

Published in final edited form as:

*Clin Exp Pharmacol Physiol*. 2010 July ; 37(7): 719–725. doi:10.1111/j.1440-1681.2010.05380.x.

## Anti-oxidant properties of high-density lipoprotein and atherosclerosis

**Eugene A Podrez**

Department of Molecular Cardiology, Lerner Research Institute, Cleveland Clinic, Cleveland, OH, USA

### SUMMARY

1. High-density lipoprotein (HDL) is one of the major carriers of cholesterol in the blood. It attracts particular attention because, in contrast with other lipoproteins, many physiological functions of HDL influence the cardiovascular system in favourable ways unless HDL is modified pathologically.
2. The best known function of HDL is the capacity to promote cellular cholesterol efflux from peripheral cells and deliver cholesterol to the liver for excretion, thereby playing a key role in reverse cholesterol transport. The functions of HDL that have recently attracted attention include anti-inflammatory and anti-oxidant activities. High anti-oxidant and anti-inflammatory activities of HDL are associated with protection from cardiovascular disease.
3. Atheroprotective activities, as well as a functional deficiency of HDL, ultimately depend on the protein and lipid composition of HDL. Conversely, these activities are compromised in many pathological states associated with inflammation.
4. The focus of the present review is on the anti-oxidant and anti-inflammatory functions of HDL and its individual components in relation to protection from atherosclerosis.

### Keywords

anti-oxidant; atherosclerosis; high-density lipoprotein; inflammation; oxidative stress

### INTRODUCTION

High-density lipoprotein (HDL) is a plasma lipoprotein heterogeneous in origin, size, composition and function. It is one of the major carriers of cholesterol in the blood. In contrast with other lipoproteins, many physiological functions of HDL influence the cardiovascular system in favourable ways unless HDL is modified pathologically. The protective role of HDL in cardiovascular disease has been documented in multiple clinical and animal studies. High-density lipoprotein is best known as a key player in reverse cholesterol transport. In this process, HDL and its major apolipoprotein, apolipoprotein (apo) A-I, promote cholesterol efflux from peripheral tissues; HDL then carries cholesterol in the circulation and delivers it to the liver so that it can be either reutilized for assembly of very low-density lipoprotein (VLDL) or excreted as free cholesterol or bile acids. This function of HDL is central to its atheroprotective role. The other atheroprotective functions

of HDL that have more recently attracted attention include its anti-inflammatory, anti-oxidant and vasodilatory properties.<sup>1–3</sup> In addition, HDL has been shown to possess anti-apoptotic, antithrombotic and anti-infectious functions among other actions.<sup>1</sup> Particular attention has focused on the small dense HDL, which is the most potent promoter of cholesterol efflux and has the highest anti-oxidative and anti-inflammatory activities. Some of these multiple functions are clearly overlapping. Moreover, recent progress in the identification of minor components in HDL indicates that not all functions of HDL are currently known.

## COMPOSITION OF HDL

### Proteins

Apolipoprotein A-I is the most abundant protein in HDL, accounting for approximately 70% of the HDL protein mass, with apoA-II being the second most abundant protein, accounting for 15–20%.<sup>1</sup> The remaining protein mass is made up of minor amphipathic proteins, such as apoC, apoE, apoD, apoM and apoA-IV, enzymes such as paraoxonase (PON) 1 and platelet-activating factor acetylhydrolase (PAF-AH), glutathione peroxidase (GPx) 1, and lipid transfer proteins such as lecithin:cholesterol acyl transferase (LCAT) and cholesteryl ester transfer protein (CETP).<sup>1</sup> The total number of minor proteins associated with HDL reaches as many as 75, as demonstrated in recent proteomic studies.<sup>4–7</sup> The molar concentration of most of these proteins in the plasma is lower than that of HDL, suggesting that specific proteins may be present only in particular HDL subclasses.

### Lipids

In addition to free and esterified cholesterol, a number of other lipids are detected in HDL, including various phospholipids, such as phosphatidyl choline, phosphatidyl ethanolamine, lysophosphatidyl choline, plasmalogen, free fatty acids, mono-, di- and triacylglycerols, and different sphingolipids, such as ceramide, sphingomyelin species, sphingosine-1-phosphate (S1P), lysosulphatide and sphingosylphosphorylcholine.<sup>8</sup>

## VARIABILITY IN HDL

Multiple, functionally distinct populations of HDL exist that differ in size, shape, protein and lipid composition. Surprisingly, although the heterogeneity of HDL particles is well known, in many pharmacological studies HDL is still regarded as a single entity whose plasma levels are reflected by HDL-cholesterol. Recent studies have addressed the hypothesis that the multiple biological functions of HDL are mediated by distinct particle subspecies defined by specific cluster(s) of bound proteins. It has been shown that proteins identified in HDL can be separated into several clusters according to their metabolic function, such as lipid transport and metabolism, markers of inflammation, protease inhibition, immune system and complement factors, coagulation and hormone binding.<sup>6,7</sup> In a more recent study, plasma HDL was subfractionated into five physicochemically defined particle subpopulations by isopycnic density gradient ultracentrifugation and the protein composition of the subfractions was assessed by mass spectrometry.<sup>9</sup> That study revealed that HDL-associated proteins were distributed in distinct patterns across the HDL particle subpopulations, including the dense HDL3 particle, known to be the most potent anti-oxidant HDL subfraction.<sup>10</sup> Proteins that were preferentially associated with the dense HDL3 subpopulation included apoL-I, apoF, PON1/2, phospholipid transfer protein (PLTP), apoJ, PON3,  $\alpha_1$ -antitrypsin (A1AT), apoA-IV, apoM, apoD, serum amyloid  $\alpha$  (SAA), albumin, fibrinogen, haptoglobin-related protein (Hrp), platelet basic protein (PBP) and transthyretin. The correlation of the relative abundance of each protein with anti-oxidative functionality of HDL revealed that apoL-I, PON1 and PON3 showed the most significant

correlations, whereas PLTP, apoM, apoJ, SAA4 and apoD showed correlations that were less significant.<sup>9</sup>

