

Disruption of the Postsynaptic Density in Alzheimer's Disease and Other Neurodegenerative Dementias

American Journal of Alzheimer's Disease & Other Dementias®
25(7) 547-555
© The Author(s) 2010
Reprints and permission:
sagepub.com/journalsPermissions.nav
DOI: 10.1177/1533317510382893
<http://aja.sagepub.com>



Yuesong Gong, PhD¹, and Carol F. Lippa, MD¹

Abstract

The most common causes of neurodegenerative dementia include Alzheimer's disease (AD), dementia with Lewy bodies (DLB), and frontotemporal dementia (FTD). We believe that, in all 3, aggregates of pathogenic proteins are pathological substrates which are associated with a loss of synaptic function/plasticity. The synaptic plasticity relies on the normal integration of glutamate receptors at the postsynaptic density (PSD). The PSD organizes synaptic proteins to mediate the functional and structural plasticity of the excitatory synapse and to maintain synaptic homeostasis. Here, we will discuss the relevant disruption of the protein network at the PSD in these dementias and the accumulation of the pathological changes at the PSD years before clinical symptoms. We suggest that the functional and structural plasticity changes of the PSD may contribute to the loss of molecular homeostasis within the synapse (and contribute to early symptoms) in these dementias.

Keywords

Alzheimer's disease, dementia with Lewy bodies, frontotemporal dementia, postsynaptic density

Introduction

Dementia involves progressive impairment of memory, cognition, language, and behavior. Common neurodegenerative diseases include Alzheimer's disease (AD), dementia with Lewy bodies (DLB), and frontotemporal dementia (FTD). Alzheimer's disease is the most frequent cause of neurodegenerative dementia, accounting for approximately 60% of patients with dementia. Dementia with Lewy bodies is arguably the second most common type of neurodegenerative dementia, which accounts for 10% to 20% of dementia cases. Frontotemporal dementia is another common type of neurodegenerative dementia, dominated by frontal lobe degeneration of the non-AD type.¹ Frontotemporal dementia includes Pick's disease, primary progressive aphasia, and dementia in motor neuron disease.^{2,3}

The pathological characterizations of AD, DLB, and FTD are complicated and overlap. β -Amyloid ($A\beta$) plaques and hyperphosphorylated tau-containing neurofibrillary tangles are the pathological hallmarks of AD. Most of α -synuclein inclusions, called Lewy bodies (LB), are demonstrated in the cortex and subcortical regions in DLB. However, aggregates of α -synuclein are also present in the Lewy body variant of AD. Also, $A\beta$ plaques and hyperphosphorylated tau are frequent in DLB cases. Tau is a microtubule-associated protein stabilizing microtubules as tracks for axonal transport.⁴ The pathological hyperphosphorylation of tau is also linked to FTD development. Although these aggregates of misfolded proteins overlap, evidence suggests that dysfunction and loss of the synapse might be a common pathological mechanism

underlying the cognitive decline and memory loss in these neurodegenerative diseases.⁵⁻¹¹

Most excitatory synapses terminate on dendritic spines for memory formation.¹² Spine size is directly related to synaptic strength, and this size is proportional to the area of the postsynaptic density (PSD).¹²⁻¹⁴ The PSD is an electron-dense thickening of membrane comprised of a proteinaceous network including glutamate receptors, adhesion molecules, scaffolding proteins, cytoskeletal proteins and associated signaling molecules. It is involved in a number of signaling pathways controlling synaptic plasticity and maintaining synaptic homeostasis. Synapses are continuously formed, eliminated, and remodeled throughout adulthood including synaptic protein synthesis, degradation, and modification. The stabilization of a new synaptic protrusion is associated with an increase in the size of the spine head and correlates with activity-driven PSD proteinaceous network formation. At this proteinaceous network, postsynaptic density protein 95 (PSD-95) is believed to play a role in synapse maturation, as it is one of the earliest detectable proteins in the PSD.¹⁵ PSD-95 induces clustering

¹ Department of Neurology, Drexel University College of Medicine, Philadelphia, PA, USA

Corresponding Author:

Yuesong Gong, Department of Neurology, Drexel University College of Medicine, 245 N 15th Street, Philadelphia, PA 19102, USA
Email: Yuesong.Gong@DrexelMed.edu

of a number of neurotransmitter receptors and scaffolding proteins.^{15,16} PSD-95 is coupled to Shank by guanylate kinase-associated protein (GKAP). Shank proteins directly regulate the formation and morphology changes of spine.¹⁷ The formation of PSD95-GKAP-Shank complex is the earliest event at the formation of the new postsynaptic site.¹⁸ The elimination of synapse is associated with a decrease in the size of the spine head and correlates with activity-driven PSD proteinaceous network degradation.¹⁹

In AD, preferential loss of postsynaptic compared with presynaptic elements has been suggested based on decreases in drebrin, a postsynaptic actin-binding protein.^{20,21} Here, the pathological changes of the postsynaptic region, PSD, are discussed in the brains of patients with AD, DLB, and FTD. The dynamic rearrangement of PSD could be the structural basis for the synaptic plasticity, the assembling and elimination of PSD components might lead to a fast alteration of synaptic structures underlying normal function or pathological changes of synapse.²²⁻²⁴ It is possible that the destruction of PSD is common mechanism at selective brain regions for these neurodegenerative diseases. Currently, the soluble oligomers of A β have been considered the initiator of synaptic dysfunction in AD.^{25,26} The pathogenic soluble oligomers of A β exist at damaged synapses in AD²⁷ and associate with PSD *in vivo*.²⁸ After PSDs were isolated and were analyzed by proteomics, glutamate receptors, PSD-95, Shank3, and synGap were shown to be dramatically altered in the frontal cortical tissues of patients with AD.¹¹ The pathological changes of these proteins might directly or indirectly affect the dendritic spine and synapse function and homeostasis.

The proteins at PSD can be subdivided into classes of (1) membrane-bound receptors and channels, (2) scaffolding and adaptor proteins, (3) cytoskeletal proteins, (4) cell-adhesion proteins, (5) modulatory enzymes including kinases/phosphatases, and others.²⁹⁻³¹ Here, we focus to discuss the pathological changes of structure-relevant proteins at PSD in these neurodegenerative dementias.

Synaptic Receptors and Channels

Glutamate Receptors

Glutamate receptors predominantly control synaptic plasticity and memory function. The various subtypes of glutamate receptors, ionotropic glutamate receptors (*N*-methyl *D*-aspartate [NMDA] and alpha-amino-3-hydroxy-5-methyl-4- isoxazole-propionic acid [AMPA]), and metabotropic glutamate receptors (mGluR) are integrated at the PSD in dendritic spines.

