Lysis of aminobisphosphonate-sensitized MCF-7 breast tumor cells by V γ 9V δ 2 T cells

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Aminobisphosphonates are drugs administered for the treatment of bone resorption. They can indirectly activate peripheral $\gamma\delta\,T$ cells and render tumor cells susceptible to lysis by $V\gamma 9V\delta 2$ T cells. We have investigated the molecules involved in conjugate formation and killing of aminobisphosphonate-treated MCF-7 breast tumor cells by Vy9V82 T cells. Lysis of aminobisphosphonate (Pamidronate and Zoledronate)-treated MCF-7 tumor cells by $V\gamma 9V\delta 2$ T cells was assessed by chromium release assays and time-lapse video microscopy. MCF-7 breast cancer cells were chosen as aminobisphosphonates are employed to alleviate bone resorption in this malignancy. Cell cycle profile and expression of MICA, ICAM-I and FasL on aminobisphosphonate-sensitized MCF-7 breast tumor cells was confirmed by flow cytometry. Involvement of $\gamma\delta$ TCR and NKG2D in mediating cytotoxicity of aminobisphosphonate-treated MCF-7 breast tumor cells by $V\gamma 9V\delta 2$ T cells was assessed using blocking antibodies in chromium release assays. MCF-7 tumor cells pretreated with Pamidronate and Zoledronate were efficiently lysed by Vy9Vo2 T cells. Pamidronate and Zoledronate treatment of MCF-7 cells induced S phase arrest and did not alter expression of MICA, ICAM-I and FasL. Blocking $\gamma\delta$ TCR and NKG2D on V γ 9V δ 2 T cells inhibited lysis of Pamidronate and Zoledronate-treated MCF-7 cells. Inhibiting the perforin-granzyme pathway in V γ 9V δ 2 T cells using concanamycin A reduced their ability to lyse aminobisphosphonate-treated MCF-7 cells. V γ 9V δ 2 T cells form strong conjugates with aminobisphosphonate-treated MCF-7 breast tumor cells. $\gamma\delta$ TCR, NKG2D and perforin-granzyme pathway are involved in the lysis of MCF-7 breast tumor cells treated with aminobisphosphonates by $V\gamma 9V\delta 2$ T cells.

<u>Keywords:</u> cultured tumor cells, breast cancer, aminobisphosphonate, T lymphocytes, immunologic cytotoxicity

Introduction

Human $\gamma\delta$ T lymphocytes constitute approximately 1-5% of the total T cell population. Peripheral blood $\gamma\delta$ (V γ 9V δ 2) T cells recognize non-peptide phosphoantigens such as isopentenyl pyrophosphate (IPP) and its structural analogs aminobisphosphonates (1). Bisphosphonates have direct antitumor effects via inhibition of tumor cell adhesion, invasion, proliferation and induction of apoptosis (2).

Studies in myeloma patients revealed that Pamidronate infusion can induce selective expansion of V γ 9V δ 2 T cells (3). *In vivo* administration of Zoledronate in cancer patients with bone metastases was reported to induce maturation of V γ 9V δ 2 T cells to an IFN- γ producing effector phenotype which could potentiate their anti-tumor responses (4). Recent studies have elucidated that Zoledronic acid can induce potent anti-tumor activity via activation of V γ 9V δ 2 T cells in colorectal and hepatocellular carcinomas (5), hormone-refractory prostate cancer (6) and multiple myeloma (7).

Substantial evidence indicates that aminobisphosphonate treatment of tumor cells augments the anti-tumor activity of V γ 9V δ 2 T cells (8-10). Aminobisphosphonates inhibit the farnesyl pyrophosphate synthase (FPPS) enzyme in the mammalian mevalonate pathway (11), allowing accumulation of endogenous IPP in tumor cells and resulting in activation of V γ 9V δ 2 T cells (12).

However, the mechanism(s) involved in the lysis of aminobisphosphonate-treated tumor cells by V γ 9V δ 2 T cells are incompletely understood. This study attempts to investigate the molecules involved in the lysis of aminobisphosphonate-treated MCF-7 breast tumor cells by Vy9V82 T cells. In the present study, representative cell lines of breast, prostate and bone cancers were chosen as aminobisphosphonates are a standard modality of treatment for skeletal metastasis seen frequently in such malignancies. We specifically analyzed the MCF-7 breast tumor cell line for our further studies as a high incidence of bone metastasis is reported in breast cancers and aminobisphosphonates are administered to inhibit bone resorption (13). We report that Pamidronate and Zoledronate treatment of MCF-7 tumor cells sensitizes them to efficient lysis by V γ 9V δ 2 T cells which is mediated by the $\gamma\delta$ TCR and partially by the NKG2D receptor. In addition, the perforingranzyme pathway is also involved in the lysis of aminobisphosphonate-treated tumor cells by V γ 9V δ 2 T cells.

