

RESEARCH ARTICLE

Open Access

# Induction of cell cycle arrest at G1 and S phases and cAMP-dependent differentiation in C6 glioma by low concentration of cycloheximide

Xijun Liu<sup>1,5</sup>, Jin-Ming Yang<sup>2,4</sup>, Samuel S Zhang<sup>1,3</sup>, Xin-Yuan Liu<sup>5,6</sup>, David X Liu<sup>1,3,4\*</sup>

## Abstract

**Background:** Differentiation therapy has been shown effective in treatment of several types of cancer cells and may prove to be effective in treatment of glioblastoma multiforme, the most common and most aggressive primary brain tumor. Although extensively used as a reagent to inhibit protein synthesis in mammalian cells, whether cycloheximide treatment leads to glioma cell differentiation has not been reported.

**Methods:** C6 glioma cell was treated with or without cycloheximide at low concentrations (0.5-1 µg/ml) for 1, 2 and 3 days. Cell proliferation rate was assessed by direct cell counting and colony formation assays. Apoptosis was assessed by Hoechst 33258 staining and FACS analysis. Changes in several cell cycle regulators such as Cyclins D1 and E, PCNA and Ki67, and several apoptosis-related regulators such as p53, p-JNK, p-AKT, and PARP were determined by Western blot analysis. C6 glioma differentiation was determined by morphological characterization, immunostaining and Western blot analysis on upregulation of GFAP and o p-STAT3 expression, and upregulation of intracellular cAMP.

**Results:** Treatment of C6 cell with low concentration of cycloheximide inhibited cell proliferation and depleted cells at both G2 and M phases, suggesting blockade at G1 and S phases. While no cell death was observed, cells underwent profound morphological transformation that indicated cell differentiation. Western blotting and immunostaining analyses further indicated that changes in expression of several cell cycle regulators and the differentiation marker GFAP were accompanied with cycloheximide-induced cell cycle arrest and cell differentiation. Increase in intracellular cAMP, a known promoter for C6 cell differentiation, was found to be elevated and required for cycloheximide-promoted C6 cell differentiation.

**Conclusion:** Our results suggest that partial inhibition of protein synthesis in C6 glioma by low concentration of cycloheximide induces cell cycle arrest at G1 and M phases and cAMP-dependent cell differentiation.

## Background

Glioblastoma multiforme (GBM) is the most common central nervous system malignancy, whose highly invasive and diffuse nature leaves rare opportunity for cure. Despite aggressive surgical approaches, optimized radiation therapy regimens and the application of cytotoxic chemotherapies, the median survival rates from time of diagnosis range from 12-15 months [1].

Differentiation, as apoptosis, is a defense mechanism by which mammalian cells guard against tumorigenesis.

Differentiation therapy, using agents that promote cancer cell differentiation, has been shown to be effective in vitro and in vivo in treatment of several types of cancer cells [2]. Thus, neuroblastoma cells were shown to undergo terminal differentiation upon elevation of intracellular adenosine 3', 5'-cyclic monophosphate (cAMP) concentration after treatment with cyclic nucleotide phosphodiesterase inhibitors or adenylate cyclase activators [3-7]. Cholera toxin was reported to induce malignant glioma cell differentiation via the PKA/CREB pathway [8]. Differentiation of glioma cells can also be achieved by treatment of cells with *Datura stramonium* agglutinin [9]. Notably, *all-trans-retinoic acid* has been used as an agent to induce cell differentiation in clinical

\* Correspondence: dliu@psu.edu

<sup>1</sup>Department of Neural and Behavioral Sciences, Penn State University College of Medicine, 500 University Drive, Hershey, PA 17033, USA  
Full list of author information is available at the end of the article

treatment of acute promyelocytic leukemia (APL) [10,11], demonstrating the remarkable efficacy of differentiation therapy in treatment of cancers.

Cycloheximide (CHX), originally isolated from *Streptomyces griseus*, has been widely used as an experimental tool to analyze the contribution of protein synthesis to the intracellular signaling and associated cellular function. It inhibits protein translation in mammalian cells at multiple steps, with a particular potency on translation initiation [12]. Protein synthesis consumes enormous energy and resources in cells especially in fast growing cancer cells, and inhibition of protein synthesis by CHX, even in a relatively mild situation, may constitute a major blockade to the cellular functions such as cell proliferation. Indeed, inhibition of synthesis of specific proteins by CHX, such as c-Myc [13], p27 [14] has been interpreted as evidence for the necessity of those cell cycle regulators in either cell proliferation or cell cycle arrest. Similarly, CHX has been shown to either block [15-17] or promote [18-20] gene expression-dependent cell death in different types of cells and experimental paradigms. Surprisingly, a possible biological effect of CHX in cancer therapy, to our knowledge, has never been investigated.

In the present study, we show that low concentration of CHX (LCC) potently inhibited C6 cell proliferation and depleted cells at both G2 and M phases, suggesting blockade at G1 and S phases during which massive protein accumulation is required. Surprisingly, treatment of C6 cell with LCC also induced dramatic morphological transformation that is indicative of cellular differentiation; this change was accompanied by upregulation of differentiation marker glial fibrillary acidic protein (GFAP). In addition, we found that intracellular cAMP levels were dramatically increased in C6 cells treated with LCC, and that this elevated intracellular cAMP is responsible for C6 cell differentiation and upregulation of GFAP.

