# Allelopathy of the moss *Rhynchostegium pallidifolium* and 3-hydroxy- $\beta$ -ionone

Hisashi Kato-Noguchi\* and Takahiro Seki

Department of Applied Biological Science; Faculty of Agriculture; Kagawa University; Miki, Kagawa Japan

**Key words:** allelopathy, growth inhibitor, 3-hydroxy-β-ionone, phytotoxicity, *Rhynchostegium pallidifolium* 

Submitted: 02/20/10

Accepted: 02/21/10

#### Previously published online: www.landesbioscience.com/journals/psb/ article/11642

\*Correspondence to: Hisashi Kato-Noguchi; Email: hisashi@ag.kagawa-u.ac.jp

The moss Rhynchostegium pallidifo*lium* (Mitt.) A. Jaeger, which often forms large pure colonies on soils and rocks, inhibited the hypocotyls and root growth of cress (Lepidium sativum L.) seedlings when R. pallidifolium and cress were incubated together on agar medium. The inhibition of cress was greater at the close position from the moss than at the far position from the moss. 3-Hydroxy- $\beta$ -ionone was found in the medium and concentration of 3-hydroxy- $\beta$ -ionone in the medium was greater at the close position than at the far position from R. pallidifolium, suggesting that R. pallidifolium may secrete 3-hydroxy- $\beta$ -ionone into the medium. Exogenously applied 3-hydroxy-β-ionone inhibited the growth of hypocotyls and roots of cress at concentrations greater than 1 and 3 µM, respectively. Considering the growth inhibitory activity and concentrations found in the medium, 3-hydroxy-β-ionone was estimated to be able to cause 46-64% of the observed growth inhibition of cress hypocotyls and roots by R. pallidifolium. Therefore, 3-hydroxy-\beta-ionone may play an important role in the allelopathic activity of R. pallidifolium and may help competition with neighboring plants resulting in the formation of pure colonies.

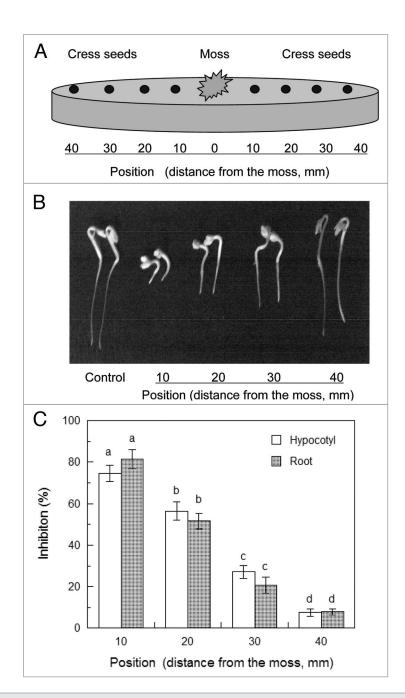
Bryophytes are almost free from attack by micro-organism and insects, and their herbarium specimens usually do not need special treatment against insects and micro-organism. In addition, many bryophyte species have their own particular odors and tastes.<sup>1</sup> These bryophyte characteristics are probably attributed to chemical constituents inherent in their structures. In fact, many biologically active substances, such as phenolics and terpenoides, have been isolated from bryophytes.<sup>2-5</sup>

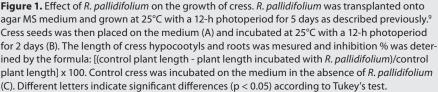
Several higher plants can not grow well in places where some bryophytes occurred. Some bryophytes dominate plant communities and form large pure colonies on soils and rocks on sunny places of lowland to upland areas including marshy places.<sup>1,6,7</sup> Therefore, allelopathic chemical interactions may play an important role in the domination of bryophytes in these plant communities. In contrast to higher plants, however, there only was a preliminary study on allelopathy of bryophytes. The moss Rhynchostegium pallidifolium (Mitt.) A. Jaeger, which belongs to Brachytheciaceae family of Bryopsida (moss) class, Bryophyta division, also forms large pure colonies and possesses strong allelopathic activity. An allelopathic substance of the moss was recently isolated and identified as 3-hydroxy-\beta-ionone.8

## Allelopathic Activity of *R. pallidifolium*

When cress seeds were grown on agar MS medium with *R. pallidifolium*, the growth of cress hypocotyls and roots was inhibited (Fig. 1). The inhibition was the greatest at the position of 10 mm from *R. pallidifolium* and the smallest at the position of 40 mm. Cress seedlings may grow with *R. pallidifolium* without competition for nutrients, because nutrients are considered to be unnecessary during the germination stage of seeds where most nutrients are withdrawn from seed reserves.<sup>9</sup> In addition, no significant pH