Results

Pamidronate and Zoledronate treatment of MCF-7 tumor cells sensitizes them to increased lysis mediated by $V\gamma 9V\delta 2$ T cells

After immunomagnetic separation of $\gamma\delta$ T cells, the purity of $\gamma\delta$ T cells was assessed by flow cytometry (Figure 1A). The percentage of purified V γ 9 and V δ 2 T cells was 96% and 95% respectively (Figure 1B). The ability of purified V γ 9V δ 2 T cells to lyse breast tumor cell line MCF-7 cells before and after treatment with the aminobisphosphonates Pamidronate and Zoledronate (100 µM) was assessed using ⁵¹Cr release assay. Further, we analyzed lysis mediated by V γ 9V δ 2 T cells against Pamidronate- and Zoledronate-sensitized PC-3 prostate carcinoma cells and SaOS-2 osteosarcoma cells, other malignancies which report frequent bone metastasis. Treatment of tumor cells with Pamidronate and Zoledronate significantly augmented the lysis of MCF-7 and PC-3 cells mediated by V γ 9V δ 2 T cells when tested at E:T ratios ranging from 30:1 to 7.5:1 (Figure 2, panels A and B). It was observed that SaOS-2 osteosarcoma cells were also efficiently primed by Pamidronate





Analysis of $\gamma\delta$ T cells in the negative and positive fractions isolated by magnetic-activated cell sorting (MACS) using single color flow cytometry. (A) Each overlay represents $\gamma\delta$ T cells stained with FITC-conjugated mAb against $\gamma\delta$ TCR (grey) and FITC-conjugated mAb against isotype IgG (black) in negative and positive fractions. The positive fraction shows the presence of 98% $\gamma\delta$ T cells. (B) Subset analysis of immunomagnetically purified $\gamma\delta$ T cells by dual color flow cytometry. Purified $\gamma\delta$ T cells were stained with fluorochrome (FITC or PE as indicated)-conjugated mAb against V γ 9, V δ 2 and CD3. The quadrants were established on the basis of staining of $\gamma\delta$ T cells with IgG isotype control FITC- and PE-conjugated antibodies. Figures indicate the percentage of double positive cells.

and Zoledronate for lysis by V γ 9V δ 2 T cells (Figure 2C). The efficiency of lysis was reduced when tumor cells were treated with lower concentrations of Pamidronate and Zoledronate (10 μ M and 50 μ M, data not shown).

We corroborated our findings of augmented lysis of aminobisphosphonate-treated MCF-7 cells by $V\gamma 9V\delta 2$ T cells using time-lapse video microscopy. When $V\gamma 9V\delta 2$ T cells were co-cultured with untreated MCF-7 cells, it was observed that $\gamma\delta$ T cells crawled on the surface of untreated tumor targets but were unable to form stable conjugates and lyse tumor cells (Figure 3, panels a-c, and Supplementary Video 1). Even after 4 hours of co-culture, there was no visible killing of the untreated MCF-7 cells by the $V\gamma 9V\delta 2$ T cells (Figure 3d).

However, when V γ 9V δ 2 T cells were co-cultured with MCF-7 cells pretreated with Zoledronate (100 μ M), it was observed that a number of $\gamma\delta$ T cells surrounded the MCF-7 tumor cell forming tight conjugates, subsequently killing the tumor cell within 10 seconds. V γ 9V δ 2 T cells constantly scanned the surface of Zoledronate-treated MCF-7 tumor cells and lysed them (Figure 4, panels a-h, and Supplementary Videos 2 and 3) unlike the crawling movement of $\gamma\delta$ T cells on untreated tumor cells. Similar observations were recorded with Pamidronate-treated tumor cells (data not shown).

Pamidronate and Zoledronate treatment induces a cell cycle arrest in MCF-7 tumor cells

In order to understand the direct effects of Pamidronate and Zoledronate on MCF-7 tumor cells, cell cycle analysis was performed. It was observed that treatment of MCF-7 cells with

Figure 2



Pamidronate and Zoledronate treatment sensitizes tumor cells to lysis by V γ 9V δ 2 T cells. MCF-7 breast carcinoma (A), PC-3 prostate carcinoma (B) and SaOS-2 osteosarcoma (C) cells were treated with Pamidronate (black rectangles, 100 μ M) and Zoledronate (black-filled circles, 100 μ M) or left untreated (open circles) for 16-18 hours. ⁵¹Cr-labeled tumor cells were co-cultured with V γ 9V δ 2 T cells for 4 hours at E:T ratios ranging from 30:1 to 1.8:1. Error bars indicate % cytotoxicity±SE. Statistical significance: @, P < 0.005; *, P < 0.0001; #, P < 0.002. Shown are the means of three independent experiments performed using $\gamma\delta$ T cells from 3 different donors against each target cell line.