## Methods

### Cell culture and reagents

Rat C6 glioma cell (ATCC cat. number CCL-107) was cultured in DMEM (Invitrogen) containing 10% FBS (Atlanta Biologicals). Cycloheximide (C4859, CHX), 8-(4-Chlorophenylthio) adenosine 3', 5'-cyclic monophosphate sodium salt (C3912, 8-pCPT-cAMP) and Adenosine (A9251, Ado) were obtained from Sigma. Sources of the antibodies and dilutions are as follows: GFAP (#3670, 1:1000 for both Western and immunostaining), p-Akt (#9271, 1:1000), Cleaved Caspase-3 (Asp175) (1:1000) were from Cell signaling Technology. PARP (P76420, 1:1000) from BD Biosciences. p-JNK (sc-6254, 1:1000), p-Stat3 (sc-7993, 1:1000), Cyclin D1(R-124, 1:1000), Cyclin E (sc-481, 1:750), PCNA (sc-56, 1:500), p-I $\kappa$ B- $\alpha$  (sc-101713, 1:1000), p53 (sc-98, 1:1000), and

Ki67 (sc-7846, 1:500) from Santa Cruz Biotechnology. The materials and experimental procedures used in this research were approved by the Biosafety and Recombinant DNA Committee (BRDC) at Penn State University Hershey Medical Center.

### Morphological observations

C6 cells were plated in 6-well plates at a density of  $1 \times 10^5$  per well. Cells were incubated in culture medium containing CHX and/or other reagents at indicated concentrations. Photographs of cells were taken 24 h later.

### Proliferation assay

C6 cells were seeded into 6-well plates at a density of  $5 \times 10^5$  per well and were treated as indicated. Cells were collected after trypsin digestion and counted with a hemocytometer. Values in proliferation charts are expressed as means  $\pm$  SEM (n = 3).

### Colony formation assay

C6 cells were seeded in 35 mm culture dish at 1,000 cells per dish. After indicated treatment, cells were stained with 0.5% crystal violet solution (made in 25% methanol and stored at room temperature) for 10 min and photographed.

### Flow cytometry

Cell flow cytometry analysis was done as described previously [21]. At the indicated time, cells were trypsinized, washed twice with  $1 \times$  PBS, and pelleted by low speed centrifugation. Pellet was resuspended with 70% ethanol for 30 min at 4°C. Cells were spun down and were incubated with the DNA-binding dye propidium iodide (PI) solution [0.1% sodium citrate (w/v), 0.1% Triton X-100 (v/v), and 50 mg/L PI in deionized water] for 1 hour at room temperature. Finally, cells were analyzed by a FACS caliber flow cytometer.

### Immunocytochemical staining and confocal microscopy

Experiments were carried out as in [17]. Briefly, cells grown on coverslips were treated as indicated and then fixed in methyl alcohol at -20°C for 10 min. Samples were blocked for 1 h with 5% BSA and then incubated for 1 h at 37°C with the Anti-GFAP antibody. A goat anti-rabbit IgG-TR (Santa Cruz) secondary antibody was used for specific detection of anti-GFAP. Nuclei were stained with Hoechst 33258.

### SDS-PAGE and immunoblotting

Cells in the culture dish were washed with ice-cold PBS and lysed in ice-cold RIPA buffer [50 mM Tris-HCl, pH 7.5, 150 mM NaCl, 0.5% sodium deoxycholate, 0.1% SDS, 1% Nonidet P40]. Cell extracts preparation and Western blotting analysis were done as previously [22].

### Determination of intracellular cAMP level

C6 cells ( $2 \times 10^5$ ) were seeded in 24-well. After overnight culturing, the cells were treated with CHX, CHX and Ado, at indicated concentrations, and vehicle control for 24 hours. Cells were lysed in 200  $\mu$ l of 0.1 M HCl and cell extracts were collected after removing cell debris by centrifugation. Quantification of cAMP was carried out using a kit (BioVision) following manufacturer's instruction. Three independent measurements were done for each treatment. cAMP level in untreated C6 cells was arbitrarily set at 1. Data are presented as means  $\pm$  SEM (n = 3).

## Results

### Potent inhibition of C6 glioma cell proliferation by LCC

To see how C6 cell responds to CHX treatment, we performed a dose-response analysis in 6-well plates using CHX concentration ranging from 0 to 100  $\mu$ g/ml (1  $\mu$ g/ml = 3.5  $\mu$ M). High concentration of CHX (50-100  $\mu$ g/ml) prompted cell death in a few hours (data not shown). LCC (0.1-10  $\mu$ g/ml) did not cause cell death, permitting us to examine C6 cell response over longer period of time. As shown in Figure 1A, dramatic morphological transformation was observed among the C6 cells after treatment with 1  $\mu$ g/ml CHX for 24 h; cells were no longer flat, but rather were spindle-shaped with processes (Figure 1A). No cell death was detectable above the background for several days as indicated by Hoechst 33258 staining (data not shown), although the nuclei of the C6 cells treated with CHX were uniformly smaller (Figure 1A). These CHX-treated cells seemed to cease proliferation completely; a cell proliferation assay showed that the normally fast-growing C6 cell no longer proliferated when treated with LCC (Figure 1B). We additionally performed colony-formation assay showing that LCC was extremely potent in inhibiting C6 cell proliferation (Figure 1C). The experiments described in Figure 1B and 1C also demonstrated a dose-dependent response and indicated that the  $IC_{50}$  for CHX to inhibit C6 proliferation was less than 0.5  $\mu$ g/ml.

