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Integrins and bone metastasis: Integrating tumor cell and stromal cell interactions

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Abstract

Integrins on both tumor cells and the supporting host stromal cells in bone (osteoclasts, new blood vessels, inflammatory cells, platelets and bone marrow stromal cells) play key roles in enhancing bone metastasis. Tumor cells localize to specific tissues through integrin-mediated contacts with extracellular matrix and stromal cells. Integrin expression and signaling are perturbed in cancer cells, allowing them to “escape” from cell-cell and cell matrix tethers, invade, migrate and colonize within new tissues and matrices. Integrin signaling through $\alpha\beta3$ and VLA-4 on tumor cells can promote tumor metastasis to and proliferation in the bone microenvironment. Osteoclast (OC) mediated bone resorption is a critical component of bone metastasis and can promote tumor growth in bone and $\alpha\beta3$ integrins are critical to osteoclast function and development. Tumors in the bone microenvironment can recruit new blood vessel formation, platelets, pro-tumor immune cells and bone marrow stromal cells that promote tumor growth and invasion in bone. Integrins play critical roles in platelet aggregation ($\alpha\beta3$ and $\alpha\text{IIb}\beta3$), hematopoietic cell mobilization (VLA-4, osteopontin), neoangiogenesis ($\alpha\beta3$, $\alpha\beta5$, $\alpha6\beta4$, $\beta1$ integrin) and stromal function (osteopontin, VLA-4). Integrins are involved in the pathogenesis of bone metastasis at many levels and further study to define integrin dysregulation by cancer will yield new therapeutic targets for the prevention and treatment of bone metastasis.

Introduction

The development of bone metastasis is common in many cancers, occurring in virtually all patients with multiple myeloma, in 65%–75% of patients with advanced breast and prostate cancers, and in 30%–40% of patients with lung cancer[1–3]. The consequences of bone metastases are often devastating and can cause pain, pathologic fractures, spinal cord and other nerve-compression syndromes and life-threatening hypercalcemia[4]. Both osteolytic lesions and osteoblastic bone metastases are associated with increased osteoclast (OC) activity and disrupted bone micro-architecture[5,6]. In the bone microenvironment, tumor cells secrete soluble factors that promote bone remodeling resulting in the release of additional bone matrix-bound growth factors which further activates OCs and osteoblasts

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(OB) and tumor growth[3,4,7–16]. Anti-resorptive therapy, e.g. with bisphosphonates or denosumab, significantly decreases skeletal complications of cancer and is a standard of care for patients with bone metastases[4,8,17–19]. Beyond their effects on bone, tumors in the bone microenvironment recruit new blood vessel formation, platelets, immune cells and stromal cells that promote tumor growth and invasion in bone. Integrin-mediated cell signaling plays a critical role in many of these processes during bone metastasis, including platelet aggregation ($\alpha\text{IIb}\beta\text{3}$), hematopoietic/immune cell mobilization (VLA-4, osteopontin), neoangiogenesis ($\alpha\text{v}\beta\text{3}$, $\alpha\text{v}\beta\text{5}$, $\alpha\text{6}\beta\text{4}$, β1 integrin) and stromal function (osteopontin, VLA-4) (see Figure 1). For these reasons, the mechanisms by which integrin signaling mediate the pathogenesis of bone metastasis has been an area of active research.

Integrin structure, activation and signaling

Integrins are heterodimeric transmembrane glycoproteins that facilitate cell-cell and cell-extracellular matrix (ECM) adhesion and cell migration[20]. Integrins recruit many intracellular signaling molecules and can activate survival, proliferation, and motility signaling pathways[21]. There are 8 beta and 18 alpha integrin subunits that assemble into 24 different known combinations in different cell types, each characterized by distinct ligand binding specificities (including collagen, osteopontin, fibronectin, laminin, and others, depending on the integrin family), signaling abilities, and regulatory mechanisms[22]. Integrins are activated by conformational changes in the integrin extracellular domains (“inside-out” signaling). When the integrin α and β subunit cytoplasmic and transmembrane domains remain closely juxtaposed, the extracellular domains are held in a closed conformation. Activation by intracellular signals to the cytoplasmic tails results in separation of the α and β cytoplasmic and transmembrane domains and exposure of the extracellular ligand binding domain[23] (“inside-out” signaling). The open conformation, facilitates high affinity ligand binding and triggers integrin-mediated cell signaling cascades (outside-in signaling)[24,25].

Many proteins play critical roles in the activation of specific integrins, but two cytoplasmic proteins, talin and kindlin, are necessary for inside-out signaling required for the activation of all integrin subtypes[23,26–29]. Talin binds to the proximal end of the beta cytoplasmic tail via a phosphotyrosine-binding (PTB) domain within its FERM domain[27] and links the integrin to the actin cytoskeleton[23]. Kindlin 1, 2, or 3, are necessary for talin-induced integrin activation[26,30,31]. Kindlin, like talin, also interacts with intracellular proteins including focal adhesion kinase resulting in cytoskeleton reorganization and adhesion[32]. G-protein coupled receptors such as the ADP receptor P2Y₁₂, also play critical roles in the inside-out signaling required for integrin activation[25,33,34]. Structure-function analyses on β3 integrins have shown that a “membrane-proximal” region is important for “inside-out” signaling required [28,35–40].

In addition to activation by inside-out signaling, ligand binding and integrin clustering can be significantly modulated by growth factor receptor interactions and other integrin interacting proteins, as reviewed in[22,23,41]. For example, integrin associated protein, CD47, augments integrin activation and affects the ability of $\alpha\text{v}\beta\text{3}$ integrin to cluster upon ligand binding[42]. Ligation of the integrin then stimulates outside-in signaling that leads to the activation of numerous signals critical for growth, migration, survival, and other functions, including FAK phosphorylation, ERK signaling, and NF- κB activation. Thus, integrin signaling in cancer cells and in associated stromal, endothelial and hematopoietic cells can be influenced by intracellular signaling proteins, growth factors, chemokines and other receptors that participate in regulating integrin function through effects on integrin activation, ligand binding, ligand affinity and integrin clustering.

Maintaining adhesion to the ECM, in part through integrin signaling, is critical to cell survival[43]. Altered cell-cell or cell-matrix interactions can result in disruption of downstream survival signaling and anchorage-dependent non-transformed cells undergo anoikis[43]. Under normal conditions, because each cell type expresses a unique set of integrins that recognize underlying ECM ligands, this form of apoptosis ensures that detached cells do not colonize inappropriate locations[43]. Cells that resist anoikis, such as metastatic cells, take advantage of several different mechanisms, including aberrant integrin expression so that the cell can adhere to a novel ECM[44], constitutive activation of molecules usually activated via integrin signaling including FAK[45], EGFR[46], and SRC[47], and lack of activation of pro-apoptotic pathways[48], among others. The integrin family of adhesion receptors link extracellular matrix to the cytoskeleton through a complex and regulated network of activation, interaction with numerous growth factor, GPCR, chemokine and cytokine receptors and induction of complex signaling cascades.