Pamidronate and Zoledronate induced an S phase arrest wherein the percentage of cells in S phase increased from 50% in untreated (Figure 5A) to 53.9% in Pamidronate- (Figure 5B) and 81.1% in Zoledronate (Figure 5C)-treated MCF-7 cells. Zoledronate was more potent in inducing S phase arrest as compared to Pamidronate. In Pamidronate-treated MCF-7 cells, the percentage of cells in G0-G1 increased to 49.91% (Figure 5B) from 32% in untreated MCF-7 cells (Figure 5A). Interestingly, both Pamidronate and Zoledronate treatment of MCF-7 cells resulted in an absent G2-M phase. In addition, there was no apoptosis in Pamidronate- and Zoledronatetreated MCF-7 cells.

Involvement of $\gamma\delta$ TCR and NKG2D in cytotoxicity of aminobisphosphonate-treated tumor cells by V γ 9V δ 2 T cells

In order to identify the molecules involved in aminobisphosphonate-mediated tumor cell susceptibility to $V\gamma 9V\delta 2$ T cell lysis, MCF-7 cells pretreated with Pamidronate and Zoledronate were co-cultured with $V\gamma 9V\delta 2$ T cells treated with blocking antibodies against pan $\gamma\delta$ TCR, $V\gamma 9V\delta 2$ subset

Figure 3



Time-lapse imaging of untreated MCF-7 cells co-cultured with $V\gamma$ **9** $V\delta$ **2 T cells.** Untreated MCF-7 cells were co-cultured with $V\gamma$ **9** $V\delta$ **2 T** cells at a 1:2 ratio for 4 hours at 37°C. Snapshots of continuous time-lapse imaging on a LSM510 Meta Zeiss confocal microscope taken during the last 1 hour are shown. The images were taken at 30 seconds interval (a-d). $V\gamma$ **9** $V\delta$ **2 T** cells (black arrows) were unable to lyse the tumor cells (yellow arrows) even at the end of 4 hours (d). Representative images from one of four independent experiments are shown.

Figure 4



Time-lapse imaging of Zoledronate-treated MCF-7 cells co-cultured with $V\gamma 9V\delta 2$ T cells. MCF-7 cells were pretreated with Zoledronate (100 μ M) for 16-18 hours and co-cultured with $V\gamma 9V\delta 2$ T cells at a 1:2 ratio for 4 hours at 37°C. Snapshots from continuous time-lapse imaging taken during the last 1 hour are shown. The images were taken at 30 seconds interval (a-h). The yellow arrow indicates a single MCF-7 cell being targeted by $V\gamma 9V\delta 2$ T cells (black arrow). Changes in tumor cell morphology are observed (e-h) finally leading to lysis of the tumor cell by $V\gamma 9V\delta 2$ T cells (g-h). Representative images from one of four independent experiments are shown.

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Figure 6



Pamidronate and Zoledronate induce S-phase arrest in MCF-7 cells. Tumor cells were either left untreated (A) or pretreated with 100 μ M Pamidronate (B) or 100 μ M Zoledronate (C) for 16-18 hours. Cells were fixed in chilled 70% ethanol and stained with 5 μ g/ml of propidium iodide. Ten thousand events were acquired on a FACS Calibur and cell cycle analysis was performed. Data representative of two independent experiments are shown.

and NKG2D in a 4-hour ⁵¹Cr release assay. We observed that $V\gamma 9V\delta 2$ T cells showed expression of the C-type lectin receptor, NKG2D (Figure 6A).

Lysis of Pamidronate-treated MCF-7 cells was inhibited by blocking V γ 9V δ 2 T cells with anti- $\gamma\delta$ TCR mAb (29.7%±4% lysis, 41% inhibition), subset-specific anti-V γ 9 mAb (5%±0.5% lysis, 95% inhibition) and anti-NKG2D mAb (26%±7% lysis, 32% inhibition)(Figure 7A). Treatment of V γ 9V δ 2 T cells with a combination of either anti- $\gamma\delta$ mAb and anti-NKG2D mAb or anti-V γ 9 mAb and anti-NKG2D did not reduce the cytotoxicity below that observed with only anti- $\gamma\delta$ mAb or anti-V γ 9 mAb treatment (Figure 7A). V γ 9V δ 2 T cells incubated with isotype IgG control antibody did not inhibit lysis of Pamidronatetreated MCF-7 cells by V γ 9V δ 2 T cells (Figure 7A), indicating the specificity of the inhibition obtained with the mAbs used in the assay.