## ANTI-OXIDANT FUNCTIONS OF HDL

The anti-oxidant properties of HDL *in vivo* can be separated into direct and indirect actions. High-density lipoprotein can directly inhibit oxidation of low-density lipoprotein (LDL; or other targets containing phospholipids). One example of a direct effect is the transfer of oxidation products from LDL to HDL so that HDL serves as a 'sink' for oxidized lipids.<sup>11</sup> In addition, inhibition of oxidative events and oxidative stress *in vivo* may be achieved indirectly via other functions of HDL, such as induction of cholesterol efflux and, in general, via 'anti-inflammatory' functions of HDL.<sup>12</sup> It has been shown that circulating HDL accumulates oxidized phospholipids, such as hydroperoxides, lysophosphatidylcholine (lyso-PC) and F2-isoprostanes.<sup>11,13</sup> The transfer of oxidized phospholipids serves several purposes. First, the transfer of lipid hydroperoxide from LDL, so-called 'seeding molecules', prevents the initiation of a free radical chain reaction of oxidation.<sup>14</sup> Second, some of the more advanced products of phospholipid oxidation serve as ligands for scavenger receptor type B (SR-B) and promote uptake of modified lipoproteins by macrophages as well as prothrombotic effects mediated by platelet scavenger receptor CD36.<sup>15,16</sup> Removal of these phospholipids will prevent these pathological activities. Interestingly, when oxidized phospholipids are present in HDL (in oxidized HDL), the receptors that recognize the particle may be different and the effects may be opposite to those of oxidized LDL.<sup>17</sup> Third, some of the products of oxidation are chemically reactive and can modify lysine residues in apoB, rendering the particle a ligand for SR-A, scavenger receptor expressed by endothelial cell (SREC) and some of the products of lipid oxidation (e.g. 4-hydroxynonenal, malondialdehyde and levuglandin).<sup>18,19</sup>

The transfer of oxidized phospholipids to HDL may be followed by subsequent degradation of oxidation products by HDL enzymes or by delivery to the liver for degradation. Hydrolysis of oxidized phospholipids in HDL leads to the destruction of pathological activity. In addition, in this process, chemically reactive fragments of esterified fatty acids are released from lipoproteins so that the lipoprotein is not modified and is not converted into a pro-atherosclerotic particle.<sup>20</sup>

Normal functional HDL has high levels of anti-oxidants and active anti-oxidant proteins and enzymes with high anti-oxidant potential and has anti-inflammatory activity. However, when anti-oxidant and anti-inflammatory functions of HDL are overwhelmed by pathological processes, such as inflammation, HDL is converted into a 'dysfunctional' pro-inflammatory particle.<sup>2,12</sup> This dysfunctional HDL is characterized by decreased levels and activities of anti-inflammatory and anti-oxidant factors, such as apoA-I and PON1. The dysfunctional HDL contains oxidized phospholipids and lysophospholipids, as well as pro-inflammatory proteins, such as serum amyloid A and ceruloplasmin. Functionally, it cannot promote cholesterol efflux effectively or prevent LDL oxidation. Many pathological processes associated with systemic inflammation are characterized by the presence of dysfunctional or pro-inflammatory HDL, including chronic heart disease, metabolic syndrome, chronic kidney disease, infections and some rheumatological diseases.<sup>12</sup> In addition to compositional changes to HDL that occur under pathological conditions, the protein or lipid components of HDL can be modified chemically.<sup>21</sup> As a result, HDL may lose its normal function and acquire pathological functions. High-density lipoprotein is susceptible to oxidation/modification *in vitro* by a variety of oxidants, such as metal ions, peroxy and hydroxyl radicals, aldehydes, various myeloperoxidase (MPO)-generated oxidants, lipoxygenase, phospholipase A<sub>2</sub>, elastase, non-enzymatic glycation and homocysteinylation.<sup>21</sup> Recent studies have shown that MPO can selectively target apoA-I as

a result of direct binding and that it catalyses apoA-I nitration and halogenation within human atheroma.<sup>22,23</sup> *In vitro* studies have demonstrated that MPO-catalysed oxidative modification of HDL or apoA-I leads to the loss of ABCA1-dependent cholesterol efflux function of the lipoprotein and converts HDL into a pro-inflammatory particle.<sup>22,23</sup>

## ANTI-OXIDANT AND ANTI-INFLAMMATORY PROPERTIES OF HDL COMPONENTS

A number of *in vitro* and *in vivo* assays have been developed that allow quantification of the anti- and pro-inflammatory properties of HDL. One cell-free assay uses the fluorescent probe 2',7'-dichlor-fluorescein, which is sensitive to oxidized phospholipids, to measure the ability of HDL to prevent the formation of oxidized phospholipids in LDL.<sup>24</sup> This assay can also be used to evaluate the ability of HDL to degrade oxidized phospholipids that are already formed.<sup>24</sup> Cell-based assays can evaluate the capacity of HDL to inhibit monocyte chemotaxis in response to oxidized LDL<sup>25</sup> or to prevent upregulation of cell adhesion molecules on endothelial cells.<sup>26</sup> Finally, the capacity of HDL to promote cholesterol efflux from cultured cells *in vitro* and reverse cholesterol transport *in vivo* can also be measured.<sup>27</sup>

