# Pathogenesis and treatment of immune-mediated neuropathies

#### Helmar C. Lehmann, Gerd Meyer zu Horste, Bernd C. Kieseier and Hans-Peter Hartung

**Abstract:** Immune-mediated neuropathies represent a heterogeneous spectrum of peripheral nerve disorders that can be classified according to time course, predominant involvement of motor/sensory fibers, distribution of deficits and paraclinical parameters such as electrophysiology and serum antibodies. In the last few years, significant advances have been achieved in elucidating underlying pathomechanisms, which made it possible to identify potential therapeutic targets. In this review, we discuss the latest development in pathogenesis and treatment of immune-mediated neuropathies.

*Keywords*: immune neuropathies, Guíllan-Barré syndrome, cidp, paraproteinemic neuropathies, treatments

#### Introduction

Immune-mediated neuropathies represent a heterogeneous group of peripheral nerve disorders, which can be classified according to clinical symptoms and signs, time course and paraclinical parameters. Over the last few years, significant advances in the development of preclinical models for these diseases have allowed us to gain a deeper insight into the molecular and cellular mechanisms resulting in immune-mediated injury of the peripheral nervous system. This progress went along with the discovery of new promising therapeutic targets, which may have the potential to be further evaluated in future clinical trials. In this review we summarize current knowledge of pathogenesis, clinical course and treatment of the most frequent forms of immune-mediated neuropathies. These include the Guillain-Barré syndrome (GBS), as a prototype of an acute, immune-mediated peripheral neuropathy, chronic inflammatory demyelinating polyradiculoneuropathy (CIDP), multifocal motor neuropathy (MMN) and the paraproteinaemic polyneuropathies.

#### Guillain-Barré syndrome

The Guillain–Barré syndrome (GBS) is the prototype of an immune-mediated, monophasic inflammatory polyneuropathy with acute disease onset [Kieseier *et al.* 2006; Hughes and Cornblath, 2005; Kieseier and Hartung, 2003].

Several subtypes of GBS exist, which can be differentiated on the basis of their clinical course, electrophysiological criteria and underlying pathology. The reported incidence rates (1-2)per 100 000) of GBS are comparable in different geographical regions of the world [Lehmann et al. 2007a; Govoni and Granieri, 2001; Cheng et al. 2000; van Koningsveld et al. 2000; Rees et al. 1998; Hughes and Rees, 1997; Sedano et al. 1994]; however, the distribution of GBS subtypes differs markedly between different regions. By far the most frequent form (90-95%) in Europe and North America is the acute inflammatory demyelinating polyneuropathy (AIDP), whereas the axonal GBS forms (acute motor axonal neuropathy [AMAN] and acute motor sensory axonal neuropathy [AMSAN]) are rare in these regions but can reach 30–40% in China, Japan and South America [Yuki, 2005; Ho et al. 1995a; Ramos-Alvarez et al. 1969]. Apart from these subtypes, there are atypical variants such as the Miller-Fisher syndrome and the cervico-brachialoropharyngeal weakness [Halstead et al. 2005; Overell and Willison, 2005; Willison, 2005; O'Leary et al. 1996].

#### Pathogenesis

Several lines of evidence suggest that in GBS, triggers such as an infection of the respiratory or gastrointestinal tract generate an aberrant immune response, which subsequently leads to a breakdown of the blood-nerve barrier and to a

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Gerd Meyer zu Horste Bernd C. Kieseier Department of Neurology, Heinrich-Heine-University, Düsseldorf, Germany destruction of myelin sheaths and/or axons [Mever zu Horste et al. 2007; Kieseier et al. 2004]. Apart from bacterial and viral infections, several other triggers have been reported for example vaccinations (including influenza) [Haber et al. 2004; Lasky et al. 1998] and events such as surgery, which may lead to an activation of the immune system [van Doorn et al. 2008]. Pathologically, AIDP is characterized by multifocal segmental demyelination and the presence of inflammatory infiltrates. Demyelinated axons can be found within the spinal roots and the peripheral nerves and often, signs of secondary axonal degeneration accompany the severe demyelinative process. Inflammatory infiltrates contain T cells and macrophages [Prineas, 1981; Asbury et al. 1969]. CD3+T cells are the dominating lymphocyte population, whereas B cells are less frequently detected. Apart from cellular infiltrates, deposition of activated complement and the membrane attack complex on Schwann cells have been reported [Hafer-Macko et al. 1996], which supports the notion that both the cellular and the humoral immune system contribute to the pathogenesis in GBS. A recently published study sugcomplement deposition gests that on demyelinated fibers is mainly found during the acute stage of GBS, whereas CD8+ cells and granzyme B-positive lymphocytes predominate the subacute stage of the disease [Wanschitz et al. 2003]. In AIDP, the autoantigens targeted by T cells and/or autoantibodies are largely unknown. A GBS-like disease can be induced by immunization of rodents with different myelin preparations, the so-called experimental allergic neuritis (EAN) [Lonigro and Devaux, 2009; Bechtold et al. 2005; Kieseier et al. 2004; Felts et al. 2002; Hadden et al. 2002; Gold et al. 2000; Kieseier et al. 2000]. The observation that EAN can be induced with the myelin proteins P0, P2 and PMP22, and by passive transfer of P0 or P2 specific CD4+ T cells points to a role of these proteins as potential autoantigens in GBS. However, only a small proportion of patients with AIDP elicit immune reactivity against those myelin proteins [Makowska et al. 2008]. More recently it has been suggested that neurofascin and gliomedin, two cell adhesion molecules, which are involved in clustering of voltage-gated sodium channels at the nodes of Ranvier may be targeted in EAN [Lonigro and Devaux, 2009]. The occurrence of IgG autoantibodies directed against these nodal proteins was associated with a more severe disease course and demyelinating neurophysiology in one EAN model.

In contrast to the demyelinating forms of GBS, the presumed targets of a pathologic autoantibody response in the axonal GBS variants and in the Miller-Fisher syndrome are much better defined [Willison and Yuki, 2002]. Clinical studies over the last two decades have shown that antibodies against several gangliosides can be detected in serum of patients with AMAN [Willison, 2002; Khalili-Shirazi et al. 1999; Ho et al. 1995b; Illa et al. 1995; Yuki et al. 1990]. These include antibodies against the major gangliosides GM1 and GD1a, and against GalNAc-GD1a and GD1b. The best correlation between antiganglioside antibodies and a clinical syndrome, however, can be found in patients with Miller-Fisher syndrome. In up to 90% of cases, antibodies against GQ1b can be detected [Overell and Willison, 2005; Willison, 2005, 2002]. Antiganglioside antibodies have shown to exert a variety of different pathogenic effects in various in vivo and in vitro models [Buchwald et al. 2007; Lehmann et al. 2007c; Susuki et al. 2007; Goodfellow et al. 2005; Halstead et al. 2004; Zhang et al. 2004; Buchwald et al. 2002]. Based on these studies it has been suggested that the nodes of Ranvier and the motor nerve terminals are the preferential targets of antiganglioside antibodies, due to high concentrations of complex gangliosides located there and the easy accessibility of 'axonal' targets within the myelinated fibers. It has been demonstrated that antiganglioside antibodies which bind to gangliosides at the nodes or at the level of the neuromuscular junction are able to induce conduction block and lead to injury to perisynaptic Schwann cells [Goodfellow et al. 2005; Halstead et al. 2004; O'Hanlon et al. 2003]. These effects are dependent on the activation of complement. Further complement-independent effects include an inhibition of the evoked quantal release at motor nerve terminals and an inhibition of axonal regeneration by passive transfer of anti-GD1a antibodies [Lehmann et al. 2007c; Buchwald et al. 1998].

It is believed that in these GBS variants, the mechanism of so-called molecular mimicry essentially contributes to the induction of an autoimmune response against peripheral nerve tissue [Yuki and Odaka, 2005]. The observation that microbes express carbohydrate epitopes that resemble glycolipid antigens on the surface of peripheral nerves is the basis for the hypothesis of postinfectious molecular mimicry in GBS. This is best documented for the gram-negative

rod *Campylobacter jejuni*, which can be identified as cause of a preceding infection in up to 60% of all GBS cases. This enteropathogen contains several ganglioside-like moieties on the outer leaflet of its lipooligosaccarides [Yuki, 1997; Yuki *et al.* 1994, 1993].

# Clinical course

Early symptoms of GBS are pain, limb paresthesia and weakness. These symptoms progress typically within several days from distal to proximal and reach a nadir within 2-4 weeks. Cranial nerve involvement is common; the facial nerves in particular are often affected (bilateral facial paresis). In up to two-thirds of the patients autonomic dysfunction occurs which may present as bradycardia, arrhythmia, arterial hypertension, blood pressure fluctuation, sweating abnormalities and bowel/bladder disturbances [Flachenecker et al. 1997a, 1997b]. Pain is another frequent symptom, which can occur at early stages in sensory and motor forms of GBS and may present as backache or interscapular pain [Ruts et al. 2008, 2007]. Functional recovery in GBS is usually much slower than the initial clinical deterioration and may be complicated by relapses. Although GBS has an overall good prognosis, around 5-10% of patients die due to complications and around 20-30% of the patients remain disabled after 1 year [Hughes and Cornblath, 2005]. Currently no treatment is known to enhance the recovery of GBS patients.

# Treatment

Generally the two mainstays of treatment in GBS are (1) supportive care and (2) immunomodulatory treatment. Supportive care is crucial to prevent complications that may occur due to the immobilization, such as thrombembolism and infections. Pneumonia is common, especially in patients that have significant bulbar weakness. Prophylactic anticoagulation, together with careful monitoring and early antibiotic treatment may help to reduce the risk for such complications. Whenever possible, GBS patients should be admitted to an ICU which is experienced in the management of GBS. Approximately one-third of patients need ventilatory assistance. Monitoring of vital capacity and respiration frequency can identify impending respiratory failure. When autonomic fibers are involved and brady-arrhythmia is observed the temporary use of pacemakers might be required. Further measures of supportive care include physiotherapy and early rehabilitation.

Currently there are two 'causal' treatments available for GBS (Table 1). Plasma exchange and intravenous immunoglobulins (IVIg) have shown beneficial effects in large randomized GBS trials. Although both treatments are considered equal in terms of efficacy, IVIg is currently favored in most centers due to its easier availability, handling and more favorable side-effect profile.

Several randomized trials have demonstrated the efficacy of plasma exchange in comparison with supportive care. In the first North American trial 245 patients were randomized to receive plasma exchange or conventional supportive therapy. The PE group showed an improvement on the disability scale at 4 weeks, time to improve one clinical grade and outcome at 6 months [Guillain-Barré Syndrome Study Group, 1985]. In the study by the French Cooperative Group on Plasma Exchange in Guillain-Barré Syndrome [1987] 220 patients were randomized to plasma exchange or supportive care alone. The PE group showed a statistically significant faster time to recover the ability to ambulate with assistance, which served as primary endpoint. Secondary parameters also favored PE treatment (reduction of patients needing assisted ventilation, time to walk with and without assistance). Further benefit of plasma exchange compared with supportive care alone was also documented after 1 year as the percentage of patients with a recovery of full muscle strength in a follow-up study [French Cooperative Group on Plasma Exchange in Guillain-Barré Syndrome, 1992]. Based on the results from another large GBS plasma exchange trial from France, the number of plasma exchanges can be adjusted to the disease severity of the patient. In this trial 556 GBS patients were included and randomized according to their disease severity [French Cooperative Group on Plasma Exchange in Guillain-Barré Syndrome, 1997]. Mildly affected patients received either no or two plasma exchange sessions, whereas patients who were unable to walk but did not require assisted ventilation received either two or four sessions. GBS patients who required assisted ventilation received either four or six plasma exchange sessions. Mildly affected patients had more benefit from treatment with two plasma exchange sessions than supportive care alone. Four plasma exchange sessions were more beneficial than two in the group of the moderately affected patients. However, more than four treatments did not vield any additional benefit in severely affected patients who required

