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Misfolding of CasBrE SU is reversed by interactions with 4070A Env: implications for gammaretroviral neuropathogenesis

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Abstract

Background: CasBrE is a neurovirulent murine leukemia virus (MLV) capable of inducing paralytic disease with associated spongiform neurodegeneration. The neurovirulence of this virus has been genetically mapped to the surface expressed subunit (SU) of the *env* gene. However, CasBrE SU synthesized in the absence of the transmembrane subunit (TM) does not retain ecotropic receptor binding activity, indicating that folding of the receptor binding domain (RBD) requires this domain. Using a neural stem cell (NSC) based viral *trans* complementation approach to examine whether misfolded CasBrE SU retained neurovirulence, we observed CasBrE SU interaction with the “non-neurovirulent” amphotropic helper virus, 4070A which restored functional activity of CasBrE SU.

Results: Herein, we show that infection of NSCs expressing CasBrE SU with 4070A (CasES+4070A-NSCs) resulted in the redistribution of CasBrE SU from a strictly secreted product to include retention on the plasma membrane. Cell surface cross-linking analysis suggested that CasBrE SU membrane localization was due to interactions with 4070A Env. Viral particles produced from CasES+4070A-NSCs contained both CasBrE and 4070A gp70 Env proteins. These particles displayed ecotropic receptor-mediated infection, but were still 100-fold less efficient than CasE+4070A-NSC virus. Infectious center analysis showed CasBrE SU ecotropic transduction efficiencies approaching those of NSCs expressing full length CasBrE Env (CasE; SU+TM). In addition, CasBrE SU-4070A Env interactions resulted in robust ecotropic superinfection interference indicating near native intracellular SU interaction with its receptor, mCAT-1.

Conclusions: In this report we provided evidence that 4070A Env and CasBrE SU physically interact within NSCs leading to CasBrE SU retention on the plasma membrane, incorporation into viral particles, restoration of mCAT-1 binding, and capacity for initiation of TM-mediated fusion events. Thus, heterotropic Env-SU interactions facilitates CasBrE SU folding events that restore Env activity. These findings are consistent with the idea that one protein conformation acts as a folding scaffold or nucleus for a second protein of similar primary structure, a process reminiscent of prion formation. The implication is that template-based protein folding may represent an inherent feature of neuropathogenic proteins that extends to retroviral Envs.

Introduction

Certain MLVs are capable of causing severe progressive non-inflammatory spongiform motor neuron disease when inoculated into susceptible neonatal mice. The prototypic virus of this class, referred to as CasBrE, was first isolated from wild mice and shown to cause a paralytic wasting disease with low incidence and a long

incubation period, highly reminiscent of certain prion diseases and amyotrophic lateral sclerosis (ALS) [1-3]. Genetic mapping studies have indicated that the primary viral neurovirulence determinants reside within the viral *env* gene, specifically to within the region encoding the SU subunit (cf., [4-10]). As a result, it has been speculated that CNS expression of neurovirulent MLV Env protein alone might be sufficient for inducing neurodegeneration [4]. Attempts to address this issue through the generation of transgenic or NSC-based chimeric

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mice expressing neurovirulent MLV Envs have yet to provide a clear resolution to this question [11-14].

We have previously demonstrated that over expression of the CasBrE Env protein, either SU or SU/TM, from engrafted NSCs does not induce acute spongiform pathology. Moreover, we have shown that dissemination of the CasBrE *env* gene from transplanted packaging/producer NSCs also does not cause spongiosis, despite Env expression within host microglia, a major CNS MLV target. Therefore, we were interested in examining whether additional retroviral proteins might also need to be delivered to host cells to induce neuropathogenic changes. To address this question, we have been exploring an NSC-based strategy wherein a “non-neurovirulent” amphotropic virus, 4070A, is used to pseudotype and *trans* complement the packagable CasBrE *env* vector, CasE, which encodes both the SU and TM subunits. The expectation being that if CasBrE *env* alone is not sufficient for disease, supplying Gag-pol elements in *trans* will provide the missing components. To address whether Env needed to be membrane associated, as has been reported for prions [15], we also investigated the effects of pseudotyping CasES, a vector that encodes CasBrE SU without the Env TM subunit. However, it is important to note that CasES expression from NSCs results in SU protein that is not able to efficiently bind to its cognate receptor, mCAT-1, a murine cationic amino acid transporter [16], when assessed by superinfection interference [11]. Why the CasBrE SU in the absence of the TM subunit had lost receptor binding activity despite the preservation of the RBD sequences in the N-terminal half of SU was not clear. Nonetheless, the loss of receptor binding might allow us to also investigate whether this activity could also be important for neuropathogenesis. However, before we could address these questions *in vivo*, we needed to examine whether 4070A Env and the CasBrE SU interacted within the engineered NSCs, as previous reports have demonstrated that the Env RBD domain could bind and *trans* complement Env proteins whose RBDs had been deleted or replaced with an alternative receptor specificity [17]. This possibility becomes increasingly important when considering how viral elements engineered into transplanted NSCs and delivered to the CNS could interact to facilitate acute spongiform neuropathogenesis. In this report, we show that CasBrE SU and 4070A proteins physically interact upon synthesis. This interaction results in CasBrE SU expression on the cell surface and facilitates its incorporation into viral particles. In addition, the interaction results in the restoration of CasBrE SU-mCAT-1 binding activity capable of establishing superinfection interference and facilitating ecotropic receptor-dependent virus infection. The finding that SU folding can be affected by

protein-protein interactions has significant implications for understanding how NSC-mediated viral pseudotyping and *trans* complementation in the brain act to precipitate spongiform neurodegeneration.

Results

CasBrE SU is expressed on the cell surface upon NSC infection with an amphotropic helper virus

To assess the feasibility of using a “non-neurovirulent” amphotropic helper virus, to disseminate and *trans* complement CasBrE *env* vectors within the brain by transplanted NSCs, C17.2 NSCs [18] were transduced with retroviral vectors encoding CasBrE SU (pCasES), CasBrE SU/TM (pCasE), or humanized *Renilla* green fluorescent protein (phrGFP), and subsequently infected with the 4070A retrovirus (Figure 1). The resulting cells were initially characterized by fluorescence activated cell sorting (FACS) for cell surface CasBrE Env expression (Figure 2). Importantly, no CasBrE Env immunostaining was observed in control NSCs with or without 4070A infection; however, CasE-NSCs, showed significant CasBrE Env expression both in the presence and absence of 4070A infection, with a small but reproducible increase in cell surface CasBrE Env after 4070A infection. Surprisingly, 4070A infection of CasES-NSCs resulted in the appearance of CasBrE Env on the cell surface, rather than being simply secreted into the medium. This result suggested that 4070A proteins interact with and *trans* complement CasBrE SU in a way that restores the native cellular distribution observed when the protein is made as the SU/TM polyprotein.

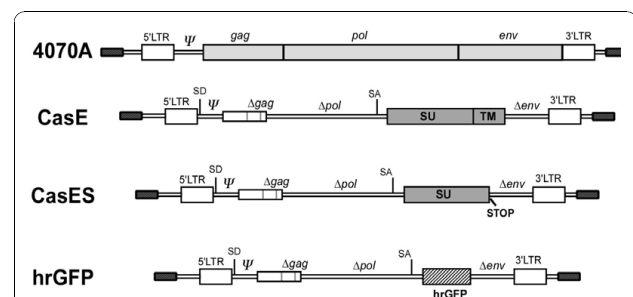
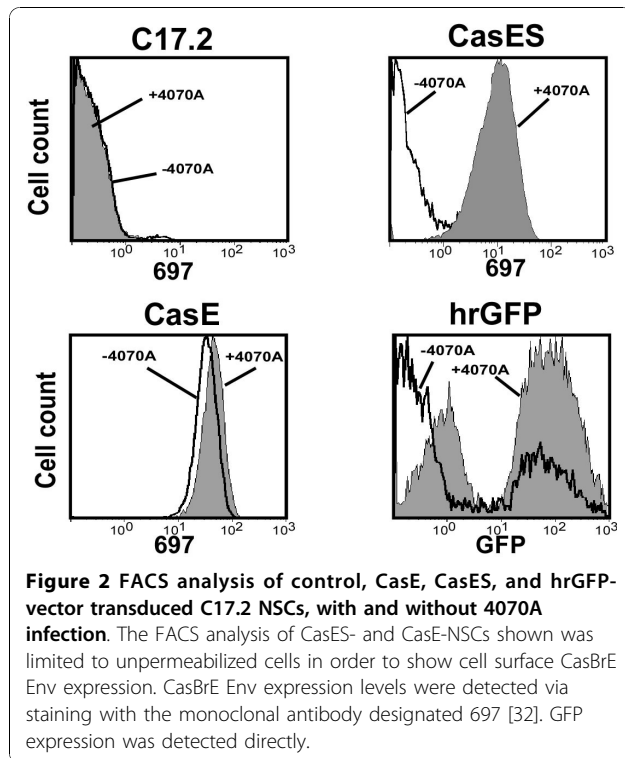