***N*-methyl *D*-aspartate receptor.** The alterations of glutamatergic synapses have been shown to be one of the earliest events and have long been considered the best pathological correlate of cognitive decline in AD.³² The evidence accumulated from the cellular to the clinical level demonstrated the glutamate receptors are dysfunctional in the initial stages of AD. *In vitro*, the oligomers of A β suppress NMDA

receptor-mediated long-term potentiation (LTP), a synaptic mechanism underlying memory and cognitive processing.³³ Interestingly, the NMDA receptor not only plays a critical role to regulate synaptic function but also has effects on amyloid precursor protein (APP) processing to release A β . Sublethal NMDA receptor activation increases the production and secretion of A β .³⁴ The oligomers of A β may reduce NMDAR-dependent Ca influx into the spine head.³³ The decrease of calcium influx through the synaptic NMDA receptors may inhibit nonamyloidogenic alpha-secretase-mediated APP processing and increase the production of A β .³⁵ Therefore, the deregulation of the glutamatergic neurotransmission may change the expression of APP and increase A β production at synapse. Deregulation of APP metabolism may exaggerate the dysfunction of glutamatergic receptors in early AD development.

Alpha-amino-3-hydroxy-5-methyl-4- isoxazole-propionic acid receptor. The oligomers of A β also disrupt the function of the AMPA receptor and reduce its expression on the synapse by Ca²⁺/calmodulin-dependent protein kinase II (CaMKII). This kinase is also altered in AD.³⁶ *In vivo*, the early reports show a decrease of AMPA binding sites in AD brain.³⁷ Recently, reports show NMDA receptor and AMPA receptor dramatically reduced at the PSD,^{11,38} which suggests the AMPA receptor (at the PSD) could also be an effective target along with the NMDA receptor for AD intervention. Interestingly, the AMPA receptor is also significantly reduced in the FTD brain, indicating that the AMPA receptor could also play an important role in synaptic dysfunction in FTD.³⁹

Metabotropic glutamate receptor. mGluRs are G-protein-coupled receptors, which are classified into 3 groups on the basis of signal transduction pathways and their pharmacological profiles. Group I mGluRs comprise mGluR1 and mGluR5. These subtypes are localized at the PSD area. The antagonists of group I mGluRs are neuroprotective. mGluR1 and mGluR5 antagonists also protect neuron in response to the administration of NMDA.^{40,41} However, the neurotoxicity of A β is exacerbated by application of the mGluR1 antagonist (RS)-1-aminoindan-1,5-dicarboxylic acid (AIDA),⁴² which implies the oligomers of A β might affect multiple targets. The dysfunction of group I mGluR has been hypothesized to be similar between AD and DLB.⁴³ Unfortunately, after the synapse was fractionated, the protein level of mGluR1 is not reduced in AD.¹¹

Acetylcholine Receptors

In addition to dysfunction of glutamate receptors, the early symptoms of AD and DLB also appear to correlate with dysfunction of cholinergic synapses.^{44,45} Acetylcholine receptors are classified as muscarinic acetylcholine receptors (mAChR) and nicotinic acetylcholine receptors (nAChR). Nicotinic acetylcholine receptors are known as "ionotropic" acetylcholine receptors. The nAChRs are ligand-gated ion channels. The clustering of α 7nAChR at PSD is retained through interactions with PSD components. Acetylcholine receptors can aid in

activating glutamatergic synapses and work together with AMPA receptors to mediate postsynaptic excitation throughout life.⁴⁶ In the AD brain, oligomers of A β may block the activity of α 7nAChR effecting on the endocytosis of NMDA receptors.⁴⁷

Dopamine Receptor

Dopamine is an important neurotransmitter in cognitive function, and the multiple dopamine receptor subtypes contribute to different aspects of learning and memory.⁴⁸ The D2R-NR2B interaction effects on the association of CaMKII with NR2B at PSD.⁴⁹ In AD, the loss of the D2 receptor-enriched modules contributes to disturbances in information processing in these high-order association cortices and in the hippocampus and may promote the cognitive and noncognitive impairments.^{50,51} Striatal D2/D3 receptors are increased in patients with AD having delusions.⁵² In DLB, D2 receptors may alter regulation of the striatal projection neurons,⁴⁴ and the reduction of D2 receptors are correlated with cognitive decline of DLB.⁵³

Serotonin receptor

Serotonin (5-hydroxytryptamine; 5-HT) regulates spine density in the hippocampus in both developing and adult animals.⁵⁴ 5-HT_{2A} receptors target spines of pyramidal neurons,⁵⁵ and 5-HT_{2A} receptors are colocalized with the NMDA and AMPA receptor subunits: NR1 and GluR2 in the hippocampal dentate gyrus and are colocalized with PSD-95 and with multiple PDZ protein-1 (MUPP1) in PSD.⁵⁶ PSD-95 profoundly modulates 5-HT_{2A} and 5-HT_{2C} receptor function.⁵⁷ Activation of 5-HT_{2A/C} receptors involves in the functional regulation of NMDA receptors at pyramidal neurons of prefrontal cortex to control cognitive and emotion.⁵⁸ The activation of 5-HT_{2A} receptors can induce a transient increase in dendritic spine size and phosphorylation of p21-activated kinase (PAK). p21-Activated kinase show pathological relocation at synapse in AD brain.⁵⁵ The protein level of 5-HT_{2A} receptors is also profoundly reduced in patients with AD,⁵⁹ in patients with mild cognitive impairment (MCI),⁶⁰ and in patients with FTD.⁶¹ The reduction of 5-HT_{2A} receptors may be correlated with the cognitive decline among these diseases.⁶²

Insulin Receptor

Insulin receptor is a tyrosine kinase, many of its actions require accessory molecules known as insulin receptor substrates (eg, IRS-1, IRS-2, and IRS-3).⁶³ Insulin receptor is concentrated at synapses and is a component of the PSD. The insulin signaling plays an important role in synaptic function.⁶⁴ The insulin receptor tyrosine kinase substrate p53 (IRSp53) is highly enriched in PSD fraction in brain. Although the soluble oligomers of A β caused major downregulation of plasma membrane insulin receptors (IRs), via a mechanism sensitive to CaMKII and casein kinase II (CK2) inhibition,⁶⁵ however, the protein level of IRSp53 did not show pathological change at PSD in AD.¹¹