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Ireatment	Irial	Year	I rial design	Patients (n)	Uutcome
<b>Effective treatment</b> Plasma exchange	nt The Guillain-Barré syndrome Study	1985	PE versus supportive care, single-blind	245	Improvement at 4 weeks, time to improve one clinical grade, time to independent walking,
	eroup French Cooperative Group on Plasma Exchange in Guillain-Barré	1987	PE [4×] with albumin $(n=57)$ versus PE [4×] with fresh frozen plasma $(n=52)$ versus no PE $(n=111)$ , non-blind	220	Shorter time to recover walking with assistance (30 d versus 44 d, $p < 0.01$ ) in PE group. Reduced number of patients requiring assisted ventilation. shorter time to onset of motor
	syndrome The French Cooperative Group on Plasma Exchange in Guillain- Barré Syndrome	1997	3 groups: 'mild' affected: 0 <i>versus</i> 2 PE treatments; 'moderate' affected: 2 <i>versus</i> 4 PEs; 'severe' affected: 4 <i>versus</i> 6 PEs. non-blind	556	2 PEs more effective than 0 for time to onset of motor recovery (4 d versus 8d, $p = 0.0002$ ) in mild group. 4 PEs superior as 2 PEs for time to walk with assistance (20 versus 24 d; $p = 0.04$ ) in moderate aroun. No difference between $\Delta$
IVIg	The Dutch Guillain-Barré Study Group	1992	IVIG (0.4 g/kg × d, 5×), <i>versus</i> PE (5×) non-blind, bias-controlled	150	Improvement one or more group. The severe group. Improvement one or more points on functional score $34\%$ in PE-group <i>versus</i> $53\%$ in IVIG-group ( $p = 0.024$ ). Time to improvement by one grade 41 d <i>versus</i> 27 d ( $p = 0.05$ ). Both treat-
	Plasma Exchange/ Sandog lobulin Guillain-Barré Syndrome Trial Group	1997	PE $5 \times [n = 121]$ versus IVIG [0.4 g/kg, 5 d] $[n = 130]$ versus PE $[5 \times ] + IVIG$ [0.4 g/kg, 5 d], single-blind	383	ments are of equal efficacy. No significant difference in major outcome mea- sure limprovement on disability scale after 4 weeks and in secondary outcome measures [time to recovery of unaided walking, time to discontinuation of ventilation]
Uncertain benefit Combination of PE and IVIg	Uncertain benefit or resumably ineffective treatment (compared to PE or IVIg) Combination of Plasma Exchange/ 1997 PE $5 \times (n = 121)$ versus PE and IVIg Sandog Lobulin $5 \text{ d}$ ( $n = 130$ ) versus I Guillain-Barré ( $0.4 \text{ g/kg}, 5 \text{ d}$ ), single-Syndrome Trial Group	eatment 1997	<b>[compared to PE or IVIg]</b> PE $5 \times (n = 121)$ versus IVIG $[0.4 \text{ g/kg}, 5 \text{ d}]$ $(n = 130)$ versus PE $[5 \times] + \text{IVIG}$ [0.4  g/kg, 5  d], single-blind	383	No significant difference in major outcome mea- sure limprovement on disability scale after 4 weeks and in secondary outcome measures litime to recovery of unaided walking, time to
Combination of IVIg and Methylpredn- isolone	Konigsveld et al., Dutch GBS trial group	2004	IVIg $[0.4 \text{ g/kg}, 5 \text{ d}]$ + methylprednisolone [n = 112] versus IVIg $[0.4  g/kg, 5  d]$ + placebo $[n = 113]$ , double-blind, randomized-controlled	225	discontinuation of ventilation) No significant difference between number of patients improved by 1 disability grade after 4 weeks. No significant differences in secondary outcome measures (ability to walk unaided
Intravenously Combination of IVIg, methyl- prednisolone and mycophe- nolate mofetil	Garssen <i>et al.</i>	2007	IVIg $[0.4 \text{ g/kg/5} d + 500 \text{ mg methylpred-nisolone i.v. } 5 d + mycophenolate mofetil (2000 mg/d 6 weeks) (n = 26) compared to historical control (Koningsveld et al.) IVIg [0.4 \text{ g/kg},$	26	arter & weeks or time to walk independently No statistical differences in primary endpoint (improvement by at least one grade on the GBS disability score after 4 weeks)
Combination of IVIg, and inter- feron beta 1a	Pritchard <i>et al.</i>	2003	5 d) + methylprednisolone $(n = 112)$ , open-labeled pilot study NIg + interferon beta 1a (22 µg 1 week then 44 µg up to 24 weeks) $(n = 13)$ versus IVIg + placebo $(n = 6)$	19	No statistical differences between the two groups
PE, plasma exchang	PE, plasma exchange; IVIg, intravenous immunoglobulins;		NDS, neurological disability score; i.v., intravenously.		

assisted ventilation. In this regard, it is important to consider autoantibody kinetics during plasma exchange. It has been demonstrated that at least two plasma exchange sessions are required to significantly reduce the amount of circulating immunoglobulins in GBS [Yuki *et al.* 1998].

Besides plasma exchange, IVIg is currently the only proven immunomodulatory treatment in GBS [Hughes et al. 2006]. To date, both treatments are considered to be comparable in terms of outcome and efficacy, whereas side effects tend to be lower in those patients who receive IVIg [Lehmann et al. 2006]. Evidence for the efficacy of IVIg in adult GBS patients stems from several multicenter trials that compared IVIg with plasma exchange [Diener et al. 2001; Plasma Exchange/Sandoglobulin Guillain-Barré Syndrome Trial Group, 1997; Bril et al. 1996; van der Meche and Schmitz, 1992]. The combination of IVIg treatment followed by plasma exchange does not yield any additional benefit for GBS patients, therefore this strategy is not generally recommended [Plasma Exchange/Sandoglobulin Guillain-Barré Syndrome Trial Group, 1997]. In childhood GBS, at least two trials have shown some beneficial effects (faster recovery) of IVIg compared with no treatment [Hughes et al. 2007; Korinthenberg et al. 2005; Gürses et al. 1995]. However, the primary study endpoint (disease severity) in the trial by Korinthenberg et al. [2005] did not show a difference between the treatment groups.

The standard dose of IVIg (0.4 g/kg/day over 5 days) used is empirically based on experience from the treatment of other autoimmune diseases [Imbach *et al.* 1981]. Alternative treatment regimens of IVIg delivering the drug over a shorter period of days have been tested in smaller trials, without showing a clear benefit for the shorter treatment interval [Korinthenberg *et al.* 2005].

Corticosteroids given either orally or intravenously are ineffective in GBS. This is based on the results of several smaller trials that found no differences when corticosteroids were compared with placebo or supportive care alone [Singh and Gupta, 1996; Goodall *et al.* 1974]. One trial compared intravenous methylprednisolone with placebo, but also failed to demonstrate any beneficial effects of corticosteroid treatment [Guillain–Barré Syndrome Steroid Trial Group, 1993]. In 2004 van Koningsveld and colleagues published a double-blind, randomized controlled trial in which 225 GBS patients received either methylprednisolone (500 mg/day for 5 days) or placebo in addition to standard IVIg treatment [van Koningsveld *et al.* 2004]. However no significant differences were observed in the primary outcome (improvement in GBS disability score of at least one grade) after 4 weeks.

More recently, Garssen et al. [2007] reported the results of an open-label pilot study of mycophenolate mofetil in addition to a combination of IVIg and methylprednisolone in GBS. Mycophenolate mofetil is an immunosuppressive agent acting on B- and T-lymphocytes, which has been shown to be beneficial as add-on therapy in other diseases of autoimmune etiology [Moore and Derry, 2006]. Twenty-six GBS patients received IVIg (0.4 g/kg/ day) and 500 mg methylprednisolone (MP) intravenously on 5 consecutive days. In addition patients received mycophenolate mofetil at a dose of 2000 mg/day for 6 weeks. This group was compared to a historical control group treated with IVIg and MP in the Dutch IVIg-MP trial [van Koningsveld et al. 2004]. There were no statistically significant differences regarding the primary endpoint, improvement by at least one grade on the GBS disability score after 4 weeks. The authors concluded that mycophenolate mofetil is probably of limited value in the treatment of GBS. Similarly, interferons are ineffective in GBS based on a randomized controlled study that failed to demonstrate a beneficial effect of interferon beta as add-on therapy to IVIg in GBS [Pritchard et al. 2003].

# Future directions in the treatment of GBS

Despite treatment with plasma exchange or IVIg during the acute phase of GBS, a proportion of patients show a poor recovery with significant disability, which emphasizes the need for more effective therapies. Over the past years, new therapeutic approaches have been explored mainly in preclinical studies. These include the selective depletion of pathogenic autoantibodies, and the use of agents that prevents complementdependent nerve injury [Halstead et al. 2008; Bullens et al. 2005; Andersen et al. 2004]. Antiganglioside antibodies can be selectively removed by synthetic molecules, which bear the terminal trisaccharide structure of gangliosides [Willison et al. 2004]. More recently, it has been shown that eculizumab, a humanized monoclonal antibody preventing formation of the terminal membrane attack complex C5b-9, can protect neuromuscular junctions from antiganglioside

antibody mediated, complement dependent injury in an *in vivo* model of Miller–Fisher syndrome [Halstead *et al.* 2008; Lehmann and Hartung, 2008]. These findings may provide a rationale for future clinical trials with eculizumab in MFS and perhaps GBS.

# Chronic inflammatory demyelinating polyradiculoneuropathy (CIDP)

Chronic inflammatory demyelinating polyradiculoneuropathy (CIDP) is an acquired, immunemediated polyradiculoneuropathy, clinically characterized by proximal and distal motor/ sensory deficits and a relapsing or progressive disease course [Köller *et al.* 2005]. Reported prevalence rates of CIDP are similar in different geographical regions and range 1–3/100 000. The disease may affect children and elderly individuals [Iijima *et al.* 2008; Chiò *et al.* 2007; Lunn *et al.* 1999; McLeod *et al.* 1999].

#### Pathogenesis

Similar to GBS, it is believed that CIDP is caused by an aberrant immune response which is mediated by autoantibodies, complement, lymphocytes and macrophages [Köller et al. 2005]. Current concepts assume that the upregulation of cellular adhesion molecules allow autoreactive T cells and B cells to transmigrate through the blood-nerve barrier. Within the endoneurium, T cells can activate residing macrophages, which lead to enhanced phagocytosis and production of proinflammatory noxious molecules, such as nitric oxide, reactive oxygen species, proteases and proinflammatory cytokines [Lehmann et al. 2007b; Kieseier et al. 2004]. This view is supported by numerous studies demonstrating the presence of inflammatory infiltrates consisting of T cells and macrophages [Illés et al. 2004; Winer et al. 2002; Illés et al. 2000; Schmidt et al. 1996] in nerve biopsies from CIDP patients. In addition, the observation of immunoglobulin and complement deposits on myelinated nerve fibers strongly suggests that autoantibodies contribute to the process of demyelination and axonal injury by complement activation or antibody-dependent cellular cytotoxicity [Dalakas and Engel, 1980]. This is further supported by passive transfer experiments in which serum or purified IgG from CIDP patients was able to induce a conduction block and demyelination [Yan et al. 2000]. Despite intensive research, the target antigens of the aberrant immune responses in CIDP are still unknown. Experimental studies suggested that glycolipids and myelin proteins may be potential candidates; however, autoantibodies against those myelin components are detectable in a minority of CIDP patients only [Sanvito *et al.* 2009, Yan *et al.*, 2001].

#### Clinical course

Clinical symptoms of CIDP include symmetrical proximal and distal muscle weakness and diminished tendon reflexes which progress over a period of more than 8 weeks, as well as sensory deficits in a similar distribution and diminished tendon reflexes. Cranial nerve involvement is only rarely seen. Further features that support the diagnosis are elevated cerebrospinal fluid protein levels ( $\sim 90\%$  of patients) and signs of demyelination in nerve conduction studies [Köller et al. 2005]. Magnetic resonance imaging can show increased gadolinium enhancement and/or enlargement of nerve roots, cauda equina or the plexuses [Duggins et al. 1999; Midroni et al. 1999]. A sural nerve biopsy can provide further diagnostic support especially if features are present such as endoneurial edema, mononuclear cell infiltrates and signs of demyelination [Köller et al. 2005]. In general, CIDP has a variable time course and can present with a relapsing or progressive disease course.

# Treatment

Current treatment concepts in CIDP are aimed to modulate and/or suppress inflammation so as to prevent injury of nerve fibers and subsequent axonal degeneration. Generally there is consensus that due to the heterogeneity of the disease course, treatment of CIDP has to be customized to the special needs of the individual patient. Low disease incidence explains the relatively small patient numbers in the currently available trials. Corticosteroids, IVIg and plasma exchange are the mainstays of CIDP treatment [Köller et al. 2005]. Their benefit has been documented in randomized controlled trials (Table 2). In circa 80-90% of patients, a satisfactory initial treatment response can be achieved by one of those three treatment options [Tackenberg et al. 2007 Chan et al. 2006; Köller et al. 2005].