### Apolipoprotein AI

Evidence is accumulating that apoA-I is the major anti-atherogenic and anti-oxidant factor in HDL. Apolipoprotein AI is a major player in reverse cholesterol transport. However, in addition, apoA-I can remove oxidized phospholipids from oxidized LDL as well as from cells.<sup>14,25</sup> Human HDL can also directly reduce cholesteryl ester hydroperoxides and phosphatidylcholine hydroperoxides via Met residues 112 and 148 of apoA-I.<sup>28</sup> The finding that recombinant HDL containing only apoA-I and 1-palmitoyl-2-oleoyl-phosphatidylcholine (POPC) and no enzymes was as effective in preventing LDL oxidation as complete HDL supports a key anti-oxidant role for apoA-I.<sup>29</sup> That interesting study did not support significant roles for PAF-AH, PON1 or LCAT in the prevention of LDL oxidation by HDL.<sup>29</sup> Multiple *in vivo* studies have demonstrated that apoA-I is a potent anti-oxidative, anti-inflammatory and anti-atherosclerotic agent.<sup>30–35</sup> Recent clinical studies have demonstrated that although very high HDL-cholesterol and large-size HDL particles are associated with a twofold increase in cardiovascular risk,<sup>36</sup> a high concentration of apoA-I is an independent negative predictor of cardiovascular risk.<sup>36,37</sup>

### Apolipoprotein A-II

It has been shown that apoA-II-enriched HDL from mice transgenic for human apoA-II protected VLDL from oxidation more efficiently than control HDL.<sup>38</sup> Moreover, basal lipoprotein oxidation in transgenic mice was not reduced, suggesting that human apoA-II-rich HDL may maintain adequate anti-oxidant potential. High-density lipoprotein enriched in human apoA-II supports effective reverse cholesterol transport from macrophages.<sup>39</sup> However, apoA-II was found to displace PON1 and PAF-AH from HDL.<sup>38</sup> This was suggested as a mechanism underlying the increased atherosclerosis seen in dyslipidaemic mice overexpressing either human or murine apoA-II.<sup>40,41</sup> Conversely, a recent prospective study demonstrated that plasma apoA-II concentrations are an independent factor negatively correlated with the risk of coronary artery disease (CAD).<sup>37</sup>

### Apolipoprotein E

The anti-atherosclerotic activity of apoE related to receptor-mediated uptake of lipoprotein is well known. However, both murine and human apoE can inhibit the development of atherosclerosis in mice without any significant effect on hypercholesterolaemia.<sup>42–44</sup> Moreover, apoE has been shown to have allele-specific anti-oxidant activity.<sup>45</sup>

Apolipoprotein E2 stimulates endothelial nitric oxide (NO) release and has anti-inflammatory activities.<sup>46</sup> In contrast, apoE4 is pro-inflammatory.<sup>47</sup>

### **Apolipoprotein J**

Apolipoprotein J is associated with a subset of small HDL containing PON1. It has been reported that HDL-associated apoJ can inhibit LDL oxidation by artery wall cells.<sup>48</sup> In addition, at low physiological levels apoJ has been reported to be cytoprotective.<sup>49</sup>

### **Apolipoprotein A-IV**

Apolipoprotein A-IV has multiple activities related to lipid and lipoprotein metabolism. It can participate in reverse cholesterol transport by promoting cholesterol efflux via ABCA1<sup>50</sup> and by activating LCAT.<sup>51</sup> In addition, it has been demonstrated that ApoA-IV has anti-oxidant, anti-inflammatory and anti-atherosclerotic actions *in vivo*.<sup>52–54</sup>

### **Paraoxonases 1 and 3**

Paraoxonase 1 is an HDL-associated enzyme with anti-oxidant properties. There are data suggesting that the direct anti-oxidant effect of HDL on LDL oxidation, measured as a reduction in lipid peroxides, is likely mediated by PON1.<sup>55–57</sup> However, it should be noted that the role of paraoxonases in the direct protection of LDL from oxidative stress is not firmly established at present.<sup>1</sup> Interestingly, it has been shown that PON1 enhances cholesterol efflux from macrophages by promoting HDL binding mediated by ABCA1.<sup>58</sup> This effect of PON1 can indirectly reduce pro-inflammatory signalling in cells *in vivo* and contribute to the anti-atherosclerotic effects of PON1.<sup>59,60</sup> Multiple *in vivo* studies support the hypothesis that PON1 protects against atherosclerosis. For example, in mice deficient in PON1 and on a high-fat diet, the average lesion area in the proximal aorta was almost twice that in control mice;<sup>59</sup> however, overexpression of human PON1 confers significant protection against the development of atherosclerosis in mice.<sup>60</sup> Moreover, signs of oxidative stress, vascular inflammation and thrombotic tendencies have been observed in PON1-deficient mice.<sup>61</sup> In human studies, higher PON1 activity is associated with a lower incidence of major cardiovascular events.<sup>62</sup> However, it remains uncertain whether this relationship is causal or correlational. A recent prospective study failed to demonstrate a causal relationship between PON1 activity and the risk of future CAD.<sup>63</sup> Pathological conditions associated with oxidative stress, such as chronic renal failure, rheumatoid arthritis and Alzheimer's disease, are frequently associated with reduced activity of PON1.<sup>62</sup> Furthermore, a pro-atherosclerotic high-fat diet leads to reduced PON1 activity.<sup>62</sup> In general, it has been shown that multiple factors (genetic, pathological, physiological, pharmacological and lifestyle) can change the PON1 activity. Taking into account the growing evidence of the protective role of PON1 in CAD, this enzyme could be an important target for pharmacological regulation.

