524. [PubMed: 19491289]
- Li Y, Sarkar FH. Inhibition of nuclear factor kappaB activation in PC3 cells by genistein is mediated via Akt signaling pathway. Clinical Cancer Research 2002a;8:2369–2377. [PubMed: 12114442]
- Li Y, Sarkar FH. Gene expression profiles of genistein-treated PC3 prostate cancer cells. Journal of Nutrition 2002b;132:3623–3631. [PubMed: 12468598]
- Li Q, Verma IM. NF-kappaB regulation in the immune system. *Nature Reviews*. Immunology 2002;2:725–734. [PubMed: 12360211]
- Li Y, Li X, Sarkar FH. Gene expression profiles of I3C- and DIM-treated PC3 human prostate cancer cells determined by cDNA microarray analysis. Journal of Nutrition 2003;133:1011–1019. [PubMed: 12672912]
- Li Y, Wang Z, Kong D, Murthy S, Dou QP, Sheng S, Reddy GP, Sarkar FH. Regulation of FOXO3a/ beta-catenin/GSK-3beta signaling by 3,3' -diindolylmethane contributes to inhibition of cell proliferation and induction of apoptosis in prostate cancer cells. Journal of Biological Chemistry 2007;282:21542–21550. [PubMed: 17522055]
- Li Y, Wang Z, Kong D, Li R, Sarkar SH, Sarkar FH. Regulation of Akt/FOXO3a/GSK-3beta/AR signaling network by isoflavone in prostate cancer cells. Journal of Biological Chemistry 2008;283:27707–27716. [PubMed: 18687691]
- Limpens J, van Weerden WM, Kramer K, Pallapies D, Obermuller-Jevic UC, Schroder FH. Re: prostate carcinogenesis in *N*-methyl-*N*-nitrosourea (NMU)-testo-sterone-treated rats fed tomato powder, lycopene, or energy-restricted diets. Journal of the National Cancer Institute 2004;96:554–555. [PubMed: 15069119]
- Lin A, Karin M. NF-kappaB in cancer: a marked target. Seminars in Cancer Biology 2003;13:107–114. [PubMed: 12654254]
- Lippman SM, Klein EA, Goodman PJ, Lucia MS, Thompson IM, Ford LG, Parnes HL, Minasian LM, Gaziano JM, Hartline JA, et al. Effect of selenium and vitamin E on risk of prostate cancer and other

cancers: the Selenium and Vitamin E Cancer Prevention Trial (SELECT). Journal of the American Medical Association 2009;301:39–51. [PubMed: 19066370]