Integrin expression and signaling on tumor cells that metastasize to bone

Tumor progression, invasion, and eventual metastasis require the activity of many adhesion proteins, including the integrin superfamily. At each stage of cancer progression, subsets of integrin heterodimers are activated, providing the necessary signaling pathways for adhesion, migration, and cell survival. Metastatic tumor cells show differential integrin heterodimerization and activation compared to non-metastatic tumor cells that enable the cell to home to and colonize in a metastatic site, such as the bone marrow cavity [49,50]. In order for primary epithelial cancers to metastasize, the tumor cells must become resistant to anoikis and detach from the primary tumor site ECM, enter the vasculature, and eventually colonize a distant site. Upon reaching a successful metastatic site, however, tumor cells use both anoikis and anoikis-resistance to their advantage, in some cases forming micro-metastases that are resistant to cancer treatment via integrin binding to the underlying bone ECM as reviewed in[51]. In addition to evading apoptosis, tumor cells must also form interactions between the tumor cell and bone stroma to establish and maintain skeletal metastasis. Many integrins have been implicated in tumor cell-host bone stroma interactions during bone metastasis and tumor growth in bone (Figure 1, Table 1), including the $\beta 1$ and $\beta 3$ integrin family members.

$\alpha v \beta 3$ is receptor for osteopontin, fibronectin, and vitronectin, ECM proteins that are important bone matrix proteins, and $\alpha v \beta 3$ has been identified as a critical integrin in breast cancer and prostate cancer skeletal metastasis[50,52–56]. Interestingly, although $\alpha v \beta 3$ has been shown to bind to fibronectin in other locations with high affinity, tumor $\alpha v \beta 3$ integrins do not bind fibronectin in bone marrow stroma, indicating that $\alpha v \beta 3$ -expressing tumor cells bind to the bone stromal ligands vitronectin and osteopontin[57]. In breast cancer, $\alpha v \beta 3$ binding of host osteopontin is necessary for tumor cell colonization to bone[58]. Bone metastatic cells have a higher expression of $\alpha v \beta 3$ than the primary tumor[53], promoting adherence to the bone matrix by binding osteopontin expressed by bone stromal cells[58]. Breast cancer cells that overexpress $\alpha v \beta 3$ have increased levels of bone metastasis and associated tumor burden and osteolysis[52,59–62]. This overexpression of $\alpha v \beta 3$ in the tumor cells leads to increased tumor cell adhesion, migration, and invasion to bone as well as enhanced OC recruitment within the bone microenvironment[60,61], implicating a role of tumor-specific $\alpha v \beta 3$ expression in breast cancer metastasis to bone as well as tumor-associated osteolysis. Likewise, in prostate cancer cells, active $\alpha v \beta 3$ is necessary for the adherence and migration to bone matrix proteins at early stages of skeletal metastasis. This tumor cell $\alpha v \beta 3$ integrin expression allows cancer cells to adhere to the bone matrix and interact directly with the native bone cells, osteoblasts and osteoclasts, as well as with the bone matrix itself [59].

The $\beta 1$ family member, $\alpha 5\beta 1$, has been identified as the primary integrin receptor for fibronectin on human bone marrow stroma[57]. $\alpha 5\beta 1$ expression on leukemia, prostate and breast cancer cells facilitates interaction with bone stroma[57,63–65]. Antibody inhibition of $\alpha 5$, $\beta 1$, or fibronectin block prostate cancer tumor cell binding to bone stroma, indicating necessary roles for both integrin $\alpha 5\beta 1$ on tumor cells and fibronectin on bone marrow stromal cells[57]. In breast cancer skeletal metastasis, the interaction between malignant cell $\alpha 5\beta 1$ and host stromal cell fibronectin contributes to the survival of growth-arrested tumor cells, a potential mechanism through which tumor cells can become sequestered and “dormant” within the bone marrow cavity and may later begin to proliferate to establish a skeletal metastasis[64]. Upon FGF-2 growth factor stimulation, breast cancer cells undergo growth arrest and up regulate $\alpha 5\beta 1$ expression. In most cases, these cells die, but cells that bind fibronectin via $\alpha 5\beta 1$ and initiate cell survival signaling cascades survive[64].

Another $\beta 1$ family member, $\alpha 2\beta 1$, a collagen type I receptor, is expressed by prostate tumor cells, and its activity promotes invasion and adherence to the bone stroma. The presence of collagen I, the most abundant protein in bone, significantly increases prostate epithelial cell adhesion in culture, and antibody inhibition of integrin subunits $\alpha 2$ and $\beta 1$ significantly inhibits tumor cell binding to stroma[66]. Hall et al showed that a skeletal metastatic prostate cancer cell line, but not cell lines that are metastatic to other organs, binds to collagen I and that this collagen I binding is $\alpha 2\beta 1$ dependent in vivo [67]. Interestingly, stromal expression of collagen I does not increase tumor growth, but instead promotes tumor cell migration[67]. Tumor cell $\alpha 2\beta 1$ binding of host bone marrow stromal collagen I activates RhoC GTPase which instigates a signaling cascade responsible for cytoskeleton reorganization, migration, and, eventually, collagen-stimulated invasion and preferential skeletal metastasis[68].

$\alpha 4\beta 1$ /vascular cell adhesion molecule-1 (VCAM-1) binding has been identified as important for cell-cell contact between $\alpha 4\beta 1$ expressing myeloma cells and VCAM-1 expressing bone marrow stroma[69]. This interaction contributes to bone tumor growth, OC stimulation and resultant osteolysis[69,70]. Likewise, epithelial tumor cells (CHO) that overexpress $\alpha 4\beta 1$ developed significantly more bone metastases than mice inoculated with CHO cells alone[71]. Bone metastases, but not other metastases, were inhibited by antibodies against $\alpha 4$ and/or VCAM-1, suggesting a role for $\alpha 4\beta 1$ /VCAM-1 binding in the skeletal metastases of solid tumors[71]. The role of integrins and chemokine cross talk in tumor cell homing to bone will be discussed below. While many aspects of tumor–bone stromal interactions remain unknown, it is clear that specific interactions between tumor cell integrins and bone stromal cell ligands are essential for successful homing and metastasis to bone.

Integrin expression and signaling and osteoclast function and bone metastasis

Bone invading metastatic tumor cells co-opt integrin signaling pathways that enhance osteoclast (OC) function and recruitment. As part of bone remodeling, OC bind to the bone matrix, form an actin ring mediated sealing zone, secrete enzymes and acid to degrade bone, and then migrate to a new site. Each of these functions is regulated in part by integrins located on the membrane surface of the OC, interacting with neighboring cells and with the extracellular matrix[72].

Several integrins are involved in OC binding to bone, including $\alpha v\beta 3$ (osteopontin, vitronectin, bone sialoprotein), $\alpha v\beta 5$ (fibronectin), and $\alpha 2\beta 1$ (collagen) [73,74]. Of these, $\alpha v\beta 3$ is the predominant integrin found on OCs, and antibody inhibition of $\alpha v\beta 3$ inhibits OC attachment to the bone matrix as well as OC mediated bone resorption[75]. In addition, mice with targeted disruption of $\beta 3$ integrin ($\beta 3^{-/-}$) have defective osteoclast function [76] and

are protected from tumor associated osteolysis[77]. $\alpha\beta3$ is responsible for mediating OC-bone recognition [53,75,78,79] and subsequent attachment to the bone matrix[75,80], signaling to create the characteristic resorptive ruffled membrane, regulation of osteoclast spreading, and overall organization of the cytoskeleton[76,81]. Activation of $\alpha\beta3$ regulates OC adhesion and migration on osteopontin, important for OC polarization and bone resorption[82]. Osteopontin ligand binding of $\alpha\beta3$ causes a reduction of OC cytosolic calcium, inducing podosome formation and subsequent resorption[83]. In addition, $\alpha\beta3$ is critical for the activation of c-Src, c-Cbl, and GTPases Rho and Rac, signaling that is necessary for the cytoskeletal reorganization important in OC function [81,84,85].