To see whether cessation of C6 cell proliferation is accompanied with changes in cell proliferation markers and cell cycle regulators in response to LCC, we prepared cell extracts from C6 cells untreated or treated with CHX (0.5  $\mu$ g/ml) for 24 or 48 h and carried out western blotting analysis. As shown in Figure 1D, cell proliferation markers Ki67 and PCNA and cell proliferation regulators cyclin D1 and cyclin E were all downregulated. Taken together, these data showed that LCC suppresses the expression of several cell cycle markers and stimulators in C6 cell and potently block C6 cell proliferation.

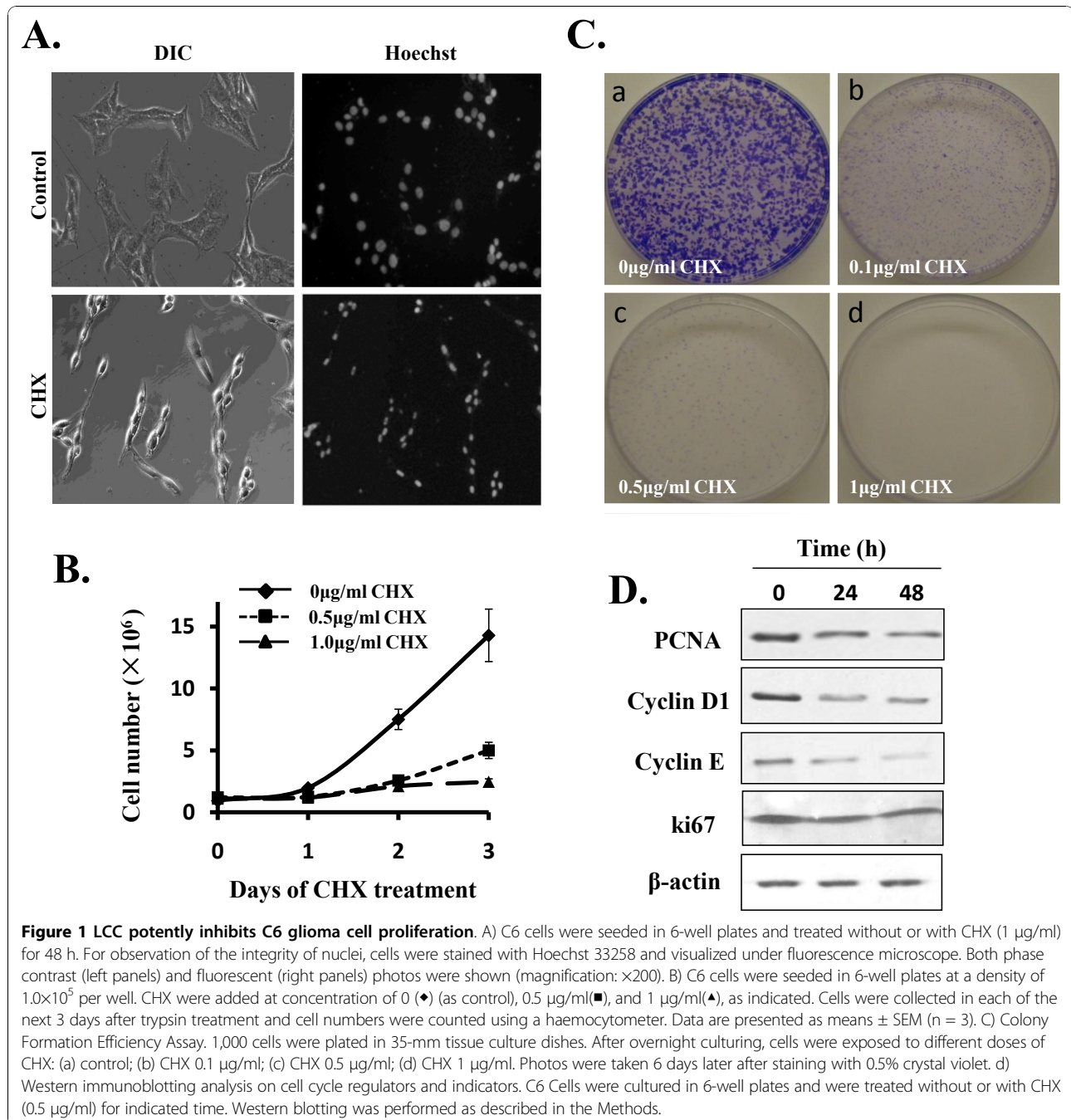
### LCC does not induce apoptosis in C6 cell but depletes cells at G2 and M phases

CHX has been shown to either block [15-17] or induce [18-20] gene expression-dependent cell death in different types of cells and experimental paradigms. In contrast to high concentration of CHX (50-100  $\mu$ g/ml), which caused death of C6 cells within 6 hours, LCC (0.1-10  $\mu$ g/ml) did not show any impact on C6 cell survival (Figure 1A); a direct count of the percentage of cell death showed no difference between control and LCC-treated C6 cells (data not shown). We next carried out FACS analysis on C6 cells treated with various LCC. As shown in Figure 2A-D, no sub-G1 population was observed in cells treated with up to 10  $\mu$ g/ml CHX, further confirming the non-apoptotic effect of LCC on C6 cell. While CHX treatment did not alter the percentage of cells in G1 and S phases, it surprisingly depleted the G2/M population in a dose-dependent manner. We interpret the results to indicate that LCC blocked C6 cell proliferation at both G1 and S phases, which is consistent with the expectation that progression of both G1 and S phases require high level of protein synthesis.