Clinical presentation, predominant affection of motor or sensory fibers, side effects and concomitant diseases are parameters, which can influence the decision about the first choice treatment for each individual patient. A positive response can be monitored as increase in muscle strength either clinically or by isokinetic dynamometry

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Treatment	Trial	Year	Trial design	Patients ( <i>n</i> )	Outcome
<b>Effective treatmen</b> Corticosteroids	it compared to Dyck <i>et al.</i>	<mark>supportive</mark> 1982	Effective treatment compared to supportive care or placebo alone Corticosteroids Dyck <i>et al.</i> 1982 Prednisone (120 mg/2d tapered to 0 mg 12 weeks) $[n = 14$ ) <i>versus</i> supportive care. Randomized controlled trial no blinding	28	Significant better outcome in changes in NDS and nerve conduction studies after 12 weeks.
Plasma exchange	Dyck <i>et al</i> .	1986	PE (6×1) versus sham-treatment, double- blind	29	Significant better outcome in measurements of nerve conduction (total, motor, proximal, velocity,
	Hahn <i>et al</i> .	1996	PE (10×1) <i>versus</i> sham-treatment, double- blind, crossover	18	significant implement in PE group in clinical out- come measures (NDS) and in most electrophy- ciological measures for the sector of the secto
IVIg, plasma exchange	Dyck <i>et al.</i>	1994	PE (7×) <i>versus</i> IVIG (0.4 g/kg, 1 per 3 weeks then 0.2 g/kg, 1 per 3 weeks) crossover in case of no improvement or worsening, sincle-blind	20	No significant differences between the two groups in clinical outcome (NDS) and electrophysiological parameter.
IVIg	Van Doorn	1990	IVIg (0.4 g/kg) 5 d) versus placebo, rando-	7	Clinical response in 7 of 7 patients after IVIg
IVIg	<i>et al.</i> Vermeulen <i>et al.</i>	1993	mizea, double-buna, crossover triat IVIg $0.4 g/kg$ , 5 d ( $n = 15$ ) versus placebo ( $n = 13$ ), double-blind, parallel group randomized controlled trial	28	treatment. No differences in clinical improvement defined as decrease on Rankin scale.
IVIg	Hahn et al. 1996	1996	IVIg [0.4 g/kg, 5 d] <i>versus</i> placebo, rando- mized, double-blind, crossover trial	25	Improvement in neurological scores (NDS); clinical grade (CG), grip strength (GS) after IVIg treatment.
IVIg	Mendell et al. 2001	2001	IVIg (1 g/kg on d 1, 2, 21) ( <i>n</i> =30) <i>versus</i> placebo ( <i>n</i> =23), randomized, controlled, double-blind	53	Improvement in primary outcome measure (change in muscle strength from baseline to day 42) and secondary outcome measures (vital capacity, nerve conduction studies)
IVIg, corticoste- roids	Hughes et al.	2001	IVIg (2.0 g/kg) ( $n$ = 12) versus prednisolone (60 mg/d 2 weeks, 40 mg/d, 1 week, 30 mg/d 1 week, 20 mg/d 1 week, 10 mg/d 1 week) ( $n$ = 12), randomized, double-	24	Significant improvement in changes in an 11-point disability scale after 2 weeks in both treatment arms.
IVIg	The ICE Trial	2008	NIMU, crossover trat NIG (2g/kg, 2-4 days then 1g/kg every 3 weeks for up to 24 weeks) <i>versus</i> pla- cebo, randomized, double-blind, pla- cebo-controlled, response-conditional crossover trial. Two periods (response- conditional crossover (rescue) period, extension phase)	117	54% participants treated with IVIg improved in adjusted INCAT disability score that was maintained through to week 24 compared to 21% of patients who received placebo ( $p$ =0.0002). Longer time to relapse during the extension phase in IVIg treated patients ( $p$ =0.011).
Treatments with uncertain or no benefit Azathioprine Dyck <i>et al.</i> 1985	<b>ncertain or no</b> Dyck <i>et al</i> .	benefit 1985	Azathioprine 2 mg/kg + prednisone $[n = 14]$ versus prednisone $[n = 13]$ . Randomized	27	No significant differences between the two groups after 9 months.
Interferon- beta 1a	Hadden <i>et al.</i>	1999	trial, parattet group design, not blind. IFN-ß (3 MIU for 2 weeks then 6 MIU for 10 weeks) s.c. 3 × weekly <i>versus</i> placebo, controlled double-blind, crossover trial.	10	No significant difference between the treatment arms in any of clinical or neurophysiological measures.
PE, plasma exchange; IVIg, intravenous immunoglobulins;	; IVIg, intravenou	ıs immunoglu	obulins; NDS, neurological disability score.		

[Harbo *et al.* 2009]. In case of insufficient or lack of response, treatment needs to be readjusted, whereas an effective initial treatment should be continued to achieve a maximum improvement. For maintenance therapy the dose of corticosteroids can be subsequently reduced to minimize side effects. Should a dose increase be required, addition of a further immunosuppressant needs to be considered. Several other immunosuppressants (azathioprine, cyclophosphamide) have been used as maintenance therapy for CIDP; however, none of those has been tested in larger controlled trials.

A beneficial effect of oral daily prednisone has been documented in a smaller trial with 28 CIDP patients, in which prednisone was given (120 mg every other day) over 12 weeks [Dyck et al. 1982]. More recently, the efficiency of pulsed oral methylprednisolone has been explored in an open-labeled prospective study [Muley et al. 2008]. In ten CIDP patients, who were followed up for at least 22 months, this steroid regimen was found to significantly improve the weakness in all patients, while treatment was well tolerated. Similarly, also the treatment regimen of high-dose intermittent methylprednisolone is often used as initial therapy for CIDP and its efficacy seems comparable to that of IVIg and oral prednisone, according to a recently published retrospective study [Lopate et al. 2005].

Two randomized controlled double-blind studies have demonstrated beneficial effects of plasma exchange in CIDP [Mehndiratta and Singh, 2007; Hahn *et al.* 1996b; Dyck *et al.* 1986]. In the first study, 29 patients with CIDP were randomized to receive six cycles of plasma exchange or sham exchange over three weeks [Dyck *et al.* 1986]. In the study by Hahn and co-workers, 18 CIDP patients were randomized to plasma exchange or sham treatment for 4 weeks before they were assigned to receive the alternate treatment in a crossover design [Hahn *et al.* 1996b].

One controlled, observer-blind study compared IVIg to plasma exchange in CIDP. In this study 20 patients with CIDP were randomized to receive either of the two treatments for 6 weeks, before they received the other treatment in a crossover design after a washout period. Both treatments improved the primary outcome measures consisting of a clinical score and summed compound muscle action potentials of motor nerves [Dyck *et al.* 1994]. Pooled data from four double-blind randomized controlled trials with totally 113 patients provided evidence for a short-term benefit effect of IVIg over placebo in CIDP [Mendell *et al.* 2001; Hahn *et al.* 1996a; Vermeulen *et al.* 1993; van Doorn *et al.* 1990]. A meta-analysis of these studies concluded that a significantly higher proportion of patients improved after IVIg treatment [Van Schaik *et al.* 2002]. Compared to oral prednisolone, IVIg has a comparable short-term efficacy according to a randomized, crossover trial that compared these two treatments in 24 patients [Hughes *et al.* 2001].

More recently, the ICE (Intravenous Immune Globulin for the Treatment of Chronic Inflammatory Demyelinating Polyradiculoneuropathy) study was published which investigated the long-term efficacy of IVIg in CIDP [Hughes *et al.* 2008]. This randomized, double-blind, placebo-controlled, response-conditional cross-over trial was designed to explore the shortand long-term benefits of 10% caprylate chromatography-purified IVIg in patients with CIDP. The percentage of patients who maintained an improvement from baseline in adjusted INCAT disability score of at least one point over 24 weeks served as primary outcome measure.

A total of 117 CIDP patients were randomized to receive either placebo (0.1% albumin; n = 58) or IVIg (n = 59) at a baseline dose of 2 g/kg over 2–4 days followed by a 1 g/kg over 1-2 days every 3 weeks for up to 24 weeks. Patients were considered as nonresponders and were assigned to the alternate treatment arm if the adjusted INCAT disability score worsened by at least one point relative to baseline or remained unchanged during the first 6 weeks, or if the score improved but returned to baseline or lower during week 6 to week 24. Treatment responders during this first period as assessed by the adjusted INCAT disability score were eligible to be randomly reassigned to IVIg or placebo in a double-blind extension phase for an additional 24 weeks. Seventy-four patients were re-randomized for the trial's extension phase, of which 43 received IVIg and 31 patients received placebo. In the first phase of the trial 32 patients (54%) who received IVIg were adjusted-INCAT responders compared with 12 patients (21%) who received placebo (p=0.0002). Although there was only a nonsignificant trend in the extension phase in several efficacy outcome measures that favor IVIg treatment, patients who continued to receive IVIg had a significantly longer time to relapse as compared to placebo treated patients (p = 0.011). This demonstrated that patients who improved under IVIg treatment were able to maintain this benefit under ongoing IVIg treatment, whereas the withdrawal of IVIg resulted in an increased risk of relapse.

This trial provided evidence that IVIg treatment has significant short-term and long-term benefits in CIDP patients. Furthermore, this study showed that patients benefit from IVIg maintenance therapy if they responded during an initial treatment with IVIg [Nobile-Orazio, 2008; Vermeulen, 2008]. The same group also reports improvement and maintenance of outcome measures regarding the health-related quality of life during the trial's treatment period [Merkies *et al.* 2008].

Immunosuppressive treatments that are widely used in CIDP include cyclophosphamide, azathioprine and cyclosporine. Azathioprine has been tested in a small, randomized controlled trial, but did not yield any additional benefit when added to prednisone [Dyck et al. 1985]. Cyclophosphamide has never been evaluated in randomized controlled trials, but beneficial effects have been reported in CIDP with an intravenous pulse therapy (1g/m<sup>2</sup> a month for up to six months) [Good et al. 1998]. According to one case series, high dose cyclophosphamide treatment can also be beneficial in CIDP patients, but there are obviously increased risks related to this treatment, such as neutropenic infections, transient renal insufficiency and alopecia [Brannagan et al. 2002]. The same group also reported a long-term benefit after a median of 2.9 years in clinical and electrophysiological parameters [Gladstone et al. 2005]. Further positive effects have been reported in case studies and smaller series for mycophenolate mofetil [Gorson et al. 2004], cyclosporine [Odaka et al. 2005; Ryan et al. 2000], alemtuzumab [Hirst et al. 2006], methotrexate [Fialho et al. 2006], autologous hematopoietic stem-cell transplantation [Reményi et al. 2007] and a combination of high dose cyclophosphamide and autologous hematopoietic stem cell transplantation [Axelson et al. 2008b]. In contrast, Interferon-beta fails to generate clinical improvement, according to a randomized controlled trial in ten CIDP patients [Hadden et al. 1999]. Similarly, intramuscular interferon beta-1a failed to significantly reduce IVIg maintenance therapy in patients with IVIgdependent CIDP, according to the results of a randomized double-blind, placebo-controlled study [Gorson *et al.* 2008].

#### Multifocal motor neuropathy

Multifocal motor neuropathy (MMN) is an immune-mediated neuropathy characterized by a slowly progressive, mainly distal muscle weakwithout sensory deficits [European ness Federation of Neurological Societies, 2006; Van Asseldonk et al. 2005; Nobile-Orazio, 2001]. Clinically it may resemble a motor neuron disease, which is the most important differential diagnosis. The exact incidence and prevalence rates of MMN are unknown, but it is believed to be a rare disease with estimated prevalence rates of 1-2/100 000 [Nobile-Orazio et al. 2005].

# Pathogenesis

Observations that patients with MMN can improve by treatment with immunomodulatory agents, especially IVIg [Léger et al. 2001], and the frequent occurrence of serum IgM antiganglioside antibodies [Willison and Yuki, 2002] suggest an immune-mediated pathogenesis of the disease. Pathological studies of motor nerves have reported loss of axons and signs of axonal degeneration at sites where a conduction block was present [Taylor et al. 2004]. In contrast, prominent changes in myelin morphology were not observed in this study, suggesting a primarily axonal pathology as underlying cause of the disease. Others have reported signs of demyelination with abnormally thin myelin and small onion-bulb formations [Kaji et al. 1993; Auer et al. 1989].