Lipoprotein Receptors

Apolipoprotein E (apoE) is a cholesterol transport protein. Apolipoprotein E is a genetic risk factor for late-onset AD.^{66,67} Apolipoprotein E receptors have recently been recognized as pivotal components of the neuronal signalling machinery.⁶⁸ Apolipoprotein E receptors effect on intraneuronal signaling cascades through NMDA-type glutamate receptors.⁶⁹ The effects of apoE receptors on NMDA receptor signaling may be mediated by the interaction between apoE receptors (LRP1 or apoER2) and PSD-95.⁷⁰ At the postsynaptic membrane, LRP1 interacts with the PSD-95⁷¹ and might be part of a large postsynaptic density protein complex where LRP1 would modulate the conductance of neuronal ion channels.⁷²

The receptors of apoE, very-low-density lipoprotein (VLDL) receptor and apoE receptor 2 (apoER2), are also receptors for reelin. The location of reelin in spines, PSD, and terminals suggests that reelin has a role in synaptic remodeling and in LTP formation.⁷³ Reelin is upregulated in the brain and cerebrospinal fluid (CSF) in several neurodegenerative diseases.⁷⁴ Reelin-mediated signaling may contribute to neuronal dysfunction associated with AD.⁷⁵ Reelin interacts with APP, potentially having important effects on neurite development.⁷⁶ Interestingly, many of APP-interacting proteins also interact with the family proteins of apoE receptors. Both APP and apoE receptor affect neuronal migration and synapse formation in brain.⁷⁷ Apolipoprotein E receptor may play important roles in the dysfunction of the synapse and in β -amyloid formation in AD development.

Neurotrophic Factor Receptor

Neurotrophins regulate the survival and differentiation of afferent neurons. The neurotrophic factors, including BDNF, NGF, bFGF, and IGF1, activate receptors that possess intrinsic tyrosine kinase activity. Among these receptors, trkB, a high-affinity receptor for BDNF, is identified at PSD. The presence of trkB at the PSD is consistent with a role for neurotrophins in the regulation of synaptic activity via direct postsynaptic mechanisms.⁷⁸ Synaptic actions of BDNF are "gated" by cyclic AMP (cAMP). Cyclic AMP regulates BDNF function in mature hippocampal neurons by modulating the trafficking of TrkB to dendritic spines, possibly by promoting its interaction with PSD-95.⁷⁹ In amyloid-transgenic mice, BDNF can reverse synapse loss, and improve cell signaling and restore learning and memory, which implies the BDNF may have potential to reverse neuronal atrophy (and cognitive impairment) in AD.^{80,81} The interesting question is whether BDNF normalizes protein synthesis at the spine and repairs the function and structure of the proteinaceous network at the PSD in neurodegenerative dementias.

Synaptic Scaffolding and Adaptor Proteins

Scaffold proteins assemble neurotransmitter receptors, signal transduction components, actin-based cytoskeleton, adhesion

molecules, and modulatory enzymes at PSD. The major scaffolding molecules are membrane-associated guanylate kinase (MAGUK) protein PSD-95 and “master organizing” molecule Shank protein. The pathological changes of these scaffold proteins may be involved in the synaptic dysfunction in neurodegenerative dementias.

Membrane-Associated Guanylate Kinase Proteins

Membrane-associated guanylate kinase proteins belong to a family of synaptic proteins homologous to the product of the *Drosophila* gene Disc Large and include PSD-95, SAP97, chapsyn-110/PSD-93, and SAP102. Modifications of MAGUK proteins in the glutamatergic synapse are common events in several neurodegenerative disorders.⁸² PSD-95 is a critical component in the family of MAGUK proteins. PSD-95 organizes ionotropic glutamate receptors and their associated signaling proteins to regulate the strength of synaptic activity. Although the level of PSD-95 protein is sometimes changed in the brains of patients with AD, results have been inconsistent among various laboratories. An increase of PSD-95 protein levels occurs in the brain tissues of learning-impaired rats⁸³ and human frontal cortex, both at the synapse⁸⁴ and in the PSD fraction.¹¹ However, a decrease of PSD-95 protein levels occurs in the AD temporal cortex,⁸⁵ which suggests that the synaptic proteins could show different changes at selective regions in the brains of patients with neurodegenerative disease.

PSD-95 regulates protein trafficking and clustering of cell surface receptors and ion channels.⁸⁶ PSD-95 has been shown to influence surface expression of NMDA receptors and to regulate AMPA receptor insertion and retention at the synapse.⁸⁷ Mice lacking PSD-95 show normal NMDA receptor clustering and function but reduced AMPA receptor function.⁸⁸ The pathological distribution of PSD-95 could alter function of NMDA receptor and AMPA receptor at PSD in AD,¹¹ which may provide a possible means to alter synaptic strength.

In addition to modulating ion channel clustering and function, PSD-95 interacts with GKAP and may influence the recruitment of Shank, a molecule coupled to the actin-binding protein cortactin and the metabotropic glutamate receptor-interacting protein Homer to control spine morphology. Shank multimers assemble in large, sheet-like structures and may serve as a platform foundation for many PSD structures.⁸⁹

Shank

Shanks are the products of 3 genes, *Shank1*, *Shank2*, and *Shank3*. Shank proteins form the postsynaptic platform in PSD and organize NMDA receptor complex, AMPA receptor complex, and mGlu receptor complex.²⁹ Shanks play a critical role in integrating the various postsynaptic membrane proteins, cell-adhesion molecules, signal components, other scaffolding proteins, and actin-based cytoskeleton at the PSD protein network. These interactions place Shanks in the heart of the deeper

layer of PSD proteins.¹⁵ The dynamic rearrangement of PSD seems to be the structural basis for the synaptic regulation and synaptic plasticity that may be involved in memory formation.²⁴ In AD brain, oligomers of A β attack the postsynaptic region²⁶ and associate with PSD,²⁸ leading to inappropriate activity of NMDA-receptor at PSD⁹⁰; NMDA receptor activity is also dependent on integrity of its subunit composition and activation of its downsignal pathway and adjacent non-NMDA glutamate receptors. The total NMDA receptor (NR1) and AMPA receptor (GluR2) are significantly lost at PSD in AD cases.¹¹ The complex of NR1 and/or GluR2-PSD 95 is linked to Shank proteins. Shanks show dramatic pathological change in AD.¹¹ Shanks interact with several actin-binding proteins, including α -fodrin/spectrin, cortactin, actin-binding protein 1 (Abp1), and IRSp53.²⁹ The pathological changes of Shank proteins may directly/indirectly effect on the dynamics of actin in synaptic spine.