OC targeted therapy is a standard of care for the treatment of bone metastasis and myeloma bone disease. Tumor cells recruit OCs resulting in bone destruction and pain[3,86,87]. Because of its known role in OC function and its high expression in skeletal metastatic tumors as discussed above, much research has focused on $\alpha\beta3$ integrin and its ligands. An important characteristic of $\alpha\beta3$ cell adhesion, both in OCs and tumor cells, is the requirement of osteopontin, an $\alpha\beta3$ ligand[58]. Osteopontin is a non-collagenous bone matrix protein that is produced by osteoblasts, OCs, and macrophages and is found in the extracellular matrix adjacent to calcified bone [88–90]. Expression of osteopontin in both the tumor cell and in the bone microenvironment can promote skeletal metastasis[91,92]. Osteopontin-deficient mice have reduced bone metastasis and tumor induced osteolysis than wild type controls in a mouse model of tumor metastasis using syngeneic B16 melanoma cells[93,94], confirming a role for host cell osteopontin expression during bone metastasis. Recombinant osteopontin induces cell migration of B16 cells that is inhibited by repressing the ERK/MAPK pathway, suggesting that the ERK/MAPK pathway regulates bone microenvironment osteopontin levels[91]. Overexpression of osteopontin in B16 melanoma cells increases cell proliferation and migration, indicating that the ligand also plays an important role in the tumor cell itself [91]. It has been demonstrated using a prostate cancer cell line over-expressing osteopontin that tumor-cell osteopontin regulates MMP-9 secretion and subsequent CD44/MMP-9 interaction, important for the migration of prostate cancer cells, contributing to metastatic potential[95]. Osteopontin-producing tumor cells enhance osteopontin production by osteoblasts[96] and OCs[97], stimulating osteoclastogenesis, OC adherence, migration, and bone resorption via host $\alpha\beta3$ binding[88,98]. Osteopontin activation of $\alpha\beta3$ integrin leads to downstream activation of FAK, c-Src kinase, and Ras-ERK, among other signaling molecules, resulting in cytoskeletal reorganization, focal adhesion formation, basolateral membrane differentiation, and osteoclastic resorption[59,99].

CD47, integrin associated protein, is expressed constitutively and interacts with integrins, including $\alpha\beta3$, as part of inside-out signaling cascades and also operates in an integrin-independent manner. CD47 plays a role in osteoclast and macrophage biology and CD47^{-/-} mice have decreased OC number and function[100,101] which can be rescued in vitro by inhibiting nitric oxide synthase[101]. CD47^{-/-} mice have decreased bone metastases and tumor-associated osteolysis compared to wild type[101]. During the early stages of osteoclastogenesis, namely, macrophage fusion, CD47 binds with SIRP1 α , a molecule that is transiently induced in myeloid cells and that likely participates in early fusion events[102]. In the event of tumor cell metastasis to bone, however, it has been reported that cancer cells may utilize this macrophage self-recognition signaling to fuse with macrophages[103], leading to mature OCs with tumor cell nuclei and subsequent overexpression of OC stimulation factors, thus leading to increased OC function[104].

These data underscore the importance of integrins, especially $\alpha\beta3$, and its adaptor proteins in OC biology and bone metabolism and point to the role of osteoclast integrins in regulating growth of cancer cells in the bone.

Integrins and tumor neovasculature and bone metastasis

Tumor neovascularization is essential for tumor cell invasion and metastasis. Access to the host blood supply provides the tumor cells with nutrients and connects the tumor to the circulation, facilitating the dissemination of metastatic cells. The angiogenic process begins with the de-stabilization and de-differentiation of local vessels, followed by activation of endothelial cells (EC), EC migration and proliferation into the tumor extracellular matrix (ECM), and finally organization of ECs into functional vessels. The ability of tumor cells to activate the normally quiescent vasculature is proposed to be controlled by an “angiogenic switch” mechanism, whereby tumor or stromal cells induce changes in the relative balance of inducers (e.g. vascular endothelial growth factor (VEGF) or TGF β , PDGF, TNF α , bFGF) and inhibitors (e.g. thrombospondin-1 [TSP-1]) of angiogenesis reviewed in [41,105–109]. Activated platelets, tumor cells, and fibroblasts secrete many of these pro-angiogenic factors. It has recently been appreciated that macrophage lineage cells play important roles in promoting tumor-associated angiogenesis [110–113]. Bone metastasis and bone residing tumors like myeloma also modify and recruit endothelial cells to enhance neoangiogenesis [114,115].

Many integrin heterodimers have been implicated in tumor-associated angiogenesis [41,105–109,116]. The first integrin found to regulate angiogenesis, $\alpha v\beta 3$, is expressed at high levels on tumor-associated vasculature [117,118] and tumor-associated angiogenesis can be inhibited by $\beta 3$ integrin neutralizing antibodies [119–122]. $\alpha v\beta 3$ has been specifically implicated in the angiogenesis associated with prostate cancer bone metastases; antibody inhibition of $\alpha v\beta 3$ decreases tumor-associated blood vessels in mice [123]. Interestingly, Reynolds et al. demonstrated enhanced (not reduced) tumor-associated angiogenesis in subcutaneous tumors in $\beta 3^{-/-}$ mice [124]. Elevated levels of VEGFR2 were found on tumor-associated blood vessels in $\beta 3^{-/-}$ mice, and a VEGFR2 inhibitor could block the enhanced blood vessel formation [125]. It should be noted that an inhibitor of integrin binding and signaling might have different consequences than loss of integrin expression. For example, apoptotic machinery is activated in certain cells expressing integrins that are not ligand-bound [126–129]. Recent reports that low dose integrin antagonists can increase tumor growth and angiogenesis while higher doses suppress tumor growth and angiogenesis [130] underscore the complexity of targeting $\beta 3$ integrins for angiogenesis and cancer therapy.

Another αv integrin, $\alpha v\beta 5$, also shows increased expression on tumor-associated vasculature, and $\alpha v\beta 5$ antibodies inhibit VEGF-induced tumor-associated angiogenesis [131]. In contrast, the $\beta 3/\beta 5^{-/-}$ double knock out mice show enhanced tumor-associated angiogenesis, as was seen in $\beta 3^{-/-}$ mice [125]. Several hypotheses have been proposed that reconcile the contradictory results involving the αv integrin family that outline the roles of the integrins as pro-angiogenic, anti-angiogenic, and/or working through different pathways as reviewed in [41,108]. It is clear, however, that $\alpha v\beta 3$ and $\alpha v\beta 5$ have distinct roles in regulation of tumor-associated angiogenesis and associated metastasis. The bone targeted bisphosphonate, zoledronic acid, alters endothelial cell integrin-mediated adhesion by reduced expression of $\alpha v\beta 3$ and $\alpha v\beta 5$ integrin on endothelial cells in vitro in one observation [132]. This observation provides a possible mechanism for osteoclast-independent anti-tumor actions for bisphosphonates that have been observed in some animal models [133–135] and clinically [136–139]. Evaluation of the effects of bisphosphonates on integrin signaling in the tumor bone microenvironment are underway.