We additionally examined expression of several molecules involved in cell survival and death regulation. As shown in Figure 2E, most of these molecules such as p-Akt and p53 either did not change or underwent changes that in fact favor cell survival. Therefore, we concluded that LCC is not apoptotic to C6 cell, instead it blocks C6 cell cycle progression at the G1 and S phases.

### LCC promotes reversible differentiation of C6 glioma cell

The morphological transformation (Figure 1A) and cessation of proliferation (Figure 1B, C) of C6 cells in response to LCC suggested that C6 cells might undergo a process of differentiation. Indeed, as shown in Figure 3A, the smaller and spindle-shaped cells formed after treatment of CHX were remarkably similar to those treated with 8-CPT-cAMP, an analog of cAMP that is known to induce C6 cell differentiation [23,24]. Immunostaining of C6 cells with GFAP, a reliable marker for gliogenesis, indicated a dramatic increase in GFAP production in C6 cells treated with CHX (1  $\mu$ g/ml) (Figure 3B). This increased expression of GFAP was observed with CHX concentrations ranging from 0.5-10  $\mu$ g/ml and at comparable levels to 8-CPT-cAMP (cAMP) treatment (Figure 3C), and with highest expression observed at 1 d post treatment (Figure 3D). These data thus showed that C6 cell differentiation induced by LCC is similar to that induced by cAMP. In addition, we demonstrated that the increase of GFAP in C6 cells treated with CHX was accompanied with a measurable increase in p-STAT3 (Figure 2E), which is an obligatory transcription activator for GFAP [25,26].