- Liu X, Allen JD, Arnold JT, Blackman MR. Lycopene inhibits IGF-I signal transduction and growth in normal prostate epithelial cells by decreasing DHT-modulated IGF-I production in co-cultured reactive stromal cells. Carcinogenesis 2008;29:816–823. [PubMed: 18283040]
- Liu P, Cheng H, Roberts TM, Zhao JJ. Targeting the phosphoinositide 3-kinase pathway in cancer. *Nature Reviews*. Drug Discovery 2009;8:627–644. [PubMed: 19644473]
- Lu QY, Hung JC, Heber D, Go VL, Reuter VE, Cordon-Cardo C, Scher HI, Marshall JR, Zhang ZF. Inverse associations between plasma lycopene and other carotenoids and prostate cancer. Cancer Epidemiology, Biomarkers & Prevention 2001;10:749–756.
- Maillard I, Pear WS. Notch and cancer: best to avoid the ups and downs. Cancer Cell 2003;3:203–205. [PubMed: 12676578]
- Marberger M. Drug insight: 5alpha-reductase inhibitors for the treatment of benign prostatic hyperplasia. *Nature Clinical Practice*. Urology 2006;3:495–503. [PubMed: 16964191]
- McCormick DL, Rao KV, Johnson WD, Bosland MC, Lubet RA, Steele VE. Null activity of selenium and vitamin E as cancer chemopreventive agents in the rat prostate. Cancer Prevention Research 2010;3:381–392. [PubMed: 20145190]
- Medina V, Calvo MB, Díaz-Prado S, Espada J. Hedgehog signalling as a target in cancer stem cells. Clinical & Translational Oncology 2009;11:199–207. [PubMed: 19380296]
- Meyer MB, Goetsch PD, Pike JW. A downstream intergenic cluster of regulatory enhancers contributes to the induction of CYP24A1 expression by 1{alpha},25-dihydroxyvitamin D3. Journal of Biological Chemistry 2010;285:15599–15610. [PubMed: 20236932]
- Miele L, Miao H, Nickoloff BJ. NOTCH signaling as a novel cancer therapeutic target. Current Cancer Drug Targets 2006;6:313–323. [PubMed: 16848722]
- Miquel J, Bernd A, Sempere JM, az-Alperi J, Ramirez A. The curcuma antioxidants: pharmacological effects and prospects for future clinical use. A review. Archives of Gerontology and Geriatrics 2002;34:37–46. [PubMed: 14764309]
- Montgomery JS, Price DK, Figg WD. The androgen receptor gene and its influence on the development and progression of prostate cancer. Journal of Pathology 2001;195:138–146. [PubMed: 11592091]
- Morgan TM, Koreckij TD, Corey E. Targeted therapy for advanced prostate cancer: inhibition of the PI3K/Akt/mTOR pathway. Current Cancer Drug Targets 2009;9:237–249. [PubMed: 19275762]
- Musquera M, Fleshner NE, Finelli A, Zlotta AR. The REDUCE trial: chemoprevention in prostate cancer using a dual 5alpha-reductase inhibitor, dutasteride. Expert Review of Anticancer Therapy 2008;8:1073–1079. [PubMed: 18588452]
- Nachshon-Kedmi M, Fares FA, Yannai S. Therapeutic activity of 3,3'-diindolylmethane on prostate cancer in an *in vivo* model. Prostate 2004;61:153–160. [PubMed: 15305338]
- Nahum A, Hirsch K, Danilenko M, Watts CK, Prall OW, Levy J, Sharoni Y. Lycopene inhibition of cell cycle progression in breast and endometrial cancer cells is associated with reduction in cyclin D levels and retention of p27(Kip1) in the cyclin E-cdk2 complexes. Oncogene 2001;20:3428–3436. [PubMed: 11423993]
- Neshat MS, Mellinghoff IK, Tran C, Stiles B, Thomas G, Petersen R, Frost P, Gibbons JJ, Wu H, Sawyers CL. Enhanced sensitivity of PTEN-deficient tumors to inhibition of FRAP/mTOR. PNAS 2001;98:10314–10319. [PubMed: 11504908]
- Nho CW, Jeffery E. Crambene, a bioactive nitrile derived from glucosinolate hydrolysis, acts via the antioxidant response element to upregulate quinone reductase alone or synergistically with indole-3-carbinol. Toxicology and Applied Pharmacology 2004;198:40–48. [PubMed: 15207647]
- Ohtsu H, Xiao Z, Ishida J, Nagai M, Wang HK, Itokawa H, Su CY, Shih C, Chiang T, Chang E, et al. Antitumor agents. 217. Curcumin analogues as novel androgen receptor antagonists with potential as anti-prostate cancer agents. Journal of Medicinal Chemistry 2002;45:5037–5042. [PubMed: 12408714]
- Orlowski RZ, Baldwin AS Jr. NF-kappaB as a therapeutic target in cancer. Trends in Molecular Medicine 2002;8:385–389. [PubMed: 12127724]