Figure 1 Structures of the amphotropic virus and retroviral vectors introduced into NSCs for pseudotyping and *trans* complementation analysis of CasBrE *env* genes. C17.2 NSCs

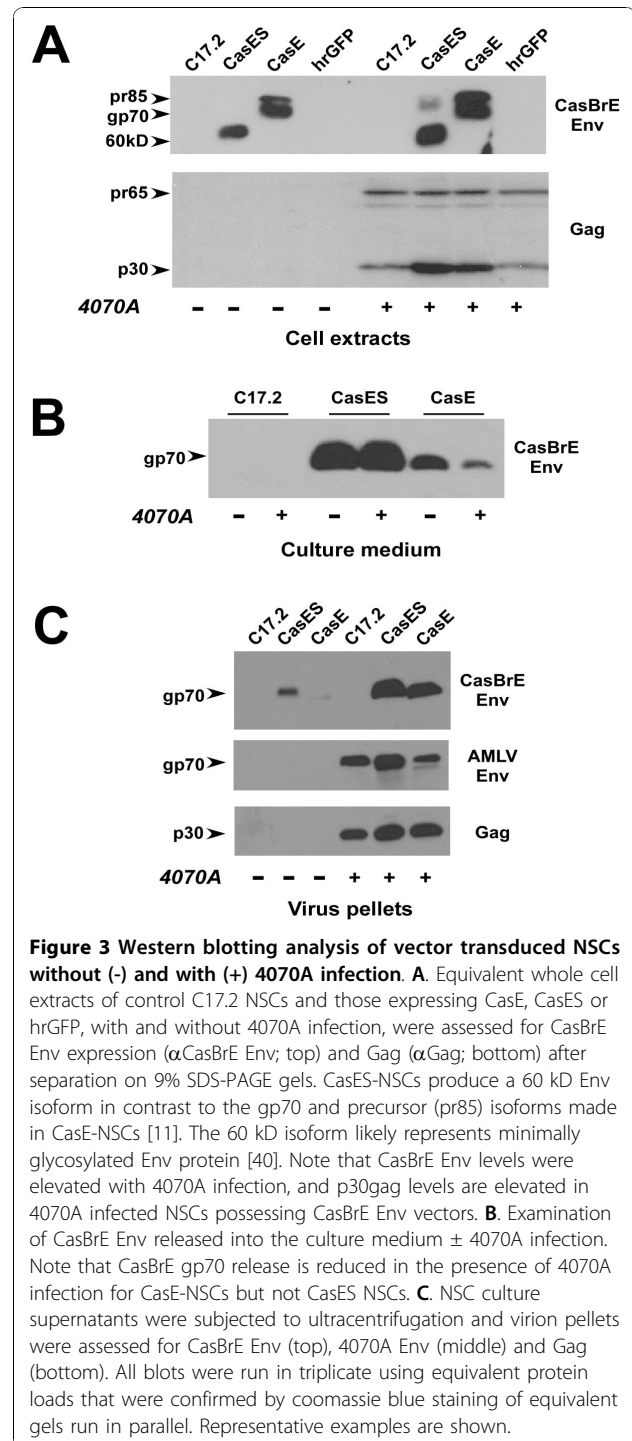
were transduced with the pSFF-based retroviral vectors encoding CasBrE SU/TM (CasE), CasBrE SU (CasES), or the humanized *renilla* green fluorescent protein (hrGFP; striped) followed by infection with the amphotropic virus 4070A. The complementing viral structural elements are shown as filled regions for the 4070A virus and CasE and CasES vectors. The hrGFP vector served as a vector control that could be followed by fluorescence microscopy. Vector elements designated with a Δ represent deleted retroviral structural elements. Ψ indicates the presence of retroviral packaging sequences. SD, Splice donor. SA, Splice acceptor; LTR, Long terminal repeat.



For vector pseudotyping controls, C17.2 NSCs were transduced with vectors encoding GFP. However, due to a lack of stable GFP expression, they could not be directly compared with CasBrE Env encoding vectors and their utility *in vivo* would be similarly limited. In this regard, GFP-sorted NSCs transduced with hrGFP (from *Victoria*; Stratagene) showed that within 10 days after isolation less than 50% of this population exhibited persistent GFP expression (Figure 2, hrGFP, -4070A). The loss of GFP expressing cells reoccurred even after repeated (3x) FACS selection. Surprisingly, infection of these cells with 4070A resulted in a higher level of cells persistently expressing GFP (+4070A), which was maintained through multiple cell passages without reselection. In parallel experiments with NSCs transduced with *Aequorea* EGFP, we were unable to generate NSCs that stably expressed GFP, regardless of whether they were infected with 4070A.

4070A infection of CasBrE Env expressing NSCs influences Env and Gag expression levels

To characterize the potential influence of 4070A virus infection on CasBrE Env expression in NSCs, equivalent cell extracts, culture supernatants, and pelleted culture supernatants were analyzed by Western blotting for CasBrE Env and 4070A proteins (Figure 3). This analysis showed that 4070A infection resulted in a reproducible increase in the levels of cell-associated CasBrE Env



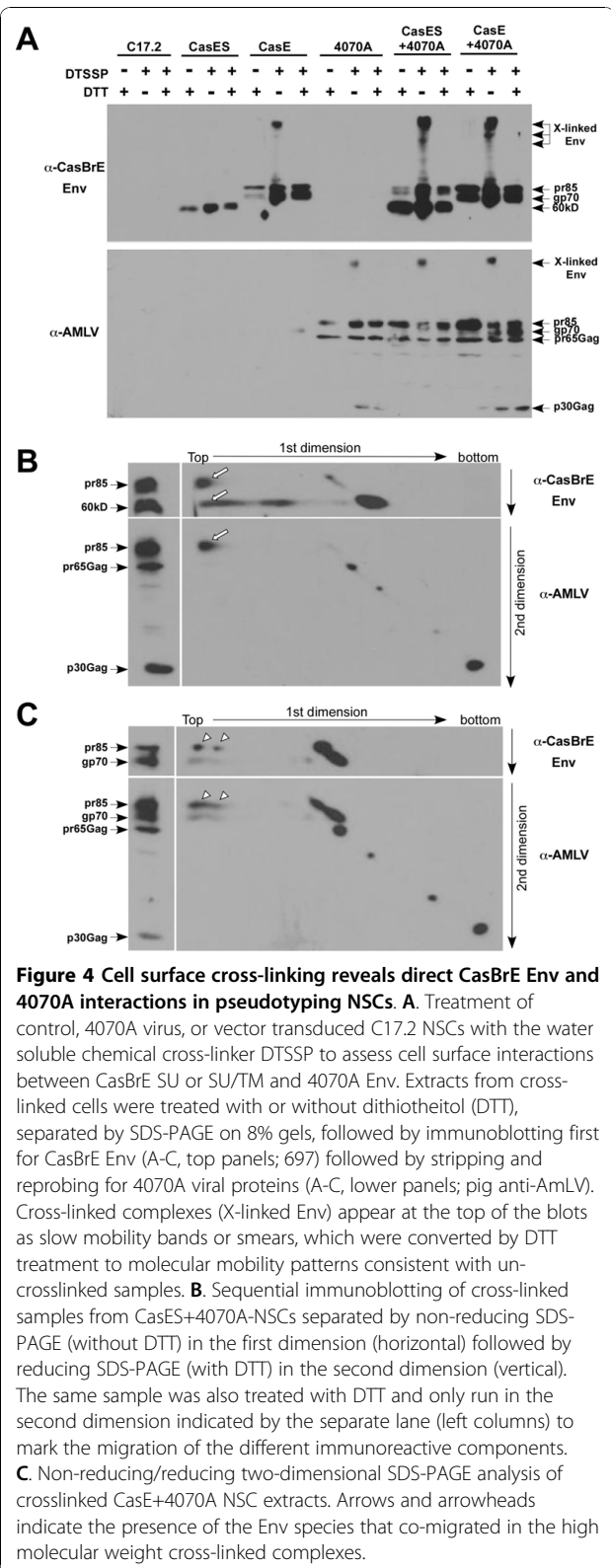
protein (Figure 3A). Specifically, NSCs expressing CasE showed an increase in precursor (pr85) and processed (gp70) Env isoforms, while CasES expressing NSCs showed a similar increase in the 60kD Env isoform that is associated with these cells {cf. [11]}. In addition, CasES NSCs infected with 4070A expressed a small amount of higher apparent molecular weight Env