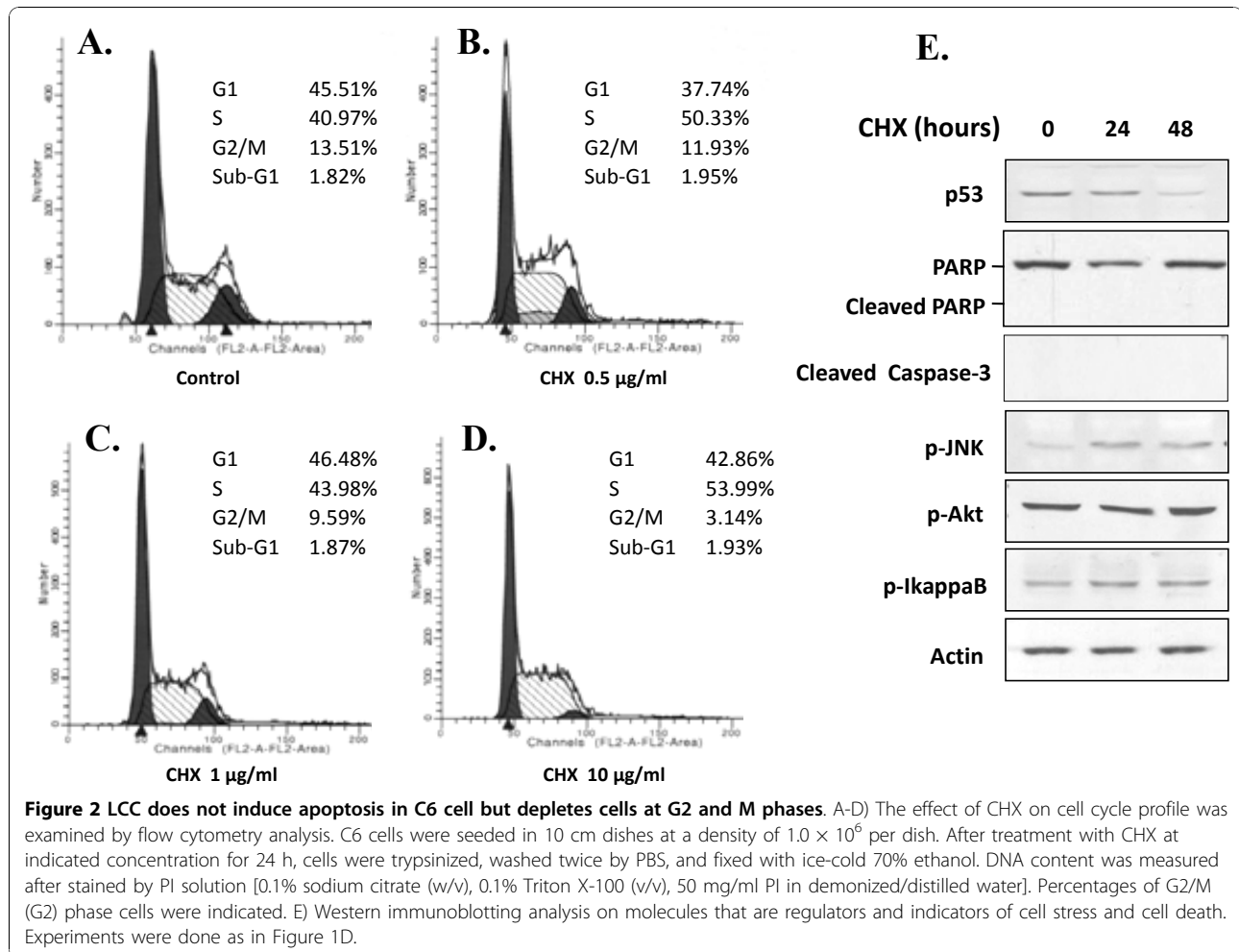


**Figure 1 LCC potently inhibits C6 glioma cell proliferation.** A) C6 cells were seeded in 6-well plates and treated without or with CHX (1  $\mu\text{g/ml}$ ) for 48 h. For observation of the integrity of nuclei, cells were stained with Hoechst 33258 and visualized under fluorescence microscope. Both phase contrast (left panels) and fluorescent (right panels) photos were shown (magnification:  $\times 200$ ). B) C6 cells were seeded in 6-well plates at a density of  $1.0 \times 10^5$  per well. CHX were added at concentration of 0 (◆) (as control), 0.5  $\mu\text{g/ml}$  (■), and 1  $\mu\text{g/ml}$  (▲), as indicated. Cells were collected in each of the next 3 days after trypsin treatment and cell numbers were counted using a haemocytometer. Data are presented as means  $\pm$  SEM ( $n = 3$ ). C) Colony Formation Efficiency Assay. 1,000 cells were plated in 35-mm tissue culture dishes. After overnight culturing, cells were exposed to different doses of CHX: (a) control; (b) CHX 0.1  $\mu\text{g/ml}$ ; (c) CHX 0.5  $\mu\text{g/ml}$ ; (d) CHX 1  $\mu\text{g/ml}$ . Photos were taken 6 days later after staining with 0.5% crystal violet. d) Western immunoblotting analysis on cell cycle regulators and indicators. C6 Cells were cultured in 6-well plates and were treated without or with CHX (0.5  $\mu\text{g/ml}$ ) for indicated time. Western blotting was performed as described in the Methods.

We further examined whether CHX-induced C6 cell differentiation can be reversed upon CHX withdrawal. We found that differentiated C6 cells reversed their differentiation phenotype and resumed proliferation within 1 d of CHX removal (data not shown). Taken together, the evidence indicated that LCC is able to block cell proliferation and induce reversible differentiation in C6 cell.

**CHX-promoted C6 cell differentiation is associated with an increase in intracellular cAMP and can be blocked by adenosine**

Since CHX has been shown to induce increases in intracellular cAMP in several types of cells including neuroblastoma and glioma [27], and cAMP or its analogs are effective inducers of differentiation in C6 glioma cell [23,24](Figure 3A), we hypothesized that LCC promote

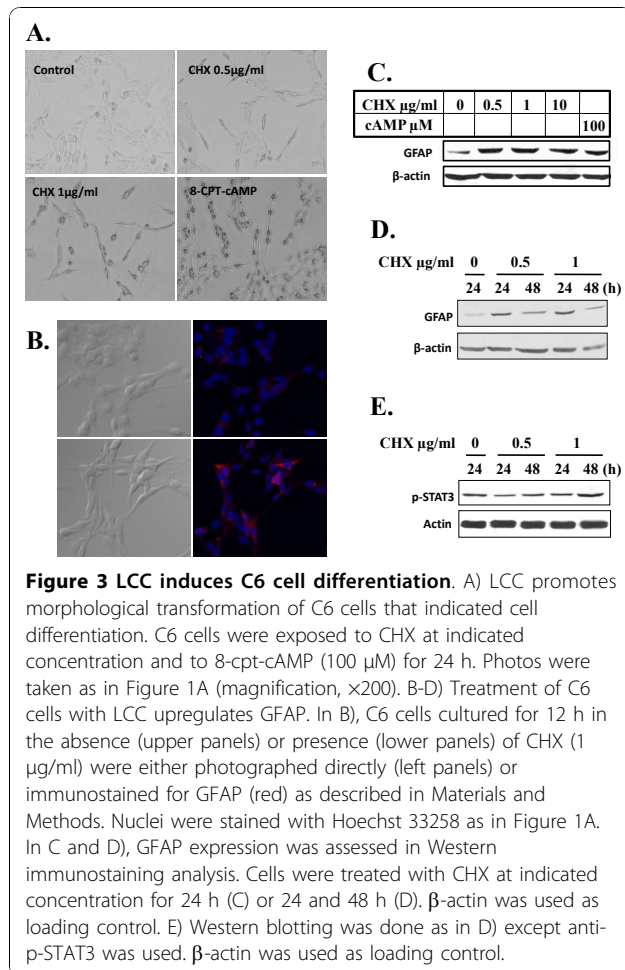


C6 cell differentiation by its ability to elevate intracellular cAMP in C6 cells. To test this hypothesis, we analyzed the intracellular cAMP in C6 cells untreated or treated with CHX (10 µg/ml). Our data indicated that C6 cell intracellular cAMP concentration increased 5-fold in response to CHX treatment (Figure 4A). To see whether CHX-induced intracellular cAMP increase is responsible for CHX-promoted differentiation transformation, we cotreated C6 cells with CHX and adenosine (Ado) (100 µM), a known inhibitor for cAMP synthesis [28], to reduce intracellular cAMP. As shown in Figure 4A, B, cotreatment of C6 cells with Ado blocked CHX-induced increase in intracellular cAMP in C6 cells (Figure 4A) and prevented CHX-promoted C6 cell phenotypic change of differentiation (Figure 4B). Interestingly, Ado alone did not cause any discernible effect on C6 cell; and it did not block CHX-promoted cessation of C6 cell proliferation (Figure 4B and data not shown). We additionally found that CHX-induced GFAP expression could be effectively blocked by Ado treatment (Figure 4C) and CHX-induced cell proliferation was not