While much of the research in integrin-mediated angiogenesis has been focused on the αv integrins, there is evidence that other heterodimers play a role in angiogenic regulation, particularly the $\beta 1$ and $\beta 4$ families. The $\beta 1$ integrin family ($\alpha 1\beta 1$, $\alpha 2\beta 1$, $\alpha 5\beta 1$, $\alpha 4\beta 1$) has a

critical role in angiogenesis with $\beta 1^{-/-}$ mice having severe vascular defects. $\alpha 1\beta 1$ (a collagen receptor) and $\alpha 2\beta 1$ (a laminin receptor) have been shown to be important for mediating cell adhesion in VEGF-stimulated endothelial cells[140]. In vivo, function-blocking antibodies to $\alpha 1$ and $\alpha 2$ significantly inhibited VEGF-induced angiogenesis, indicating a positive regulatory role for $\alpha 1\beta 1$ and $\alpha 2\beta 1$ expression in tumor-associated angiogenesis [141]. Genetic data further support a role for the integrin $\alpha 1\beta 1$ as a positive regulator of angiogenesis as $\alpha 1$ -deficient mice show reduced angiogenesis[142].

Fibronectin receptor $\alpha 5\beta 1$ has also been implicated as a positive regulator of angiogenesis: $\alpha 5\beta 1$ antagonists inhibit tumor-associated angiogenesis in mice by promoting endothelial cell migration and regulating proliferation and apoptosis[143,144]. Importantly, the $\alpha 5\beta 1$ antagonists did not inhibit angiogenesis induced by VEGF, indicating that the integrin $\alpha 5\beta 1$ (together with $\alpha v\beta 3$) may act in a VEGF-independent pathway[144]. $\alpha 4\beta 1$, together with its ligand, VCAM-1, expressed in vessel mural cells, plays an important role in adhesion of endothelial cells and vascular smooth muscle cells during blood vessel formation[145]. Both anti- $\alpha 4\beta 1$ antibodies and anti-VCAM-1 antibodies inhibit angiogenesis in vivo. Another integrin, laminin receptor $\alpha 6\beta 4$ is reported to regulate several aspects of tumor angiogenesis. Genetic studies reveal that $\alpha 6\beta 4$ promotes endothelial cell migration in culture; in addition, the integrin is involved in the translational regulation of VEGF, having a pro-angiogenic effect[146,147].

In many cases, integrins influence angiogenesis through their interaction with the integrin ligand, thrombospondin 1 (TSP-1). Mice with a TSP-1 deficiency have increased tumor burden and tumor-associated vasculature, both in capillary size and number, while mice that over-express TSP-1 have delayed or absent tumor growth and reduced tumor-associated vasculature[148]. These data indicate that TSP-1 can contribute to tumor burden via negative regulation of angiogenesis. In contrast, in a human breast cancer cell line, TSP-1 stimulation up-regulates both integrin subunit $\alpha 6$ mRNA levels and protein levels which leads to elevated adhesion to ECM protein laminin in vitro, suggesting that TSP-1 facilitates pathogenic angiogenesis[149]. TSP-1 also interacts with $\alpha 9\beta 1$ via its N-terminal domain and has a positive effect on proliferation and motility in culture and on angiogenesis in vivo that can be reduced by $\alpha 9\beta 1$ inhibitors. This binding of the microvasculature-associated integrin in endothelial cells with TSP-1 activates signaling cascades including ERK and paxillin. Thus, TSP-1 can play both pro- and anti-angiogenic roles, depending on its specific integrin interaction.

The roles of integrins in tumor-associated angiogenesis are complex, not only involving integrin-ligand interactions and associated signaling pathways, but also specific temporal regulation and indirect effects through proteins such as TSP-1, and are important for the progression of angiogenesis and eventual metastasis.

Integrins and hematopoietic and tumor-induced mobilization and modulation of bone marrow cells

The bone marrow is the primary site of hematopoiesis in the adult. Osteoblasts and bone marrow stromal cells regulate hematopoietic stem cell (HSC) growth, differentiation and bone marrow retention through numerous signaling pathways including integrin VLA-4/VCAM[150], chemokine SDF-1/CXCR4, BMPs and Notch[151–156]. Hematopoietic progenitors and stem cells express the integrin, VLA-4 and the chemokine receptor, CXCR4. Osteoblast and bone marrow stromal cells produce VCAM-1, SDF-1 and osteopontin, all important components of the “hematopoietic stem cell niche”[157–159]. Integrin and chemokine signaling work in concert to promote HSC and progenitor cell homing and mobilization in the bone marrow [160]. Disruption of VLA-4/VCAM-1 and

SDF-1/CXCR4 result in mobilization of HSC into the circulation[159]. G-CSF mobilization of HSC acts in part through disruption of VLA-4/VCAM-1 and CXCR4/SDF1[158,161]. Osteoclast resorption can also regulate HSC mobilization and the stem cell niche [162].

Diverse integrins are expressed on hematopoietic progenitor cells in specific patterns and at distinct time points[163]. Integrins not only mediate the binding of normal progenitor cells to stroma and matrix molecules, but may also regulate expansion, maturation and differentiation of those cells[164], [165]. For example, $\alpha 4\beta 1$ integrin regulates hematopoietic progenitor cell fate through changes in integrin expression and activity levels during cell maturation and differentiation into erythrocytes and neutrophils[165–167]. $\alpha 4$ containing integrins mediate adhesion of hematopoietic progenitors to stromal cells likely through binding to matrix components such as fibronectin[168] or cellular receptors such as VCAM-1[169]. The integrin subunits $\alpha 5$, $\alpha 6$ and $\alpha 9$ have also been shown to be expressed by progenitor cells[170–172]. Studies using blocking antibodies demonstrated that $\alpha 6$ subunit cooperates in collaboration with the $\alpha 4$ subunit is regulating homing of progenitor cells[171]. $\alpha 9\beta 1$ integrin is also important for adhesion of progenitor cells to osteoblasts in the bone marrow[172], illustrating the fact that hematopoiesis takes place in three dimensional matrices, the so-called bone marrow niches. These niches are located either at the endosteum near osteoblasts and also in the vascular niche close to marrow blood vessels [173].

Tumor cells both in the bone microenvironment and at distant sites can modulate and mobilize hematopoietic progenitor and immune cells to promote bone and visceral metastasis and local tumor growth. Tumor-induced mobilization of VEGFR+ and Sca+kit-bone marrow derived cells have been implicated in enhancing distant tumor and metastatic growth[174]. These mobilized VEGFR+ cells also express $\alpha 4\beta 1$ and can migrate to sites of increased synthesis of matrix components such as fibronectin and establish a “pre-metastatic niche” that can favor tumor metastasis and growth[174]. $\beta 2$ integrins on bone marrow derived endothelial progenitors can also mediate the adhesion and VEGF-induced migration of the progenitors to the mature endothelium of actively remodeling vasculature[175].

Tumor cells from a primary lesion can act at a distance to influence bone marrow hematopoiesis through secreted factors such as the integrin ligand, osteopontin[176]. Primary epithelial tumors can “instigate” growth of indolent tumors through modulation of the bone marrow microenvironment and mobilization and recruitment of bone marrow cells to distant tumor sites[176,177]. McAllister et al. found that tumor secretion of osteopontin is necessary but not sufficient in xenograft models to modulate the bone microenvironment and promote bone marrow cell recruitment to tumor metastasis[176]. Pazolli et al. found that osteopontin secreted by senescent fibroblasts promoted tumorigenesis in animal models of skin cancer[178].

Thus tumors cells both in bone and at distant sites can modulate hematopoiesis in part through osteopontin and bone marrow cell integrins resulting in the mobilization and recruitment of bone marrow derived cells that will enhance local and metastatic tumor growth.