Paraoxonase 3 is also associated with HDL and has been shown to prevent the oxidation of LDL *in vitro*.<sup>64,65</sup> Furthermore, human PON3 transgenic mice have been shown to be protected from the development of atherosclerosis, without any significant changes in plasma lipoprotein cholesterol, triglyceride or glucose levels.<sup>66</sup> However, additional studies are needed to determine the role of *PON3* gene polymorphisms in cardiovascular disease.

### **Platelet-activating factor acetylhydrolase**

Platelet-activating factor acetylhydrolase is another enzyme that is found in HDL that can hydrolyse oxidized phospholipids. There are data suggesting that PAF-AH, rather than PON1, is the major hydrolase in HDL responsible for the hydrolysis of oxidized phospholipids.<sup>67,68</sup> Lipoproteins isolated from mice expressing human PAF-AH are more

resistant to oxidative stress.<sup>69</sup> Moreover, murine HDL with human PAF-AH has been shown to inhibit foam cell formation and facilitate cholesterol efflux in macrophages.<sup>69</sup> Adenoviral gene transfer of human PAF-AH in apoE<sup>-/-</sup> mice increased circulating PAF-AH activity by 50% and was associated *in vivo* with reduced monocyte adhesion to the endothelium.<sup>70</sup> The b-migrating (b) VLDL isolated from these mice had a low capacity to induce monocyte adhesion *ex vivo*. In experimental atherosclerosis, gene transfer of *PAF-AH* inhibited lesion formation in apoE-deficient mice.<sup>71</sup> In arteries of non-hyperlipidaemic rabbits, local expression of PAF-AH reduced the accumulation of oxidatively modified LDL without changing plasma levels of PAF-AH and reduced the expression of endothelial cell adhesion molecules.<sup>72</sup> In humans, PAF-AH deficiency is associated with increases in cardiovascular disease; for example, missense mutation of the gene of the PAF-AH gene is an independent risk factor for coronary artery disease in Japanese men.<sup>73</sup> At the same time, circulating levels of PAF-AH serve as an independent marker of the risk of CAD.<sup>74</sup> Whether PAF-AH plays a causal role in cardiovascular disease or simply is increased in response to the chronic inflammatory environment needs to be established. It is believed that PAF-AH in the necrotic core of coronary lesions may contribute to inflammation and plaque vulnerability. For example, prolonged inhibition of PAF-AH with the oral inhibitor darapladib prevented necrotic core expansion in patients with CAD.<sup>75</sup> In conclusion, even though the data from animal studies suggest that PAF-AH is anti-atherosclerotic, the role of PAF-AH in human physiology is far from being completely understood.<sup>73</sup>

### Glutathione peroxidase 1

Glutathione peroxidase 1, another enzyme detected in HDL, can reduce lipid hydroperoxides to corresponding hydroxides and thereby detoxify them.<sup>76,77</sup> Multiple clinical studies suggest an atheroprotective role for GPx-1.<sup>78–82</sup> Furthermore, two studies in apoE<sup>-/-</sup> mice have demonstrated that GPx-1 deficiency leads to increased atherosclerosis when mice are further challenged by a Western-type diet or made diabetic.<sup>83,84</sup> However, it should be noted that no protection from atherosclerosis has been found in GPx-1<sup>-/-</sup> mice fed a high-fat diet.<sup>85</sup> It seems that the role of GPx-1 in the development of atherosclerosis is particularly prominent under conditions of significant oxidative stress.

### Lecithin: cholesterol acyltransferase

Lecithin:cholesterol acyltransferase has been shown to directly hydrolyse oxidized polar phospholipids.<sup>86</sup>

### Sphingosine-1-phosphate

High-density lipoprotein is the most prominent plasma carrier of S1P. Moreover, many biological effects of HDL, such as the induction of endothelial NO production, vasodilation, survival and cardioprotection, are partially mediated by S1P. Approximately half the effects of HDL on the induction of endothelial NO synthase occur via S1P receptors.<sup>87</sup> In general, multiple effects of HDL on endothelial cells, such as migration, proliferation, endothelial integrity and angiogenesis, are mediated in part by S1P in HDL.<sup>8</sup> Furthermore, S1P in HDL inhibits pro-inflammatory responses, such as the generation of reactive oxygen species, activation of NAD(P)H oxidase and the production of monocyte chemoattractant protein-1, in vascular smooth muscle cells and the aorta.<sup>88</sup> In endothelial cells, S1P in HDL inhibits tumour necrosis factor- $\alpha$ -induced expression of cell adhesion molecules and promotes prostacyclin production.<sup>89</sup> It should be noted that in its free form (i.e. not associated with HDL), S1P is also involved in pro-inflammatory processes.<sup>88</sup>

In summary, high anti-oxidant and anti-inflammatory activities of HDL are associated with protection from cardiovascular disease. These activities depend on the protein and lipid composition of HDL and are the highest in small dense HDL. These activities are

compromised in many pathological states associated with inflammation. The functional deficiency of HDL is intimately associated with changes in HDL composition. The search is continuing to find the best approaches for the prevention of the loss or the restoration of the anti-oxidant and anti-inflammatory potential of HDL.

## Acknowledgments

The author's work described herein was supported by National Institutes of Health grants (HL077213, RO1HL077213-05S1, HL053315 and P01HL073311).