proteins resolving at 80-85 kD. Analysis of CasBrE Env protein released into the medium from the engineered/infected NSCs (Figure 3B) showed that those cells expressing CasES released significant levels of gp70, which was not appreciably affected by 4070A infection. In contrast, 4070A-infected CasE-NSCs showed a reproducible reduction in CasBrE gp70 release into the medium post infection. It was not specifically explored whether this decrease resulted from the increase in cell-associated CasBrE Env protein noted above. Parallel analysis of Gag expression in the engineered NSCs with and without 4070A infection (Figure 3A, lower panel) showed p30 Gag expression in all 4070A-infected NSC cultures. Notably, after infection with 4070A, p30Gag appeared to be expressed at higher levels in CasE- and CasES-NSCs. Increased Gag levels appeared to positively correlate with CasBrE Env protein, rather than simply the pSFF vector, as hrGFP-NSCs infected with 4070A showed levels of Gag protein indistinguishable from 4070A-infected control NSCs.

To assess whether both CasBrE and 4070A Env proteins were incorporated into viral particles, culture supernatants from the control, CasES- and CasE-NSCs with and without 4070A, were subjected to centrifugation to pellet virus. The sedimented fractions were assessed for CasBrE and 4070A Env by immunoblot. Figure 3C, **far right panels** show that both 4070A and CasBrE gp70 s were associated with p30Gag in the pellets indicating that both 4070A and CasBrE Envs were being incorporated into virions. This is especially noteworthy for the CasES+4070A-NSCs as CasBrE SU has no inherent membrane interacting domain and thus was likely interacting with 4070A proteins to achieve virion incorporation. The small amount of CasBrE SU noted after sedimentation of the CasES supernatant without 4070A represents trace amounts of medium containing high levels of SU rather than the sedimentation of SU aggregates.

CasBrE SU physically interacts with 4070A on the plasma membrane of NSCs

The appearance of CasBrE SU on the cell surface and in virions after NSC infection with 4070A suggested that the 4070A Env interacts with CasBrE SU to tether the latter protein to the plasma membrane. To provide support for this idea, NSCs were exposed to DTSSP, a water soluble, thiol-cleavable crosslinker, to stabilize cell surface protein-protein interactions. Cellular extracts were then subject to Western blotting using both CasBrE Env and amphotropic virus specific antibodies to investigate whether Env hetero-oligomers were formed.



As shown in Figure 4A, low mobility (high apparent molecular weight) CasBrE Env and amphotropic virus-specific bands were detected in CasE-, 4070A-, CasES+4070A-, and CasE+4070A-NSC cultures in the presence of the DTSSP, which were eliminated by reduction with dithiothreitol (DTT). Importantly, no crosslinked CasBrE SU complexes were detected in DTSSP-treated CasES-NSC extracts, consistent with the idea that CasBrE SU multimers or mixed protein complexes were absent from the cell membrane. Because the cross-linked CasES+4070A low mobility bands observed with CasBrE-specific and AMLV-specific antibodies were observed to localize to the same spots when the blots were overlaid, the results suggested that CasBrE SU and 4070A Env were inter-cross-linked on the cell membrane. Similar findings were observed for low mobility bands for CasE+4070A-NSCs, however, the potential for interaction here are less clear here since complexes with low mobility can be seen with CasE- and 4070A-NSCs alone.

To further examine what proteins were contained in the low mobility (high molecular weight), DTSSP-cross-linked bands from CasES+4070A- and CasE+4070A-NSCs, extracts were subjected to non-reducing SDS-PAGE in the first dimension followed by reducing SDS-PAGE in the second dimension, with subsequent analysis by Western blotting for CasBrE Env and 4070A viral proteins. Two CasBrE immunoreactive bands corresponding to pr85 and 60 kD proteins, and one amphotropic pr85 Env band (Figure 4B, **arrows**) were readily detected in the low mobility cross-linked bands obtained from CasES+4070A cells; again, suggesting that CasBrE SU and 4070A Env proteins may directly interact. Similar results were observed for CasE+4070A NSCs with both pr85 and gp70 bands being found in the low mobility bands (Figure 4C), however, because CasE-NSCs and 4070A-NSCs alone also show low mobility complexes after DTSSP treatment, it was not possible to distinguish between homo- versus hetero-oligomers upon dissociation in the presence of reducing agent.

4070A helper virus-infected NSCs efficiently transduce viral and vector genomes

We next examined whether the NSC-generated virions were capable of transducing viral and vector genes to naïve targets. As shown in Figure 5, 4070A-infected NSCs generated total virus titers (**white bars**) greater than 10^6 focus forming units per ml (FFU/ml) for all control or vector engineered NSCs, except those containing hrGFP. The presence of CasES or CasE in the NSCs did not appear to alter infectious virus production from these cells, in seeming contrast to the presence of hrGFP. Infected NSCs examined for the production of virus encoding CasBrE-SU/TM, -SU, or hrGFP showed that all three vectors were

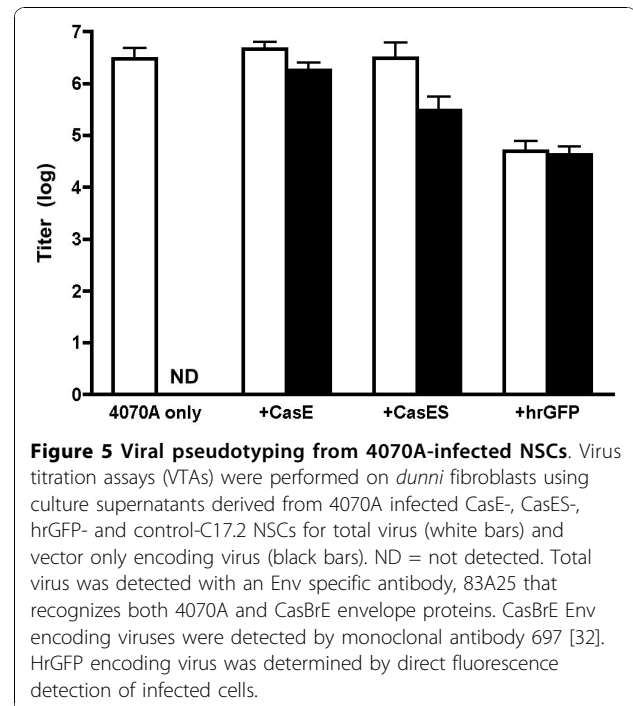
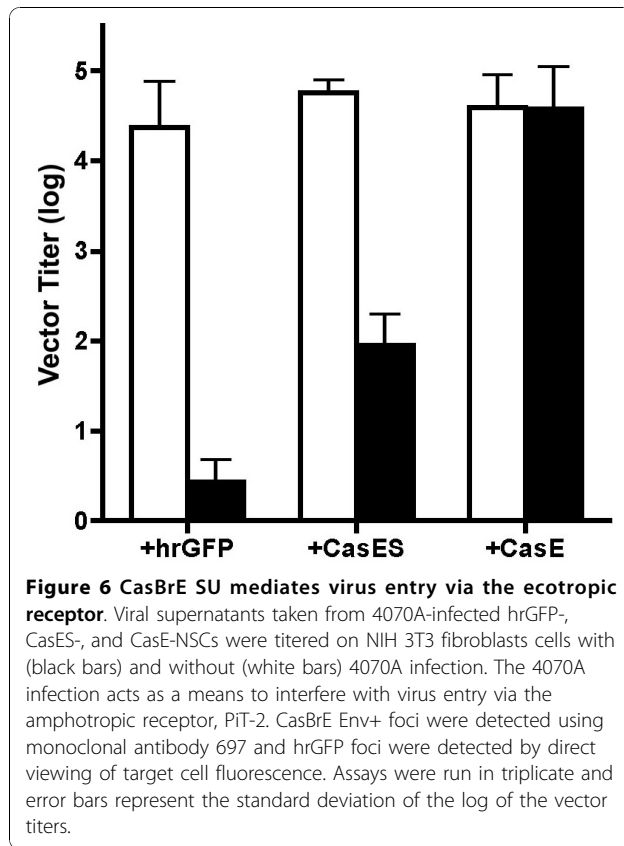