Cytoskeletal Proteins

The ability of globular actin (G-actin) to rapidly assemble and disassemble into filaments (F-actin) is critical to many cell behaviors. At the synapse, the dynamics of actin determine spine architecture by the GTPase-activating protein SynGAP, which has recently been shown to regulate both steady-state and activity-dependent cofilin phosphorylation,⁹¹ and showed loss at PSD in human AD brain.¹¹ F-actin is particularly associated by drebrin A at dendritic spines, postsynaptic sides of excitatory glutamatergic synapses. Drebrin A regulates the change of spine morphology, size, and density, presumably via regulation of actin cytoskeleton remodeling and dynamics.⁹² The disappearance of drebrin may contribute to the spine loss in AD.²⁰ The actin-rich dendritic spine architecture is also modulated by neuronal cofilin.⁹³ The concentration of cofilin at PSD may regulate the efficacy of synaptic NMDA receptor⁹⁴ and dynamic reorganization of actin cytoskeleton in space and time.⁹⁵ A β peptides can induce the AD-like pathological change of ADF/cofilin-actin rods in vitro⁹⁶ and tau phosphorylation. Although, hyperphosphorylated tau is viewed as the major cytoskeletal protein pathology in AD, the phosphorylated neurofilament is another constituent of aggregates in AD brain.⁹⁷ Similar phosphorylated neurofilaments also occur in the neural inclusions, Lewy bodies, in DLB disease.^{97,98}

Synaptic Cell-Adhesion Proteins

Synaptic adhesion proteins are not merely static structural components but are often dynamic regulators of synaptic function, participate in the formation, maturation, function, and plasticity of synapse, and control the number, location, and type of synapse. These cell-adhesion molecules (CAMs) include neuroligin, integrin, cadherin, Ephrins-EPH receptor, Nectin, NCAM, SynCAM, LI-CAM, and protocadherin. The known AD-relevant CAMs are summarized below.

Neuroigin

Neurexin and neuroigin are synaptic cell-adhesion proteins. The neurexin and neuroigin junction connects presynaptic and postsynaptic neurons to form the synapse, mediates signaling across the synapse, and shapes the properties of neural network by specifying synaptic function. The neurexin and neuroigin junction selects binding to postsynaptic proteins PSD-95, GKAP, and Shank.⁹⁹ The protein level of postsynaptic neuroigin-1 is affected by apoE4, the major genetic risk factor for AD and other neurodegenerative diseases. The lower level of apoE4 protein could lead to the loss of postsynaptic neuroigin-1 in mice model.¹⁰⁰

Integrin

Integrin-mediated communication regulates many cell physiological processes including cell cycles. The fibrils of A β effect on integrin/focal adhesion (FA) signaling pathways that mediate cell-cycle activation and cell death. The development of AD includes an extremely complicated change at the synaptic protein level. The pathological alteration of integrin/FAK/FA signaling pathway induced by fibrils of A β may alter neuronal viability and synaptic plasticity during the course of AD.¹⁰¹

N-Cadherin

Cadherins are transmembrane cell adhesion proteins. N-cadherin homophilic interactions connect pre- and postsynaptic membranes together.^{102,103} N-cadherin, NMDA receptor, AMPA receptor GluR2/3, and PSD-95 and other synaptic proteins are integrated at spine; the dynamic alteration of N-cadherin modifies the plasticity of synaptic structure and function.¹⁰⁴⁻¹⁰⁶ This dynamic change of N-cadherin is also mediated by the activities of NMDA receptor and presenilin-1 (PS1).^{107,108} PS1 binds cadherins and stabilizes the cadherin/catenin cell adhesion complexes at the plasma membrane.¹⁰⁹

Ephrins-Eph Receptors

Ephrins and Eph receptors regulate excitatory neurotransmission and play a role in the morphological change of dendritic spine remodeling. The pathological changes of Ephrins and Eph receptors could be related to the AD development. EphA4 and EphB2 receptors were lost in the hippocampus at early stage of APP mutation AD mouse. A similar pathological change in Eph receptor levels was also observed in postmortem hippocampal tissue from incipient AD patients.¹¹⁰

Modulatory Enzymes

The PSD consists of hundreds of proteins beside the above reminded proteins. The PSD-associated protein kinase/phosphatase and other modulatory enzymes play multiple roles in the spine loss in AD development. These proteins include nonreceptor protein tyrosine kinase, Fyn¹¹¹; CaMKII^{33,112}; protein kinase C (PKC)¹¹³; mitogen activated kinase, Cdk5,

GSK3 β , Rho kinase (ROCK)¹¹⁴; p21-activated kinase¹¹⁵; calcineurin^{116,117}; protein phosphatase I¹¹⁸; protein phosphatase 2A¹¹⁹; neuronal nitric oxide synthase¹²⁰; ubiquitin proteasome system¹¹; O-GlcNAc modification relevant enzymes¹²¹; and peptidyl-prolyl isomerase Pin1.¹²² Although relevant, the current manuscript will not further discuss how these enzymes might affect the PSD structure and function in AD development.

Summary

The PSD is complicated and the current manuscript only summarizes some key points. The spatial-temporal pathological changes at the PSD protein network are likely to occur in early AD. The accumulation of these pathological changes could impact synaptic homeostasis leading to negative synaptic formation and synaptic loss and dysfunction. Animal AD models are limited because they do not show the full spectrum of pathological change seen in human AD. Most current studies of AD pathogenesis use human postmortem tissues and there is need for studies of the PSD in cases who die with early stage disease.

Declaration of Conflicting Interests

The author(s) declared no conflicts of interest with respect to the authorship and/or publication of this article.

Funding

The author(s) disclosed receipt of the following financial support for the research and/or authorship of this article: NIH1R21AG031388 (YG) and Potamkin Foundation (CFL).