## References

1. Kontush A, Chapman MJ. Functionally defective high-density lipoprotein. A new therapeutic target at the crossroads of dyslipidemia, inflammation, and atherosclerosis. *Pharmacol Rev* 2006;58:342–74. [PubMed: 16968945]
2. Navab M, Reddy ST, Van Lenten BJ, Anantharamaiah GM, Fogelman AM. The role of dysfunctional HDL in atherosclerosis. *J Lipid Res* 2009;50(Suppl):S145–9. [PubMed: 18955731]
3. Sviridov D, Mukhamedova N, Remaley AT, Chin-Dusting J, Nestel P. Antiatherogenic functionality of high density lipoprotein: How much versus how good. *J Atheroscler Thromb* 2008;15:52–62. [PubMed: 18385533]
4. Heller M, Stalder D, Schlappritzi E, Hayn G, Matter U, Haeberli A. Mass spectrometry-based analytical tools for the molecular protein characterization of human plasma lipoproteins. *Proteomics* 2005;5:2619–30. [PubMed: 15892164]
5. Karlsson H, Leanderson P, Tagesson C, Lindahl M. Lipoproteomics II. Mapping of proteins in high-density lipoprotein using two-dimensional gel electrophoresis and mass spectrometry. *Proteomics* 2005;5:1431–45. [PubMed: 15761960]
6. Rezaee F, Casetta B, Levels JH, Speijer D, Meijers JC. Proteomic analysis of high-density lipoprotein. *Proteomics* 2006;6:721–30. [PubMed: 16419016]
7. Vaisar T, Pennathur S, Green PS, et al. Shotgun proteomics implicates protease inhibition and complement activation in the antiinflammatory properties of HDL. *J Clin Invest* 2007;117:746–56. [PubMed: 17332893]
8. Sattler K, Levkau B. Sphingosine-1-phosphate as a mediator of high-density lipoprotein effects in cardiovascular protection. *Cardiovasc Res* 2009;82:201–11. [PubMed: 19233866]
9. Davidson WS, Silva RA, Chantepie S, Lagor WR, Chapman MJ, Kontush A. Proteomic analysis of defined HDL subpopulations reveals particle-specific protein clusters: Relevance to antioxidative function. *Arterioscler Thromb Vasc Biol* 2009;29:870–6. [PubMed: 19325143]
10. Kontush A, Chantepie S, Chapman MJ. Small, dense HDL particles exert potent protection of atherogenic LDL against oxidative stress. *Arterioscler Thromb Vasc Biol* 2003;23:1881–8. [PubMed: 12920049]
11. Bowry VW, Stanley KK, Stocker R. High density lipoprotein is the major carrier of lipid hydroperoxides in human blood plasma from fasting donors. *Proc Natl Acad Sci USA* 1992;89:10316–20. [PubMed: 1332045]
12. Ansell BJ, Fonarow GC, Fogelman AM. The paradox of dysfunctional high-density lipoprotein. *Curr Opin Lipidol* 2007;18:427–34. [PubMed: 17620860]
13. Proudfoot JM, Barden AE, Loke WM, Croft KD, Puddey IB, Mori TA. HDL is the major lipoprotein carrier of plasma F2-isoprostanes. *J Lipid Res* 2009;50:716–22. [PubMed: 19050315]
14. Navab M, Hama SY, Cooke CJ, et al. Normal high density lipoprotein inhibits three steps in the formation of mildly oxidized low density lipoprotein: Step 1. *J Lipid Res* 2000;41:1481–94. [PubMed: 10974056]
15. Podrez EA, Poliakov E, Shen Z, et al. Identification of a novel family of oxidized phospholipids that serve as ligands for the macrophage scavenger receptor CD36. *J Biol Chem* 2002;277:38503–16. [PubMed: 12105195]
16. Valiyaveetil M, Podrez EA. Platelet hyperreactivity, scavenger receptors and atherothrombosis. *J Thromb Haemost* 2009;7(Suppl 1):218–21. [PubMed: 19630804]