Detection of antibodies against various gangliosides (GM1, GD1a, GD1b) in up to 60% of all patients with MMN [Slee et al. 2007; Van Asseldonk et al. 2006, 2005; Willison and Yuki, 2002; Pestronk et al. 1988] provides hints for a role of these antibodies in the pathogenesis of MMN. Gangliosides, which are sialic acid containing glycolipids, can be found in most mammalian tissues, but are enriched in the nervous system. The gangliosides mentioned above belong to the most abundant types in the nervous system. It is noteworthy that the ganglioside content of motor and sensory nerves is similar [Gong et al. 2002; Ogawa-Goto and Abe, 1998; Svennerholm et al. 1994, 1992; Ogawa-Goto et al. 1990], which argues against the hypothesis that differences in ganglioside distribution are mainly responsible for the selective motor nerve damage in MMN and other antiganglioside

antibody associated neuropathies. Rather, other factors, such as antibody affinity and antigen accessibility probably play an important role. GM1 and GD1a are localized at the nodes of Ranvier and in motor nerve terminals [Gong et al. 2002; Sheikh et al. 1999]. There is evidence that under certain circumstances antiganglioside antibodies may exert various pathogenic effects. This has been shown in different experimental paradigms, which demonstrated that antiganglioside antibodies can induce conduction block, evoke demyelination, and inhibit axon regeneration [Buchwald et al. 2007; Lehmann et al. 2007c; Goodfellow et al. 2005; Halstead et al. 2004]. However, other factors are likely to contribute to the pathogenesis of MMN, since almost half of the MMN patients do not have circulating antibodies against GM1 and passive transfer studies have failed to replicate the disease in rodents [Harvey et al. 1995].

#### Clinical course

Clinically, MMN is characterized by a slowly progressive weakness with an asymmetrical distribution of motor nerve involvement. It is usually more pronounced distally than proximally, and often starts in the upper limbs. Apart from muscle weakness, progressive muscle atrophy, fasciculations and muscle cramps can occur. The age of onset is between 20 and 65 years and men tend to be more frequently affected than women [Van Asseldonk et al. 2005]. Although MMN is a slowly progressive disease, it can be associated with substantial disability in the later stages of the disease. The involvement of the upper limbs often affects manual tasks, which are important for daily living (use of keys, turn on switches, etc.) [Lange et al. 2006; Van Asseldonk et al. 2005]. A hallmark of the disease is the presence of proximal partial motor conduction block, defined as reduction in proximally stimulated muscle evoked response compared with the distally stimulated response. This can be found in 70% of patients. However, although conduction block is an electrophysiological hallmark of the disease, it remains controversial if it is essential for the diagnosis of MMN to be established [Chaudhry and Swash, 2006]. This is supported by the notion that cases of MMN without conduction block may be otherwise undistinguishable in terms of clinical course and response to IVIg [Delmont et al. 2006]. A further characteristic feature is the presence of progressive axonal degeneration, which may account for the disability in the later stages of the disease.

#### Treatment

In contrast to motor neuron diseases, MMN is a treatable disorder. It has been reported that patients with MMN improve by treatment with several immunomodulatory agents; however, the best evidence is available for IVIg [Léger et al. 2001; Federico et al. 2000] (Table 3). The efficacy of IVIg has been demonstrated in four randomized trials [Léger et al. 2001; Federico et al. 2000; Van den Berg et al. 1997; Azulay et al. 1994]. Despite clinical improvement observed in all, the data are somewhat conflicting with regard to reversibility of conduction block by IVIg treatment [Kieseier et al. 2008]. GM1 antibody titers do not correlate with treatment response [Léger et al. 2001], whereas younger patients and those with conduction blocks may benefit more from IVIg treatment. Maintenance therapy is usually required, and if it is stopped, the weakness may progress. Long-term follow-up studies revealed that the effectiveness of IVIg tends to decrease over time [Kieseier et al. 2008; Léger et al. 2008; Terenghi et al. 2004; Taylor et al. 2000]. In contrast to other chronic immune-mediated neuropathies, corticosteroids and plasma exchange are considered ineffective, and may even worsen the clinical course [Nobile-Orazio et al. 2005; Specht et al. 2000].

In a recent retrospective case series the effects of plasma exchange in MMN were studied [Lehmann et al. 2008]. Two out of seven patients responded to plasma exchange in the early phase of the disease, whereas the majority of patients did not show any clinical response and electrophysiological follow-up examinations demonstrated further worsening. This suggests that the therapeutic value of plasma exchange in MMN is limited. However, it is possible that carefully selected patients may benefit from plasma exchange treatment; for example, those with high titers of antiganglioside antibodies. Since antiganglioside antibodies are only transiently reduced, plasma exchange is probably only useful as an adjunctive treatment in combination with long-term immunosuppression.

Other agents that have been proposed as treatment for MMN include cyclophosphamide [Chaudhry *et al.* 1993; Feldman *et al.* 1991; Pestronk *et al.* 1988] and azathioprine [Hausmanowa-Petrusewicz *et al.* 1991]. According to one case report cyclophosphamide and autologous stem cell transplantation are ineffective in MMN [Axelson *et al.* 2008a]. Beneficial effects have

Treatment	Trial	Year	Trial design	Patients (n)	Outcome
Effective treatme	ent				
IVIg	Léger <i>et al.</i>	2001	IVIg (500 mg/kg/day, 5 d 1 × month for 3 months <i>versus</i> placebo, double-blind, randomized, controlled crossover trial	18	7/9 patients who received IVIg responded compared with 2/9 patients who received placebo ( $p=0.03$ ). No dif- ferences in MRC score, electrophysiological studies and changes in anti-GM1 antibody titers.
IVIg	Federico et al.	2000	IVIg (0.4 g/kg/day, 5 d) <i>versus</i> pla- cebo, double-blind, randomized, controlled crossover trial	16	Improvement in NDS with IVIg treatment compared to placebo ( $p = 0.038$ ) after 28 d. Improvement in grip strength ( $p = 0.0021$ ) and conduction block ( $p = 0.037$ ) with IVIg
IVIg	Azulay et al.	1994	IVIg (0.4 g/kg/day, 5 d) <i>versus</i> pla- cebo, double-blind, randomized, controlled crossover trial	12	treatment. Improvement in muscle strength after IVIg treatment.
IVIg	Van den Bergh et al.	1997	IVIg $(2 \times 0.4 \text{ g/kg/day}, 5 \text{ d})$ $(n = 4)$ and placebo; IVIg $(1 \times 0.4 \text{ g/kg/}$ day, 5 d) and placebo $(n = 2)$ , double-blind, randomized, controlled crossover trial	6	Clinical improvement in 5/6 patients with IVIg but not placebo.
Presumably ineff	ective treatm	ent			
Combination of IVIg and mycopheno- late mofetil	Pieper <i>et al.</i>	2007	Standard dose IVIg every 2–5 weeks + 1000 mg/d mycophe- nolate mofetil first week then 2000 mg/d <i>versus</i> placebo for 12 months, randomized, double- blind, placebo-controlled study	28	No statistical differences in primary endpoint (IVIg dose reduction of 50% during adjunctive treatment).

Table 3. Treatment of	' multifocal motor	r neuropathy based	l on evidence o	t clinical trials.

PE, plasma exchange; IVIg, intravenous immunoglobulins; NDS, neurological disability score; MIU, million international units, s.c., subcutaneously.

been observed in patients treated with the monoclonal anti-CD20 antibody rituximab, [Ruegg *et al.* 2004]; however, other groups were unable to confirm this [Gorson *et al.* 2007; Rojas-García *et al.* 2003]. More recently, a randomized controlled trial evaluated the efficacy of mycophenolate mofetil as adjunctive therapy for MMN [Piepers *et al.* 2007]. Twenty-eight patients who received IVIg were randomized to receive in addition to IVIg either mycophenolate mofetil or placebo for a period of 12 months. A reduction of the IVIg dose to 50% served as the primary endpoint. After 1 year, there was no significant difference between the two groups, suggesting that mycophenolate mofetil is probably ineffective in MMN.

# Polyneuropathy associated with monoclonal gammopathy

In approximately 10% of patients with idiopathic polyneuropathies a monoclonal immunoglobulin

can be detected in serum or urine [Kelly et al. 1981]. Of those, monoclonal gammopathy of undetermined significance (MGUS) is the most frequent form, whereas lymphoproliferative disorders such as Waldenström's macroglobulinemia represent rare causes of monoclonal gammopathies [Ropper and Gorson, 1998]. Occasionally, a paraprotein occurs as part of the POEMS syndrome (peripheral neuropathy organomegaly - endocrinopathy - myeloma skin disease) [Dispenzieri et al. 2003]. Polyneuropathies associated with monoclonal gammopathy can be very heterogeneous in terms of clinical course, electrophysiology and response to treatment. The best-defined paraprotein- associated neuropathic condition is the IgM paraprotein associated demyelinating polyneuropathy in which the IgM reacts with myelin-associated glycoprotein (MAG) [Nobile-Orazio, 2004; Vital, 2001]. Whether axonal changes noticed in the context of a monoclonal

gammopathy are a coincidental finding or part of the disease spectrum [Allen *et al.* 2007; Lunn and Nobile-Orazio, 2006] remains a matter of debate, since both polyneuropathy and monoclonal gammopathy are common in the elderly.

#### Pathogenesis

Due to the heterogeneous disease spectrum of IgA and IgG paraprotein associated polyneuropathies, no consistent pathogenesis has been established for these neuropathic conditions. In contrast, the finding that up to 50% of IgM paraproteins react with MAG suggests a role of these antibodies in the pathogenesis of this subgroup. Pathologic studies have demonstrated segmental demyelination and signs of secondary axonal degeneration in sural nerve biopsies [Willison and Yuki, 2002]. By immunohistochemistry deposits of IgM and complement can be found on dermal myelinated fibers [Lombardi et al. 2005]. Another pathological feature is abnormally spaced myelin, which can be detected by electron microscopy [Ritz et al. 1999; Jacobs and Scadding 1990]. Lunn and colleagues reported in an electron microscopic study an altered distance of neighboring neurofilaments in axons of sural nerves from patients with anti-MAG paraproteinaemic neuropathies [Lunn et al. 2002]. These findings implicate a pathogenic effect of the anti-MAG antibody by alteration of MAG dependent control of neurofilament spacing in peripheral nerves.

# Clinical course

Anti-MAG paraproteinaemic neuropathy is clinically characterized by a predominantly sensory demyelinating neuropathy [Nobile-Orazio, 2004]. It is slowly progressive and has an overall better prognosis than other chronic immunemediated neuropathies. Sometimes an upper limb postural neuropathic tremor may occur.

# Treatment

Treatment in patients with paraproteinaemic neuropathy is aimed at targeting the presumably pathogenic paraprotein or abnormal B-cell clone. An adequate hematological treatment is the mainstay in cases in which a lymphoproliferative disorder can be identified as cause for the gammopathy [Nobile-Orazio, 2004]. In those cases patients can be referred to a specialized hematology service, which establish the primary treatment strategy in cooperation with the treating neurologist, who can be consulted in case of any questions that relates to the accompanying neuropathy. For example in POEMS syndrome, high-dose chemotherapy with autologous hematopoietic stem-cell transplantation is a treatment strategy, which can result in remission of hematological manifestations and marked improvement of the neuropathy [Créange *et al.* 2008].

Generally, controlled trials are rare for paraproteinaemic neuropathies, which makes it difficult to recommend any particular evidence-based immunotherapy [Lunn and Nobile-Orazio, 2003] (Table 4). In a randomized double-blind placebo-controlled crossover trial IVIg was not effective in the majority of 11 patients with IgM paraproteinemic demyelinating neuropathy [Dalakas et al. 1996]. Another double-blind crossover trial of 22 patients with IgM paraproteinemic demyelinating neuropathy demonstrated a clinical benefit at 4 weeks [Comi et al. 2002]. These results indicate that IVIg may only have a short-term benefit in the treatment of IgM paraproteinemic neuropathy [Lunn and Nobile-Orazio, 2003].

According to one randomized, controlled, double-blind trial, patients with MGUS associated polyneuropathy can benefit from plasma exchange [Dyck *et al.* 1991]. In this study, 39 patients received either plasma exchange twice weekly for 3 weeks or sham treatment. The primary outcome measures, the neuropathy disability score and summed compound muscle action potentials of motor nerves showed a beneficial effect in the plasma exchange group. A subgroup analysis further demonstrated that patients with IgM gammopathy are less responsive to plasma exchange than those with IgG or IgA gammopathy.