References

1. Arvanitakis Z. Update on frontotemporal dementia. *Neurologist*. 2010;16(1):16-22.
2. Ballatore C, Lee VM, Trojanowski JQ. Tau mediated neurodegeneration in Alzheimer's disease and related disorders. *Nat Rev Neurosci*. 2007;8(9):663-672.
3. Vessel KA, Miller BL. New approaches to the treatment of frontotemporal lobar degeneration. *Curr Opin Neurol*. 2008;21(6):708-716.
4. Mandelkow E, von Bergen M, Biernat J, Mandelkow EM. Structural principles of tau and the paired helical filaments of Alzheimer's disease. *Brain Pathol*. 2007;17(1):83-90.
5. Terry RD, Masliah E, Salmon DP, et al. Physical basis of cognitive alterations in Alzheimer's disease: synapse loss is the major correlate of cognitive impairment. *Ann Neurol*. 1991;30(4):572-580.
6. Brun A, Passant U. Frontal lobe degeneration of non-Alzheimer type. Structural characteristics, diagnostic criteria and relation to other frontotemporal dementias. *Acta Neurol Scand Suppl*. 1996;168:28-30.
7. Lippa CF, Pulaski-Salo D, Dickson DW, Smith TW. Alzheimer's disease, Lewy body disease and aging: a comparative study of the perforant pathway. *J Neurol Sci*. 1997;147(2):161-166.
8. Lipton AM, Cullum CM, Satumira S, et al. Contribution of asymmetric synapse loss to lateralizing clinical deficits in frontotemporal dementias. *Arch Neurol*. 2001;58(8):1233-1239.

9. Lippa CF. Synaptophysin immunoreactivity in Pick's disease: comparison with Alzheimer's disease and dementia with Lewy bodies. *Am J Alzheimers Dis Other Demen.* 2004;19(6):341-344.
10. Revuelta GJ, Lippa CF. Dementia with Lewy bodies and Parkinson's disease dementia may best be viewed as two distinct entities. *Int Psychogeriatr.* 2009;21(2):213-216.
11. Gong Y, Lippa CF, Zhu J, Lin Q, Rosso AL. Disruption of glutamate receptors at Shank-postsynaptic platform in Alzheimer's disease. *Brain Res.* 2009;1292:191-198.
12. Knott GW, Holtmaat A, Wilbrecht L, Welker E, Svoboda K. Spine growth precedes synapse formation in the adult neocortex in vivo. *Nat Neurosci.* 2006;9(9):1117-1124.
13. Harris KM, Stevens JK. Dendritic spines of CA 1 pyramidal cells in the rat hippocampus: serial electron microscopy with reference to their biophysical characteristics. *J Neurosci.* 1989;9(8):2982-2997.
14. Wallace W, Bear MF. A morphological correlate of synaptic scaling in visual cortex. *J Neurosci.* 2004;24(31):6928-6938.
15. Kim E, Sheng M. PDZ domain proteins of synapses. *Nat Rev Neurosci.* 2004;5(10):771-781.
16. El-Husseini AE, Schnell E, Chetkovich DM, Nicoll RA, Brecht DS. PSD-95 involvement in maturation of excitatory synapses. *Science.* 2000;290(5495):1364-1368.
17. Roussignol G, Ango F, Romorini S, et al. Shank expression is sufficient to induce functional dendritic spine synapses in aspiny neurons. *J Neurosci.* 2005;25(14):3560-3570.
18. Gerrow K, Romorini S, Nabi SM, Colicos MA, Sala C, El-Husseini A. A preformed complex of postsynaptic proteins is involved in excitatory synapse development. *Neuron.* 2006;49(4):547-562.
19. Ehlers MD. Activity level controls postsynaptic composition and signaling via the ubiquitin-proteasome system. *Nat Neurosci.* 2003;6(3):231-242.
20. Harigaya Y, Shoji M, Shirao T, Hirai S. Disappearance of actin-binding protein, drebrin, from hippocampal synapses in Alzheimer's disease. *J Neurosci Res.* 1996;43(1):87-92.
21. Shim KS, Lubec G. Drebrin, a dendritic spine protein, is manifold decreased in brains of patients with Alzheimer's disease and Down syndrome. *Neurosci Lett.* 2002;324(3):209-212.
22. Hering H, Sheng M. Dendritic spines: structure, dynamics and regulation. *Nat Rev Neurosci.* 2001;2(12):880-888.
23. Yuste R, Bonhoeffer T. Morphological changes in dendritic spines associated with long-term synaptic plasticity. *Annu Rev Neurosci.* 2001;24:1071-1089.
24. Carlisle HJ, Kennedy MB. Spine architecture and synaptic plasticity. *Trends Neurosci.* 2005;28(4):182-187.
25. Gong Y, Chang L, Viola KL, et al. Alzheimer's disease-affected brain: presence of oligomeric A beta ligands (ADDLs) suggests a molecular basis for reversible memory loss. *Proc Natl Acad Sci U S A.* 2003;100(18):10417-10422.
26. Lacor PN, Buniel MC, Chang L, et al. Synaptic targeting by Alzheimer's-related beta-amyloid oligomers. *J Neurosci.* 2004;24(45):10191-10200.
27. Takahashi RH, Almeida CG, Kearney PF, et al. Oligomerization of Alzheimer's beta-amyloid within processes and synapses of cultured neurons and brain. *J Neurosci.* 2004;24(14):3592-3599.
28. Koffie RM, Meyer-Luehmann M, Hashimoto T, et al. Oligomeric amyloid beta associates with postsynaptic densities and correlates with excitatory synapse loss near senile plaques. *Proc Natl Acad Sci U S A.* 2009;106(10):4012-4007.
29. Boeckers TM. The postsynaptic density. *Cell Tissue Res.* 2006;326(2):409-422.
30. Kennedy MB. Signal-processing machines at the postsynaptic density. *Science.* 2000;290(5492):750-754.
31. Sheng M. Molecular organization of the postsynaptic specialization. *Proc Natl Acad Sci U S A.* 2001;98(13):7058-7061.
32. Coleman PD, Yao PJ. Synaptic slaughter in Alzheimer's disease. *Neurobiol Aging.* 2003;24(8):1023-1027.
33. Shankar GM, Bloodgood BL, Townsend M, Walsh DM, Selkoe DJ, Sabatini BL. Natural oligomers of the Alzheimer amyloid-beta protein induce reversible synapse loss by modulating an NMDA-type glutamate receptor-dependent signaling pathway. *J Neurosci.* 2007;27(11):2866-2875.
34. Lesné S, Ali C, Gabriel C, et al. NMDA receptor activation inhibits alpha-secretase and promotes neuronal amyloid-beta production. *J Neurosci.* 2005;25(41):9367-9377.
35. Hoey SE, Williams RJ, Perkinson MS. Synaptic NMDA receptor activation stimulates alpha-secretase amyloid precursor protein processing and inhibits amyloid-beta production. *J Neurosci.* 2009;29(14):4442-4460.
36. Gu Z, Liu W, Yan Z. Beta-Amyloid impairs AMPA receptor trafficking and function by reducing Ca²⁺/calmodulin-dependent protein kinase II synaptic distribution. *J Biol Chem.* 2009;284(16):10639-10649.
37. Armstrong DM, Ikonomic MD, Sheffield R, Wenthold RJ. AMPA-selective glutamate receptor subtype immunoreactivity in the entorhinal cortex of non-demented elderly and patients with Alzheimer's disease. *Brain Res.* 1994;639(2):207-216.
38. Gasparini L, Dityatev A. Beta-amyloid and glutamate receptors. *Exp Neurol.* 2008;212(1):1-4.
39. Procter AW, Qurne M, Francis PT. Neurochemical features of frontotemporal dementia. *Dement Geriatr Cogn Disord.* 1999;10(suppl 1):80-84.
40. Mukhin AG, Ivanova SA, Faden AI. mGluR modulation of post-traumatic neuronal death: role of NMDA receptors. *Neuroreport.* 1997;8(11):2561-2566.
41. O'Leary DM, Movsesyan V, Vicini S, Faden AI. Selective mGluR5 antagonists MPEP and SIB-1893 decrease NMDA or glutamate-mediated neuronal toxicity through actions that reflect NMDA receptor antagonism. *Br J Pharmacol.* 2000;131(7):1429-1437.
42. Allen JW, Eldadah BA, Faden AI. Beta-Amyloid-induced apoptosis of cerebellar granule cells and cortical neurons: exacerbation by selective inhibition of group I metabotropic glutamate receptors. *Neuropharmacology.* 1999;38(8):1243-1252.
43. Albasanz JL, Dalfó E, Ferrer I, Martín M. Impaired metabotropic glutamate receptor/phospholipase C signaling pathway in the cerebral cortex in Alzheimer's disease and dementia with Lewy bodies correlates with stage of Alzheimer's-disease-related changes. *Neurobiol Dis.* 2005;20(3):685-693.
44. Piggott MA, Owens J, O'Brien J, et al. Muscarinic receptors in basal ganglia in dementia with Lewy bodies, Parkinson's