Integrins and tumor cell homing/colonization of bone

The site of metastasis is tumor cell specific depending on their integrin, chemokine receptor and cytokine/receptor expression profiles[50,179–181]. At the metastatic site, normal physiology is changed towards increased secretion of cytokines and activation of integrins to support recruitment, survival and growth of tumor cells. Metastasizing cancer cells can co-opt the same mechanisms used in physiological hematopoietic progenitor cell homing to bone through expression of integrins and chemokines [150,152,153]. CXCR4 expressed on

cancer cells can direct those cells to bone[181–186]. The migration of myeloma cells to and across bone marrow stromal cells is in part regulated by SDF-1 α /CXCR4 ligation and up-regulation of α 4 β 1 (VLA-4) resulting in adhesion of myeloma cells to the underlying bone marrow stroma[187]. Likewise, CXCR4 ligation can increase α v β 3 expression and aggressiveness of metastatic prostate cancer cells and disruption of CXCR4 can inhibit prostate cancer bone metastases[183–185].

It has been recently shown that β 3 integrin activity on circulating CXCR4 positive bone marrow derived cells is important for their migration and recruitment to sites of angiogenesis. In mice with mutated tyrosine residues “knocked in” to the β 3 integrin locus to inhibit proper phosphorylation (DiYF mice)[188], CXCR4 positive bone marrow derived cells were higher in number and defective in recruitment to subcutaneously implanted tumors or wounds, where SDF-1 levels were also lower[189]. These data demonstrate that β 3 integrin on bone marrow derived cells may be critical for the CXCR4/SDF-1 gradient, and thus maybe important for localization of tumor cells to the bone microenvironment and also localization of myeloid/endothelial cells to tumors. Interestingly, CXCR4 deletion on bone marrow cells can enhance osteoclast activity which could counteract some of the beneficial effects of CXCR4 inhibition on bone metastases[9]. Integrins expressed by tumor cells, in concert with bone microenvironment chemokine secretion and further integrin activation, determines the osteotropic characteristics of metastasizing cancer cells and represent an ideal target for skeletal metastatic cancer therapy.

Integrins and myeloid/immune cell function during tumor growth in bone

Myeloid cell integrins are involved in tumor evasion from immune responses and tumor induced angiogenesis. Bone marrow derived myeloid cells (macrophages, monocytes, myeloid derived suppressor cells, myeloid dendritic cells) migrate to tumors and contribute to in tumor growth, invasion, and angiogenesis[190–194]. Macrophages within tumors are called tumor-associated-macrophages (TAM) [127], and are recruited by chemoattractants secreted by the tumor such as MCP-1[195] and then differentiate into tissue macrophages[196]. The anti-tumor M1 phenotype, represents a classical activation that is induced by pathogens, lipopolysaccharides (LPS) or interferon gamma resulting in secretion of proinflammatory cytokines such as tumor necrosis factor α (TNF α), interleukin 1 β (IL-1 β) and others. M1 macrophages can act in an anti-tumor fashion by secretion of cytotoxic cytokines and antigen presentation to lymphocytes[197]. The pro-tumor M2 phenotype, represents alternative activation induced by IL-4 or IL-10[198]. M2 polarized macrophages, promote tumor cell proliferation and survival, suppress immune responses, and drive tumor neoangiogenesis[197,199–201]. Studies have shown that the TAM content of tumors and prognosis of patients are inversely correlated[192,202,203].

β 2 integrins are involved in monocyte/myeloid cell migration through endothelium and in phagocytosis, while β 1 integrins mediate adhesion to matrix proteins and the induction of inflammatory genes[204]. α 4 β 1 and α v β 3 integrins have been implicated in myeloid cell homing, adhesion, and migration to tumors. α 4 β 1 promotes endothelial progenitor cells and monocyte homing and adhesion to sites of active pathological angiogenesis[205]. Inhibition of α 4 β 1 leads to suppressed monocyte and macrophage colonization of tumors and associated vasculature and decreased angiogenesis[194].

The α v β 3 integrin is downregulated during differentiation of bone marrow myeloid progenitor cells to monocytes, but induced in macrophages during inflammation[206,207]. α v β 3 promotes myeloid homing and adhesion and migration of bone marrow derived cells through the endothelium to sites of tumor angiogenesis[189]. β 3 integrins are involved in phagocytosis of apoptotic cells[208,209] and limit the secretion of inflammatory

mediators[207]. Defective macrophage tumor infiltration is observed in TAM from $\beta 3$ integrin $^{-/-}$ knockout bone marrow, myeloid specific $\beta 3$ KOM $^{-/-}$ mice and in the signaling defective *DiYF* $\beta 3$ knock-in mice (knock-in mice with two mutated tyrosine residues) [111,189,210–213], suggesting that defective cytoskeletal (re)organization or lack of appropriately polarized macrophages[212] within tumors may be due to $\beta 3$ integrin deficiency.

Myeloid derived suppressor cell (MDSC)[214] are a subpopulation of immature myeloid cells that are roughly characterized by GR1+ and by the $\alpha M\beta 2$ (CD11b) integrin adhesion marker[214]. The MDSC suppress T-cell antigen receptor mediated immune responses[190]. MDSC can promote TAM M2 polarization[215] MDSC from myeloma bearing mice had a greater capacity to become bone resorbing cells compared to MDSC from control mice[191]. The role of integrins in MDSC differentiation, recruitment and function is underway. Thus, integrins are involved in monocyte/macrophage differentiation and recruitment to tumors and can influence local and metastatic tumor growth.

Integrins and tumor recruited platelets and bone metastasis

Cancer cells co-exist with platelets and mononuclear hematopoietic cells in thrombi located throughout the organs of patients with metastatic cancer[216], [217,218]. Platelet aggregation and activation enhances tumor growth and metastasis to bone[77,219]. Platelets are anuclear, metabolically active cells that are formed from bone marrow megakaryocytes. Platelet aggregation is stimulated by soluble factors such as ADP and thromboxane (TXA₂), or membrane proteins collagen or von Willebrand factor that are produced by injured endothelial cells, inflammatory cells and tumor cells. $\alpha_{IIb}\beta 3$ plays a central role in the initiation of arterial thrombosis and platelet aggregation[220,221]. $\alpha_{IIb}\beta 3$ integrins are expressed on the surface of megakaryocytes and platelets and are undetectable on any other non-cancerous cell type. Mice globally deficient for the $\beta 3$ integrin have prolonged bleeding times, defects in platelet aggregation and clot retraction, cutaneous and gastrointestinal bleeding, all characteristics of Glanzmann's thrombasthenia, [222] a disease characterized by functional reduction or absence of $\alpha_{IIb}\beta 3$ in humans. Targeting $\beta 3$ integrins by monoclonal antibodies to the receptor (abciximab/Reopro) or by inhibiting the binding of the ligand fibrinogen to the receptor (tirofiban/Integrilin) are used in patients with acute coronary and cerebral vascular syndromes but have significant bleeding risks that prevent their usefulness for chronic uses such as cancer.