- Ozes ON, Mayo LD, Gustin JA, Pfeffer SR, Pfeffer LM, Donner DB. NF-kappaB activation by tumour necrosis factor requires the Akt serine-threonine kinase. Nature 1999;401:82–85. [PubMed: 10485710]
- Palozza P, Sheriff A, Serini S, Boninsegna A, Maggiano N, Ranelletti FO, Calviello G, Cittadini A. Lycopene induces apoptosis in immortalized fibroblasts exposed to tobacco smoke condensate through arresting cell cycle and down-regulating cyclin D1, pAKT and pBad. Apoptosis 2005;10:1445–1456. [PubMed: 16215689]
- Peifer M, Polakis P. Wnt signaling in oncogenesis and embryogenesis a look outside the nucleus. Science 2000;287:1606–1609. [PubMed: 10733430]
- Pike JW, Meyer MB, Martowicz ML, Bishop KA, Lee SM, Nerenz RD, Goetsch PD. Emerging regulatory paradigms for control of gene expression by 1,25-dihydroxyvitamin D(3). Journal of Steroid Biochemistry and Molecular Biology. 2010
- Pinski J, Wang Q, Quek ML, Cole A, Cooc J, Danenberg K, Danenberg PV. Genistein-induced neuroendocrine differentiation of prostate cancer cells. Prostate 2006;66:1136–1143. [PubMed: 16652383]
- Qin J, Xie LP, Zheng XY, Wang YB, Bai Y, Shen HF, Li LC, Dahiya R. A component of green tea, (—)epigallocatechin-3-gallate, promotes apoptosis in T24 human bladder cancer cells via modulation of the PI3K/Akt pathway and Bcl-2 family proteins. Biochemical and Biophysical Research Communications 2007;354:852–857. [PubMed: 17266926]
- Ren F, Zhang S, Mitchell SH, Butler R, Young CY. Tea polyphenols down-regulate the expression of the androgen receptor in LNCaP prostate cancer cells. Oncogene 2000;19:1924–1932. [PubMed: 10773882]
- Renehan AG, Zwahlen M, Minder C, O'Dwyer ST, Shalet SM, Egger M. Insulin-like growth factor (IGF)-I, IGF binding protein-3, and cancer risk: systematic review and meta-regression analysis. Lancet 2004;363:1346–1353. [PubMed: 15110491]
- Reya T, Clevers H. Wnt signalling in stem cells and cancer. Nature 2005;434:843–850. [PubMed: 15829953]
- Rizzo P, Osipo C, Foreman K, Golde T, Osborne B, Miele L. Rational targeting of Notch signaling in cancer. Oncogene 2008;27:5124–5131. [PubMed: 18758481]
- Rochette-Egly C. Nuclear receptors: integration of multiple signalling pathways through phosphorylation. Cellular Signalling 2003;15:355–366. [PubMed: 12618210]
- Romashkova JA, Makarov SS. NF-kappaB is a target of AKT in anti-apoptotic PDGF signalling. Nature 1999;401:86–90. [PubMed: 10485711]
- Ryu MJ, Cho M, Song JY, Yun YS, Choi IW, Kim DE, Park BS, Oh S. Natural derivatives of curcumin attenuate the Wnt/beta-catenin pathway through down-regulation of the transcriptional coactivator p300. Biochemical and Biophysical Research Communications 2008;377:1304–1308. [PubMed: 19000900]
- Saez C, Gonzalez-Baena AC, Japon MA, Giraldez J, Segura DI, Miranda G, Rodriguez-Vallejo JM, Gonzalez-Esteban J, Torrubia F. Regressive changes in finasteride-treated human hyperplastic prostates correlate with an upregulation of TGF-beta receptor expression. Prostate 1998;37:84–90. [PubMed: 9759702]
- Santagata S, Demichelis F, Riva A, Varambally S, Hofer MD, Kutok JL, Kim R, Tang J, Montie JE, Chinnaiyan AM, et al. JAGGED1 expression is associated with prostate cancer metastasis and recurrence. Cancer Research 2004;64:6854–6857. [PubMed: 15466172]
- Sarkar FH, Li Y, Wang Z, Kong D. NF-kappaB signaling pathway and its therapeutic implications in human diseases. International Reviews of Immunology 2008;27:293–319. [PubMed: 18853341]
- Sarker D, Reid AH, Yap TA, de Bono JS. Targeting the PI3K/AKT pathway for the treatment of prostate cancer. Clinical Cancer Research 2009;15:4799–4805. [PubMed: 19638457]
- Sato N, Gleave ME, Bruchovsky N, Rennie PS, Goldenberg SL, Lange PH, Sullivan LD. Intermittent androgen suppression delays progression to androgen-independent regulation of prostate-specific antigen gene in the LNCaP prostate tumour model. Journal of Steroid Biochemistry and Molecular Biology 1996;58:139–146. [PubMed: 8809195]