- disease and Alzheimer's disease. *J Chem Neuroanat.* 2003;25(3):161-173.
45. Jellinger KA. Morphological substrates of mental dysfunction in Lewy body disease: an update. *J Neural Transm Suppl.* 2000;59:185-212.
46. Levy RB, Aoki C. Alpha7 nicotinic acetylcholine receptors occur at postsynaptic densities of AMPA receptor-positive and -negative excitatory synapses in rat sensory cortex. *J Neurosci.* 2002;22(12):5001-5015.
47. Snyder EM, Nong Y, Almeida CG, et al. Regulation of NMDA receptor trafficking by amyloid-beta. *Nat Neurosci.* 2005;8(8):1051-1058.
48. El-Ghundi M, O'Dowd BF, George SR. Insights into the role of dopamine receptor systems in learning and memory. *Rev Neurosci.* 2007;18(1):37-66.
49. Liu XY, Chu XP, Mao LM, et al. Modulation of D2R-NR2B interactions in response to cocaine. *Neuron.* 2006;52(5):897-909.
50. Joyce JN, Myers AJ, Gurevich E. Dopamine D2 receptor bands in normal human temporal cortex are absent in Alzheimer's disease. *Brain Res.* 1998;784(1-2):7-17.
51. Kempainen N, Laine M, Laakso MP, et al. Hippocampal dopamine D2 receptors correlate with memory functions in Alzheimer's disease. *Eur J Neurosci.* 2003;18(1):149-154.
52. Reeves S, Brown R, Howard R, Grasby P. Increased striatal dopamine (D2/D3) receptor availability and delusions in Alzheimer disease. *Neurology.* 2009;72(6):528-534.
53. Piggott MA, Ballard CG, Rowan E, et al. Selective loss of dopamine D2 receptors in temporal cortex in dementia with Lewy bodies, association with cognitive decline. *Synapse.* 2007;61(11):903-911.
54. Hajszan T, MacLusky NJ, Leranth C. Short-term treatment with the antidepressant fluoxetine triggers pyramidal dendritic spine synapse formation in rat hippocampus. *Eur J Neurosci.* 2005;21(5):1299-1303.
55. Jones KA, Srivastava DP, Allen JA, Strachan RT, Roth BL, Penzes P. Rapid modulation of spine morphology by the 5-HT2A serotonin receptor through kalirin-7 signaling. *Proc Natl Acad Sci U S A.* 2009;106(46):19575-19580.
56. Peddie CJ, Davies HA, Colyer FM, Stewart MG, Rodriguez JJ. Colocalisation of serotonin2A receptors with the glutamate receptor subunits NR1 and GluR2 in the dentate gyrus: An ultrastructural study of a modulatory role. *Exp Neurol.* 2008;211(2):561-573.
57. Abbas AI, Yadav PN, Yao WD, et al. PSD-95 is essential for haloperidol and atypical antipsychotic drug actions at serotonin receptors. *J Neurosci.* 2009;29(22):7124-7136.
58. Yuen EY, Jiang Q, Chen P, Feng J, Yan Z. Activation of 5-HT2A/C receptors counteracts 5-HT1A regulation of n-methyl-D-aspartate receptor channels in pyramidal neurons of prefrontal cortex. *J Biol Chem.* 2008;283(25):17194-17204.
59. Blin J, Baron JC, Dubois B, et al. Loss of brain 5-HT2 receptors in Alzheimer's disease. In vivo assessment with positron emission tomography and [¹⁸F]setoperone. *Brain.* 1993;116(pt 3):497-510.
60. Hasselbalch SG, Madsen K, Svarer C, et al. Reduced 5-HT2A receptor binding in patients with mild cognitive impairment. *Neurobiol Aging.* 2008;29(12):1830-1838.
61. Bowen DM, Procter AW, Mann DM, et al. Imbalance of a serotonergic system in frontotemporal dementia: implication for pharmacotherapy. *Psychopharmacology (Berl).* 2008;196(4):603-610.
62. Lai MK, Tsang SW, Alder JT, et al. Loss of serotonin 5-HT2A receptors in the postmortem temporal cortex correlates with rate of cognitive decline in Alzheimer's disease. *Psychopharmacology (Berl).* 2005;179(3):673-677.
63. White MF, Yenush L. The IRS-signaling system: a network of docking proteins that mediate insulin and cytokine action. *Curr Top Microbiol Immunol.* 1998;228:179-208.
64. Abbott MA, Wells DG, Fallon JR. The insulin receptor tyrosine kinase substrate p58/53 and the insulin receptor are components of CNS synapses. *J Neurosci.* 1999;19(17):7300-7308.
65. De Felice FG, Vieira MN, Bomfim TR, et al. Protection of synapses against Alzheimer's-linked toxins: insulin signaling prevents the pathogenic binding of Aβ oligomers. *Proc Natl Acad Sci U S A.* 2009;106(6):1971-1976.
66. Strittmatter WJ, Weisgraber KH, Huang DY, et al. Binding of human apolipoprotein E to synthetic amyloid beta peptide: isoform-specific effects and implications for late-onset Alzheimer disease. *Proc Natl Acad Sci U S A.* 1993;90(17):8098-8102.
67. Coon KD, Myers AJ, Craig DW, et al. A high-density whole-genome association study reveals that apoE is the major susceptibility gene for sporadic late-onset Alzheimer's disease. *J Clin Psychiatry.* 2007;68(4):613-618.
68. Herz J, Chen Y. Reelin, lipoprotein receptors and synaptic plasticity. *Nat Rev Neurosci.* 2006;7(11):850-859.
69. Hoe HS, Harris DC, Rebeck GW. Multiple pathways of apolipoprotein E signaling in primary neurons. *J Neurochem.* 2005;93(1):145-155.
70. Hoe HS, Pocivavsek A, Chakraborty G, et al. Apolipoprotein E receptor 2 interactions with the N-methyl-D-aspartate receptor. *J Biol Chem.* 2006;281(6):3425-3431.
71. Gotthardt M, Trommsdorff M, Nevitt MF, et al. Interactions of the low density lipoprotein receptor gene family with cytosolic adaptor and scaffold proteins suggest diverse biological functions in cellular communication and signal transduction. *J Biol Chem.* 2000;275(33):25616-25624.
72. May P, Rohlmann A, Bock HH, et al. Neuronal LRP1 functionally associates with postsynaptic proteins and is required for normal motor function in mice. *Mol Cell Biol.* 2004;24(20):8872-8883.
73. Roberts RC, Xu L, Roche JK, Kirkpatrick B. Ultrastructural localization of reelin in the cortex in post-mortem human brain. *J Comp Neurol.* 2005;482(3):294-308.
74. Botella-López A, Burgaya F, Gavín R, et al. Reelin expression and glycosylation patterns are altered in Alzheimer's disease. *Proc Natl Acad Sci U S A.* 2006;103(14):5573-5578.
75. Knuesel I, Nyffeler M, Mormède C, et al. Age-related accumulation of Reelin in amyloid-like deposits. *Neurobiol Aging.* 2009;30(5):697-716.
76. Hoe HS, Lee KJ, Carney RS, et al. Interaction of reelin with amyloid precursor protein promotes neurite outgrowth. *J Neurosci.* 2009;29(23):7459-7473.
77. Hoe HS, Rebeck GW. Functional interactions of APP with the apoE receptor family. *J Neurochem.* 2008;106(6):2263-2271.