Similarly, cytotoxicity of V γ 9V δ 2 T cells against Zoledronatetreated MCF-7 cells was reduced when blocked with anti- $\gamma\delta$ mAb (24%±3% lysis, 57% inhibition), anti-V γ 9 mAb (5%±1% lysis, 93% inhibition) and anti-NKG2D mAb (35%±2% lysis, 42% inhibition)(Figure 7B). Combination of mAbs against TCR and NKG2D could not further lower the cytotoxicity of V γ 9V δ 2 T cells against Zoledronate-treated MCF-7 tumor cells (Figure 7B). Thus, it can be inferred that lysis of Pamidronate-

 $V\gamma 9V\delta 2$ T cells express NKG2D and intracellular granzyme and perforin. V $\gamma 9V\delta 2$ T cells were stained for NKG2D (A), intracellular granzyme B (B) and perforin (C) expression where isotype control (black) and positive staining (grey) is shown. Figures in the histograms indicate the percentage of positive cells. Data representative of three independent experiments are shown.

and Zoledronate-treated MCF-7 cells by V γ 9V δ 2 T cell is mediated by the $\gamma\delta$ TCR and is partially dependent on the NKG2D receptor expressed on V γ 9V δ 2 T cells.

Lysis of aminobisphosphonate-treated MCF-7 cells by V γ 9V δ 2 T cells is dependent on the perforin-granzyme pathway

A high expression of granzyme B (95%, Figure 6B) and perforin (98%, Figure 6C) was observed in V γ 9V δ 2 T cells. In order to investigate the involvement of the perforin-granzyme pathway in the lysis of aminobisphosphonate-treated MCF-7 cells by V γ 9V δ 2 T cells, T cells were treated with concanamycin A (specific inhibitor of perforin release and lysosomal acidification). V γ 9V δ 2 T cells efficiently lysed Pamidronate- (30%±5% lysis, Figure 8A) and Zoledronate (50%±3% lysis, Figure 8B)-treated MCF-7 cells. Pretreatment of V γ 9V δ 2 T cells with concanamycin A significantly inhibited the cytotoxicity of V γ 9V δ 2 T cells against Pamidronate- (17%±4% lysis, 40% inhibition, Figure 8B)-treated MCF-7 cells.

Discussion

Aminobisphosphonates are known to stimulate $V\gamma 9V\delta 2$ T cells (1) and concomitantly have direct anti-tumor effects (14). In the present study, we have attempted to understand how treatment with aminobisphosphonates alters the conjugate formation between $V\gamma 9V\delta 2$ T cells and MCF-7 breast tumor





Cytotoxicity of V γ 9V δ 2 T cells against Pamidronate- and Zoledronate-treated MCF-7 is mediated by $\gamma\delta$ TCR and NKG2D. Pamidronate- (A) and Zoledronate (B)treated MCF-7 cells were incubated with V γ 9V δ 2 T cells at an E:T ratio of 30:1 and the percentage of specific lysis determined by ⁵¹Cr release assay. V γ 9V δ 2 T cells were untreated (Unt) or treated with anti- $\gamma\delta$ TCR (10 µg/ml), anti-V γ 9 (10 µg/ml) and anti-NKG2D (20 µg/ml) mAb or a combination of anti- $\gamma\delta$ TCR (10 µg/ml) and anti-NKG2D (20 µg/ml) or anti-V γ 9 (10 µg/ml) and anti-NKG2D (20 µg/ml) mAbs. As control, V γ 9V δ 2 T cells treated with isotype specific antibodies were used as effectors and co-cultured with tumor targets. Error bars indicate % cytotoxicity±SE. Statistical significance: #, *P* < 0.002; @, *P* < 0.003; *, *P* < 0.0001. Data is representative of two independent experiments.

cells by time-lapse video microscopy and correlated these observations using cytotoxicity assays. We further demonstrated the role of molecules such as NKG2D, perforin and granzyme in the lysis of aminobisphosphonate-treated tumor cells by V γ 9V δ 2 T cells.

Our studies reveal that $V\gamma 9V\delta 2$ T cells efficiently lysed Pamidronate- and Zoledronate-treated MCF-7 breast cancer cells, PC-3 prostate cancer cells, and SaOS-2 osteosarcoma cells. Other investigators have demonstrated that aminobisphosphonate treatment of tumor cells augments lysis of myeloma (15), non-small cell lung carcinoma (16), breast and prostate adenocarcinoma (17) and colon cancer stem cells (18) by Vγ9Vδ2 T cells. A comparison of cytotoxicity of Vγ9Vδ2 T cells against Pamidronate- and Zoledronate-treated THP-1 (myelomonocytic cell line) and AW8507 (oral carcinoma cell line) cell lines at E:T 30:1 revealed differential susceptibility to lysis by V γ 9V δ 2 T cells (data not shown). These data demonstrate that aminobisphosphonates can differentially prime tumor cells of diverse origin to lysis by $V\gamma 9V\delta 2$ T cells.