Figure 5 Viral pseudotyping from 4070A-infected NSCs. Virus titration assays (VTAs) were performed on *dunni* fibroblasts using culture supernatants derived from 4070A infected CasE-, CasES-, hrGFP- and control-C17.2 NSCs for total virus (white bars) and vector only encoding virus (black bars). ND = not detected. Total virus was detected with an Env specific antibody, 83A25 that recognizes both 4070A and CasBrE envelope proteins. CasBrE Env encoding viruses were detected by monoclonal antibody 697 [32]. HrGFP encoding virus was determined by direct fluorescence detection of infected cells.

incorporated into infectious virus (Figure 5, **black bars**). While the data suggest that the CasE vector was most efficiently pseudotyped by 4070A, the differences between groups were not statistically significant when analyzed by one way ANOVA at the 95% confidence interval.

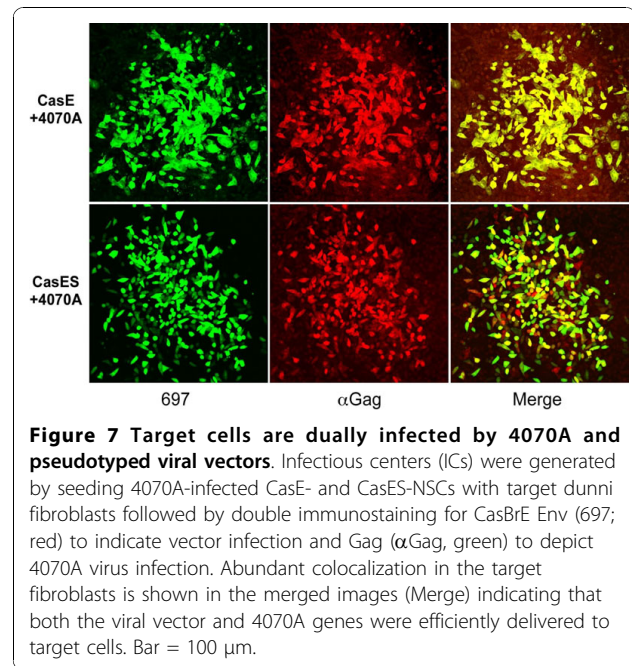
CasBrE SU-4070A Env interactions facilitate virus entry via the ecotropic receptor

To assess whether virion-associated CasBrE SU was capable of initiating virus entry via interactions with the ecotropic receptor, mCAT-1, rather than via the amphotropic receptor, PiT-2, supernatants from hrGFP+4070A-, CasES+4070A- and CasE+4070A-NSCs were examined in a virus titration assay using target fibroblasts (3T3 cells) that had been previously infected with 4070A (3T3Am) to establish amphotropic virus superinfection interference. Foci were scored based on the expression of the vector encoded gene products, CasBrE Env or hrGFP. As shown in Figure 6, supernatants from all three NSC lines exhibited vector titers of approximately 5 logs on normal 3T3 targets (white bars). The vector titer was not detectably diminished when supernatants from CasE+4070A-NSCs were tittered on 3T3Am targets, indicating that CasBrE SU/TM facilitates vector transmission even when the amphotropic receptor was blocked. In contrast, titers from CasES+4070A- and hrGFP+4070A-NSC supernatants were reduced by at least 3 orders of magnitude when assayed on 3T3Am cells (or *dunni*Am cells; not shown). Because hrGFP+4070A-NSC-derived virions do not possess an ecotropic Env protein to overcome amphotropic



viral interference the reduced viral titers were expected, however, with CasES+4070A NSCs, the results suggest that 4070A Env-CasBrE SU interactions responsible for particle incorporation were inadequate to facilitate efficient free virus entry through mCAT-1.

Because retroviral SU-TM interactions are generally considered to be meta-stable in the absence of a disulfide cross-link, and in the case of CasBrE SU-4070A SU/TM, hetero-oligomeric interactions may be even less stable, we asked whether 4070A Env-CasBrE SU



interactions might facilitate more efficient retroviral vector transfer via CasBrE Env-mCAT-1 interactions under conditions of direct cell-cell contact. Therefore, the 4070A-infected viral vector expressing NSCs were examined by infectious center assay on target fibroblasts with and without amphotropic virus infection. As shown in Table 1, the CasE+4070A, CasES+4070A-, and hrGFP+4070A-NSCs scored very efficiently as infectious centers on naïve dunnii fibroblasts (cf., Figure 7), and suggested that essentially all the helper virus-infected NSCs appeared capable of efficiently delivering retroviral vectors to susceptible target cells *in vitro*, despite the variability in plating efficiency. Importantly, when these same NSCs were seeded onto

Table 1 CasBrE Env-NSC mediated superinfection of amphotropic-restricted targets through cell-cell contact

NSCs	Infectious Centers ^a		Plating Efficiency ^b	Infectious Center Efficiency ^c	
	Dunnii	DunniiAm		Dunnii	DunniiAm
hrGFP+4070A	10.5 \pm 1.3	< 0.05*	2.5 \pm 1	420%	<2%
CasES+4070A	67 \pm 8.3	18 \pm 3	16.5 \pm 3	406%	109%
CasE+4070A	49 \pm 5.0	64.7 \pm 7.2	16.2 \pm 3.8	302%	400%

a- Mitomycin C-treated 4070A-NSCs expressing GFP, CasE or CasES were counted and serial dilutions were seeded with 10^5 dunnii fibroblasts. The number of foci expressing the viral vector once the Dunnii cells reached confluence was evaluated and expressed as foci/per 100 NSCs seeded.

b- To determine the % plating efficiency, the mitomycin C-treated NSCs were counted and seeded as in "a" in the absence of dunnii cells and the percentage of vector expressing cells present on these plates, versus the number seeded, was assessed at 48 hours.

c- IC efficiency is simply the number of foci/100 NSCs seeded divided by the plating efficiency expressed as a percentage.

*-The detection of an infectious center is limited by the maximum number of NSCs seeded per plate of targets. For these experiments it was limited to \leq 2000 NSCs/plate.

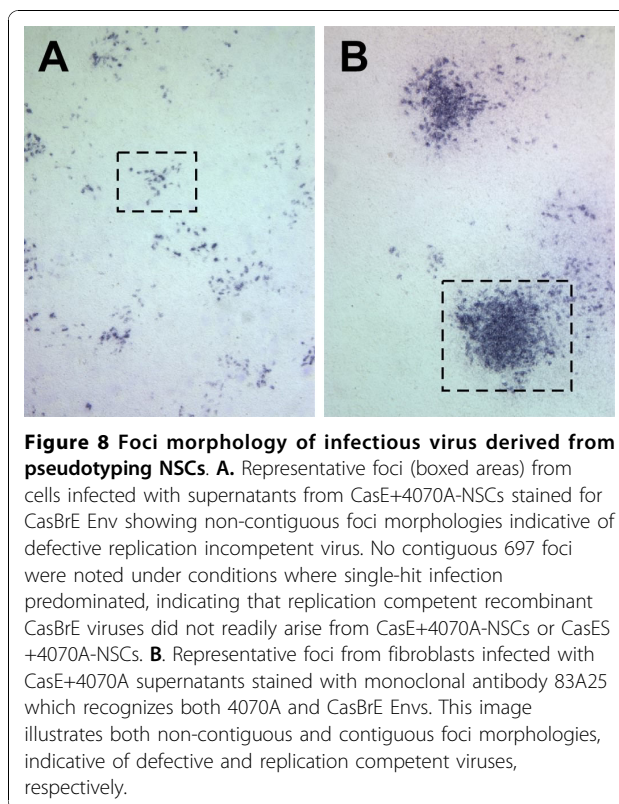
4070A-infected durni fibroblasts (durniAm), only hrGFP+4070A-NSCs failed to generate viral vector positive foci. CasES+4070A-NSC ICs were readily observed in durniAm cultures, although the numbers were reduced from their value in naïve durni cell cultures. In contrast, CasE+4070A-NSCs showed no decrease in IC titers on durniAm cells. These results indicate that in the presence of the 4070A Env, CasBrE SU facilitates ecotropic entry, but not with the same efficiency as native CasBrE SU/TM.