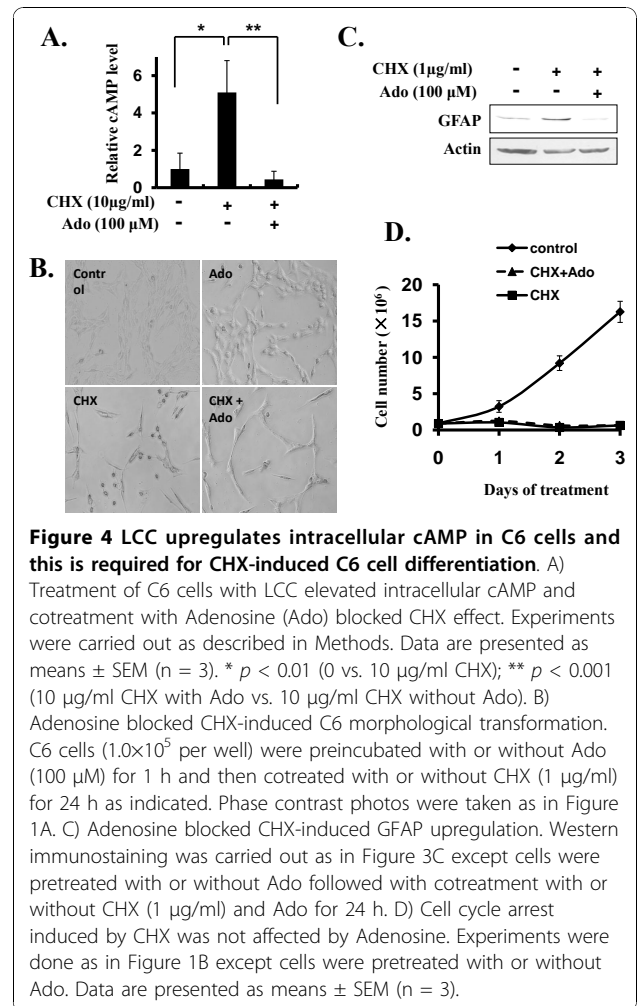
reversed with cotreatment of Ado (Figure 4D). Taken together, these data indicated that CHX-induced increase in intracellular cAMP in C6 cells is responsible for CHX-promoted C6 cell differentiation and that the underlining mechanism for CHX-induced cell cycle arrest and that for CHX-induced cell differentiation are independently regulated and separable.

### Discussion

CHX is a potent inhibitor of protein translation in mammalian cells and is widely used in probing the molecular mechanism of various biological processes. While high concentration of CHX (100 µM) was shown to induce apoptosis in hepatocytes both *in vitro* [20] and *in vivo* [19], and we found that high concentration of CHX (50-100 µg/ml, or 180-360 µM) kills C6 cells in a few hours (data not shown), LCC (0.1-10 µg/ml), with an IC<sub>50</sub> at below 0.5 µg/ml, arrested C6 cell proliferation (Figure 1 and 2) and promoted cell differentiation (Figure 3). Although it was not the focus of this study to identify the critical proteins that are involved in each of



the biological processes, the more-than-100-fold disparity in CHX concentrations required for induction of C6 cell death and for C6 cell cycle arrest and differentiation indicated a fundamental difference in the requirement for C6 cell survival, proliferation, and differentiation. We speculated that when C6 cell encounters LCC that shut off the production of a fraction of protein population in the cell, the first to be affected and thus most sensitive to CHX inhibition are the ones that are for cell proliferation. In this situation, C6 cell even increases the production of selective proteins such as GFAP (Figure 3) and *c-fos* and *c-jun* [29] that may be needed for differentiation. With increasing concentration of CHX, however, synthesis of proteins whose function involves cell survival, which is more fundamental and strategic to the cell in the long run, will be affected. Thus, the differential response of C6 cell to low and high concentration of CHX may indicate the presence of a priority hierarchy for protein production in C6 cell. Interestingly, we also found that CHX induces U251 human glioma differentiation (data not shown). However, the CHX concentration that is required for U251 to



differentiate (10 µg/ml) is about 10 times higher, possibly due to cellular difference in CHX sensitivity or in permeability.

We observed downregulation of a number of cell cycle regulators in C6 cell treated with LCC (Figure 1D), which is generally consistent with the outcome of cell cycle arrest. In addition, we found that p53 is downregulated in C6 cell treated with LCC (Figure 2E). This may explain the fact that apoptosis is not activated in these cells. With regard to differentiation, we saw strong increases in GFAP, a reliable indicator for glial differentiation, both in immunostaining and in Western blotting analyses (Figure 3B, C and 3D). In addition, the increase of GFAP was accompanied with an increase in p-STAT3 (Figure 3E), which is an obligatory transcription activator for GFAP [25,26]. These data convincingly demonstrated that partial blockade of protein synthesis drives forward the differentiation process in C6 glioma cell. Previously, it was shown that C6 cell differentiates when treated with cholera toxin [8] and that other neuroblastomas and glioma also differentiate after treated with



cyclic nucleotide phosphodiesterase inhibitors or adenylate cyclase activators [3-7]. These are in line with our findings that LCC upregulated intracellular concentration of cAMP by 5-fold (Figure 4A) and that the increase in intracellular cAMP is required for CHX-induced C6 cell differentiation (Figure 4B, C). These results argue that CHX employs the classical cAMP pathway to inhibit C6 cell proliferation and promote cell differentiation and that intracellular cAMP is a major intermediate that governs glioma cell proliferation and differentiation.