A recent study evaluated the effects of rituximab in 26 patients with anti-MAG-antibody positive polyneuropathy. The patients were randomized to receive either four weekly infusions of  $375 \text{ mg/m}^2$ rituximab or placebo. As primary outcome served a change of 1 or more on the INCAT leg disability scores after 8 months. In the rituximab group 4 out of 13 patients improved, whereas none of the patients in the placebo group showed an improvement [Dalakas et al. 2007]. This study was preceded by several case series and open trials that evaluated the therapeutic potential of rituximab on IgM paraproteinaemic neuropathies with and without anti-MAG reactivity [Benedetti et al. 2008, 2007; Gorson et al. 2007; Renaud et al. 2006, 2003;

Treatment	Trial	Year	Characteristics: design	Patients ( <i>n</i> )	Outcome
Effective treatme	nt				
IVIg	Comi <i>et al.</i>	2002	IVIg (2 g/kg) ( <i>n</i> = 12) <i>versus</i> placebo, double-blind, randomized, controlled crossover trial	22	Decrease in overall disability during IVIg treatment (p=0.001). Improvement in secondary outcome measures (Rankin scale, time to walk 10 meters, grip strength) during IVIg treatment.
IVIg	Dalakas et al.	1996	IVIg (2 g/kg 1 × /month for 3 months) <i>versus</i> placebo, double-blind, randomized, controlled crossover trial	11	Improvement only in 2/11 patients during IVIg treatment. No sig- nificant benefit of IVIg treatment
Plasma exchange	Dyck <i>et al</i> .	1991	PE (6×) <i>versus</i> sham-treatment, double-blind	39	Marked improvement in disability score. More benefit in the IgG and IgA gammopathy subgroup as compared with IgM gammo- pathy population.
Presumably ineff	ective treatment				
Cyclophospha- mide with prednisone	Niermeijer <i>et al.</i>	2007	Oral cyclophosphamide (500 mg/d, 4 d + oral prednisone 60 mg/d 5 d <i>versus</i> placebo every 28 d (6×)	35	No difference in functional scale after 6 months. Improvement in several secondary outcomes.
Interferon-alpha	Mariette <i>et al.</i>	2000	Interferon- $\alpha$ (4.5 MIU 3 × week for 6 months) <i>versus</i> placebo, rando- mized, double blind trial	24	No clinical improvement com- pared to placebo.

Table 4. Treatment of paraproteinemic neuropathies (with and without anti-MAG antibodies) based on evidence of clinical trials.

PE, plasma exchange; IVIg, intravenous immunoglobulins; NDS, neurological disability score; MIU, million international units, s.c., subcutaneously.

Levine and Pestronk, 1999]. In a mixed cohort of 21 patients with different forms of polyneuropathies associated with antiganglioside or anti-MAG antibodies the majority showed an improvement in muscle strength after one and two years of treatment, IgM levels and antibody titers decreased [Pestronk et al. 2003]. In a case series by Rojas-Garcia et al. [2003] rituximab had no favorable effect on two patients (one with chronic motor neuropathy and antiganglioside IgM antibodies, one with an IgM monoclonal gammopathy). In contrast Renaud and colleagues reported a salutary effect of rituximab (dose  $375 \text{ mg/m}^2$ ) in nine patients with antiassociated polyneuropathy. MAG Clinical improvement was correlated with diminished IgM levels and antibody titers [Renaud et al. 2003]. The same group also reported a followup study in which eight patients were treated with a higher dose  $(750 \text{ mg/m}^2)$ . The higher dosage was well tolerated and four patients experienced marked clinical improvement, which went in parallel with a decrease in anti-MAG antibody titers and serum IgM levels.

One randomized, double-blind trial investigated the effect of interferon-alpha in 24 patients with polyneuropathy with monoclonal IgM but could not demonstrate any benefit [Mariette *et al.* 2000]. Likewise, oral cyclophosphamide combined with prednisone did not show a benefit in primary outcome measures compared with placebo treatment in a double-blind, randomized study in 35 patients with IgM MGUS polyneuropathy [Niermeijer *et al.* 2007].

#### **Conflict of interest statement**

None declared

#### References

Allen, D., Lunn, M.P., Niermeijer, J. and Nobile-Orazio, E. (2007) Treatment for IgG and IgA paraproteinaemic neuropathy. *Cochrane Database Syst Rev* CD005376.

Andersen, S.M., Ling, C.C., Zhang, P., Townson, K., Willison, H.J. and Bundle, D.R. (2004) Synthesis of ganglioside epitopes for oligosaccharide specific immunoadsorption therapy of Guillian-Barré syndrome. Org Biomol Chem 2: 1199–1212.

Asbury, A.K., Arnason, B.G. and Adams, R.D. (1969) The inflammatory lesion in idiopathic polyneuritis. *Medicine* 4: 173–215.

Auer, R., Bell, R. and Lee, M. (1989) Neuropathy with onion bulb formations and pure motor manifestations. *Can J Neurol Sci* 16: 194–197.

Axelson, H., Oberg, G. and Askmark, H. (2008a) No benefit of treatment with cyclophosphamide and autologous blood stem cell transplantation in multifocal motor neuropathy. *Acta Neurol Scand* 117: 432–434.

Axelson, H., Oberg, G. and Askmark, H. (2008b) Successful repeated treatment with high dose cyclophosphamide and autologous blood stem cell transplantation in CIDP. *J Neurol Neurosurg Psychiatry* 79: 612–614.

Azulay, J., Blin, O., Pouget, J., Boucraut, J., Billé-Turc, F., Carles, G. and Serratrice, G. (1994) Intravenous immunoglobulin treatment in patients with motor neuron syndromes associated with anti-GM1 antibodies: a double-blind, placebo-controlled study. *Neurology* 44: 429–432.

Bechtold, D.A., Yue, X., Evans, R.M., Davies, M., Gregson, N.A. and Smith, K.J. (2005) Axonal protection in experimental autoimmune neuritis by the sodium channel blocking agent flecainide. *Brain* 128: 18–28.

Benedetti, L., Briani, C., Franciotta, D., Carpo, M., Padua, L., Zara, G. *et al.* (2008) Long-term effect of rituximab in anti-mag polyneuropathy. *Neurology* 71: 1742–1744.

Benedetti, L., Briani, C., Grandis, M., Vigo, T., Gobbi, M., Ghiglione, E. *et al.* (2007) Predictors of response to rituximab in patients with neuropathy and anti-myelin associated glycoprotein immunoglobulin M. *J Peripher Nerv Syst* 12: 102–107.

Brannagan, T.R., Pradhan, A., Heiman-Patterson, T., Winkelman, A., Styler, M., Topolsky, D. *et al.* (2002) High-dose cyclophosphamide without stem-cell rescue for refractory CIDP. *Neurology* 58: 1856–1858.

Bril, V., Ilse, W.K., Pearce, R., Dhanani, A., Sutton, D. and Kong, K. (1996) Pilot trial of immunoglobulin versus plasma exchange in patients with Guillain– Barré syndrome. *Neurology* 46: 100–103.

Buchwald, B., Ahangari, R. and Toyka, K.V. (2002) Differential blocking effects of the monoclonal anti-GQ1b IgM antibody and alpha-latrotoxin in the absence of complement at the mouse neuromuscular junction. *Neurosci Lett* 334: 25–28.

Buchwald, B., Toyka, K.V., Zielasek, J., Weishaupt, A., Schweiger, S. and Dudel, J. (1998) Neuromuscular blockade by IgG antibodies from patients with Guillain–Barré syndrome: a macro-patch-clamp study. *Ann Neurol* 44: 913–922. Buchwald, B., Zhang, G., Vogt-Eisele, A.K., Zhang, W., Ahangari, R., Griffin, J.W. *et al.* (2007) Anti-ganglioside antibodies alter presynaptic release and calcium influx. *Neurobiol Dis* 28: 113–121.

Bullens, R.W., Halstead, S.K., O'Hanlon, G.M., Veitch, J., Molenaar, P.C., Willison, H.J. and Plomp, J.J. (2005) Concanavalin a inhibits pathophysiological effects of anti-ganglioside GQ1b antibodies at the mouse neuromuscular synapse. *Muscle Nerve*.

Chan, Y., Allen, D., Fialho, D., Mills, K. and Hughes, R. (2006) Predicting response to treatment in chronic inflammatory demyelinating polyradiculoneuropathy. *J Neurol Neurosurg Psychiatry* 77: 114–116.

Chaudhry, V., Corse, A., Cornblath, D., Kuncl, R., Drachman, D., Freimer, M., Miller, R. and Griffin, J. (1993) Multifocal motor neuropathy: response to human immune globulin. *Ann Neurol* 33: 237–242.

Chaudhry, V. and Swash, M. (2006) Multifocal motor neuropathy: is conduction block essential?. *Neurology* 67: 558–559.

Cheng, Q., Jiang, G.X., Fredrikson, S., Link, H. and De Pedro-Cuesta, J. (2000) Incidence of Guillain– Barré syndrome in Sweden 1996. *Eur J Neurol* 7: 11–16.

Chiò, A., Cocito, D., Bottacchi, E., Buffa, C., Leone, M., Plano, F. *et al.* (2007) Idiopathic chronic inflammatory demyelinating polyneuropathy: an epidemiological study in Italy. *J Neurol Neurosurg Psychiatry* 78: 1349–1353.

Comi, G., Roveri, L., Swan, A., Willison, H., Bojar, M., Illa, I. *et al.* (2002) A randomised controlled trial of intravenous immunoglobulin in IgM paraprotein associated demyelinating neuropathy. *J Neurol* 249: 1370–1377.

Créange, A., Chater, A., Brouet, J., Jaccard, A., Rahmouni, A., Lefaucheur, J. *et al.* (2008) A case of POEMS syndrome treated by autologous hematopoietic stem-cell transplantation. *Nat Clin Pract Neurol* 4: 686–691.

Dalakas, M. and Engel, W. (1980) Immunoglobulin and complement deposits in nerves of patients with chronic relapsing polyneuropathy. *Arch Neurol* 37: 637–640.

Dalakas, M.C., Quarles, R.H., Farrer, R.G., Dambrosia, J., Soueidan, S., Stein, D.P. *et al.* (1996) A controlled study of intravenous immunoglobulin in demyelinating neuropathy with IgM gammopathy. *Ann Neurol* 40: 792–795.

Dalakas, M.C., Rakocevic, G., Salajegheh, M.K., Dambrosia, J., Hahn, A., Raju, R. *et al.* (2009) A double-blind, placebo-controlled study of rituximab in patients with anti-MAG antibody-demyelinating polyneuropathy. *Ann Neurol*, in press.

Delmont, E., Azulay, J., Giorgi, R., Attarian, S., Verschueren, A., Uzenot, D. et al. (2006)

Multifocal motor neuropathy with and without conduction block: a single entity?. *Neurology* 67: 592–596.

Diener, H.C., Haupt, W.F., Kloss, T.M., Rosenow, F., Philipp, T., Koeppen, S. *et al.* (2001) A preliminary, randomized, multicenter study comparing intravenous immunoglobulin, plasma exchange, and immune adsorption in Guillain–Barré syndrome. *Eur Neurol* 46: 107–109.

Dispenzieri, A., Kyle, R., Lacy, M., Rajkumar, S., Therneau, T., Larson, D. *et al.* (2003) POEMS syndrome: definitions and long-term outcome. *Blood* 101: 2496–2506.

Duggins, A., McLeod, J., Pollard, J., Davies, L., Yang, F., Thompson, E. *et al.* (1999) Spinal root and plexus hypertrophy in chronic inflammatory demyelinating polyneuropathy. *Brain* 122: 1383–1390.

Dyck, P., O'Brien, P., Oviatt, K., Dinapoli, R., Daube, J., Bartleson, J. *et al.* (1982) Prednisone improves chronic inflammatory demyelinating polyradiculoneuropathy more than no treatment. *Ann Neurol* 11: 136–141.

Dyck, P., O'Brien, P., Swanson, C., Low, P. and Daube, J. (1985) Combined azathioprine and prednisone in chronic inflammatory-demyelinating polyneuropathy. *Neurology* 35: 1173–1176.

Dyck, P.J., Daube, J., O'Brien, P., Pineda, A., Low, P.A., Windebank, A.J. *et al.* (1986) Plasma exchange in chronic inflammatory demyelinating polyradiculoneuropathy. *N Engl J Med* 314: 461–465.

Dyck, P.J., Litchy, W.J., Kratz, K.M., Suarez, G.A., Low, P.A., Pineda, A.A. *et al.* (1994) A plasma exchange versus immune globulin infusion trial in chronic inflammatory demyelinating polyradiculoneuropathy. *Ann Neurol* 36: 838–845.

Dyck, P.J., Low, P.A., Windebank, A.J., Jaradeh, S.S., Gosselin, S., Bourque, P. *et al.* (1991) Plasma exchange in polyneuropathy associated with monoclonal gammopathy of undetermined significance. *N Engl J Med* 325: 1482–1486.

European Federation of Neurological Societies (2006) European Federation of Neurological Societies/ Peripheral Nerve Society Guideline on management of multifocal motor neuropathy. Report of a joint task force of the European Federation of Neurological Societies and the Peripheral Nerve Society, *J Peripher Nerv Syst* 11: 1–8.

Federico, P., Zochodne, D., Hahn, A., Brown, W. and Feasby, T. (2000) Multifocal motor neuropathy improved by IVIg: randomized, double-blind, placebocontrolled study. *Neurology* 55: 1256–1262.