17. Valiyaveettil M, Kar N, Ashraf MZ, Byzova TV, Febbraio M, Podrez EA. Oxidized high-density lipoprotein inhibits platelet activation and aggregation via scavenger receptor BI. *Blood* 2008;111:1962–71. [PubMed: 17993610]
18. Holvoet P. Oxidative modification of LDL in atherothrombosis. *Acta Cardiol* 1998;53:253–60. [PubMed: 9922802]
19. Hoppe G, O'Neil J, Sayre LM, Hoff HF. Non-conventional modification of LDL: Chemical models for macrophage recognition of oxidized LDL. *Biochim Biophys Acta* 1997;1362:103–8. [PubMed: 9540840]
20. Heinecke JW. Free radical modification of low-density lipoprotein: Mechanisms and biological consequences. *Free Radic Biol Med* 1987;3:65–73. [PubMed: 3040538]
21. Ferretti G, Bacchetti T, Nègre-Salvayre A, Salvayre R, Dousset N, Curatola G. Structural modifications of HDL and functional consequences. *Atherosclerosis* 2006;184:1–7. [PubMed: 16157342]
22. Bergt C, Pennathur S, Fu X, et al. The myeloperoxidase product hypo-chlorous acid oxidizes HDL in the human artery wall and impairs ABCA1-dependent cholesterol transport. *Proc Natl Acad Sci USA* 2004;101:13032–7. [PubMed: 15326314]
23. Zheng L, Nukuna B, Brennan ML, et al. Apolipoprotein A-I is a selective target for myeloperoxidase-catalyzed oxidation and functional impairment in subjects with cardiovascular disease. *J Clin Invest* 2004;114:529–41. [PubMed: 15314690]
24. Navab M, Hama SY, Hough GP, Subbanagounder G, Reddy ST, Fogelman AM. A cell-free assay for detecting HDL that is dysfunctional in preventing the formation of or inactivating oxidized phospholipids. *J Lipid Res* 2001;42:1308–17. [PubMed: 11483633]
25. Navab M, Hama SY, Anantharamaiah GM, et al. Normal high density lipoprotein inhibits three steps in the formation of mildly oxidized low density lipoprotein: Steps 2 and 3. *J Lipid Res* 2000;41:1495–508. [PubMed: 10974057]
26. Nicholls SJ, Lundman P, Harmer JA, et al. Consumption of saturated fat impairs the anti-inflammatory properties of high-density lipoproteins and endothelial function. *J Am Coll Cardiol* 2006;48:715–20. [PubMed: 16904539]
27. Wang X, Collins HL, Ranalletta M, et al. Macrophage ABCA1 and ABCG1, but not SR-BI, promote macrophage reverse cholesterol transport *in vivo*. *J Clin Invest* 2007;117:2216–24. [PubMed: 17657311]
28. Garner B, Waldeck AR, Witting PK, Rye KA, Stocker R. Oxidation of high density lipoproteins. II. Evidence for direct reduction of lipid hydroperoxides by methionine residues of apolipoproteins AI and AII. *J Biol Chem* 1998;273:6088–95. [PubMed: 9497326]
29. Zerrad-Saadi A, Therond P, Chantepie S, et al. HDL3-mediated inactivation of LDL-associated phospholipid hydroperoxides is determined by the redox status of apolipoprotein A-I and HDL particle surface lipid rigidity: Relevance to inflammation and atherogenesis. *Arterioscler Thromb Vasc Biol* 2009;29:2169–75. [PubMed: 19762782]
30. Plump AS, Scott CJ, Breslow JL. Human apolipoprotein A-I gene expression increases high density lipoprotein and suppresses atherosclerosis in the apolipoprotein E-deficient mouse. *Proc Natl Acad Sci USA* 1994;91:9607–11. [PubMed: 7937814]
31. Pászty C, Maeda N, Verstuyft J, Rubin EM. Apolipoprotein AI trans-gene corrects apolipoprotein E deficiency-induced atherosclerosis in mice. *J Clin Invest* 1994;94:899–903. [PubMed: 8040345]
32. Burger D, Dayer J-M. High-density lipoprotein-associated apolipoprotein A-I: The missing link between infection and chronic inflammation? *Autoimmun Rev* 2002;1:111–17. [PubMed: 12849067]
33. Rader DJ. Molecular regulation of HDL metabolism and function: Implications for novel therapies. *J Clin Invest* 2006;116:3090–100. [PubMed: 17143322]
34. Nissen SE, Tsunoda T, Tuzcu EM, et al. Effect of recombinant apoA-I Milano on coronary atherosclerosis in patients with acute coronary syndromes: A randomized controlled trial. *JAMA* 2003;290:2292–2300. [PubMed: 14600188]
35. Getz GS, Wool GD, Reardon CA. Apoprotein A-I mimetic peptides and their potential antiatherogenic mechanisms of action. *Curr Opin Lipidol* 2009;20:171–5. [PubMed: 19373084]



36. van der Steeg WA, Holme I, Boekholdt SM, et al. High-density lipoprotein cholesterol, high-density lipoprotein particle size, and apo-lipoprotein A-I: Significance for cardiovascular risk: The IDEAL and EPIC-Norfolk studies. *J Am Coll Cardiol* 2008;51:634–42. [PubMed: 18261682]
37. Birjmohun RS, Dallinga-Thie GM, Kuivenhoven JA, et al. Apolipoprotein A-II is inversely associated with risk of future coronary artery disease. *Circulation* 2007;116:2029–35. [PubMed: 17923573]
38. Boisfer E, Stengel D, Pastier D, et al. Antioxidant properties of HDL in transgenic mice overexpressing human apolipoprotein A-II. *J Lipid Res* 2002;43:732–41. [PubMed: 11971944]
39. Rotllan N, Ribas V, Calpe-Berdiel L, Martin-Campos JM, Blanco-Vaca F, Escola-Gil JC. Overexpression of human apolipoprotein A-II in transgenic mice does not impair macrophage-specific reverse cholesterol transport *in vivo*. *Arterioscler Thromb Vasc Biol* 2005;25:E128–32. [PubMed: 15994442]
40. Escola-Gil JC, Marzal-Casacuberta A, Julve-Gil J, et al. Human apolipoprotein A-II is a pro-atherogenic molecule when it is expressed in transgenic mice at a level similar to that in humans: Evidence of a potentially relevant species-specific interaction with diet. *J Lipid Res* 1998;39:457–62. [PubMed: 9580110]
41. Warden CH, Hedrick CC, Qiao JH, Castellani LW, Lusis AJ. Atherosclerosis in transgenic mice overexpressing apolipoprotein A-II. *Science* 1993;261:469–72. [PubMed: 8332912]
42. Raffai RL, Loeb SM, Weisgraber KH. Apolipoprotein E promotes the regression of atherosclerosis independently of lowering plasma cholesterol levels. *Arterioscler Thromb Vasc Biol* 2005;25:436–41. [PubMed: 15591220]
43. Tangirala RK, Pratico D, FitzGerald GA, et al. Reduction of isoprostanes and regression of advanced atherosclerosis by apolipoprotein E. *J Biol Chem* 2001;276:261–6. [PubMed: 11024044]
44. Thorngate FE, Rudel LL, Walzem RL, Williams DL. Low levels of extrahepatic nonmacrophage ApoE inhibit atherosclerosis without correcting hypercholesterolemia in ApoE-deficient mice. *Arterioscler Thromb Vasc Biol* 2000;20:1939–45. [PubMed: 10938015]
45. Miyata M, Smith JD. Apolipoprotein E allele-specific antioxidant activity and effects on cytotoxicity by oxidative insults and beta-amyloid peptides. *Nat Genet* 1996;14:55–61. [PubMed: 8782820]
46. Sacre SM, Stannard AK, Owen JS. Apolipoprotein E (apoE) isoforms differentially induce nitric oxide production in endothelial cells. *FEBS Lett* 2003;540:181–7. [PubMed: 12681505]
47. Ophir G, Amariglio N, Jacob-Hirsch J, Elkon R, Rechavi G, Michaelson DM. Apolipoprotein E4 enhances brain inflammation by modulation of the NF-kappaB signaling cascade. *Neurobiol Dis* 2005;20:709–18. [PubMed: 15979312]
48. Navab M, Hama-Levy S, Van Lenten BJ, et al. Mildly oxidized LDL induces an increased apolipoprotein J/paraoxonase ratio. *J Clin Invest* 1997;99:2005–19. [PubMed: 9109446]
49. Trougakos IP, Lourda M, Agiostratidou G, Kletsas D, Gonos ES. Differential effects of clusterin/apolipoprotein J on cellular growth and survival. *Free Radic Biol Med* 2005;38:436–49. [PubMed: 15649646]
50. Remaley AT, Stonik JA, Demosky SJ, et al. Apolipoprotein specificity for lipid efflux by the human ABCA1 transporter. *Biochem Biophys Res Commun* 2001;280:818–23. [PubMed: 11162594]
51. Steinmetz A, Utermann G. Activation of lecithin: Cholesterol acyltransferase by human apolipoprotein A-IV. *J Biol Chem* 1985;260:2258–64. [PubMed: 3918999]
52. Ostos MA, Conconi M, Vergnes L, et al. Antioxidative and antiatherosclerotic effects of human apolipoprotein A-IV in apolipoprotein E-deficient mice. *Arterioscler Thromb Vasc Biol* 2001;21:1023–8. [PubMed: 11397714]
53. Recalde D, Ostos MA, Badell E, et al. Human apolipoprotein A-IV reduces secretion of proinflammatory cytokines and atherosclerotic effects of a chronic infection mimicked by lipopolysaccharide. *Arterioscler Thromb Vasc Biol* 2004;24:756–61. [PubMed: 14751811]
54. Vowinkel T, Mori M, Krieglstein CF, et al. Apolipoprotein A-IV inhibits experimental colitis. *J Clin Invest* 2004;114:260–9. [PubMed: 15254593]
55. Mackness MI, Arrol S, Durrington PN. Paraoxonase prevents accumulation of lipoperoxides in low-density lipoprotein. *FEBS Lett* 1991;286:152–4. [PubMed: 1650712]