Helper virus-infected NSCs deliver both virus and vector to naïve target cells

To assess whether the 4070A-infected vector-expressing NSCs could deliver both virus and vector to individual *durni* fibroblasts to facilitate *trans* complementation within those targets, infectious center foci were assessed for CasBrE Env and 4070 Gag (Figure 7). Examination of the merged images showed that virtually all the Env+ cells (green) were also Gag+ (red) with CasE+4070A-NSCs. In foci generated from CasES+4070A NSCs approximately 60% of cells were double positive, whereas, approximately 15% were singly positive for CasBrE SU, and 25% singly positive for Gag. Analysis of hrGFP+4070A NSCs gave results similar to those generated by the CasES vector with between 50-70% of cells in foci expressing GFP and Gag (not shown).

4070A helper virus infection of CasBrE Env expressing NSCs does not result in the generation of recombinant replication competent CasBrE virus

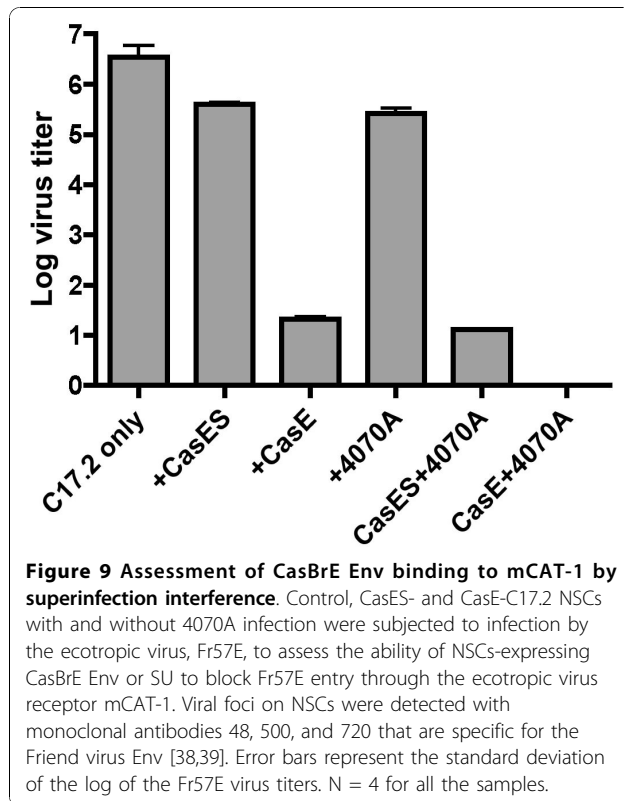
Despite findings indicating that MLVs infrequently co-package retroviral genomes arising from different DNA transcriptional templates [19], the potential still existed for possible recombination between the helper virus and the retroviral vectors containing CasBrE Env that could be amplified. Thus, NSC supernatants were evaluated for the presence of replication competent CasBrE Env-encoding virus by examining foci contiguity at virus dilutions where single-hit kinetics predominate as previously outlined [14]. As shown by the example in Figure 8, **left panel**, all CasBrE Env positive (697+) foci appeared non-contiguous (small box), indicating that the initial virus infectious event did not spread cell-to-cell from the primary target, but rather expanded due to target cell division and migration. The lack of detectable contiguous foci suggests that if recombination did occur it did so at a very low frequency, given that more than 1000 foci were examined. Similar results were observed for supernatants from infected NSCs expressing CasES (not shown). In contrast, examination of parallel assays at the same time, with antibody reactive to both 4070A and CasBrE Envs (83A25), showed abundant contiguous foci (**box area, right panel**) indicative



of cell-to-cell spread of the replication competent 4070A virus.

CasBrE SU-4070A Env interactions facilitate efficient binding and blocking of mCAT-1 in NSCs

The interactions between CasBrE SU and 4070A Env appear to at least partially restore CasBrE SU ecotropic receptor binding and fusion activity, so we further explored the extent to which this interaction could influence superinfection interference by binding to mCAT-1 within the NSCs and prevent infection by a second ecotropic virus, Fr57E. The results, shown in Figure 9, indicate that CasES expression alone within NSCs only reduced superinfection by the ecotropic virus, Fr57E, by ~1 log, whereas, CasE expression reduced infectivity by ~5 logs compared to control NSCs. 4070A infection of NSCs also reduced the Fr57E titer by ~1 log, however, 4070A infection of CasES- and CasE-NSCs reduced Fr57E titers by 5 and 6 logs respectively. The 4 log increase in ecotropic retrovirus superinfection interference when 4070A and CasBrE SU are co-expressed suggests that 4070A Env-CasBrE SU interactions result in restoration of near native CasBrE SU-mCAT-1 binding interactions analogous to CasBrE SU/TM. Whether such intracellular Env-receptor interactions play a role in retroviral neuropathogenesis has not been specifically investigated, but in the helper virus



paradigm being explored herein, CasBrE SU appeared capable of carrying out the same functions as SU/TM, albeit with more limited specific activity, and only in the presence of the *trans* complementing protein from 4070A.

Discussion

In this report we examined the capacity of the 4070A amphotropic virus to *trans* complement and pseudotype CasBrE *env* vectors when co-expressed in NSCs as a prelude to transplanting these cells in the CNS in order to define what retroviral elements and interactions were required to induce spongiform neurodegeneration. This analysis revealed that the engineered NSCs could indeed deliver both the *env* vector and complementing 4070A virus to target cells in culture, in the absence of detectable recombinant virus. Given these results, the potential exists for exploiting this capacity for NSC-mediated pseudotyping and *trans* complementation to reveal insights into the elements and activities involved in retroviral neurovirulence and their capacity to target certain CNS cell types. However, our *in vitro* analysis herein, also revealed how viral protein-protein interactions might be expected to alter the interpretation of the *in vivo* experiments where these tools are employed. Specifically, we showed that direct heteromeric interactions between 4070A Env and the protein product of

the CasES vector, CasBrE SU, restored CasBrE SU's capacity to associate with the plasma membrane, become incorporated into viral particles, bind to the ecotropic receptor and trigger virus fusion.

We previously reported that when CasBrE SU is made in the absence of TM subunit sequences, the protein possessed little ecotropic receptor binding activity and was secreted from the cell [11]. Presumably the absence of TM sequences in the Env precursor protein prevented the normal folding of the CasBrE receptor binding domain (RBD) found within SU. The loss of receptor binding activity for CasBrE SU was not predicted given the findings of Davey et al. showing that eukaryotic cell synthesis of the N-terminal one third of the Friend virus SU results in receptor binding activity equivalent to the full length protein (SU/TM) [20]. Spatial modeling of the CasBrE RBD sequence, based on the crystal structure defined for the RBD from the Friend virus [21], predicts a similar structure for CasBrE RBD (unpublished observations). Thus, sequences in the latter two thirds of SU may negatively influence the proper folding of the CasBrE SU in the absence of TM sequences. It is clear from studies by Lavillette et al., that there are critical interactions between the C-terminal domain of SU and the RBD that affect various Env entry activities [22]. Thus, the results here suggest that, at least for CasBrE Env, sequences within TM may help to facilitate the proper folding of these domains in CasBrE SU.