78. Wu K, Xu JL, Suen PC, et al. Functional trkB neurotrophin receptors are intrinsic components of the adult brain postsynaptic density. *Brain Res Mol Brain Res*. 1996;43(1-2):286-290.
79. Ji Y, Pang PT, Feng L, Lu B. Cyclic AMP controls BDNF-induced TrkB phosphorylation and dendritic spine formation in mature hippocampal neurons. *Nat Neurosci*. 2005;8(2):164-172.
80. Nagahara AH, Merrill DA, Coppola G, et al. Neuroprotective effects of brain-derived neurotrophic factor in rodent and primate models of Alzheimer's disease. *Nat Med*. 2009;15(3):331-337.
81. Simmons DA, Rex CS, Palmer L, et al. Up-regulating BDNF with an ampakine rescues synaptic plasticity and memory in Huntington's disease knockin mice. *Proc Natl Acad Sci U S A*. 2009;106(12):4906-4911.
82. Gardoni F. MAGUK proteins: new targets for pharmacological intervention in the glutamatergic synapse. *Eur J Pharmacol*. 2008;585(1):147-152.
83. Nyffeler M, Zhang WN, Feldon J, Knuesel I. Differential expression of PSD proteins in age-related spatial learning impairments. *Neurobiol Aging*. 2007;28(1):143-155.
84. Leuba G, Savioz A, Vernay A, et al. Differential changes in synaptic proteins in the Alzheimer frontal cortex with marked increase in PSD-95 postsynaptic protein. *J Alzheimers Dis*. 2008;15(1):139-151.
85. Gyls KH, Fein JA, Yang F, Wiley DJ, Miller CA, Cole GM. Synaptic changes in Alzheimer's disease: increased amyloid-beta and gliosis in surviving terminals is accompanied by decreased PSD-95 fluorescence. *Am J Pathol*. 2004;165(5):1809-1817.
86. Christopherson KS, Hillier BJ, Lim WA, Brecht DS. PSD-95 assembles a ternary complex with the N-methyl-D-aspartate receptor and a bivalent neuronal NO synthase PDZ domain. *J Biol Chem*. 1999;274(39):27467-27473.
87. Lin Y, Jover-Mengual T, Wong J, Bennett MV, Zukin RS. PSD-95 and PKC converge in regulating NMDA receptor trafficking and gating. *Proc Natl Acad Sci U S A*. 2006;103(52):19902-19907.
88. Ehrlich I, Klein M, Rumpel S, Malinow R. PSD-95 is required for activity-driven synapse stabilization. *Proc Natl Acad Sci U S A*. 2007;104(10):4176-4181.
89. Baron MK, Boeckers TM, Vaida B, et al. An architectural framework that may lie at the core of the postsynaptic density. *Science*. 2006;311(5760):531-535.
90. Roselli F, Tirard M, Lu J, et al. Soluble beta-amyloid1-40 induces NMDA-dependent degradation of postsynaptic density-95 at glutamatergic synapses. *J Neurosci*. 2005;25(48):11061-11070.
91. Carlisle HJ, Manzerra P, Marcora E, Kennedy MB. SynGAP regulates steady-state and activity-dependent phosphorylation of cofilin. *J Neurosci*. 2008;28(50):13673-13683.
92. Ivanov A, Esclapez M, Ferhat L. Role of drebrin A in dendritic spine plasticity and synaptic function: implications in neurological disorders. *Commun Integr Biol*. 2009;2(3):268-270.
93. Hotulainen P, Llano O, Smirnov S, et al. Defining mechanisms of actin polymerization and depolymerization during dendritic spine morphogenesis. *J Cell Biol*. 2009;185(2):323-339.
94. Morishita W, Marie H, Malenka RC. Distinct triggering and expression mechanisms underlie LTD of AMPA and NMDA synaptic responses. *Nat Neurosci*. 2005;8(8):1043-1050.
95. Lee-Hoeflich ST, Causing CG, Podkowa M, Zhao X, Wrana JL, Attisano L. Activation of LIMK1 by binding to the BMP receptor, BMPRII, regulates BMP-dependent dendritogenesis. *EMBO J*. 2004;23(24):4792-4801.
96. Davis RC, Maloney MT, Minamide LS, Flynn KC, Stonebraker MA, Bamberg JR. Mapping cofilin-actin rods in stressed hippocampal slices and the role of cdc42 in amyloid-beta-induced rods. *J Alzheimers Dis*. 2009;18(1):35-50.
97. Shepherd CE, McCann H, Thiel E, Halliday GM. Neurofilament-immunoreactive neurons in Alzheimer's disease and dementia with Lewy bodies. *Neurobiol Dis*. 2002;9(2):249-257.
98. Hansen LA, Samuel W. Criteria for Alzheimer's disease and the nosology of dementia with Lewy bodies. *Neurology*. 1997;48(1):126-132.
99. Futai K, Kim MJ, Hashikawa T, Scheiffele P, Sheng M, Hayashi Y. Retrograde modulation of presynaptic release probability through signaling mediated by PSD-95-neurologin. *Nat Neurosci*. 2007;10(2):186-195.
100. Zhong N, Scarce-Levie K, Ramaswamy G, Weisgraber KH. Apolipoprotein E4 domain interaction: synaptic and cognitive deficits in mice. *Alzheimers Dement*. 2008;4(3):179-192.
101. Caltagarone J, Jing Z, Bowser R. Focal adhesions regulate Abeta signaling and cell death in Alzheimer's disease. *Biochim Biophys Acta*. 2007;1772(4):438-445.
102. Togashi H, Abe K, Mizoguchi A, Takaoka K, Chisaka O, Takeichi M. Cadherin regulates dendritic spine morphogenesis. *Neuron*. 2002;35(1):77-89.
103. Okabe T, Nakamura T, Nishimura YN, et al. RICS, a novel GTPase-activating protein for Cdc42 and Rac1, is involved in the beta-catenin-N-cadherin and N-methyl-D-aspartate receptor signaling. *J Biol Chem*. 2003;278(11):9920-9927.
104. Fannon AM, Colman DR. A model for central synaptic junctional complex formation based on the differential adhesive specificities of the cadherins. *Neuron*. 1996;17(3):423-434.
105. Benson DL, Tanaka H. N-cadherin redistribution during synaptogenesis in hippocampal neurons. *J Neurosci*. 1998;18(17):6892-6904.
106. Silverman JB, Restituito S, Lu W, Lee-Edwards L, Khatri L, Ziff EB. Synaptic anchorage of AMPA receptors by cadherins through neural plakophilin-related arm protein AMPA receptor-binding protein complexes. *J Neurosci*. 2007;27(32):8505-8516.
107. Marambaud P, Wen PH, Dutt A, et al. A CBP binding transcriptional repressor produced by the PS1/epsilon-cleavage of N-cadherin is inhibited by PS1 FAD mutations. *Cell*. 2003;114(5):635-645.
108. Uemura K, Kihara T, Kuzuya A, et al. Characterization of sequential N-cadherin cleavage by ADAM10 and PS1. *Neurosci Lett*. 2006;402(3):278-283.
109. Serban G, Kouchi Z, Baki L, et al. Cadherins mediate both the association between PS1 and beta-catenin and the effects of PS1 on beta-catenin stability. *J Biol Chem*. 2005;280(43):36007-36012.
110. Simón AM, de Maturana RL, Ricobaraza A, et al. Early changes in hippocampal eph receptors precede the onset of memory decline in mouse models of Alzheimer's disease. *J Alzheimers Dis*. 2009;17(4):773-786.
111. Shirazi SK, Wood JG. The protein tyrosine kinase, fyn, in Alzheimer's disease pathology. *Neuroreport*. 1993;4(4):435-437.