Tumor cell lines have been shown to induce platelet aggregation and adhesion in vitro through mechanisms involving $\alpha_{IIb}\beta 3$ integrin, ADP, thrombin, von Willebrand factor, and selectins[77,223–229]. The metastatic potential of tumor cell lines is markedly diminished in mice with defective platelet aggregation ($\beta 3$ integrin $^{-/-}$, *Gaq* $^{-/-}$, *Par4* $^{-/-}$, *NFE2* $^{-/-}$, and *fibrinogen* $^{-/-}$) [77,219,223,226,228–244]. *3* $^{-/-}$ mice are protected from bone metastasis in part through a mechanism involving defective platelet aggregation[77]. Additionally, tumor cells engineered to respond to platelet-derived lysophosphatidic acid (LPA) have enhanced bone metastatic potential in mice[219]. Platelets also represent a significant source of proangiogenic (VEGF) and antiangiogenic factors (TSP-1) and are recruited to sites of tumor where their aggregation could affect local tumor growth[245]. Platelet specific integrin targeting is a promising therapeutic approach for inhibiting bone metastasis, especially to prevent or slow metastasis.

On the other hand, bone marrow megakaryocytes can inhibit prostate cancer tumor growth in bone[246]. Megakaryocytes can indirectly inhibit bone resorption by inhibiting osteoclast formation[247]. The negative effect of megakaryocytes on bone resorption is likely mediated in part through the osteoclast inhibitory factor, osteoprotegerin, that is contained in

secretory granules of platelets and megakaryocytes[248,249]. Adhesion of mature polyploid megakaryocytes to fibronectin is also mediated by $\beta 1$ subunit containing integrins [250,251]. Megakaryocytes may also influence bone remodeling and resorption through effects on osteoblast proliferation that are mediated by the $\alpha 3\beta 1$, $\alpha 5\beta 1$ and glycoprotein IIb integrins[252]. Given the location of mature megakaryocytes at vascular sinusoids, they are also among the first cells to physically encounter cancer cells as they enter the bone marrow, so that direct mechanism of action involving integrin mediated signal transduction could be involved. Interestingly, bisphosphonates (BP) increase megakaryocyte proliferation and increase the platelet concentration of the anti-angiogenic integrin ligand TSP1[253–255] which suggest non-osteoclast mechanisms of bisphosphonates' impact in decreasing tumor growth in bone. Thus, platelets and their megakaryocytic precursors interact with cancer cells before, during and after metastasis to bone through interactions mainly determined by integrins and their ligands.

Integrins and bone metastasis: Therapeutic aspects

Because of the wide range of functions in physiological and pathological processes, the integrin family of adhesion receptors has been adopted as a promising target for metastatic bone diseases. Several tumor cell types express an abnormal integrin profile compared to non-tumor cells [41,51,256], providing an opportunity for specific targeting. Targeting integrins on both tumor and/or host cells has proven to be effective not only in blocking local cancer progression, but also in reducing tumor cell detachment from their primary site in preclinical models[257–259].

In recent years, integrins on the tumor cells and/or on the endothelium have been targeted by monoclonal antibodies and RGD peptides in order to reduce tumor angiogenesis[109,260]. Integrin antagonists, including humanized monoclonal antibodies, small molecule antagonists, and cyclic peptides, have been developed based on the recognition sequences of integrin physiological ligands[261]. Several compounds are already in clinical use or undergoing their clinical evaluation for various diseases.

For the future treatment of skeletal metastasis, the $\alpha v\beta 3$ integrin has become an attractive target because of its expression in tumor and angiogenic cells, its role in OC differentiation and function, and its role in tumor cell homing to bone[53,60,61,183,262–267]. The multiple expected beneficial effects on endothelial, cancer, and osteoclastic cells instigated a significant effort to develop drug candidates that target the $\alpha v\beta 3$ integrin for therapy of skeletal complications of cancer. These strategies resulted predominantly in antagonists of $\alpha v\beta 3$, $\alpha v\beta 5$, and $\alpha II\beta 3$ integrins that showed efficacy in animal models. Peptidomimetic antagonists of the $\alpha v\beta 3$ and $\alpha v\beta 5$ integrin were successfully used to inhibit OC in vitro and to reduce bone loss in a rat osteoporosis model [268]. An active nonpeptide $\alpha v\beta 3$ integrin antagonist and anti- $\alpha v\beta 3$ antibodies were shown to hinder cancer induced bone loss [79,268–270]. It is possible that the current treatment for bone metastasis, bisphosphonates, may also exert an effect on $\alpha v\beta 3$ on both endothelial cells[132] as well as OCs in a similar way.

Many drugs candidates targeting integrin $\alpha v\beta 3$ have advanced to the clinics for the treatment of osteoporosis and cancer, though none have specifically targeted patients with bone metastases. A lipophilic isoester of RGD (L000845704), developed by Merck, is effective in increasing bone mineral density (BMD) in postmenopausal women[271]. Another inhibitor, RGD-mimetic cyclic peptide Cilengitide (EMD-1219974) directed at both $\alpha v\beta 3$ and $\alpha v\beta 5$ [272] and currently produced by MerckSerono, is in advanced stages of clinical testing for the treatment of glioblastoma multiforme and is under investigation for the treatment of squamous cell carcinoma, prostate cancer, and lung cancer (Phase II).

Clinical trials of function-blocking antibodies are also ongoing, including Vitaxin (LM609), a humanized monoclonal IgG₁ antibody against the extracellular domain of the $\alpha v\beta 3$ integrin heterodimer. Vitaxin had substantial anti-angiogenic effects in preclinical models [119,262] and has shown direct anti-tumor effects as well as impaired bone resorption by inhibiting OC attachment to the bone surface [273]. Another monoclonal antibody (CNT095), directed against the αv subunit, is under development by Centocor and is in phase I–II testing for solid tumors. Two other additions to this therapeutic family are planned to be more specifically evaluated for their effects on bone metastasis [62], organic small molecule GLPG0187 [62] and peptide antagonist S247 [257].

Given the participation of the osteoclasts, blood vessels and platelets in bone metastases, it may be beneficial to block both $\alpha v\beta 3$ and $\alpha II\beta 3$ integrins on host cells. This concept of combination inhibition relies on the common RGD ligand binding domains of $\alpha v\beta 3$, $\alpha v\beta 5$, and $\alpha II\beta 3$. In fact, many of the synthetically designed $\alpha v\beta 3$ integrin inhibitors display some selectivity towards $\alpha v\beta 5$ integrin, and, in the case of Cilengitide, this dual antagonism is part of the mechanism to treat cancer by inhibiting neoangiogenesis as well as invasion [274,275]. The strategy to combine multiple targets also bears some risks with regards to the desired high therapeutic specificity and low off-target toxicity. This issue is further complicated by the differential function of the integrins as determined by their location, expression level, activation status, and ligand binding. Studies in animal models and xenograft tumor models have demonstrated that low concentrations of $\alpha v\beta 3$ integrin antagonists can act as integrin agonists [130,276,277]. Further research is necessary to identify optimal drug dosing and targeting that overcome the problem of generalized integrin inhibition to reduce or prevent skeletal metastasis.

Another area of active research in bone metastasis therapeutics is the specific targeting of integrins on hematopoietic stem cells or progenitors that prepare the metastatic niche and enhance bone marrow colonization by cancer cells which then instigate the vicious cycle of bone metastasis [278,279]. Interfering with integrin mediated homing of cancer cells to the bone or to hijack the bone cells represents an early option for intervention. siRNA against the αv integrin subunit was used to prevent the progression of prostate cancer to bone by interfering with the ECM-integrin interaction [280]. In another approach, a disintegrin and a neutralizing antibody to VCAM-1 or its receptor $\alpha 4\beta 1$ integrin reduced metastasis of melanoma cells and diminished osteolysis by decreasing osteoclast activity in a myeloma in vitro model [69,281]. These strategies, however, are not yet in clinical trials. An exciting new approach to cancer therapy takes advantage of the fact that cancer cells use CXCR4 and VLA-4 to home to and “engraft” in the marrow. HSC mobilizing agents such as AMD3100 and anti-VLA-4 targeted agents can be used to mobilize leukemia and myeloma cells into the blood from the bone marrow which leads to increased sensitivity to chemotherapy [282–284] in mice. This approach is now being tried in clinical trials.