We observed that Zoledronate treatment of tumor cells increased their susceptibility to lysis by V γ 9V δ 2 T cells greater than Pamidronate. This could be attributed to the fact that Zoledronate is a more potent aminobisphosphonate for inhibiting the FPPS enzyme in the mevalonate pathway with a half maximal inhibitory concentration (IC₅₀) value of 3 nM, as compared to 200 nM for Pamidronate (19).

Cell cycle analysis of Pamidronate- and Zoledronate-treated MCF-7 cells showed that Zoledronate induced a prominent S phase arrest while Pamidronate treatment increased the proportion of G0-G1 cells. We have included the cell cycle arrest data (Figure 5) to emphasize the fact that after exposure to Zoledronate for 16-18 hours tumor cells do not undergo apoptosis in our experimental system. There are reports which have accounted for the phenomenon of apoptosis or at least cell cycle arrest in tumor cells treated with aminobisphosphonates (20, 21) although the exposure time to bisphosphonates and the cell types used varied. Our purpose to include the cell cycle arrest data was to highlight that in our experimental protocol, overnight incubation of MCF-7 cells with

Figure 8



Cytotoxicity of $V\gamma 9V\delta 2$ T cells against Pamidronate- and Zoledronate-treated MCF-7 cells is partially dependent on the perforin-granzyme pathway. Pamidronate-(A) and Zoledronate (B)-treated MCF-7 cells were incubated with $V\gamma 9V\delta 2$ T cells at an E:T ratio of 30:1 and the percentage of specific lysis was determined by a 4-hour ⁵¹Cr release assay. $V\gamma 9V\delta 2$ T cells were either untreated or pretreated with concanamycin A (200 nM) for 1 hour at 37°C and then added to targets and further incubated for 4 hours at 37°C. Error bars indicate % cytotoxicity±SE. Statistical significance: *, P < 0.001; #, P < 0.05. Shown are means of two independent experiments.

aminobisphosphonates does not induce apoptosis of the MCF-7 cells. This is to support our data that the bisphosphonate-treated tumor cells do not die of apoptosis but are killed by $\gamma\delta$ T cells.

An earlier study has reported that the aminobisphosphonate Risedronate induced a G2 arrest in MCF-7 cells after 48 hours (22). On the other hand, Zoledronate induced a prominent S phase arrest in human osteosarcoma (23) and in cholangiocarcinoma cells (24), while a G2 arrest was reported in HCT-116 colon carcinoma cells (25). In addition, we observed that treatment of MCF-7 cells with Pamidronate and Zoledronate did not induce apoptosis as observed by Annexin V staining (data not shown). Although we do not have direct evidence that cell cycle arrest makes bisphosphonate-treated tumor cells susceptible to lysis by $\gamma\delta$ T cells, we speculate that the cell cycle arrest induced by aminobisphosphonates may predispose tumor cells to apoptosis. A recent study by Li et al. (26) showed that murine lung adenocarcinoma cells were arrested at the S/G2/M phase of the cell cycle upon Zoledronate treatment but no apoptotic cells were detected. It was suggested that Zolendronate may be a good candidate to pre-sensitize tumor cells to cytotoxic agents and radiation.

To visualize the events involved in V γ 9V δ 2 T cell-mediated lysis of Pamidronate- and Zoledronate-treated MCF-7 tumor cells, we analyzed $\gamma\delta$ T cell and MCF-7 tumor cell interaction by time-lapse confocal video microscopy, something which has not been reported to date. V γ 9V δ 2 T cells co-cultured with Zoledronate-treated MCF-7 cells clearly showed strong conjugate formation with numerous T cells moving on the surface of a single tumor cell. This culminated in membrane damage and subsequent lysis of the tumor cell within a 10 second time frame (data not shown), followed by the release of the attacking V γ 9V δ 2 T cell. $\gamma\delta$ T cells are known to participate in synaptic transfer and membrane stripping of target tumor cells (27). Using video microscopy, we observed that once the target cell was lysed, $\gamma\delta$ T cells moved out and continued scanning for other prospective tumor targets. This is reminiscent of an earlier study where CTLs were shown to maintain sustained synapses with targets and polarized the release of lytic granules (27).