Feldman, E.L., Bromberg, M.B., Albers, J.W. and Pestronk, A. (1991) Immunosuppressive treatment in multifocal motor neuropathy. *Ann Neurol* 30: 397–401.

Felts, P.A., Smith, K.J., Gregson, N.A. and Hughes, R.A. (2002) Brain-derived neurotrophic factor in experimental autoimmune neuritis. *J Neuroimmunol* 124: 62–69. Fialho, D., Chan, Y., Allen, D., Reilly, M. and Hughes, R. (2006) Treatment of chronic inflammatory demyelinating polyradiculoneuropathy with methotrexate. *J Neurol Neurosurg Psychiatry* 77: 544–547.

Flachenecker, P., Hartung, H.P. and Reiners, K. (1997a) Power spectrum analysis of heart rate variability in Guillain–Barré syndrome. A longitudinal study. *Brain* 120(Pt 10): 1885–1894.

Flachenecker, P., Wermuth, P., Hartung, H.P. and Reiners, K. (1997b) Quantitative assessment of cardiovascular autonomic function in Guillain–Barré syndrome. *Ann Neurol* 42: 171–179.

French Cooperative Group on Plasma Exchange in Guillain–Barré syndrome. (1987) Efficiency of plasma exchange in Guillain–Barré syndrome: role of replacement fluids, *Ann Neurol* 22: 753–761.

French Cooperative Group on Plasma Exchange in Guillain–Barré Syndrome. (1992) Plasma exchange in Guillain–Barré syndrome: one-year follow-up, *Ann Neurol* 32: 94–97.

French Cooperative Group on Plasma Exchange in Guillain–Barré Syndrome. (1997) Appropriate number of plasma exchanges in Guillain–Barré syndrome, *Ann Neurol* 41: 298–306.

Garssen, M., van Koningsveld, R., van Doorn, P., Merkies, I., Scheltens-de Boer, M., van Leusden, J. *et al.* (2007) Treatment of Guillain–Barré syndrome with mycophenolate mofetil: a pilot study. *J Neurol Neurosurg Psychiatry* 78: 1012–1013.

Gladstone, D., Prestrud, A. and Brannagan, T.R. (2005) High-dose cyclophosphamide results in longterm disease remission with restoration of a normal quality of life in patients with severe refractory chronic inflammatory demyelinating polyneuropathy. *7 Peripher Nerv Syst* 10: 11–16.

Gold, R., Hartung, H.P. and Toyka, K.V. (2000) Animal models for autoimmune demyelinating disorders of the nervous system. *Mol Med Today* 6: 88–91.

Gong, Y., Tagawa, Y., Lunn, M.P., Laroy, W., Heffer-Lauc, M., Li, C.Y. *et al.* (2002) Localization of major gangliosides in the PNS: implications for immune neuropathies. *Brain* 125: 2491–2506.

Good, J., Chehrenama, M., Mayer, R. and Koski, C. (1998) Pulse cyclophosphamide therapy in chronic inflammatory demyelinating polyneuropathy. *Neurology* 51: 1735–1738.

Goodall, J., Kosmidis, J. and Geddes, A. (1974) Effect of corticosteroids on course of Guillain–Barré syndrome. *Lancet* 1: 524–526.

Goodfellow, J.A., Bowes, T., Sheikh, K., Odaka, M., Halstead, S.K., Humphreys, P.D. *et al.* (2005) Overexpression of GD1a ganglioside sensitizes motor nerve terminals to anti-GD1a antibody-mediated injury in a model of acute motor axonal neuropathy. *J Neurosci* 25: 1620–1628.

Gorson, K., Amato, A. and Ropper, A. (2004) Efficacy of mycophenolate mofetil in patients with chronic

immune demyelinating polyneuropathy. *Neurology* 63: 715–717.

Gorson, K., Hughes, R., Cros, D., Pollard, J., Vallat, J., Riester, K. *et al.* (2008) Efficacy of interferon beta-1 a in patients with chronic inflammatory demyelinating polyradiculoneuropathy (CIDP) *Neurology* 70: A369.

Gorson, K., Natarajan, N., Ropper, A. and Weinstein, R. (2007) Rituximab treatment in patients with IVIg-dependent immune polyneuropathy: a prospective pilot trial. *Muscle Nerve* 35: 66–69.

Govoni, V. and Granieri, E. (2001) Epidemiology of the Guillain–Barré syndrome. *Curr Opin Neurol* 14: 605–613.

Guillain–Barré Syndrome Steroid Trial Group, (1993) Double-blind trial of intravenous methylprednisolone in Guillain–Barré syndrome, *Lancet* 341: 586–590.

Guillain–Barré Syndrome Study Group, (1985) Plasmapheresis and acute Guillain–Barré syndrome, *Neurology* 35: 1096–1104.

Gürses, N., Uysal, S., Cetinkaya, F., Işlek, I. and Kalayci, A. (1995) Intravenous immunoglobulin treatment in children with Guillain–Barré syndrome. *Scand J Infect Dis* 27: 241–243.

Haber, P., DeStefano, F., Angulo, F., Iskander, J., Shadomy, S., Weintraub, E. *et al.* (2004) Guillain– Barré syndrome following influenza vaccination. *JAMA* 292: 2478–2481.

Hadden, R.D., Gregson, N.A., Gold, R., Smith, K.J. and Hughes, R.A. (2002) Accumulation of immunoglobulin across the 'blood-nerve barrier' in spinal roots in adoptive transfer experimental autoimmune neuritis. *Neuropathol Appl Neurobiol* 28: 489–497.

Hadden, R.D., Sharrack, B., Bensa, S., Soudain, S.E. and Hughes, R.A. (1999) Randomized trial of interferon beta-1a in chronic inflammatory demyelinating polyradiculoneuropathy. *Neurology* 53: 57–61.

Hafer-Macko, C.E., Sheikh, K.A., Li, C.Y., Ho, T.W., Cornblath, D.R., McKhann, G.M. *et al.* (1996) Immune attack on the Schwann cell surface in acute inflammatory demyelinating polyneuropathy. *Ann Neurol* 39: 625–635.

Hahn, A., Bolton, C., Zochodne, D. and Feasby, T. (1996a) Intravenous immunoglobulin treatment in chronic inflammatory demyelinating polyneuropathy. A double-blind, placebo-controlled, cross-over study. *Brain* 119(Pt 4): 1067–1077.

Hahn, A.F., Bolton, C.F., Pillay, N., Chalk, C., Benstead, T., Bril, V. *et al.* (1996b) Plasma-exchange therapy in chronic inflammatory demyelinating polyneuropathy. A double-blind, sham-controlled, cross-over study. *Brain* 119: 1055–1066.

Halstead, S.K., Humphreys, P.D., Goodfellow, J.A., Wagner, E.R., Smith, R.A. and Willison, H.J. (2005) Complement inhibition abrogates nerve terminal injury in Miller Fisher syndrome. *Ann Neurol* 58: 203–210. Halstead, S.K., O'Hanlon, G.M., Humphreys, P.D., Morrison, D.B., Morgan, B.P., Todd, A.J. *et al.* (2004) Anti-disialoside antibodies kill perisynaptic Schwann cells and damage motor nerve terminals via membrane attack complex in a murine model of neuropathy. *Brain* 127: 2109–2123.

Halstead, S.K., Zitman, F.M., Humphreys, P.D., Greenshields, K., Verschuuren, J.J., Jacobs, B.C. *et al.* (2008) Eculizumab prevents anti-ganglioside antibody-mediated neuropathy in a murine model. *Brain*.

Harbo, T., Andersen, H. and Jakobsen, J. (2009) Acute motor response following a single IVIG treatment course in chronic inflammatory demyelinating polyneuropathy. *Muscle Nerve*.

Harvey, G., Toyka, K., Zielasek, J., Kiefer, R., Simonis, C. and Hartung, H. (1995) Failure of anti-GM1 IgG or IgM to induce conduction block following intraneural transfer. *Muscle Nerve* 18: 388–394.

Hausmanowa-Petrusewicz, I., Rowińska-Marcińska, K. and Kopeć, A. (1991) Chronic acquired demyelinating motor neuropathy. *Acta Neurol Scand* 84: 40–45.

Hirst, C., Raasch, S., Llewelyn, G. and Robertson, N. (2006) Remission of chronic inflammatory demyelinating polyneuropathy after alemtuzumab (Campath 1H). *J Neurol Neurosurg Psychiatry* 77: 800–802.

Ho, T.W., Mishu, B., Li, C.Y., Gao, C.Y., Cornblath, D.R., Griffin, J.W. *et al.* (1995a) Guillain–Barré syndrome in northern China. Relationship to Campylobacter jejuni infection and anti-glycolipid antibodies. *Brain* 118(Pt 3): 597–605.

Ho, T.W., Mishu, B., Li, C.Y., Gao, C.Y., Cornblath, D.R., Griffin, J.W. *et al.* (1995b) Guillain–Barré syndrome in northern China. Relationship to Campylobacter jejuni infection and anti-glycolipid antibodies. *Brain* 118(Pt 3): 597–605.

Hughes, R., Bensa, S., Willison, H., Van den Bergh, P., Comi, G., Illa, I. *et al.* (2001) Randomized controlled trial of intravenous immunoglobulin versus oral prednisolone in chronic inflammatory demyelinating polyradiculoneuropathy. *Ann Neurol* 50: 195–201.

Hughes, R., Donofrio, P., Bril, V., Dalakas, M., Deng, C., Hanna, K. *et al.* (2008) Intravenous immune globulin (10% caprylate-chromatography purified) for the treatment of chronic inflammatory demyelinating polyradiculoneuropathy (ICE study): a randomised placebo-controlled trial. *Lancet Neurol* 7: 136–144.

Hughes, R.A. and Cornblath, D.R. (2005) Guillain–Barré syndrome. *Lancet* 366: 1653–1666.

Hughes, R.A., Raphael, J.C., Swan, A.V. and van Doorn, P.A. (2006) Intravenous immunoglobulin for Guillain–Barré syndrome. *Cochrane Database Syst Rev* CD002063. Hughes, R.A. and Rees, J.H. (1997) Clinical and epidemiologic features of Guillain–Barré syndrome. J Infect Dis 176(Suppl. 2): S92–98.

Hughes, R.A., Swan, A.V., Raphael, J.C., Annane, D., van Koningsveld, R. and van Doorn, P.A. (2007) Immunotherapy for Guillain–Barré syndrome: a systematic review. *Brain* 130: 2245–2257.

Iijima, M., Koike, H., Hattori, N., Tamakoshi, A., Katsuno, M., Tanaka, F. *et al.* (2008) Prevalence and incidence rates of chronic inflammatory demyelinating polyneuropathy in the Japanese population. *J Neurol Neurosurg Psychiatry* 79: 1040–1043.

Illa, I., Ortiz, N., Gallard, E., Juarez, C., Grau, J.M. and Dalakas, M.C. (1995) Acute axonal Guillain–Barré syndrome with IgG antibodies against motor axons following parenteral gangliosides. *Ann Neurol* 38: 218–224.

Illés, Z., Kondo, T., Newcombe, J., Oka, N., Tabira, T. and Yamamura, T. (2000) Differential expression of NK T cell V alpha 24J alpha Q invariant TCR chain in the lesions of multiple sclerosis and chronic inflammatory demyelinating polyneuropathy. *7 Immunol* 164: 4375–4381.

Illés, Z., Shimamura, M., Newcombe, J., Oka, N. and Yamamura, T. (2004) Accumulation of Valpha7.2-Jalpha33 invariant T cells in human autoimmune inflammatory lesions in the nervous system. *Int Immunol* 16: 223–230.

Imbach, P., Barandun, S., d'Apuzzo, V., Baumgartner, C., Hirt, A., Morell, A., Rossi, E., Schöni, M., Vest, M. and Wagner, H. (1981) High-dose intravenous gammaglobulin for idiopathic thrombocytopenic purpura in childhood. *Lancet* 1: 1228–1231.

Jacobs, J.M. and Scadding, J.W. (1990) Morphological changes in IgM paraproteinaemic neuropathy. *Acta Neuropathol* 80: 77–84.

Kaji, R., Oka, N., Tsuji, T., Mezaki, T., Nishio, T., Akiguchi, I. and Kimura, J. (1993) Pathological findings at the site of conduction block in multifocal motor neuropathy. *Ann Neurol* 33: 152–158.

Kelly, J.J., Jr., Kyle, R.A., O'Brien, P.C. and Dyck, P.J. (1981) Prevalence of monoclonal protein in peripheral neuropathy. *Neurology* 31: 1480–1483.