Addendum to: Kato-Noguchi H, Seki T, Shigemori H. Allelopathy and allelopathic substance in the moss *Rhynchostegium pallidifolium*. J Plant Physiol 2010; 167: 468-71. PMID: 20018404; DOI: 10.1016/j. jplph.2009.10.018.





changes occurred in the medium during the period of the incubation. These results suggest that the inhibitory effect of *R. pallidifolium* on cress may not be due to competitive interference for nutrients and pH changes in the medium, but rather due to an alleloapthic effect.

#### **3-Hydroxy**-β-lonone Concentration

3-Hydroxy- $\beta$ -ionone was found in the medium, but the concentration in the medium differed at the position from *R. pallidifolium* (Table 1). The highest

<b>Table 1.</b> Concentration of 3-hydroxy-β-
ionone in growth medium

Position (mm)	Concentration ( $\mu$ M)
10	11.3 ± 1.4
20	5.6 ± 1.1
30	2.1 ± 0.4
40	0.1 ± 0.1

*R. pallidifolium* was grown on agar MS growth medium for 5 days as shown in **Figure 1A** and 3-hydroxy- $\beta$ -ionone was determined as described previously.<sup>9</sup>

concentration of 3-hydroxy- $\beta$ -ionone was at the position of 10 mm from *R. pallidifolium* and the lowest concentration was at the position of 40 mm from the moss. 3-Hydroxy- $\beta$ -ionone was also found in soils under *R. pallidifolium*.<sup>9</sup> These results suggest that 3-hydroxy- $\beta$ -ionone was probably secreted by *R. pallidifolium* into the medium.

#### Inhibitory Activity of 3-Hydroxy-β-Ionone

3-Hydroxy- $\beta$ -ionone inhibited the growth of hypocotyls and roots of cress at concentrations greater than 1 and 3 µM, respectively (Fig. 2). When inhibition of cress hypocotyls and roots was plotted against the logarithm of the concentrations of 3-hydroxy-\beta-ionone as described by Streibig,10 good logistic functions were obtained. The equations of the functions of 3-hydroxy- $\beta$ -ionone were Y = [(0.552 - $75.290)/\{1 + (X/72.372)^{0.958}\}] + 75.290;$  $(r^2 = 0.998)$  and  $Y = [(0.313 - 84.479)/{1}$ +  $(X/68.973)^{1.298}$ ] + 84.479; (r<sup>2</sup> = 0.999) for hypocotyls and roots, respectively. Y in the equations indicates the growth inhibition (%) and X indicates the concentration (μM) of 3-hydroxy-β-ionone as shown in Figure 2.

### Contribution of 3-Hydroxy-β-lonone to Allelopathy of *R. pallidifolium*

To quantify the contribution of 3-hydroxy- $\beta$ -ionone to overall allelopathy of *R. pallidifolium*, the potential growth inhibition on cress caused by secreted 3-hydroxy- $\beta$ -ionone in the medium was calculated using the derived equations of logistic functions and substituting X values with **Table 2.** Potential growth inhibition on cress by 3-hydroxy- $\beta$ -ionone found in growth medium

	Potential growth inhibition (%)		
Position (mm)	Hypocotyl	Root	
10	43.7	43.5	
20	32.3	23.9	
30	16.6	9.7	
40	4.2	4.1	

The potential growth inhibition on cress caused by 3-hydroxy- $\beta$ -ionone was calculated using the equations of logistic functions with substituting X values by 3-hydroxy- $\beta$ -ionone concentrations found in the growth medium (**Table 1**) as described in the text.

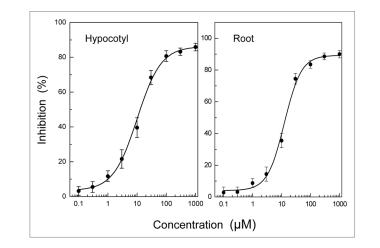
**Table 3.** Contribution of 3-hydroxy- $\beta$ -ionone to the growth inhibition by *R. pallidifolium* 

	Contribution (%)		
Position (mm)	Hypocotyl	Root	
10	63.5	53.3	
20	57.3	46.3	
30	61.3	46.8	
40	57.3	52.3	

Contribution of 3-hydroxy- $\beta$ -ionone to the growth inhibition on cress by *R. pallidifolium* was calculated by the formula: [potential growth inhibition (**Table 2**)]/[measured growth inhibition (**Fig. 1C**)] x 100.

the concentrations found in the growth medium (Table 1). These values in Table 2 indicate that 3-hydroxy- $\beta$ -ionone in the medium has the potential to inhibit 43.7 and 43.5% for cress hypocotyl and root growth, respectively, at the potion 10 mm from the moss and 4.2 and 4.1% for cress hypocotyl and root growth, respectively, at the potion 40 mm from the moss.