56. Watson AD, Berliner JA, Hama SY, et al. Protective effect of high density lipoprotein associated paraoxonase. Inhibition of the biological activity of minimally oxidized low density lipoprotein. *J Clin Invest* 1995;96:2882–91. [PubMed: 8675659]
57. Shih DM, Xia YR, Wang XP, et al. Combined serum paraoxonase knockout/apolipoprotein E knockout mice exhibit increased lipoprotein oxidation and atherosclerosis. *J Biol Chem* 2000;275:17527–35. [PubMed: 10748217]
58. Rosenblat M, Vaya J, Shih D, Aviram M. Paraoxonase 1 (PON1) enhances HDL-mediated macrophage cholesterol efflux via the ABCA1 transporter in association with increased HDL binding to the cells: A possible role for lysophosphatidylcholine. *Atherosclerosis* 2005;179:69–77. [PubMed: 15721011]
59. Shih DM, Gu L, Xia YR, et al. Mice lacking serum paraoxonase are susceptible to organophosphate toxicity and atherosclerosis. *Nature* 1998;394:284–7. [PubMed: 9685159]
60. Tward A, Xia YR, Wang XP, et al. Decreased atherosclerotic lesion formation in human serum paraoxonase transgenic mice. *Circulation* 2002;106:484–90. [PubMed: 12135950]
61. Ng DS, Chu T, Esposito B, Hui P, Connelly PW, Gross PL. Paraoxonase-1 deficiency in mice predisposes to vascular inflammation, oxidative stress, and thrombogenicity in the absence of hyperlipidemia. *Cardiovasc Pathol* 2008;17:226–32. [PubMed: 18402813]
62. Soran H, Younis NN, Charlton-Menys V, Durrington P. Variation in paraoxonase-1 activity and atherosclerosis. *Curr Opin Lipidol* 2009;20 :265–74. [PubMed: 19550323]
63. Birjmohun RS, Vergeer M, Stroes ES, et al. Both paraoxonase-1 genotype and activity do not predict the risk of future coronary artery disease: The EPIC-Norfolk Prospective Population Study. *PLoS ONE* 2009;4:E6809. [PubMed: 19710913]
64. Draganov DI, Stetson PL, Watson CE, Billecke SS, La Du BN. Rabbit serum paraoxonase 3 (PON3) is a high density lipoprotein-associated lactonase and protects low density lipoprotein against oxidation. *J Biol Chem* 2000;275:33435–42. [PubMed: 10931838]
65. Reddy ST, Wadleigh DJ, Grijalva V, et al. Human paraoxonase-3 is an HDL-associated enzyme with biological activity similar to paraoxonase-1 protein but is not regulated by oxidized lipids. *Arterioscler Thromb Vasc Biol* 2001;21:542–7. [PubMed: 11304470]
66. Shih DM, Xia YR, Wang XP, et al. Decreased obesity and atherosclerosis in human paraoxonase 3 transgenic mice. *Circ Res* 2007;100:1200–7. [PubMed: 17379834]
67. Connelly PW, Draganov D, Maguire GF. Paraoxonase-1 does not reduce or modify oxidation of phospholipids by peroxynitrite. *Free Radic Biol Med* 2005;38:164–74. [PubMed: 15607900]
68. Marathe GK, Zimmerman GA, McIntyre TM. Platelet-activating factor acetylhydrolase, and not paraoxonase-1, is the oxidized phospholipid hydrolase of high density lipoprotein particles. *J Biol Chem* 2003;278:3937–47. [PubMed: 12466264]
69. Noto H, Hara M, Karasawa K, et al. Human plasma platelet-activating factor acetylhydrolase binds to all the murine lipoproteins, conferring protection against oxidative stress. *Arterioscler Thromb Vasc Biol* 2003;23:829–35. [PubMed: 12649088]
70. Theilmeyer G, De Geest B, Van Veldhoven PP, et al. HDL-associated PAF-AH reduces endothelial adhesiveness in apoE<sup>-/-</sup> mice. *FASEB J* 2000;14:2032–9. [PubMed: 11023987]
71. Quarck R, De Geest B, Stengel D, et al. Adenovirus-mediated gene transfer of human platelet-activating factor-acetylhydrolase prevents injury-induced neointima formation and reduces spontaneous atherosclerosis in apolipoprotein E-deficient mice. *Circulation* 2001;103:2495–500. [PubMed: 11369691]
72. Arakawa H, Qian JY, Baatar D, et al. Local expression of platelet-activating factor-acetylhydrolase reduces accumulation of oxidized lipoproteins and inhibits inflammation, shear stress-induced thrombosis, and neointima formation in balloon-injured carotid arteries in nonhyperlipidemic rabbits. *Circulation* 2005;111:3302–9. [PubMed: 15956136]
73. McIntyre TM, Prescott SM, Stafforini DM. The emerging roles of PAF acetylhydrolase. *J Lipid Res* 2009;50(Suppl):S255–9. [PubMed: 18838739]
74. Garza CA, Montori VM, McConnell JP, Somers VK, Kullo IJ, Lopez-Jimenez F. Association between lipoprotein-associated phospholipase A2 and cardiovascular disease: A systematic review. *Mayo Clin Proc* 2007;82:159–65. [PubMed: 17290721]