Khalili-Shirazi, A., Gregson, N., Gray, I., Rees, J., Winer, J. and Hughes, R. (1999) Antiganglioside antibodies in Guillain–Barré syndrome after a recent cytomegalovirus infection. *J Neurol Neurosurg Psychiatry* 66: 376–379.

Kieseier, B., Meyer Zu Hörste, G., Lehmann, H., Gold, R. and Hartung, H. (2008) Intravenous immunoglobulins in the treatment of immune neuropathies. *Curr Opin Neurol* 21: 555–562.

Kieseier, B.C. and Hartung, H.P. (2003) Therapeutic strategies in the Guillain–Barré syndrome. *Semin Neurol* 23: 159–168.

Kieseier, B.C., Kiefer, R., Gold, R., Hemmer, B., Willison, H.J. and Hartung, H.P. (2004) Advances in understanding and treatment of immune-mediated disorders of the peripheral nervous system. *Muscle Nerve* 30: 131–156.

Kieseier, B.C., Krivacic, K., Jung, S., Pischel, H., Toyka, K.V., Ransohoff, R.M. *et al.* (2000) Sequential expression of chemokines in experimental autoimmune neuritis. *J Neuroimmunol* 110: 121–129.

Kieseier, B.C., Wiendl, H. and Hartung, H.P. (2006) The inflamed peripheral nervous system: update on immune therapies. *Curr Opin Neurol* 19: 433–436.

Köller, H., Kieseier, B.C., Jander, S. and Hartung, H.P. (2005) Chronic inflammatory demyelinating neuropathy. *N Engl J Med* 352: 1343–1356.

Korinthenberg, R., Schessl, J., Kirschner, J. and Mönting, J. (2005) Intravenously administered immunoglobulin in the treatment of childhood Guillain–Barré syndrome: a randomized trial. *Pediatrics* 116: 8–14.

Lange, D., Weimer, L., Trojaborg, W., Lovelace, R., Gooch, C. and Rowland, L. (2006) Multifocal motor neuropathy with conduction block: slow but not benign. *Arch Neurol* 63: 1778–1781.

Lasky, T., Terracciano, G., Magder, L., Koski, C., Ballesteros, M., Nash, D. *et al.* (1998) The Guillain– Barré syndrome and the 1992–1993 and 1993–1994 influenza vaccines. *N Engl J Med* 339: 1797–1802.

Léger, J., Viala, K., Cancalon, F., Maisonobe, T., Gruwez, B., Waegemans, T. *et al.* (2008) Intravenous immunoglobulin as short- and long-term therapy of multifocal motor neuropathy: a retrospective study of response to IVIg and of its predictive criteria in 40 patients. *J Neurol Neurosurg Psychiatry* 79: 93–96.

Léger, J.M., Chassandre, B., Musset, L., Meininger, V., Bouche, P. and Baumann, N. (2001) Intravenous immunoglobulin therapy in multifocal motor neuropathy: a double-blind, placebo-controlled study. *Brain* 124: 145–153.

Lehmann, H.C. and Hartung, H.P. (2008) Complementing the therapeutic armamentarium for Miller Fisher Syndrome and related immune neuropathies. *Brain* 131: 1168–1170.

Lehmann, H.C., Hartung, H.-P., Hetzel, G.R., Stüve, O. and Kieseier, B.C. (2006) Plasma exchange in neuroimmunological disorders. Part 2 Treatment of neuromuscular disorders. *Arch Neurol* 63: 1066–1071.

Lehmann, H.C., Hoffmann, F.R., Fusshoeller, A., Hetzel, G.R., Hartung, H.P., Schroeter, M. *et al.* (2008) The therapeutic value of plasma exchange in multifocal motor neuropathy. *J Neurol Sci* 27: 34–9.

Lehmann, H.C., Kohne, A., zu Horste, G.M. and Kieseier, B.C. (2007a) Incidence of Guillain–Barré syndrome in Germany. J Peripher Nerv Syst 12: 285.

Lehmann, H.C., Köhne, A., Meyer zu Hörste, G., Dehmel, T., Kiehl, O., Hartung, H.P., Kastenbauer, S. and Kieseier, B.C. (2007b) Role of nitric oxide as mediator of nerve injury in inflammatory neuropathies. *J Neuropathol Exp Neurol* 66: 305–312.

Lehmann, H.C., Lopez, P.H., Zhang, G., Ngyuen, T., Zhang, J., Kieseier, B.C., Mori, S. and Sheikh, K.A. (2007c) Passive immunization with anti-ganglioside antibodies directly inhibits axon regeneration in an animal model. *J Neurosci* 27: 27–34.

Levine, T. and Pestronk, A. (1999) IgM antibodyrelated polyneuropathies: B-cell depletion chemotherapy using Rituximab. *Neurology* 52: 1701–1704.

Lombardi, R., Erne, B., Lauria, G., Pareyson, D., Borgna, M., Morbin, M. *et al.* (2005) IgM deposits on skin nerves in anti-myelin-associated glycoprotein neuropathy. *Ann Neurol* 57: 180–187.

Lonigro, A. and Devaux, J. (2009) Disruption of neurofascin and gliomedin at nodes of Ranvier precedes demyelination in experimental allergic neuritis. *Brain* 132: 260–273.

Lopate, G., Pestronk, A. and Al-Lozi, M. (2005) Treatment of chronic inflammatory demyelinating polyneuropathy with high-dose intermittent intravenous methylprednisolone. *Arch Neurol* 62: 249–254.

Lunn, M., Manji, H., Choudhary, P., Hughes, R. and Thomas, P. (1999) Chronic inflammatory demyelinating polyradiculoneuropathy: a prevalence study in south east England. *J Neurol Neurosurg Psychiatry* 66: 677–680.

Lunn, M.P., Crawford, T.O., Hughes, R.A., Griffin, J.W. and Sheikh, K.A. (2002) Antimyelin-associated glycoprotein antibodies alter neurofilament spacing. *Brain* 125: 904–911.

Lunn, M.P. and Nobile-Orazio, E. (2003) Immunotherapy for IgM anti-myelin-associated glycoprotein paraprotein-associated peripheral neuropathies. *Cochrane Database Syst Rev* CD002827.

Lunn, M.P. and Nobile-Orazio, E. (2006) Immunotherapy for IgM anti-myelin-associated glycoprotein paraprotein-associated peripheral neuropathies. *Cochrane Database Syst Rev* CD002827.

Makowska, A., Pritchard, J., Sanvito, L., Gregson, N., Peakman, M., Hayday, A. *et al.* (2008) Immune responses to myelin proteins in Guillain–Barré syndrome. *J Neurol Neurosurg Psychiatry* 79: 664–671.

Mariette, X., Brouet, J., Chevret, S., Leger, J., Clavelou, P., Pouget, J. *et al.* (2000) A randomised double blind trial versus placebo does not confirm the benefit of alpha-interferon in polyneuropathy associated with monoclonal IgM. *J Neurol Neurosurg Psychiatry* 69: 279–280.

McLeod, J., Pollard, J., Macaskill, P., Mohamed, A., Spring, P. and Khurana, V. (1999) Prevalence of chronic inflammatory demyelinating polyneuropathy in New South Wales, Australia. *Ann Neurol* 46: 910–913.

Mehndiratta, M.M. and Singh, A.C. (2007) Plasmapheresis for chronic inflammatory demyelinating polyradiculoneuropathy. *Curr Allergy Asthma Rep* 7: 274–279.

Mendell, J., Barohn, R., Freimer, M., Kissel, J., King, W., Nagaraja, H. *et al.* (2001) Randomized controlled trial of IVIg in untreated chronic inflammatory demyelinating polyradiculoneuropathy. *Neurology* 56: 445–449.

Merkies, I.S.J., Bril, V., Dalakas, M.C., Deng, C.Q., Donofrio, P., Hanna, K. *et al.* (2008) Quality-of-life (QoL) improvements with immune globulin intravenous, 10% caprylate/chromatography purified (IGIV-C) in chronic inflammatory demyelinating polyneuropathy (CIDP). *Ann Neurol* 64: S4–S5.

Meyer zu Horste, G., Hartung, H.P. and Kieseier, B.C. (2007) From bench to bedside– experimental rationale for immune-specific therapies in the inflamed peripheral nerve. *Nat Clin Pract Neurol* 3: 198–211.

Midroni, G., de Tilly, L., Gray, B. and Vajsar, J. (1999) MRI of the cauda equina in CIDP: clinical correlations. *J Neurol Sci* 170: 36–44.

Moore, R. and Derry, S. (2006) Systematic review and meta-analysis of randomised trials and cohort studies of mycophenolate mofetil in lupus nephritis. *Arthritis Res Ther* 8: R182.

Muley, S., Kelkar, P. and Parry, G. (2008) Treatment of chronic inflammatory demyelinating polyneuropathy with pulsed oral steroids. *Arch Neurol* 65: 1460–1464.

Niermeijer, J.M.F., Eurelings, M., van der Linden, M.W., Lokhorst, H.M., Franssen, H., Fischer, K. *et al.* (2007) Intermittent cyclophosphamide with prednisone versus placebo for polyneuropathy with IgM monoclonal gammopathy. *Neurology* 69: 50–59.

Nobile-Orazio, E. (2001) Multifocal motor neuropathy. *J Neuroimmunol* 115: 4–18.

Nobile-Orazio, E. (2004) IgM paraproteinaemic neuropathies. *Curr Opin Neurol* 17: 599–605.

Nobile-Orazio, E. (2008) Evidence for long-term IVIg treatment of chronic inflammatory demyelinating polyradiculoneuropathy. *Nat Clin Pract Neurol* 4: 352–353.

Nobile-Orazio, E., Cappellari, A. and Priori, A. (2005) Multifocal motor neuropathy: current concepts and controversies. *Muscle Nerve* 31: 663–680.

O'Hanlon, G.M., Humphreys, P.D., Goldman, R.S., Halstead, S.K., Bullens, R.W., Plomp, J.J. *et al.* (2003) Calpain inhibitors protect against axonal degeneration in a model of anti-ganglioside antibody-mediated motor nerve terminal injury. *Brain* 126: 2497–2509.

O'Leary, C.P., Veitch, J., Durward, W.F., Thomas, A.M., Rees, J.H. and Willison, H.J. (1996) Acute oropharyngeal palsy is associated with antibodies to GQ1b and GT1a gangliosides. *J Neurol Neurosurg Psychiatry* 61: 649–651. Odaka, M., Tatsumoto, M., Susuki, K., Hirata, K. and Yuki, N. (2005) Intractable chronic inflammatory demyelinating polyneuropathy treated successfully with ciclosporin. *J Neurol Neurosurg Psychiatry* 76: 1115–1120.

Ogawa-Goto, K. and Abe, T. (1998) Gangliosides and glycosphingolipids of peripheral nervous system myelins – a minireview. *Neurochem Res* 23: 305–310.

Ogawa-Goto, K., Funamoto, N., Abe, T. and Nagashima, K. (1990) Different ceramide compositions of gangliosides between human motor and sensory nerves. *7 Neurochem* 55: 1486–1493.

Overell, J.R. and Willison, H.J. (2005) Recent developments in Miller Fisher syndrome and related disorders. *Curr Opin Neurol* 18: 562–566.

Pestronk, A., Cornblath, D.R., Ilyas, A., Baba, H., Quarles, R.H., Griffin, J.W. *et al.* (1988) A treatable multifocal motor neuropathy with antbodies to GM1 ganglioside. *Ann Neurol* 24: 73–78.

Pestronk, A., Florence, J., Miller, T., Choksi, R., Al-Lozi, M. and Levine, T. (2003) Treatment of IgM antibody associated polyneuropathies using rituximab. *J Neurol Neurosurg Psychiatry* 74: 485–489.

Piepers, S., Van den Berg-Vos, R., Van der Pol, W., Franssen, H., Wokke, J. and Van den Berg, L. (2007) Mycophenolate mofetil as adjunctive therapy for MMN patients: a randomized, controlled trial. *Brain* 130: 2004–2010.

Plasma Exchange/Sandoglobobulin Gulillain–Barré Syndrom Trial Group (1997) Randomised trial of plasma exchange, intravenous immunoglobulin, and combined treatments in Guillain–Barré syndrome. *Lancet* 349: 225–230.

Prineas, J.W. (1981) Pathology of the Guillain–Barré syndrome. *Ann Neurol* 9(Suppl.): 6–19.

Pritchard, J., Gray, I.A., Idrissova, Z.R., Lecky, B.R., Sutton, I.J., Swan, A.V. *et al.* (2003) A randomized controlled trial of recombinant interferon-beta 1a in Guillain–Barré syndrome. *Neurology* 61: 1282–1284.