On the other hand, $V\gamma 9V\delta 2$ T cells continuously scanned the surface of untreated tumor cells but were unable to form conjugates and lyse the tumor cells even after 4 hours. These results support the possibility that aminobisphosphonatesensitized tumor targets could allow efficient tethering of $\gamma\delta$ T cells, either via interaction of a receptor-ligand complex or by altering the kinetics of known receptor-ligand interaction. For the video time-lapse microscopy experiments, the tumor cells (MCF-7) were treated overnight at 37°C with Zoledronate at a concentration of 100 μ M. On the next day the tumor cells were washed and co-incubated with $\gamma\delta$ T cells. It is therefore unlikely that there is residual bisphosphonate left in the culture medium that could stimulate the $\gamma\delta$ T cells in 4 hours. We would like to mention that purified $\gamma\delta$ T cells do not respond to bisphosphonates unless the bisphosphonates are presented to them via an antigen-presenting cell. Several papers demonstrate the requirement of monocytes to present bisphosphonates to $\gamma\delta$ T cells (10, 28). It is therefore unlikely that residual aminobisphononates in the medium can stimulate $\gamma\delta$ T cells to form the conjugates with MCF-7 cells. Since we do not use monocytes in our assay system but only pretreat the tumor cells with bisphosphonates, we believe that the increased conjugate formation which occurs between bisphosphonate-coated tumor cells and $\gamma\delta$ T cells, as compared to untreated tumor cells, is due to the binding of NKG2D expressed on $\gamma\delta$ T cells with an unidentified ligand expressed on bisphosphonate-treated tumor cells. However, if the $\gamma\delta$ T cells are independently stimulated with aminobisphosphonates using monocytes for presentation, the activated $\gamma\delta$ T cells will form conjugates with tumor cells.

To elucidate the mechanism(s) involved in the increased lysis of aminobisphosphonate-treated tumor cells by Vy9V82 T cells, we analyzed the expression of surface molecules (MICA, ICAM-I and FasL) on MCF-7 cells and studied whether these altered after aminobisphosphonate treatment are (Supplementary Figure 1). Earlier reports have suggested an important role of the adhesion molecule ICAM-I expressed on tumor cells in modulating the cytotoxicity of V γ 9V δ 2 T cells by interacting with its ligand LFA-I on V γ 9V δ 2 T cells (29, 30). In addition, FasL expression on MCF-7 cells was shown to be upregulated by treatment with IFN- γ and TNF- α , thus facilitating CTL-mediated lysis of tumor cells (31). We observed low MICA, ICAM-I and FasL expression on MCF-7 cells which was not altered after treatment with Pamidronate and Zoledronate. This suggested that aminobisphosphonate treatment of MCF-7 cells did not quantitatively increase the expression of any of the molecules responsible for V γ 9V δ 2 T cell-mediated cytotoxicity against tumor cells known so far.

The optimal activation of V γ 9V δ 2 T cells is controlled by activating NK receptors (e.g. NKG2D) and inhibitory NK receptors (e.g. NKp44, NKG2A/C, etc.) which act as costimulatory molecules on V γ 9V δ 2 T cells (32). NKG2D, a Ctype lectin receptor expressed on NK cells and $\gamma\delta$ T cells and its interaction with its cognate receptors MICA/B, ULBP-1 etc. has been reported to play a prominent role in controlling cytotoxicity (33-36). In our studies we observed that V γ 9V δ 2 T cells showed high expression of NKG2D and therefore cytotoxicity assays were performed using blocking antibodies against $\gamma\delta$ TCR and NKG2D.

Blocking the V γ 9V δ 2 T cells with subset-specific V γ 9 mAb showed 95% inhibition in Vy9V82 T cell lysis of Pamidronateand Zoledronate-treated MCF-7 cells. On the other hand, blocking the NKG2D receptor on V γ 9V δ 2 T cells reduced their lytic efficiency by almost 50%. Thus, lysis of aminobisphosphonate-treated tumor targets by $V\gamma 9V\delta 2$ T cells appears to be mediated by the $\gamma\delta$ TCR and is partially dependent on the NKG2D receptor. However, unlike earlier studies where a synergy between the $\gamma\delta$ TCR and the NKG2D receptor in controlling the lytic activity of $V\gamma 9V\delta 2$ T cells was reported (5, 36), in our study we observed that blocking the V γ 9 receptor was sufficient to abolish the lytic activity of $V\gamma 9V\delta 2$ T cells against Pamidronate- and Zoledronate-treated MCF-7 cells. A recent study has reported the involvement of DNAX accessory molecule-1 (DNAM-1) besides NKG2D in Vy9V82 T cell lysis of tumor cells (37). This explains the partial inhibition in cytotoxicity assays we observed upon blocking NKG2D.