75. Serruys PW, Garcia-Garcia HM, Buszman P, et al. Effects of the direct lipoprotein-associated phospholipase A(2) inhibitor darapladib on human coronary atherosclerotic plaque. *Circulation* 2008;118:1172–82. [PubMed: 18765397]
76. Chen N, Liu Y, Greiner CD, Holtzman JL. Physiologic concentrations of homocysteine inhibit the human plasma GSH peroxidase that reduces organic hydroperoxides. *J Lab Clin Med* 2000;136:58–65. [PubMed: 10882228]
77. Maddipati KR, Marnett LJ. Characterization of the major hydroperoxide-reducing activity of human plasma. Purification and properties of a selenium-dependent glutathione peroxidase. *J Biol Chem* 1987;262(17):398–403.
78. Hamanishi T, Furuta H, Kato H, et al. Functional variants in the glutathione peroxidase-1 (GPx-1) gene are associated with increased intima-media thickness of carotid arteries and risk of macrovascular diseases in Japanese type 2 diabetic patients. *Diabetes* 2004;53:2455–60. [PubMed: 15331559]
79. Colak E, Majkic-Singh N, Stankovic S, et al. Parameters of antioxidative defense in Type 2 diabetic patients with cardiovascular complications. *Ann Med* 2005;37:613–20. [PubMed: 16338763]
80. Blankenberg S, Rupprecht HJ, Bickel C, et al. Glutathione peroxidase 1 activity and cardiovascular events in patients with coronary artery disease. *N Engl J Med* 2003;349:1605–13. [PubMed: 14573732]
81. Winter JP, Gong Y, Grant PJ, Wild CP. Glutathione peroxidase 1 genotype is associated with an increased risk of coronary artery disease. *Coron Artery Dis* 2003;14:149–53. [PubMed: 12655278]
82. Lapenna D, de Gioia S, Ciofani G, et al. Glutathione-related antioxidant defenses in human atherosclerotic plaques. *Circulation* 1998;97:1930–4. [PubMed: 9609086]
83. Lewis P, Stefanovic N, Pete J, et al. Lack of the antioxidant enzyme glutathione peroxidase-1 accelerates atherosclerosis in diabetic apolipoprotein E-deficient mice. *Circulation* 2007;115:2178–87. [PubMed: 17420349]
84. Torzewski M, Ochsenhirt V, Kleschyov AL, et al. Deficiency of glutathione peroxidase-1 accelerates the progression of atherosclerosis in apolipoprotein E-deficient mice. *Arterioscler Thromb Vasc Biol* 2007;27 :850–7. [PubMed: 17255533]
85. de Haan JB, Witting PK, Stefanovic N, et al. Lack of the antioxidant glutathione peroxidase-1 does not increase atherosclerosis in C57BL/J6 mice fed a high-fat diet. *J Lipid Res* 2006;47:1157–67. [PubMed: 16508038]
86. Goyal J, Wang K, Liu M, Subbaiah PV. Novel function of lecithin-cholesterol acyltransferase. Hydrolysis of oxidized polar phospholipids generated during lipoprotein oxidation. *J Biol Chem* 1997;272:16231–9. [PubMed: 9195924]
87. Nofer JR, van der Giet M, Tolle M, et al. HDL induces NO-dependent vasorelaxation via the lysophospholipid receptor S1P3. *J Clin Invest* 2004;113:569–81. [PubMed: 14966566]
88. Tolle M, Pawlak A, Schuchardt M, et al. HDL-associated lysosphingolipids inhibit NAD(P)H oxidase-dependent monocyte chemoattractant protein-1 production. *Arterioscler Thromb Vasc Biol* 2008;28:1542–8. [PubMed: 18483405]
89. Kimura T, Tomura H, Mogi C, et al. Role of scavenger receptor class B type I and sphingosine 1-phosphate receptors in high density lipoprotein-induced inhibition of adhesion molecule expression in endothelial cells. *J Biol Chem* 2006;281:37457–67. [PubMed: 17046831]