- Sawaya ME, Blume-Peytavi U, Mullins DL, Nusbaum BP, Whiting D, Nicholson DW, Lotocki G, Keane RW. Effects of finasteride on apoptosis and regulation of the human hair cycle. Journal of Cutaneous Medicine and Surgery 2002;6:1–9. [PubMed: 11896416]
- Schmidt LJ, Murillo H, Tindall DJ. Gene expression in prostate cancer cells treated with the dual 5 alphareductase inhibitor dutasteride. Journal of Andrology 2004;25:944–953. [PubMed: 15477368]
- Schmidt LJ, Regan KM, Anderson SK, Sun Z, Ballman KV, Tindall DJ. Effects of the 5 alpha-reductase inhibitor dutasteride on gene expression in prostate cancer xenografts. Prostate 2009;69:1730–1743. [PubMed: 19676081]
- Shah YM, Kaul A, Dong Y, Ip C, Rowan BG. Attenuation of estrogen receptor alpha (ERalpha) signaling by selenium in breast cancer cells via downregulation of ERalpha gene expression. Breast Cancer Research and Treatment 2005;92:239–250. [PubMed: 16155795]
- Shariat SF, Anwuri VA, Lamb DJ, Shah NV, Wheeler TM, Slawin KM. Association of preoperative plasma levels of vascular endothelial growth factor and soluble vascular cell adhesion molecule-1 with lymph node status and biochemical progression after radical prostatectomy. Journal of Clinical Oncology 2004;22:1655–1663. [PubMed: 15117988]
- Shou J, Ross S, Koeppen H, de Sauvage FJ, Gao WQ. Dynamics of notch expression during murine prostate development and tumorigenesis. Cancer Research 2001;61:7291–7297. [PubMed: 11585768]
- Shukla S, MacLennan GT, Marengo SR, Resnick MI, Gupta S. Constitutive activation of PI3K-Akt and NF-kappaB during prostate cancer progression in autochthonous transgenic mouse model. Prostate 2005;64:224–239. [PubMed: 15712212]
- Slusarz A, Shenouda NS, Sakla MS, Drenkhahn SK, Narula AS, MacDonald RS, Besch-Williford CL, Lubahn DB. Common botanical compounds inhibit the Hedgehog signaling pathway in prostate cancer. Cancer Research 2010;70:3382–3390. [PubMed: 20395211]
- de Souza PL, Russell PJ, Kearsley J. Role of the Akt pathway in prostate cancer. Current Cancer Drug Targets 2009;9:163–175. [PubMed: 19275757]
- Storz P, Toker A. NF-kappaB signaling an alternate pathway for oxidative stress-responses. Cell Cycle 2003;2:9–10. [PubMed: 12695674]
- Su Y, Simmen RC. Soy isoflavone genistein upregulates epithelial adhesion molecule E-cadherin expression and attenuates beta-catenin signaling in mammary epithelial cells. Carcinogenesis 2009;30:331–339. [PubMed: 19073877]
- Su Y, Simmen FA, Xiao R, Simmen RC. Expression profiling of rat mammary epithelial cells reveals candidate signaling pathways in dietary protection from mammary tumors. Physiological Genomics 2007;30:8–16. [PubMed: 17341692]
- Suh J, Payvandi F, Edelstein LC, Amenta PS, Zong WX, Gelinas C, Rabson AB. Mechanisms of constitutive NF-kappaB activation in human prostate cancer cells. Prostate 2002;52:183–200. [PubMed: 12111695]
- Syed DN, Khan N, Afaq F, Mukhtar H. Chemoprevention of prostate cancer through dietary agents: progress and promise. Cancer Epidemiology, Biomarkers & Prevention 2007;16:2193–2203.
- Syed DN, Suh Y, Afaq F, Mukhtar H. Dietary agents for chemoprevention of prostate cancer. Cancer Letters 2008;265:167–176. [PubMed: 18395333]
- Taipale J, Beachy PA. The Hedgehog and Wnt signalling pathways in cancer. Nature 2001;411:349–354. [PubMed: 11357142]
- Tang FY, Nguyen N, Meydani M. Green tea catechins inhibit VEGF-induced angiogenesis *in vitro* through suppression of VE-cadherin phosphorylation and inactivation of Akt molecule. International Journal of Cancer 2003;106:871–878.
- Tang FY, Shih CJ, Cheng LH, Ho HJ, Chen HJ. Lycopene inhibits growth of human colon cancer cells via suppression of the Akt signaling pathway. Molecular Nutrition & Food Research 2008;52:646– 654. [PubMed: 18537129]
- Thompson TA, Wilding G. Androgen antagonist activity by the antioxidant moiety of vitamin E, 2,2,5,7,8-pentamethyl-6-chromanol in human prostate carcinoma cells. Molecular Cancer Therapeutics 2003;2:797–803. [PubMed: 12939470]