The contribution of 3-hydroxy- $\beta$ ionone to the growth inhibition on cress by *R. pallidifolium* was calculated by the formula: [potential growth inhibition values (**Table 2**)]/[measured growth inhibition (Fig. 1C)] x 100. The values in **Table 3** show that 3-hydroxy- $\beta$ -ionone accounts



**Figure 2.** Effects of 3-hydroxy- $\beta$ -ionone on hypocotyl and root growth of cress seedlings. Cress seeds were incubated in aqueous solution of 3-hydroxy- $\beta$ -ionone for 2 days and the length of hypocotyls and root of cress seedlings were measured as described previously.<sup>9</sup> Inhibition % was then determined as described in **Figure 1**.

for 57.3–63.5% and 46.3–53.3% of the observed growth inhibition of cress hypocotyls and roots, respectively, by *R. pallidifolium* (Fig. 1C). Therefore, the contribution of 3-hydroxy- $\beta$ -ionone to the growth inhibition may explain about 46–64% of the allelopathic activity of *R. pallidifolium* against cress.

It is suggested that, although mechanisms of the exudation are not well understood, plants are able to secrete a wide variety of compounds from root cells by plasmalemma-derived exudation, endoplasmic-derived exudation, and proton-pumping mechanisms.12,13 Through the exudation of compounds, plants are able to regulate the soil microbial community in their immediate vicinity and inhibit the growth of competing plant species.<sup>11-13</sup> Considering the inhibitory activity of 3-hydroxy-\beta-ionone and the secretion level of 3-hydroxy-β-ionone, 3-hydroxy- $\beta$ -ionone may play a very important role in *R. pallidifolium* defense mechanism in the rhizosphere as an allelopathic substance and may help competition with neighboring plants resulting in the formation of pure colonies.

#### References

- Ando H, Matsuo A. Applied bryology. In: Cramer J, ed. Advance Bryology, Vol 2. Berlin: International Association of Bryologists 1984; 133-229.
- Asakawa Y. Biologically active substances from bryophytes. In: Chopra RN, Bhatla SC, ed. Bryophyte development: Physiology and Biochemistry. Boca Raton, Fla: CRC Press 1990; 259-87.
- Asakawa Y. Chemosystematics of the Hepaticae. Phytochemistry 2004; 65:623-69.
- Basile A, Sorbo S, López-Sáez JA, Cobianchi RC. Effects of seven pure flavonoids from mosses on germination and growth of *Tortula muralis* HEDW. (Bryophyta) and *Raphanus sativus* l. (Magnoliophyta). Phytochemistry 2003; 62:1145-51.
- 5. Li X, Wurtele ES, LaMotte CE. Abscisic acid is present in liverworts. Phytochemistry 1994; 37:625-7.
- Blime JM. The role of bryophytes in temperate forest ecosystems. Hikobia 2001; 13:267-89.
- Tsubota H, Kuroda A, Masuzaki H, Nakahara M, Deguchi H. Preliminary study on allelopathic activity of bryophytes under laboratory conditions using the sandwich method. J Hattori Bot Lab 2006; 100:517-25.
- Kato-Noguchi H, Seki T, Shigemori H. Allelopathy and allelopathic substance in the moss *Rhynchostegium pallidifolium*. J Plant Physiol 2010; 167: 468-71.
- Fuerst EP, Putnam AR. Separating the competitive and allelopathic components of interference: Theoretical principles. J Chem Ecol 1983; 8:937-44.
- Streibig JC. Herbicide bioassay. Weed Res 1988; 28:479-84.
- 11. McCully E. Roots in soil: unearthing the complexities of roots and their rhizospheres. Annu Rev Plant Physiol Plant Mol Biol 1999; 50:695-718.
- 12. Hawes MC, Gunawardena U, Miyasaka S, Zhao X. The role of root border cells in plant defense. Trends Plant Sci 2000; 5:128-33.
- Bais HP, Park S-W, Weir TL, Callaway RM, Vivanco JM. How plants communicate using the underground information superhighway. Trends Plant Sci 2004; 9:26-32.