Future perspectives

Despite the high level of complexity of the integrin family, the $\beta 3$ integrin remains a major target in the search for effective therapies for skeletal metastasis. In recent years, a steady increase in knowledge has led several interesting compounds into clinical testing. There remains, however, a lack of clarity concerning the exact roles of the integrins in different cell types. In the initiated clinical studies using $\alpha v\beta 3$ integrin antagonists, the overall effect in reducing tumor growth and pathological angiogenesis in fast progressing deadly tumors may outweigh potential undesired effects in tissues or cells other than tumor or endothelial origin. Drugs designed to tackle skeletal complications of cancer must be “targeted” to the bone microenvironment as underscored by the clinical successes of the bone matrix targeted bisphosphonates and the osteoclast targeted denosumab in treating and preventing skeletal

complications of bone metastases and myeloma. Because of the complexity of cells recruited in the tumor microenvironment and the pro-and anti-tumor effects of integrins depending on the cellular context, a detailed understanding of the role of integrin regulation in both the metastatic tumor cells and the tumor-associated stroma will allow for a more targeted and focused approach to treat bone metastases.

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References

1. Lipton A. Pathophysiology of bone metastases: how this knowledge may lead to therapeutic intervention. *J Support Oncol* 2004;2:205–13. discussion 213–4, 216–7, 219–20. [PubMed: 15328823]
2. Coleman RE. Metastatic bone disease: clinical features, pathophysiology and treatment strategies. *Cancer Treat Rev* 2001;27:165–76. [PubMed: 11417967]
3. Mundy GR. Metastasis to bone: causes, consequences and therapeutic opportunities. *Nat Rev Cancer* 2002;2:584–93. [PubMed: 12154351]
4. Roodman GD. Mechanisms of bone metastasis. *N Engl J Med* 2004;350:1655–64. [PubMed: 15084698]
5. Lipton A, Costa L, Ali S, Demers L. Use of markers of bone turnover for monitoring bone metastases and the response to therapy. *Semin Oncol* 2001;28:54–9. [PubMed: 11544577]
6. Coleman RE, Major P, Lipton A, Brown JE, Lee KA, Smith M, Saad F, Zheng M, Hei YJ, Seaman J, Cook R. Predictive value of bone resorption and formation markers in cancer patients with bone metastases receiving the bisphosphonate zoledronic acid. *J Clin Oncol* 2005;23:4925–35. [PubMed: 15983391]
7. Guise TA, Mohammad KS, Clines G, Stebbins EG, Wong DH, Higgins LS, Vessella R, Corey E, Padalecki S, Suva L, Chirgwin JM. Basic mechanisms responsible for osteolytic and osteoblastic bone metastases. *Clin Cancer Res* 2006;12:6213s–6216s. [PubMed: 17062703]
8. Hirbe A, Morgan EA, Uluckan O, Weilbaecher K. Skeletal complications of breast cancer therapies. *Clin Cancer Res* 2006;12:6309s–6314s. [PubMed: 17062720]
9. Hirbe AC, Rubin J, Uluckan O, Morgan EA, Eagleton MC, Prior JL, Piwnica-Worms D, Weilbaecher KN. Disruption of CXCR4 enhances osteoclastogenesis and tumor growth in bone. *Proc Natl Acad Sci U S A* 2007;104:14062–7. [PubMed: 17715292]
10. Bendre MS, Montague DC, Peery T, Akel NS, Gaddy D, Suva LJ. Interleukin-8 stimulation of osteoclastogenesis and bone resorption is a mechanism for the increased osteolysis of metastatic bone disease. *Bone* 2003;33:28–37. [PubMed: 12919697]
11. Pfeilschifter J, D'Souza SM, Mundy GR. Effects of transforming growth factor-beta on osteoblastic osteosarcoma cells. *Endocrinology* 1987;121:212–8. [PubMed: 3474142]
12. Kozlow W, Guise TA. Breast cancer metastasis to bone: mechanisms of osteolysis and implications for therapy. *J Mammary Gland Biol Neoplasia* 2005;10:169–80. [PubMed: 16025223]
13. Kingsley LA, Fournier PG, Chirgwin JM, Guise TA. Molecular biology of bone metastasis. *Mol Cancer Ther* 2007;6:2609–17. [PubMed: 17938257]
14. Clines GA, Guise TA. Molecular mechanisms and treatment of bone metastasis. *Expert Rev Mol Med* 2008;10:e7. [PubMed: 18321396]
15. Yin JJ, Selander K, Chirgwin JM, Dallas M, Grubbs BG, Wieser R, Massague J, Mundy GR, Guise TA. TGF-beta signaling blockade inhibits PTHrP secretion by breast cancer cells and bone metastases development. *J Clin Invest* 1999;103:197–206. [PubMed: 9916131]
16. Kakonen SM, Selander KS, Chirgwin JM, Yin JJ, Burns S, Rankin WA, Grubbs BG, Dallas M, Cui Y, Guise TA. Transforming growth factor-beta stimulates parathyroid hormone-related protein