- Thompson IM, Goodman PJ, Tangen CM, Lucia MS, Miller GJ, Ford LG, Lieber MM, Cespedes RD, Atkins JN, Lippman SM, et al. The influence of finasteride on the development of prostate cancer. New England Journal of Medicine 2003;349:215–224. [PubMed: 12824459]
- Varjosalo M, Taipale J. Hedgehog: functions and mechanisms. Genes and Development 2008;22:2454– 2472. [PubMed: 18794343]
- Verras M, Sun Z. Roles and regulation of Wnt signaling and beta-catenin in prostate cancer. Cancer Letters 2006;237:22–32. [PubMed: 16023783]
- Vezina CM, Bushman AW. Hedgehog signaling in prostate growth and benign prostate hyperplasia. Current Urology Reports 2007;8:275–280. [PubMed: 18519011]
- Vunta H, Belda BJ, Arner RJ, Channa RC, Vanden Heuvel JP, Sandeep PK. Selenium attenuates proinflammatory gene expression in macrophages. Molecular Nutrition & Food Research 2008;52:1316–1323. [PubMed: 18481333]
- Wang Z, Zhang Y, Banerjee S, Li Y, Sarkar FH. Inhibition of nuclear factor kappab activity by genistein is mediated via Notch-1 signaling pathway in pancreatic cancer cells. International Journal of Cancer 2006a;118:1930–1936.
- Wang Z, Zhang Y, Banerjee S, Li Y, Sarkar FH. Notch-1 down-regulation by curcumin is associated with the inhibition of cell growth and the induction of apoptosis in pancreatic cancer cells. Cancer 2006b;106:2503–2513. [PubMed: 16628653]
- Wang Z, Li Y, Banerjee S, Kong D, Ahmad A, Nogueira V, Hay N, Sarkar FH. Down-regulation of Notch-1 and Jagged-1 inhibits prostate cancer cell growth, migration and invasion, and induces apoptosis via inactivation of Akt, mTOR, and NF-kappaB signaling pathways. Journal of Cellular Biochemistry 2010;109:726–736. [PubMed: 20052673]
- Waters DJ, Shen S, Cooley DM, Bostwick DG, Qian J, Combs GF Jr, Glickman LT, Oteham C, Schlittler D, Morris JS. Effects of dietary selenium supplementation on DNA damage and apoptosis in canine prostate. Journal of the National Cancer Institute 2003;95:237–241. [PubMed: 12569146]
- Wen Y, Hu MC, Makino K, Spohn B, Bartholomeusz G, Yan DH, Hung MC. HER-2/neu promotes androgen-independent survival and growth of prostate cancer cells through the Akt pathway. Cancer Research 2000;60:6841–6845. [PubMed: 11156376]
- Whittemore AS, Kolonel LN, Wu AH, John EM, Gallagher RP, Howe GR, Burch JD, Hankin J, Dreon DM, West DW, et al. Prostate cancer in relation to diet, physical activity, and body size in blacks, whites, and Asians in the United States and Canada. Journal of the National Cancer Institute 1995;87:652–661. [PubMed: 7752270]
- Yamamoto Y, Gaynor RB. Role of the NF-kappaB pathway in the pathogenesis of human disease states. Current Molecular Medicine 2001;1:287–296. [PubMed: 11899077]
- Yang F, Oz HS, Barve S, de Villiers WJ, McClain CJ, Varilek GW. The green tea polyphenol (—)epigallocatechin-3-gallate blocks nuclear factor-kappa B activation by inhibiting I kappa B kinase activity in the intestinal epithelial cell line IEC-6. Molecular Pharmacology 2001;60:528–533. [PubMed: 11502884]
- Yang L, Xie G, Fan Q, Xie J. Activation of the hedgehog-signaling pathway in human cancer and the clinical implications. Oncogene 2010;29:469–481. [PubMed: 19935712]
- Yu S, Shen G, Khor TO, Kim JH, Kong AN. Curcumin inhibits Akt/mammalian target of rapamycin signaling through protein phosphatase-dependent mechanism. Molecular Cancer Therapeutics 2008;7:2609–2620. [PubMed: 18790744]
- Zardawi SJ, O'Toole SA, Sutherland RL, Musgrove EA. Dysregulation of Hedgehog, Wnt and Notch signalling pathways in breast cancer. Histology and Histopathology 2009;24:385–398. [PubMed: 19130408]
- Zeng H, Botnen JH. Selenium is critical for cancer-signaling gene expression but not cell proliferation in human colon Caco-2 cells. Biofactors 2007;31:155–164. [PubMed: 18997278]
- Zhao H, Whitfield ML, Xu T, Botstein D, Brooks JD. Diverse effects of methylseleninic acid on the transcriptional program of human prostate cancer cells. Molecular Biology of the Cell 2004;15:506–519. [PubMed: 14617803]



Figure 1.