Ramos-Alvarez, M., Bessudo, L. and Sabin, A.B. (1969) Paralytic syndromes associated with noninflammatory cytoplasmic or nuclear neuronopathy. Acute paralytic disease in Mexican children, neuropathologically distinguishable from Landry-Guillain– Barré syndrome. *JAMA* 207: 1481–1492.

Rees, J.H., Thompson, R.D., Smeeton, N.C. and Hughes, R.A. (1998) Epidemiological study of Guillain–Barré syndrome in south east England. *J Neurol Neurosurg Psychiatry* 64: 74–77.

Reményi, P., Masszi, T., Borbényi, Z., Soós, J., Siklós, L. and Engelhardt, J. (2007) CIDP cured by allogeneic hematopoietic stem cell transplantation. *Eur J Neurol* 14: e1–2.

Renaud, S., Fuhr, P., Gregor, M., Schweikert, K., Lorenz, D., Daniels, C. *et al.* (2006) High-dose rituximab and anti-MAG-associated polyneuropathy. *Neurology* 66: 742–744. Renaud, S., Gregor, M., Fuhr, P., Lorenz, D., Deuschl, G., Gratwohl, A. *et al.* (2003) Rituximab in the treatment of polyneuropathy associated with anti-MAG antibodies. *Muscle Nerve* 27: 611–615.

Ritz, M., Erne, B., Ferracin, F., Vital, A., Vital, C. and Steck, A. (1999) Anti-MAG IgM penetration into myelinated fibers correlates with the extent of myelin widening. *Muscle Nerve* 22: 1030–1037.

Rojas-García, R., Gallardo, E., de Andrés, I., de Luna, N., Juarez, C., Sánchez, P. and Illa, I. (2003) Chronic neuropathy with IgM anti-ganglioside antibodies: lack of long term response to rituximab. *Neurology* 61: 1814–1816.

Ropper, A.H. and Gorson, K.C. (1998) Neuropathies associated with paraproteinemia. *N Engl J Med* 338: 1601–1607.

Ruegg, S.J., Fuhr, P. and Steck, A. (2004) Rituximab stabilizes multifocal motor neuropathy increasingly less responsive to IVIg. *Neurology* 63: 2178–2179.

Ruts, L., Rico, R., van Koningsveld, R., Botero, J., Meulstee, J., Gerstenbluth, I., Merkies, I. and van Doorn, P. (2008) Pain accompanies pure motor Guillain–Barré syndrome. *J Peripher Nerv Syst* 13: 305–306.

Ruts, L., van Koningsveld, R., Jacobs, B. and van Doorn, P. (2007) Determination of pain and response to methylprednisolone in Guillain–Barré syndrome. *J Neurol* 254: 1318–1322.

Ryan, M., Grattan-Smith, P., Procopis, P., Morgan, G. and Ouvrier, R. (2000) Childhood chronic inflammatory demyelinating polyneuropathy: clinical course and long-term outcome. *Neuromusc Disord* 10: 398–406.

Sanvito, L., Makowska, A., Mahdi-Rogers, M., Hadden, R., Peakman, M., Gregson, N. *et al.* (2009) Humoral and cellular immune responses to myelin protein peptides in chronic inflammatory demyelinating polyradiculoneuropathy. *J Neurol Neurosurg Psychiatry* 80: 333–338.

Schmidt, B., Toyka, K.V., Kiefer, R., Full, J., Hartung, H.P. and Pollard, J. (1996) Inflammatory infiltrates in sural nerve biopsies in Guillain–Barré syndrome and chronic inflammatory demyelinating neuropathy. *Muscle Nerve* 19: 474–487.

Sedano, M.J., Calleja, J., Canga, E. and Berciano, J. (1994) Guillain–Barré syndrome in Cantabria, Spain. An epidemiological and clinical study. *Acta Neurol Scand* 89: 287–292.

Sheikh, K.A., Deerinck, T.J., Ellisman, M.H. and Griffin, J.W. (1999) The distribution of ganglioside-like moieties in peripheral nerves. *Brain* 122(Pt 3): 449–460.

Singh, N.K. and Gupta, A. (1996) Do corticosteroids influence the disease course or mortality in Guillain-Barre' syndrome? *J Assoc Physicians India* 44: 22–24.

Slee, M., Selvan, A. and Donaghy, M. (2007) Multifocal motor neuropathy: the diagnostic spectrum and response to treatment. *Neurology* 69: 1680–1687. Specht, S., Claus, D. and Zieschang, M. (2000) Plasmapheresis in multifocal motor neuropathy: a case report.  $\mathcal{J}$  *Neurol Neurosurg Psychiatry* 68: 533–535.

Susuki, K., Rasband, M.N., Tohyama, K., Koibuchi, K., Okamoto, S., Funakoshi, K. *et al.* (2007) Anti-GM1 antibodies cause complementmediated disruption of sodium channel clusters in peripheral motor nerve fibers. *J Neurosci* 27: 3956–3967.

Svennerholm, L., Bostrom, K., Fredman, P., Jungbjer, B., Lekman, A., Mansson, J.E. *et al.* (1994) Gangliosides and allied glycosphingolipids in human peripheral nerve and spinal cord. *Biochim Biophys Acta* 1214: 115–123.

Svennerholm, L., Bostrom, K., Fredman, P., Jungbjer, B., Mansson, J.E. and Rynmark, B.M. (1992) Membrane lipids of human peripheral nerve and spinal cord. *Biochim Biophys Acta* 1128: 1–7.

Tackenberg, B., Lünemann, J., Steinbrecher, A., Rothenfusser-Korber, E., Sailer, M., Brück, W. *et al.* (2007) Classifications and treatment responses in chronic immune-mediated demyelinating polyneuropathy. *Neurology* 68: 1622–1629.

Taylor, B., Dyck, P., Engelstad, J., Gruener, G. and Grant, I. (2004) Multifocal motor neuropathy: pathologic alterations at the site of conduction block. *J Neuropathol Exp Neurol* 63: 129–137.

Taylor, B., Wright, R., Harper, C. and Dyck, P. (2000) Natural history of 46 patients with multifocal motor neuropathy with conduction block. *Muscle Nerve* 23: 900–908.

Terenghi, F., Cappellari, A., Bersano, A., Carpo, M., Barbieri, S. and Nobile-Orazio, E. (2004) How long is IVIg effective in multifocal motor neuropathy? *Neurology* 62: 666–668.

Van Asseldonk, J.T., Franssen, H., Van den Berg-Vos, R.M., Wokke, J.H. and Van den Berg, L.H. (2005) Multifocal motor neuropathy. *Lancet Neurol* 4: 309–319.

Van Asseldonk, J.T.H., Van den Berg, L.H., Kalmijn, S., Van den Berg-Vos, R.M., Polman, C.H., Wokke, J.H.J. *et al.* (2006) Axon loss is an important determinant of weakness in multifocal motor neuropathy. *J Neurol Neurosurg Psychiatry* 77: 743–747.

Van den Berg, L., Franssen, H., Van Doorn, P. and Wokke, J. (1997) Intravenous immunoglobulin treatment in lower motor neuron disease associated with highly raised anti-GM1 antibodies. *J Neurol Neurosurg Psychiatry* 63: 674–677.

van der Meche, F.G. and Schmitz, P.I. (1992) A randomized trial comparing intravenous immune globulin and plasma exchange in Guillain–Barré syndrome. The Dutch Guillain–Barré Study Group. *N Engl J Med* 326: 1123–1129. van Doorn, P., Brand, A., Strengers, P., Meulstee, J. and Vermeulen, M. (1990) High-dose intravenous immunoglobulin treatment in chronic inflammatory demyelinating polyneuropathy: a double-blind, placebo-controlled, crossover study. *Neurology* 40: 209–212.

van Doorn, P.A., Ruts, L. and Jacobs, B.C. (2008) Clinical features, pathogenesis, and treatment of Guillain–Barré syndrome. *Lancet Neurol* 7: 939–950.

van Koningsveld, R., Schmitz, P.I., Meche, F.G., Visser, L.H., Meulstee, J. and van Doorn, P.A. (2004) Effect of methylprednisolone when added to standard treatment with intravenous immunoglobulin for Guillain–Barré syndrome: randomised trial. *Lancet* 363: 192–196.

van Koningsveld, R., Van Doorn, P., Schmitz, P., Ang, C. and Van der Meché, F. (2000) Mild forms of Guillain–Barré syndrome in an epidemiologic survey in The Netherlands. *Neurology* 54: 620–625.

Van Schaik, I.N., Winer, J.B., De Haan, R. and Vermeulen, M. (2002) Intravenous immunoglobulin for chronic inflammatory demyelinating polyradiculoneuropathy. *Cochrane Database Syst Rev* CD001797.

Vermeulen, M. (2008) Intravenous immunoglobulin: a first-line treatment in CIDP?. *Lancet Neurol* 7: 115–116.

Vermeulen, M., van Doorn, P., Brand, A., Strengers, P., Jennekens, F. and Busch, H. (1993) Intravenous immunoglobulin treatment in patients with chronic inflammatory demyelinating polyneuropathy: a double blind, placebo controlled study. *J Neurol Neurosurg Psychiatry* 56: 36–39.

Vital, A. (2001) Paraproteinemic neuropathies. *Brain Pathol* 11: 399–407.

Wanschitz, J., Maier, H., Lassmann, H., Budka, H. and Berger, T. (2003) Distinct time pattern of complement activation and cytotoxic T cell response in Guillain–Barré syndrome. *Brain* 126: 2034–2042.

Willison, H.J. (2002) Anti-glycolipid antibodies in the diagnosis of autoimmune neuropathies. *Rev Neurol* (*Paris*) 158: S16–20.

Willison, H.J. (2005) The immunobiology of Guillain– Barré syndromes. *J Peripher Nerv Syst* 10: 94–112.

Willison, H.J., Townson, K., Veitch, J., Boffey, J., Isaacs, N., Andersen, S.M. *et al.* (2004) Synthetic disialylgalactose immunoadsorbents deplete anti-GQ1b antibodies from autoimmune neuropathy sera. *Brain* 127: 680–691.

Willison, H.J. and Yuki, N. (2002) Peripheral neuropathies and anti-glycolipid antibodies. *Brain* 125: 2591–2625.

Winer, J., Hughes, S., Cooper, J., Ben-Smith, A. and Savage, C. (2002) Gamma delta T cells infiltrating sensory nerve biopsies from patients with inflammatory neuropathy. *J Neurol* 249: 616–621. Yan, W.X., Taylor, J., Andrias-Kauba, S. and Pollard, J.D. (2000) Passive transfer of demyelination by serum or IgG from chronic inflammatory demyelinating polyneuropathy patients. *Ann Neurol* 47: 765–775.

Yan, W.X., Archelos, J.J., Hartung, H.-P. and Pollard, J.D. (2000) PO protein is a target antigen in chronic inflammatory demyelinating polyradiculoneuropathy. *Ann Neurol* 50: 286–292.

Yuki, N. (1997) Molecular mimicry between gangliosides and lipopolysaccharides of Campylobacter jejuni isolated from patients with Guillain–Barré syndrome and Miller Fisher syndrome. *J Infect Dis* 176: S150–153.

Yuki, N. (2005) Carbohydrate mimicry: a new paradigm of autoimmune diseases. *Curr Opin Immunol* 17: 577–582.

Yuki, N. and Odaka, M. (2005) Ganglioside mimicry as a cause of Guillain–Barré syndrome. *Curr Opin Neurol* 18: 557–561.

Yuki, N., Tagawa, Y. and Hirata, K. (1998) Minimal number of plasma exchanges needed to reduce

immunoglobulin in Guillain-Barré syndrome. *Neurology* 51: 875-877.

Yuki, N., Taki, T., Inagaki, F., Kasama, T., Takahashi, M., Saito, K. *et al.* (1993) A bacterium lipopolysaccharide that elicits Guillain–Barré syndrome has a GM1 ganglioside-like structure. *J Exp Med* 178: 1771–1775.

Yuki, N., Taki, T., Takahashi, M., Saito, K., Yoshino, H., Tai, T. *et al.* (1994) Molecular mimicry between GQ1b ganglioside and lipopolysaccharides of *Campylobacter jejuni* isolated from patients with Fisher's syndrome. *Ann Neurol* 36: 791–793.

Yuki, N., Yoshino, H., Sato, S. and Miyatake, T. (1990) Acute axonal polyneuropathy associated with anti-GM1 antibodies following *Campylobacter* enteritis. *Neurology* 40: 1900–1902.

Zhang, G., Lopez, P.H., Li, C.Y., Mehta, N.R., Griffin, J.W., Schnaar, R.L. *et al.* (2004) Anti-ganglioside antibody-mediated neuronal cytotoxicity and its protection by intravenous immunoglobulin: implications for immune neuropathies. *Brain* 127: 1085–1100.

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