We observed that treatment of $V\gamma 9V\delta 2$ T cells with concanamycin A, a specific inhibitor of the perforin pathway abrogates the anti-tumor activity of $V\gamma 9V\delta 2$ T cells against Pamidronate- and Zoledronate-treated MCF-7 tumor cells by almost 50%. Further, our results are in agreement with earlier reports demonstrating high expression of granzyme B and perforin in $\gamma\delta$ T cells that are key molecules involved in granulemediated cytotoxicity (38, 39). Thus, it appears that the lysis of aminobisphosphonate-treated tumor cells by $V\gamma 9V\delta 2$ T cells also involves a functional perforin-granzyme pathway.

The physiological phosphoantigen recognized by $V\gamma 9V\delta 2$ T cells is isopentenyl pyrophosphate (IPP), an intermediate of the mevalonate pathway. Investigations have shown that aminobisphosphonates, the chemical analogs of IPP, inhibit the FPPS enzyme in the mevalonate pathway and thereby cause indirect activation of V γ 9V δ 2 T cells. To this end, recent studies have revealed that Zoledronate exposure of MCF-7 cells leads to the formation of a cytotoxic analog ApppI [triphosphoric acid 1-adenosin-5-yl ester 3-(3-methylbut-3-enyl) ester], that can induce apoptosis in these tumor cells (40). However, we speculate that in addition to the above mechanism, aminobisphosphonates could also modulate the avidity of interaction between Vy9V82 T cells and tumor cells, thus culminating in the formation of a stable $\gamma\delta$ T cell:tumor cell conjugate and eventual lysis of tumor cells. A proteomic analysis of aminobisphosphonate-treated tumor cells may help in identifying novel molecules that are involved in conjugate formation between $\gamma\delta$ T cells and tumor cells and also provide important leads in understanding mechanisms involved in the lysis of aminobisphosphonate-sensitized tumor cells by Vy9V82 T cells.

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Materials and methods

Reagents

Pamidronate (Pamifos) was purchased from Dabur, India and Zoledronate (Zoldonat) was purchased from Natco, India. Concanamycin A was obtained from Sigma-Aldrich, USA.

Expansion of $V\gamma 9V\delta 2$ T cells

Peripheral blood mononuclear cells (PBMCs) were isolated from healthy volunteers (n = 20) by Ficoll-Hypaque (Sigma-Aldrich, USA) density centrifugation. PBMCs were stimulated with anti-CD3 mAb (1 µg/ml) and rIL-2 (100 IU/ml) for 5 days in RPMI 1640 (Invitrogen Life-Technologies, USA) with 10% heat-inactivated human AB serum and further cultured for 12 days as described earlier (41, 42). $\gamma\delta$ T cells were isolated by positive separation using MACS (Miltenyi Biotec, Germany) and showed >95% expression of V γ 9V δ 2 TCR. $\gamma\delta$ T cells were left in culture medium without IL-2 for 24 hours before use in all functional assays. This study was approved by the Institutional Ethics Committee.

Tumor cell lines

MCF-7 (breast adenocarcinoma), PC-3 (prostatic carcinoma) and SaOS-2 (osteosarcoma) were obtained from the American Type Culture Collection, USA. MCF-7 cells were maintained in RPMI 1640 medium (Invitrogen Life-Technologies, USA), PC-3 cells were cultured in Ham's F-12K medium (Sigma-Aldrich, USA) and SaOS-2 cells were cultured in McCoy's 5A medium in 10% FBS (Invitrogen Life-Technologies, USA) with antibiotics.

Immunostaining

Purified $V\gamma 9V\delta 2$ T cells (0.5-1 x 10⁶) were washed in FACS buffer (1xPBS, 1% FCS, 0.01% sodium azide), fixed in 1% paraformaldehyde (Sigma-Aldrich, USA) and stained with the following FITC- or PE-conjugated antibodies: Vy9 TCR (clone B3, IgG1, BD Pharmingen USA), Vδ2 TCR (clone B6, IgG1, BD Pharmingen USA), Granzyme B (clone 351927, IgG2A, R&D Systems, USA), Perforin (clone deltaG9, IgG2b, eBioscience, USA) and NKG2D (clone 149810, IgG1, R&D Systems). Prior to intracellular staining of granzyme B and perforin, Vy9V82 T cells were permeabilized by treating with 0.1% saponin (Sigma-Aldrich, USA) in FACS buffer for 10 minutes at room temperature. Depending on the primary antibody used, staining was detected by incubating cells with either polyclonal goat antimouse FITC-conjugated antibody (Sigma-Aldrich, USA) or polyclonal goat anti-rabbit PE-conjugated antibody (Sigma-Aldrich, USA). At least 10,000 events were acquired on a FACS Calibur (Becton Dickinson, USA) and the data analyzed using the Cell Quest software (Becton Dickinson, USA).