Major cell signaling pathways altered during the development and progression of prostate cancer. The dysfunctions of AR, Akt, NF- κ B, Wnt, Hedgehog, and Notch signaling could result in the uncontrolled transcription and proliferation of prostatic epithelial cells, leading to the formation of prostate cancer. The crosstalk between AR, Akt, NF- κ B, Wnt, Hedgehog, and Notch signaling plays critical roles in prostatic carcinogenesis. Therefore, targeting these signaling pathways is important strategy for the prevention of prostate cancer.

Table 1

Chemopreventive agents and their targets in the prevention of prostate cancer

Agent	Target signaling or altered molecules	References
Finasteride	AR, AKR1B1, PTEN, NKX3.1, PMEPA1, PSA, XRCC2, Akt, caspases, XIAP, TGF-β, etc.	Saez et al. (1998), Sawaya et al. (2002), Chen et al. (2005), Li & Kim (2009)
Dutasteride	AR, TRADD, caspase-7, caspase-8, BIRC1, Wnt, VEGF, etc.	Schmidt et al. (2004, 2009), Biancolella et al. (2007)
Vitamin D	CYP24A1, osteopontin (Spp1), LRP5, TRPV6, VDR, etc.	Meyer et al. (2010), Pike et al. (2010)
Selenium	AR, ER, NF-κB, A2M, IGFBP3, HHIP, CXC L9, HSPB2, Dhcr24, Abcc4, etc.	Shah <i>et al.</i> (2005), Christensen <i>et al.</i> (2007), Zeng & Botnen (2007), Legg <i>et al.</i> (2008), Schmidt <i>et al.</i> (2009)
Vitamin E	Akt, protein kinase C, NF-κB, AP1, P450, glutathione S-transferase, MMP-1, MMP-19, IL-2, IL-4, cyclin D1, cyclin E, Bcl-2, p27, CD95, TGF-β, etc.	Azzi et al. (2004), Banks et al. (2010)
Soy isoflavone	AR, Akt, NF-κB, Wnt, Notch, VEGF, p21, Bcl-2, Bax, Src, cyclin B, p27, MMP-9, protease M, uPAR, VEGF, neuropilin, TSP, BPGF, LPA, TGF-β2, TSP-1, PAR-2, Gli1, etc.	Li & Sarkar (2002 <i>a</i> , <i>b</i>), Wang <i>et al.</i> (2006 <i>a</i>), Li <i>et al.</i> (2008), Sarkar <i>et al.</i> (2008)
Lycopene	NF-κB, AR, Akt, Wnt, MAPK, p-ERK, p-p38, p-JNK, p27, Bax, β-catenin, cyclin D1, etc.	Kim et al. (2004), Palozza et al. (2005), Huang et al. (2007), Liu et al. (2008), Tang et al. (2008)
Green tea	AR, PSA, NF-κB, PI3K/Akt, VE-cadherin, Wnt, HBP1, p53, p21 ^{WAF1} , Bax, Gli1, etc.	Ahmad et al. (2000), Gupta et al. (2000), Ren et al. (2000), Tang et al. (2003), Kim et al. (2006), Qin et al. (2007), Khan & Mukhtar (2008)
DIM	AR, Akt, Wnt, NF-KB, VEGF, uPA, etc.	Le et al. (2003), Li et al. (2003, 2007), Kong et al. (2007, 2008), Ahmad et al. (2009)
Curcumin	NF- κ B, Akt, Notch, Wnt, Hedgehog, AR, etc.	Ohtsu <i>et al.</i> (2002), Bharti <i>et al.</i> (2003), Duvoix <i>et al.</i> (2003), Ryu <i>et al.</i> (2008), Yu <i>et al.</i> (2008)

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