Given the virtual absence of CasBrE SU receptor binding activity and cell surface expression originally observed in CasES NSCs, we predicted that 4070A infection of CasBrE SU-expressing NSCs would result in the production of virus that would demonstrate only amphotropic tropism while packaging either 4070A or CasES genomes. It was therefore surprising to observe CasBrE SU on the cell surface of CasES +4070A-NSCs in the absence of any recombinant virus. The follow-up cross-linking, sedimentation, infection and superinfection analyses demonstrated that direct 4070A Env-CasBrE SU interactions increased ecotropic receptor interactions by 10,000 fold and facilitated SU incorporation into virions that could activate fusion activity.

How the lost SU activities were restored is not clear, but it is of interest to speculate that the 4070A Env may have acted as a folding template for CasBrE SU during its synthesis through the secretory pathway. In this way, Env would be analogous to the prion protein where the scrapie form is able to direct the folding and ultimate conformation of the unfolded protein through direct interactions. This idea appears to be a reoccurring theme in a diverse array of infectious, sporadic and genetic neurodegenerative diseases often characterized

by protein aggregation. While retroviral neuropathogenesis is not characterized by identifiable protein aggregates, their ability to form multimers may provide a structural link between such diseases. The possibility that template-based protein folding extends to retrovirus-induced neurodegeneration such as HTLV-1 associated myelopathy or HIV associated dementia is provocative.

An alternative to the template-based folding idea is that SU interactions with 4070A SU or TM may have facilitated more efficient or prolonged SU interactions with ER chaperones through non-covalent tethering of SU to the membranes in the secretory pathway. However, we observed that CasBrE SU synthesized alone appears to be fully glycosylated and of the correct molecular size upon its secretion into the medium, suggesting that it has been routed and processed through the requisite ER-golgi protein quality control machinery that includes lectins and sugar processing enzymes (reviewed in [23-25]). Whether the addition of a simple transmembrane domain or a glycosphosphatidyl inositol linker to CasBrE SU would result in a similar restoration of CasBrE SU binding to mCAT-1 might help distinguish between these two possibilities.

The observation that correctly folded CasBrE SU could facilitate ecotropic receptor binding and entry in the presence of amphotropic Env is consistent with the modular nature of type-C retroviral Envs described by Barnett et al. [17]. These investigators showed that the addition of soluble Friend RBD could restore infectivity when the RBD was deleted from ecotropic, amphotropic, or xenotropic Envs. Moreover, this group showed that RBD could enhance infectivity 50-fold when Env RBD sequences were replaced by erythropoietin as an alternate receptor specificity. Herein, CasBrE-SU facilitated ecotropic receptor-mediated virus fusion indicating that the CasBrE SU induced conformational changes in 4070A derived-TM trimers required for membrane fusion. Whether this involved CasBrE SU binding to functional 4070A SU/TM monomers, dimers, or trimers, or represented CasBrE SU displacement of 4070A SU to interact with TM is not clear. The cross-linking studies appear to suggest that heteromeric CasBrE-4070A SU-SU interactions were occurring, but the exact nature of the fusogenic structures is a matter of conjecture at this stage. Interestingly, a previous report by Yang et al. suggests that ecotropic and amphotropic Env hetero-trimers are not formed when co-expressed in the same cell, however, this apparent discrepancy could be accounted for by the fact that differing ecotropic viruses were employed in the two studies. In this regard, the CasBrE virus was initially co-isolated from the wild along with amphotropic viruses [26], suggesting that the two viruses likely co-evolved. Nonetheless, since we did not

observe full restoration of CasBrE Env binding and fusion activity in this complementation paradigm, it suggests that CasBrE SU-4070A TM heterodimer formation was either not highly efficient or that misfolded CasBrE SU might also compete with natively folded SU for association with 4070A SU/TM or TM proteins. It is clear that a more detailed analysis of the protein complexes being formed will be required to reveal insight into the nature of the specific protein-protein interactions taking place.

Given that previous experiments investigating prion protein neuropathogenesis have demonstrated that the glycosphosphatidyl inositol membrane anchor was required for prion protein induced neurodegeneration but not replication [15], we initially thought that NSC-mediated 4070A delivery of CasES to CNS cells would allow us to assess whether ecotropic receptor binding activity and/or Env membrane association were required for MLV-induced spongiform neurodegeneration. However, since these Env activities were restored by CasBrE SU-4070A Env interactions, these questions cannot be effectively addressed using this helper virus pseudotyping approach. Nonetheless, the capacity for *trans* complementation *in vivo* examined in the companion study was able to show that ecotropic entry is a critical step in the pathogenic process, and that the “non-neurovirulent” virus amphotropic virus contains neurovirulent elements that could be revealed by such an analysis. Perhaps more dramatically, these findings of Env-SU interactions highlight the difficulty in assessing which *in vivo* Env interactions are responsible for the induction of neuropathogenic changes given that the mouse genome, like that of other mammals, is replete with retroviral sequences. Ruling out that exogenous viral proteins interact with host retroviral elements to cause disease may be a Sisyphean task.

Conclusions

The findings presented here show that co-expression of 4070A and CasBrE SU results in a physical interaction between 4070A Env and CasBrE SU. This interaction facilitates CasBrE SU retention on the plasma membrane, incorporation into viral particles and restoration of its ability to interact with its receptor, mCAT-1. Moreover, Env-SU oligomers were capable of initiating ecotropic receptor-dependent TM-mediated fusion events, despite reduced efficiencies in free virus. Thus, the results indicate that heterotropic Env-SU interactions that occur during protein synthesis facilitate CasBrE SU folding events and interactions with TM such that ecotropic viral binding and membrane fusion events were almost fully restored. This template based folding process is reminiscent of prion formation, where one protein conformation acts as a scaffold for the folding of

a second protein of similar primary structure. These findings support the idea that protein scaffold-based protein folding represents an inherent feature of neuro-pathogenic proteins that appears to extend to retroviral Envs.

Methods

DNA Constructs

The CasBrE env expression vectors CasE (SU/TM), CasES (SU), described previously [11], contain the *env* gene derived from the CasBrE molecular clone 15.1 [27,28] (GenBank accession number X57540) inserted into the pSFF vector [29]. In order to assess effects associated with vector alone, a pSFF-based vector encoding humanized GFP from Renilla (hrGFP, Stratagene) was generated by introducing a Xho I and Sal I fragment from pFB-hrGFP (Promega) into the multiple cloning site of pSFF.

Cells

C17.2 neural stem cell lines with and without expression of CasBrE SU or SU/TM have been described previously [11,30] and were grown on Primaria dishes (Falcon) in DMEM supplemented with 10% fetal bovine serum and 100 U of penicillin and streptomycin per ml. *Mus dunni*, NIH 3T3 fibroblasts, and PT67 packaging/producer cells (Invitrogen) were maintained in DMEM with 10% newborn calf serum and 100 U of penicillin and streptomycin per ml on Nunclon D™ dishes (Nunc).

HrGFP-NSCs were generated by exposing C17.2 cells to hrGFP virus at a multiplicity of infection of 1 in the presence of 8 µg/mL polybrene. Positive NSCs were sorted by flow cytometry with the brightest 10% selected and used for subsequent analysis. C17.2 NSCs with and without SU, SU/TM, hrGFP or 15-1EIH were infected with the 4070A virus at an MOI of 1 in the presence of 8 µg/mL polybrene, and passaged 4 times prior to analysis for CasBrE Env and hrGFP expression and 4070A virus pseudotyping.

Viruses

4070A amphotropic virus stocks were generated by transfecting the permuted proviral clone p4070A [31] into NIH3T3 cells. Virus encoding hrGFP was generated by co-transfection of PT67 packaging producer cells (Invitrogen) with pCDNA3.1-hygro and phrGFP followed by collection of viral supernatants from hygromycin selected cultures. Similarly, CasE, CasES, and SFF-FE virus stocks were generated by transfection of PT67 cells with the corresponding plasmid.

FACS analysis

NSCs were removed from tissue culture plates by using 0.05% trypsin with EDTA (Gibco), quenched with DMEM+10% NCS, washed with PBS, and incubated at 4°

C for 30 minutes with the mouse monoclonal antibody 697 for detecting CasBrE Env [32], or mouse monoclonal 48 for Friend 57 Env. Cells were washed three times with PBS, and incubated with Alexa Fluor 488-conjugated goat anti-mouse IgG (Molecular Probes) for another 30 min before analysis. HrGFP expression within NSCs was analyzed directly. Analysis and sorting was carried out on an Altra Flow Cytometer (Beckman/Coulter).