112. Singh TJ, Grundke-Iqbal I, Wu WQ, et al. Protein kinase C and calcium/calmodulin-dependent protein kinase II phosphorylate three-repeat and four-repeat tau isoforms at different rates. *Mol Cell Biochem.* 1997;168(1-2):141-148.
113. Pascale A, Amadio M, Govoni S, Battaini F. The aging brain, a key target for the future: the protein kinase C involvement. *Pharmacol Res.* 2007;55(6):560-569.
114. Ma QL, Yang F, Calon F, et al. p21-activated kinase-aberrant activation and translocation in Alzheimer disease pathogenesis. *J Biol Chem.* 2008;283(20):14132-14143.
115. Salminen A, Suuronen T, Kaarniranta K. ROCK, PAK, and Toll of synapses in Alzheimer's disease. *Biochem Biophys Res Commun.* 2008;371(4):587-590.
116. Wu HY, Hudry E, Hashimoto T, et al. Beta-Amyloid induces the morphological neurodegenerative triad of spine loss, dendritic simplification, and neuritic dystrophies through calcineurin activation. *J Neurosci.* 2010;30(7):2636-2649.
117. Zhao WQ, Santini F, Breese R, et al. Inhibition of calcineurin-mediated endocytosis and alpha-amino-3-hydroxy-5-methyl-4-isoxazolepropionic Acid (AMPA) receptors prevents beta-amyloid oligomer-induced synaptic disruption. *J Biol Chem.* 2010;285(10):7619-7632.
118. Vintém AP, Henriques AG, da Cruz E, Silva OA, da Cruz E, Silva EF. PP1 inhibition by Abeta peptide as a potential pathological mechanism in Alzheimer's disease. *Neurotoxicol Teratol.* 2009;31(2):85-88.
119. Wang JZ, Grundke-Iqbal I, Iqbal K. Kinases and phosphatases and tau sites involved in Alzheimer neurofibrillary degeneration. *Eur J Neurosci.* 2007;25(1):59-68.
120. Galimberti D, Venturelli E, Gatti A, et al. Association of neuronal nitric oxide synthase C276T polymorphism with Alzheimer's disease. *J Neurol.* 2005;252(8):985-986.
121. Liu F, Shi J, Tanimukai H, et al. Reduced O-GlcNAcylation links lower brain glucose metabolism and tau pathology in Alzheimer's disease. *Brain.* 2009;132(pt 7):1820-1832.
122. Lu KP, Zhou XZ. The prolyl isomerase PIN1: a pivotal new twist in phosphorylation signalling and disease. *Nat Rev Mol Cell Biol.* 2007;8(11):904-916.