Current therapies used to treat cancer are highly toxic and often nonspecific. Differentiation therapy could be a potentially valuable strategy [2]. In this regard, *all-trans*-retinoic acid has been shown to be remarkable efficient to induce APL cell differentiation in clinical settings [10,11]. Since C6 cell is a well-established cell line for study on tumorigenesis, cancer cell proliferation and differentiation, our data showing that LCC induces C6 cell cycle arrest and cell differentiation will have important impact on the strategy of cancer differentiation therapy.

## Conclusions

Several important conclusions came out from this study. First, unlike high concentration of CHX that provokes C6 cell death in a few hours, LCC is not lethal to C6 cell even after several days of treatment; it causes cell cycle arrest at G1 and S phases during which large amount of proteins, and many of them being critical for cell cycle progression, are synthesized. Second, LCC promotes a profound morphological transformation that indicates C6 cell differentiation, which is further supported by the observation that the expression of glial differentiation marker GFAP is dramatically upregulated. Third, LCC dives forward the C6 cell differentiation process by mobilizing a classical pathway that involves upregulation of intracellular concentration of cAMP. Forth, LCC seems to illicit multiple biological effects in C6 cell, which include cell cycle arrest and cell differentiation, by mechanisms that are clearly distinct and separable. For instance, LCC-promoted C6 cell differentiation but not cell cycle arrest can be reversed by blockade of upregulation of intracellular cAMP; and Ado-induced downregulation of intracellular cAMP, *per se*, does not inhibit C6 cell proliferation. Since CHX is one of the most used reagents in molecular and cellular biology research and C6 cell is a well-established cell line to study cancer cell proliferation, apoptosis, and differentiation, our findings reported here is likely to have a wide implication to future studies in these related fields.

## Acknowledgements

This work was supported in part by grants from American Cancer Society (RSG-08-288-01), U.S. Department of Defense (BC085617), and Penn State University College of Medicine Institutional Award to D.X.L.

## Author details

<sup>1</sup>Department of Neural and Behavioral Sciences, Penn State University College of Medicine, 500 University Drive, Hershey, PA 17033, USA. <sup>2</sup>Department of Pharmacology, Penn State University College of Medicine, 500 University Drive, Hershey, PA 17033, USA. <sup>3</sup>Penn State Hershey Neuroscience Institute, Penn State University College of Medicine, 500 University Drive, Hershey, PA 17033, USA. <sup>4</sup>Penn State Cancer Institute, Penn State University College of Medicine, 500 University Drive, Hershey, PA 17033, USA. <sup>5</sup>Xinyuan Institute of Medicine and Biotechnology, School of Life Science, Zhejiang Sci-Tech University, Hangzhou, Zhejiang 310018, China. <sup>6</sup>Institute of Biochemistry and Cell Biology, Chinese Academy of Sciences, 320 Yue-Yang Road, Shanghai 200031, China.

## Authors' contributions

Conception and design of the project (XL, X-YL, DXL), acquisition, analysis and interpretation of data (XL, DXL), drafting the manuscript (DXL), revision for important intellectual content (J-MY, SSZ, DXL).

## Competing interests

The authors declare that they have no competing interests.

Received: 11 April 2010 Accepted: 15 December 2010

Published: 15 December 2010

## References

1. McLendon R, Enterline D, Tien R, Thorstad W, Bruner J: **Russell and Rubinstein's Pathology of Tumors of the Nervous System.** Edited by: Bigner D, McLendon R, Bruner J. Oxford University Press: New York; 1998: Chapter 9:307-571.
2. Leszczyniecka M, Roberts T, Dent P, Grant S, Fisher PB: **Differentiation therapy of human cancer: basic science and clinical applications.** *Pharmacology & Therapeutics* 2001, **90**:105-56.
3. Prasad KN, Kentroti S, Edwards-Prasad J, Vernadakis A, Imam M, Carvalho E, Kumar S: **Modification of the expression of cAMP-induced differentiated functions in neuroblastoma cells by beta-carotene and D-alpha-tocopheryl succinate.** *Journal of the American College of Nutrition* 1994, **13**:298-303.
4. Rudie Hovland A, Nahreini P, Andreatta CP, Edwards-Prasad J, Prasad KN: **Identifying genes involved in regulating differentiation of neuroblastoma cells.** *Journal of Neuroscience Research* 2001, **64**:302-10.
5. Nahreini P, Andreatta CP, Prasad KN, Toribara NW: **VP16CREB-induced differentiation of neuroblastoma.** *Indian Journal of Pediatrics* 2008, **75**:1009-13.
6. Prasad KN, Cole WC, Yan XD, Nahreini P, Kumar B, Hanson A, Prasad JE: **Defects in cAMP-pathway may initiate carcinogenesis in dividing nerve cells: a review.** *Apoptosis* 2003, **8**:579-86.
7. Yan XD, Hanson AJ, Nahreini P, Koustas WT, Andreatta C, Prasad KN: **Altered expression of genes regulating cell growth, proliferation, and apoptosis during cAMP- induced differentiation of neuroblastoma cells in culture.** *In Vitro Cellular & Developmental Biology Animal* 2002, **38**:529-37.
8. Li Y, Yin W, Wang X, Zhu W, Huang Y, Yan G: **Cholera toxin induces malignant glioma cell differentiation via the PKA/CREB pathway.** *Proceedings of the National Academy of Sciences of the United States of America* 2007, **104**:13438-43.
9. Sasaki T, Yamazaki K, Yamori T, Endo T: **Inhibition of proliferation and induction of differentiation of glioma cells with Datura stramonium agglutinin.** *British Journal of Cancer* 2002, **87**:918-23.
10. Huang ME, Ye YC, Chen SR, Chai JR, Lu JX, Zhao L, Gu LJ, Wang ZY: **Use of all-trans retinoic acid in the treatment of acute promyelocytic leukemia.** *Blood* 1988, **72**:567-72.
11. Wang ZY, Chen Z: **Differentiation and apoptosis induction therapy in acute promyelocytic leukaemia.** *Lancet Oncology* 2000, **1**:101-6.
12. Obrigg TG, Culp WJ, McKeehan WL, Hardesty B: **The mechanism by which cycloheximide and related glutarimide antibiotics inhibit peptide synthesis on reticulocyte ribosomes.** *Journal of Biological Chemistry* 1971, **246**:174-81.
13. Zhang L, Wali A, Ramana CV, Rishi AK: **Cell growth inhibition by okadaic acid involves gut-enriched Kruppel-like factor mediated enhanced expression of c-Myc.** *Cancer Research* 2007, **67**:10198-206.
14. Moon J, Yu SJ, Kim HS, Sohn J: **Induction of G(1) cell cycle arrest and p27 (KIP1) increase by panaxydol isolated from Panax ginseng.** *Biochemical Pharmacology* 2000, **59**:1109-16.