Western blotting

Tissue culture cell extracts were generated as previously outlined [11,33] and equivalent samples were separated by 8-10% SDS-PAGE, transferred to nitrocellulose membrane, blocked with 5% nonfat milk in TBS. To detect CasBrE Env and SU mouse monoclonal antibody 697 was used. Gag proteins were detected using rat anti-capsid R187 monoclonal antibody or rabbit anti-capsid antisera R3. Amphotropic viral proteins were detected using swine anti-AMLV serum (NCI). Primary antibodies or antisera was detected using horseradish peroxidase (HRP)-labeled goat anti-mouse IgG (Southern Biotechnologies), HRP-rabbit anti-swine IgG (RDI), HRP-donkey anti-rat IgG (Jackson Immunoresearch), or HRP-goat anti-rabbit IgG (Southern Biotechnologies), which were detected by chemiluminescence using ECL reagents (Pierce). Equivalent amounts of proteins were loaded for each tissue or cell sample and which was confirmed by Coomassie blue staining of a gel run in parallel in each experiment and by post transfer staining of the gel utilized in the transfer.

Chemical cross-linking of cell surface proteins

The water soluble chemical cross-linker dithiobis sulfo-succinimidylpropionate, DTSSP (Pierce), was dissolved in PBS (pH 7.2) to a concentration of 1 mM just before use. Full confluent cells were washed with PBS three times and then incubated with 1 mM DTSSP solution at room temperature for 30 min, followed by washing three times with Tris-buffered saline (pH 7.5). DTSSP treated cells were extracted by using lysis buffer containing 0.1% Triton X-100 as previously described [33]. Cleared crosslinked samples were analyzed for viral proteins by Western blotting after non-reducing sodium dodecylsulfate polyacrylamide gel electrophoresis (SDS-PAGE) in the first dimension and reducing SDS-PAGE in the second dimension. Briefly, equivalent samples were boiled in SDS-PAGE sample buffer without DTT and resolved on an 8% gel and analyzed by Western blotting using monoclonal 697 for CasBrE Env and swine anti-AMLV (NCI) for 4070A proteins. To assess the composition of the cross-linked viral proteins, first dimension gel strips were equilibrated with sample buffer containing 100 mM DTT at room temperature for 15 min, positioned on the top of a second 8% gel, and

then subjected to electrophoresis in the orthogonal directions and again analyzed for viral proteins by sequential immunoblotting.

Virus titration assay

Virus titers of tissue culture supernatants were determined by a focus forming assay on *dunni* cells [34]. Supernatants collected from confluent cell cultures were frozen at -80°C before analysis. Viral Env positive foci were detected by using CasBrE Env specific MAb 697 or rat MAb 83A25 [35] which recognizes an epitope common to all known endogenous and most exogenous MLV Envs. Alkaline phosphatase-labeled anti-mouse IgG or anti-rat IgG antibodies (Promega) and Western Blue stabilized substrate for alkaline phosphatase (Promega) were used. GFP positive foci were determined directly under a fluorescence microscope. Virus foci were scored based on their morphology as contiguous or non-contiguous, the latter being consistent with defective virus [14]. To determine CasBrE SU positive or Gag positive foci, cells were permeabilized with ice cold methanol for 5 min before staining.

Infectious center assay

In order to assess NSCs' ability to disseminate viral vectors by cell-to-cell contact, infectious center assays were performed as outlined by Czub et al. [36] with minor modifications. Briefly, transduced C17.2 NSCs were treated with mitomycin C (Fisher) at $10\ \mu\text{g}/\text{ml}$ in DMEM medium at 37°C for 2 hr, washed with PBS three times, trypsinized and resuspended in DMEM+10% FBS. Treated NSCs were seeded into TC-6 tissue culture plates using serial dilutions yielding from 100 to 2000 cells per well. Wells were also seeded with 1×10^5 *dunni* target cells per well. Once the *dunni* cells reach confluent, 697, R187 and MAbs 720/500/48 were used to detect CasBrE Env, amphotropic Gag and Friend Env expression respectively, followed by Alexa Fluor 488 goat anti-mouse IgG, Alex Fluor donkey 488 anti-goat IgG and Alexa Fluor 594 donkey anti-rat IgG (Molecular Probes) respectively, and foci were examined under the epifluorescence microscopy. Images were merged to colocalize CasBrE Env or SU and 4070A Gag expression in some instances. Plates without *dunni* cells were used to assess the plating efficiency of the mitomycin-treated NSCs and to normalize the number of foci generated.

To study whether CasES+4070A-NSCs were capable of delivering virus via the ecotropic receptor by cell-cell contact, a similar assay was performed in which naïve *dunni* and amphotropic 4070A virus infected *dunni* (DunniAm) cells were used as target cells. CasBrE SU positive foci were determined. As controls, CasE-4070A-NSCs and hrGFP-4070A-NSCs were analyzed in parallel.

Superinfection interference assays

To study whether CasBrE SU and Env could facilitate virus entry via the ecotropic receptor in the presence of the amphotropic 4070A virus, the superinfection interference assay was carried out as described previously [37]. Briefly, NSCs were infected with serially diluted ecotropic MLV Fr57E in the presence of polybrene ($8\ \mu\text{g}/\text{ml}$). Friend Env positive foci were evaluated by its' specific monoclonal antibodies 500, 48 and 720 [38,39], followed by Alexa Fluor 488 goat anti-mouse IgG (Molecular Probes). GFP positive foci were counted under a fluorescent microscopy directly.

To investigate whether the viruses generated by CasES+4070A-NSCs was capable of infecting targets via the ecotropic receptor, a similar method was carried out, in which 4070A virus infected-NIH 3T3 (3T3Am) cells and naïve 3T3 cells were used as target cells and SU positive foci were determined by MAb 697. As controls, viruses generated by CasE+4070A- and hrGFP+4070A-NSC culture were evaluated in parallel.

Acknowledgements

We thank Ninuan Huang and Rochelle Cutrone for technical support and Dr. Angelo DeLucia and Ms. Sandra Cardona for a critical reading of the manuscript. This work was supported in part by grants from the ALS Association and NIH grant NS37614 to WPL.

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Authors' contributions

YL carried out the majority of the experimental studies, helped design the experiments, analyzed the data and provided a preliminary draft of the manuscript. WPL conceived of the study, participated in its design and coordination, performed experiments, analyzed the data, and drafted the manuscript. Both authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

Received: 29 July 2010 Accepted: 5 November 2010

Published: 5 November 2010

References

1. Gardner MB: Retroviral spongiform polioencephalomyelopathy. *Rev Infect Dis* 1985, **7**:99-110.
2. Gardner MB, Henderson BE, Officer JE, Rongey RW, Parker JC, Oliver C, Estes JD, Huebner RJ: A spontaneous lower motor neuron disease apparently caused by indigenous type-C RNA virus in wild mice. *JNCI* 1973, **51**:1243-1254.
3. Gardner MB, Rasheed S, Klement V, Rongey RW, Brown JC, Dworsky R, Henderson BE: Lower motor neuron disease in wild mice caused by indigenous type C virus and search for a similar etiology in human amyotrophic lateral sclerosis. *UCLA Forum Med Sci* 1976, **19**:217-234.
4. DesGroseillers L, Barrette M, Jolicoeur P: Physical Mapping of the paralysis-inducing determinant of a wild mouse ecotropic neurotropic virus. *J Virol* 1984, **52**:356-363.
5. Jolicoeur P, Gravel C, Kay DG: Pathogenesis of murine spongiform myeloencephalopathy induced by a murine retrovirus. In *Molecular*