Cell cycle

Tumor cells were treated with Pamidronate and Zoledronate at 100 μ M for 16-18 hours and fixed in 70% chilled ethanol. The cells were stained with 5 μ g/ml propidium iodide (Sigma-Aldrich, USA) for 15 minutes at 37°C, 5% CO₂ in the dark and acquired on a FACS Calibur (Becton Dickinson, USA). The cell cycle profile was analyzed using ModFit software (Verity Inc., USA).

Cytotoxicity assay

In all experiments, tumor cells were treated with Pamidronate and Zoledronate (100 μ M each) for 16-18 hours at 37°C. Tumor cells (1 x 10⁶) were labeled with 50 μ Ci of ⁵¹Cr (sodium chromate, Amersham, UK) for 1 hour at 37°C. Purified V γ 9V δ 2

T cells were titrated at effector:target ($\gamma\delta$:tumor) ratios ranging from 30:1 to 1.8:1 in triplicates and added to ⁵¹Cr-labeled tumor targets (5 x 10³). Tumor cells were co-cultured with V γ 9V δ 2 T cells for 4 hours at 37°C in 96-well plates (Nunc, Denmark). After 4 hours, supernatants were collected and chromium release was quantitated in a Gamma counter (Packard, USA) and expressed as counts per minute (cpm). The maximum release of chromium was determined by lysing tumor cells with Triton X-100 while the spontaneous release was determined by incubating only tumor cells in medium for 4 hours. The % cytotoxicity of V γ 9V δ 2 T cells was calculated as follows: % cytotoxicity = [(mean experimental CPM - mean spontaneous CPM)/(mean maximum CPM - mean spontaneous CPM)]x100.

For blocking experiments, $V\gamma 9V\delta 2$ T cells were incubated with anti-pan $\gamma\delta$ mAb (10 µg/ml, clone B1, BD PharMingen, USA), anti- $V\gamma 9$ mAb (10 µg/ml, clone B3, BD Pharmingen), anti-NKG2D mAb (20 µg/ml, R&D Systems, USA) or a combination of TCR and NKG2D antibodies for 1 hour at 37°C before adding to the targets at an E:T ratio of 30:1. To analyze the perforingranzyme pathway, effectors were pretreated with concanamycin A (200 nM, Fluka, USA) for 1 hour at 37°C before adding to targets at an E:T ratio of 30:1. The % inhibition of cytotoxicity was determined as follows: % inhibition of cytotoxicity = 1-(cytotoxicity in the presence of the mAb/ cytotoxicity in the absence of the mAb)x100.

Time-lapse video microscopy

MCF-7 cells were left untreated or treated with Pamidronate and Zoledronate for 16-18 hours. Tumor targets were washed and co-cultured with V γ 9V δ 2 T cells an E:T ratio of 1:2 for 4 hours at 37°C in 35-mm plates (Nunc, Denmark). The cocultures were then monitored for the last 1 hour on a confocal microscope (LSM510 Meta, Zeiss, USA) at 37°C. Selected areas were imaged every 30 seconds and videos were recorded.

Statistical analysis

All results were analyzed using SPSS software (Version 15.0, SPSS Inc., USA). Significance was assessed by unpaired Student's *t* test. The difference between groups was considered statistically significant when the *P* value was <0.05. All values in the figures or text are represented as either % cytotoxicity or mean fluorescence intensity (MFI) \pm standard error of mean (SEM).

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Supplemental data

Supplementary Figure 1. Pamidronate and Zoledronate do not alter the expression of MICA, ICAM-I and FasL on MCF-7 cells.

Download from http://www.cancerimmunity.org/v10p10/ 100809_suppl_fig1.pdf (355 KB PDF file).

Supplementary Video 1. Time-lapse video microscopy showing that $V\gamma 9V\delta 2$ T cells are unable to kill untreated MCF-7 cells.

Download from http://www.cancerimmunity.org/v10p10/ 100809_suppl_vid1.avi (91.4 MB AVI file).

Supplementary Video 2. Time-lapse video microscopy showing that Zoledronate-treated MCF-7 cells are efficiently lysed by $V\gamma9V\delta2~T$ cells.

Download from http://www.cancerimmunity.org/v10p10/ 100809_suppl_vid2.avi (24.3 MB AVI file).

Supplementary Video 3. Time-lapse video microscopy showing that Zoledronate-treated MCF-7 cells are lysed by $V\gamma 9V\delta 2$ T cells: Note the two tumor cells being lysed by $V\gamma 9V\delta 2$ T cells.

Download from http://www.cancerimmunity.org/v10p10/ 100809_suppl_vid3.avi (27.4 MB AVI file).

Entire supplemental data set.

Download from http://www.cancerimmunity.org/v10p10/ 100809_suppl_data.zip (60.1 MB WinZip file).