15. Shih SR, Weng KF, Stollar V, Li ML: **Viral protein synthesis is required for Enterovirus 71 to induce apoptosis in human glioblastoma cells.** *Journal of Neurovirology* 2008, **14**:53-61.
16. Eriksson D, Lofroth PO, Johansson L, Riklund K, Stigbrand T: **Apoptotic signaling in HeLa Hep2 cells following 5 Gy of cobalt-60 gamma radiation.** *Anticancer Research* 2009, **29**:4361-6.
17. Liu DX, Greene LA: **Regulation of neuronal survival and death by E2F-dependent gene repression and derepression.** *Neuron* 2001, **32**:425-38.
18. Alessenko AV, Boikov P, Filippova GN, Khrenov AV, Loginov AS, Makarieva ED: **Mechanisms of cycloheximide-induced apoptosis in liver cells.** *FEBS Letters* 1997, **416**:113-6.
19. Ito K, Kiyosawa N, Kumagai K, Manabe S, Matsunuma N, Yamoto T: **Molecular mechanism investigation of cycloheximide-induced hepatocyte apoptosis in rat livers by morphological and microarray analysis.** *Toxicology* 2006, **219**:175-86.
20. Blom WM, de Bont HJ, Meijerman I, Mulder GJ, Nagelkerke JF: **Prevention of cycloheximide-induced apoptosis in hepatocytes by adenosine and by caspase inhibitors.** *Biochemical Pharmacology* 1999, **58**:1891-8.
21. Riccardi C, Nicoletti I: **Analysis of apoptosis by propidium iodide staining and flow cytometry.** *Nature Protocols* 2006, **1**:1458-61.
22. Li G, Li W, Angelastro JM, Greene LA, Liu DX: **Identification of a novel DNA binding site and a transcriptional target for activating transcription factor 5 in c6 glioma and mcf-7 breast cancer cells.** *Molecular Cancer Research: MCR* 2009, **7**:933-43.
23. Messens J, Slegers H: **Synthesis of glial fibrillary acidic protein in rat C6 glioma in chemically defined medium: cAMP-dependent transcriptional and translational regulation.** *Journal of Neurochemistry* 1992, **58**:2071-80.
24. Takanaga H, Yoshitake T, Hara S, Yamasaki C, Kunimoto M: **cAMP-induced astrocytic differentiation of C6 glioma cells is mediated by autocrine interleukin-6.** *Journal of Biological Chemistry* 2004, **279**:15441-7.
25. Yanagisawa M, Nakashima K, Taga T: **STAT3-mediated astrocyte differentiation from mouse fetal neuroepithelial cells by mouse oncostatin M.** *Neuroscience Letters* 1999, **269**:169-72.
26. Ochiai W, Yanagisawa M, Takizawa T, Nakashima K, Taga T: **Astrocyte differentiation of fetal neuroepithelial cells involving cardiotrophin-1-induced activation of STAT3.** *Cytokine* 2001, **14**:264-71.
27. Moylan RD, Brooker G: **Cycloheximide potentiation of prostaglandin E1- and cholera toxin-stimulated cAMP accumulation in NG108-CC15 neuroblastoma-glioma hybrid cells.** *Journal of Biological Chemistry* 1981, **256**:6573-6.
28. Borasio PG, Pavan B, Fabbri E, Ginanni-Corradini F, Arcelli D, Poli A: **Adenosine analogs inhibit acetylcholine release and cAMP synthesis in the guinea-pig superior cervical ganglion.** *Neuroscience Letters* 1995, **184**:97-100.
29. Kim YH, Choi MR, Song DK, Huh SO, Jang CG, Suh HW: **Regulation of c-fos gene expression by lipopolysaccharide and cycloheximide in C6 rat glioma cells.** *Brain Research* 2000, **872**:227-30.

#### Pre-publication history

The pre-publication history for this paper can be accessed here:  
<http://www.biomedcentral.com/1471-2407/10/684/prepub>

doi:10.1186/1471-2407-10-684

**Cite this article as:** Liu *et al.*: Induction of cell cycle arrest at G1 and S phases and cAMP-dependent differentiation in C6 glioma by low concentration of cycloheximide. *BMC Cancer* 2010 **10**:684.

**Submit your next manuscript to BioMed Central  
and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at  
[www.biomedcentral.com/submit](http://www.biomedcentral.com/submit)