- Neurovirology*. Edited by: Roos RP. Totowa, NJ: Humana Press Inc; 1992:199-224.
6. Portis JL: **Genetic determinants of neurovirulence of murine oncornaviruses.** *Adv Virus Res* 2001, **56**:3-38.
 7. Portis JL, Lynch WP: **Dissecting the determinants of neuropathogenesis of the murine oncornaviruses.** *Virology* 1998, **247**:127-136.
 8. Wong PKY, Yuen PH: **Molecular basis of neurologic disorders induced by a mutant ts1, of Moloney murine leukemia virus.** In *Molecular Neurovirology: Pathogenesis of Viral CNS Infections*. Edited by: Roos R. Totowa: Humana Press; 1992:161-197.
 9. Murphy SL, Honczarenko MJ, Dugger NV, Hoffman PM, Gaulton GN: **Disparate regions of envelope protein regulate syncytium formation versus spongiform encephalopathy in neurological disease induced by murine leukemia virus TR.** *J Virol* 2004, **78**:8392-8399.
 10. Paquette Y, Hanna Z, Savard P, Brousseau R, Robitaille Y, Jolicoeur P: **Retrovirus-induced murine motor neuron disease: Mapping the determinant of spongiform degeneration within the envelope gene.** *Proc Natl Acad Sci USA* 1989, **86**:3896-3900.
 11. Lynch WP, Snyder EY, Qualtieri L, Portis JL, Sharpe AH: **Late virus replication events in microglia are required for neurovirulent retrovirus-induced spongiform neurodegeneration: evidence from neural progenitor-derived chimeric mouse brains.** *J Virol* 1996, **70**:8896-8907.
 12. Kay DG, Gravel C, Pothier F, Laperriere A, Robitaille Y, Jolicoeur P: **Neurological disease induced in transgenic mice expressing the env gene of the Cas-Br-E murine retrovirus.** *Proc Natl Acad Sci USA* 1993, **90**:4538-4542.
 13. Yu YE, Choe W, Zhang W, Stoica G, Wong PK: **Development of pathological lesions in the central nervous system of transgenic mice expressing the env gene of ts1 Moloney murine leukemia virus in the absence of viral gag and pol genes and viral replication.** *Journal of Neurovirology* 1997, **3**:274-282.
 14. Lynch WP, Sharpe AH, Snyder EY: **Neural stem cells as engraftable packaging lines can mediate gene delivery to microglia: evidence from studying retroviral env-related neurodegeneration.** *J Virol* 1999, **73**:6841-6851.
 15. Chesebro B, Trifilo M, Race R, Meade-White K, Teng C, LaCasse R, Raymond L, Favara C, Baron G, Priola S, et al: **Anchorless prion protein results in infectious amyloid disease without clinical scrapie.** *Science* 2005, **308**:1435-1439.
 16. Albritton LM, Tseng L, Scadden D, Cunningham JM: **A putative murine ecotropic retrovirus receptor gene encodes a multiple membrane-spanning protein and confers susceptibility to virus infection.** *Cell* 1989, **57**:659-666.
 17. Barnett AL, Davey RA, Cunningham JM: **Modular organization of the Friend murine leukemia virus envelope protein underlies the mechanism of infection.** *PNAS* 2001, **98**:4113-4118.
 18. Snyder EY, Deitcher DL, Walsh C, Arnold-Aldea S, Hartweg E, Cepko CL: **Multipotent neural cell lines can engraft and participate in development of mouse cerebellum.** *Cell* 1992, **68**:33-51.
 19. Flynn JA, An W, King SR, Telesnitsky A: **Nonrandom dimerization of murine leukemia virus genomic RNAs.** *J Virol* 2004, **78**:12129-12139.
 20. Davey RA, Hamson CA, Healey JJ, Cunningham JM: **In vitro binding of purified murine ecotropic retrovirus envelope surface protein to its receptor, MCAT-1.** *J Virol* 1997, **71**:8096-8102.
 21. Fass D, Davey RA, Hamson CA, Kim PS, Cunningham JM, Berger JM: **Structure of a Murine Leukemia Virus Receptor-Binding Glycoprotein at 2.0 Angstrom Resolution.** *Nature* 1997, **277**:1662-1666.
 22. Lavillette D, Boson B, Russell SJ, Cosset FL: **Activation of membrane fusion by murine leukemia viruses is controlled in cis or in trans by interactions between the receptor-binding domain and a conserved disulfide loop of the carboxy terminus of the surface glycoprotein.** *J Virol* 2001, **75**:3685-3695.
 23. Anelli T, Sitia R: **Protein quality control in the early secretory pathway.** *Embo J* 2008, **27**:315-327.
 24. Helenius A, Aebi M: **Roles of N-linked glycans in the endoplasmic reticulum.** *Annu Rev Biochem* 2004, **73**:1019-1049.
 25. Vembar SS, Brodsky JL: **One step at a time: endoplasmic reticulum-associated degradation.** *Nat Rev Mol Cell Biol* 2008, **9**:944-957.
 26. Gardner MB: **Type C viruses of wild mice-Characterization and natural history of amphotropic, ecotropic, and xenotropic MuLV.** *Curr Top Microbiol Immunol* 1978, **79**:215-259.
 27. Perryman S, Nishio J, Chesebro B: **Complete nucleotide sequence of Friend murine leukemia virus, strain FB29.** *Nucleic Acids Res* 1991, **19**:6950.
 28. Portis JL, Czub S, Garon CF, McAtee FJ: **Neurodegenerative disease induced by the Wild Mouse ecotropic retrovirus is markedly accelerated by long terminal repeat and gag-pol sequences from nondefective Friend murine leukemia virus.** *J Virol* 1990, **64**:1648-1656.
 29. Kozak SL, Kabat D: **Ping-pong amplification of a retroviral vector achieves high level gene expression: human growth hormone production.** *J Virol* 1990, **64**:3500-3508.
 30. Ryder EF, Snyder EY, Cepko CL: **Establishment and characterization of multipotent neural cell lines using retrovirus vector-mediated oncogene transfer.** *Journal of Neurobiology* 1990, **21**:356-375.
 31. Miller AD, Law M-F, Verma IM: **Generation of helper-free amphotropic retroviruses that transduce a dominant-acting, methotrexate-resistant dihydrofolate reductase gene.** *Mol Cell Biol* 1985, **5**:431-437.
 32. McAtee FJ, Portis JL: **Monoclonal antibodies specific for wild mouse neurotropic retrovirus: Detection of comparable levels of virus replication in mouse strains susceptible and resistant to paralytic disease.** *J Virol* 1985, **56**:1010-1022.
 33. Lynch WP, Brown WJ, Spangrude GJ, Portis JL: **Microglial infection by a neurovirulent murine retrovirus results in defective processing of envelope protein and intracellular budding of virus particles.** *J Virol* 1994, **68**:3401-3409.
 34. Czub M, Czub S, McAtee F, Portis J: **Age-dependent resistance to murine retrovirus-induced spongiform neurodegeneration results from CNS-specific restriction of virus replication.** *J Virol* 1991, **65**:2539-2544.
 35. Evans LH, Morrison RP, Malik FG, Portis J, Britt WJ: **A neutralizable epitope common to the envelope glycoproteins of ecotropic, polytropic, xenotropic, and amphotropic murine leukemia viruses.** *J Virol* 1990, **64**:6176-6183.
 36. Czub M, McAtee FJ, Portis JL: **Murine retrovirus-induced spongiform encephalomyelopathy: host and viral factors which determine the length of the incubation period.** *J Virol* 1992, **66**:3298-3305.
 37. Miller AD, Wolgamot G: **Murine retroviruses use at least six different receptors for entry into *Mus dunni* cells.** *J Virol* 1997, **71**:4531-4535.
 38. Chesebro B, Britt W, Evans L, Wehrly K, Nishio J, Cloyd M: **Characterization of monoclonal antibodies reactive with murine leukemia viruses: use in analysis of strains of friend MCF and Friend ecotropic murine leukemia virus.** *Virology* 1983, **127**:134-148.
 39. Robertson MN, Miyazawa M, Mori S, Caughey B, Evans LH, Hayes SF, Chesebro B: **Production of monoclonal antibodies reactive with a denatured form of the Friend murine leukemia virus gp70 envelope protein: use in a focal infectivity assay, immunohistochemical studies, electron microscopy and western blotting.** *J Virol Methods* 1991, **34**:255-271.
 40. Lynch WP, Sharpe AH: **Differential glycosylation of the Cas-Br-E env protein is associated with retrovirus-induced spongiform neurodegeneration.** *J Virol* 2000, **74**:1558-1565.

doi:10.1186/1742-4690-7-93

Cite this article as: Li and Lynch: Misfolding of CasBrE SU is reversed by interactions with 4070A Env: implications for gammaretroviral neuropathogenesis. *Retrovirology* 2010 **7**:93.

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