- and osteolytic metastases via Smad and mitogen-activated protein kinase signaling pathways. *J Biol Chem* 2002;277:24571–8. [PubMed: 11964407]
17. Hamdy NA. Denosumab: RANKL inhibition in the management of bone loss. *Drugs Today (Barc)* 2008;44:7–21. [PubMed: 18301800]
 18. Body JJ, Lipton A, Gralow J, Steger GG, Gao G, Yeh H, Fizazi K. Effects of Denosumab in Patients with Bone Metastases, with and without Previous Bisphosphonate Exposure. *J Bone Miner Res.* 2009
 19. Fizazi K, Lipton A, Mariette X, Body JJ, Rahim Y, Gralow JR, Gao G, Wu L, Sohn W, Jun S. Randomized phase II trial of denosumab in patients with bone metastases from prostate cancer, breast cancer, or other neoplasms after intravenous bisphosphonates. *J Clin Oncol* 2009;27:1564–71. [PubMed: 19237632]
 20. Schwartz MA, Schaller MD, Ginsberg MH. Integrins: emerging paradigms of signal transduction. *Annu Rev Cell Dev Biol* 1995;11:549–99. [PubMed: 8689569]
 21. Hynes RO. Integrins: versatility, modulation, and signaling in cell adhesion. *Cell* 1992;69:11–25. [PubMed: 1555235]
 22. Hynes RO. Integrins: bidirectional, allosteric signaling machines. *Cell* 2002;110:673–87. [PubMed: 12297042]
 23. Shattil SJ, Kim C, Ginsberg MH. The final steps of integrin activation: the end game. *Nat Rev Mol Cell Biol* 2010;11:288–300. [PubMed: 20308986]
 24. Qin J, Vinogradova O, Plow EF. Integrin bidirectional signaling: a molecular view. *PLoS Biol* 2004;2:e169. [PubMed: 15208721]
 25. Offermanns S. Activation of platelet function through G protein-coupled receptors. *Circ Res* 2006;99:1293–304. [PubMed: 17158345]
 26. Ma YQ, Qin J, Wu C, Plow EF. Kindlin-2 (Mig-2): a co-activator of beta3 integrins. *J Cell Biol* 2008;181:439–46. [PubMed: 18458155]
 27. Tadokoro S, Shattil SJ, Eto K, Tai V, Liddington RC, de Pereda JM, Ginsberg MH, Calderwood DA. Talin binding to integrin beta tails: a final common step in integrin activation. *Science* 2003;302:103–6. [PubMed: 14526080]
 28. Vinogradova O, Velyvis A, Velyviene A, Hu B, Haas T, Plow E, Qin J. A structural mechanism of integrin alpha(IIb)beta(3) “inside-out” activation as regulated by its cytoplasmic face. *Cell* 2002;110:587–97. [PubMed: 12230976]
 29. Harburger DS, Calderwood DA. Integrin signalling at a glance. *J Cell Sci* 2009;122:159–63. [PubMed: 19118207]
 30. Montanez E, Ussar S, Schifferer M, Bosl M, Zent R, Moser M, Fassler R. Kindlin-2 controls bidirectional signaling of integrins. *Genes Dev* 2008;22:1325–30. [PubMed: 18483218]
 31. Moser M, Nieswandt B, Ussar S, Pozgajova M, Fassler R. Kindlin-3 is essential for integrin activation and platelet aggregation. *Nat Med* 2008;14:325–30. [PubMed: 18278053]
 32. Lai-Cheong JE, Parsons M, McGrath JA. The role of kindlins in cell biology and relevance to human disease. *Int J Biochem Cell Biol* 2010;42:595–603. [PubMed: 19854292]
 33. Kamae T, Shiraga M, Kashiwagi H, Kato H, Tadokoro S, Kurata Y, Tomiyama Y, Kanakura Y. Critical role of ADP interaction with P2Y12 receptor in the maintenance of alpha(IIb)beta3 activation: association with Rap1B activation. *J Thromb Haemost* 2006;4:1379–87. [PubMed: 16706985]
 34. Woulfe D, Jiang H, Mortensen R, Yang J, Brass LF. Activation of Rap1B by G(i) family members in platelets. *J Biol Chem* 2002;277:23382–90. [PubMed: 11970953]
 35. Chen YP, O’Toole TE, Ylance J, Rosa JP, Ginsberg MH. A point mutation in the integrin beta 3 cytoplasmic domain (S752-->P) impairs bidirectional signaling through alpha IIb beta 3 (platelet glycoprotein IIb-IIIa). *Blood* 1994;84:1857–65. [PubMed: 8080992]
 36. Hughes PE, Diaz-Gonzalez F, Leong L, Wu C, McDonald JA, Shattil SJ, Ginsberg MH. Breaking the integrin hinge. A defined structural constraint regulates integrin signaling. *J Biol Chem* 1996;271:6571–4. [PubMed: 8636068]
 37. O’Toole TE, Katagiri Y, Faull RJ, Peter K, Tamura R, Quaranta V, Loftus JC, Shattil SJ, Ginsberg MH. Integrin cytoplasmic domains mediate inside-out signal transduction. *J Cell Biol* 1994;124:1047–59. [PubMed: 7510712]

38. Vinogradova O, Haas T, Plow EF, Qin J. A structural basis for integrin activation by the cytoplasmic tail of the alpha IIb-subunit. *Proc Natl Acad Sci U S A* 2000;97:1450–5. [PubMed: 10677482]
39. Vinogradova O, Vaynberg J, Kong X, Haas TA, Plow EF, Qin J. Membrane-mediated structural transitions at the cytoplasmic face during integrin activation. *Proc Natl Acad Sci U S A* 2004;101:4094–9. [PubMed: 15024114]
40. Ylanne J, Huuskonen J, O'Toole TE, Ginsberg MH, Virtanen I, Gahmberg CG. Mutation of the cytoplasmic domain of the integrin beta 3 subunit. Differential effects on cell spreading, recruitment to adhesion plaques, endocytosis, and phagocytosis. *J Biol Chem* 1995;270:9550–7. [PubMed: 7721884]
41. Desgrosellier JS, Cheresh DA. Integrins in cancer: biological implications and therapeutic opportunities. *Nat Rev Cancer* 10:9–22. [PubMed: 20029421]
42. Brown EJ, Frazier WA. Integrin-associated protein (CD47) and its ligands. *Trends Cell Biol* 2001;11:130–5. [PubMed: 11306274]
43. Frisch SM, Screaton RA. Anoikis mechanisms. *Curr Opin Cell Biol* 2001;13:555–62. [PubMed: 11544023]
44. Bissell MJ, Radisky D. Putting tumours in context. *Nat Rev Cancer* 2001;1:46–54. [PubMed: 11900251]
45. Frisch SM, Vuori K, Ruoslahti E, Chan-Hui PY. Control of adhesion-dependent cell survival by focal adhesion kinase. *J Cell Biol* 1996;134:793–9. [PubMed: 8707856]
46. Demers MJ, Thibodeau S, Noel D, Fujita N, Tsuruo T, Gauthier R, Arguin M, Vachon PH. Intestinal epithelial cancer cell anoikis resistance: EGFR-mediated sustained activation of Src overrides Fak-dependent signaling to MEK/Erk and/or PI3-K/Akt-1. *J Cell Biochem* 2009;107:639–54. [PubMed: 19479902]
47. Shain KH, Landowski TH, Dalton WS. Adhesion-mediated intracellular redistribution of c-Fas-associated death domain-like IL-1-converting enzyme-like inhibitory protein-long confers resistance to CD95-induced apoptosis in hematopoietic cancer cell lines. *J Immunol* 2002;168:2544–53. [PubMed: 11859150]
48. Simpson KJ, Selfors LM, Bui J, Reynolds A, Leake D, Khvorova A, Brugge JS. Identification of genes that regulate epithelial cell migration using an siRNA screening approach. *Nat Cell Biol* 2008;10:1027–38. [PubMed: 19160483]
49. Edlund M, Miyamoto T, Sikes RA, Ogle R, Laurie GW, Farach-Carson MC, Otey CA, Zhau HE, Chung LW. Integrin expression and usage by prostate cancer cell lines on laminin substrata. *Cell Growth Differ* 2001;12:99–107. [PubMed: 11243469]
50. Yoneda T. Cellular and molecular basis of preferential metastasis of breast cancer to bone. *J Orthop Sci* 2000;5:75–81. [PubMed: 10664443]
51. Clezardin P. Integrins in bone metastasis formation and potential therapeutic implications. *Curr Cancer Drug Targets* 2009;9:801–6. [PubMed: 20025568]
52. van der P, Vloedgraven H, Papapoulos S, Lowick C, Grzesik W, Kerr J, Robey PG. Attachment characteristics and involvement of integrins in adhesion of breast cancer cell lines to extracellular bone matrix components. *Lab Invest* 1997;77:665–75. [PubMed: 9426405]
53. Liapis H, Flath A, Kitazawa S. Integrin alpha V beta 3 expression by bone-residing breast cancer metastases. *Diagn Mol Pathol* 1996;5:127–35. [PubMed: 8727100]
54. McCabe NP, De S, Vasanji A, Brainard J, Byzova TV. Prostate cancer specific integrin alphavbeta3 modulates bone metastatic growth and tissue remodeling. *Oncogene* 2007;26:6238–43. [PubMed: 17369840]
55. Townsend PA, Villanova I, Uhlmann E, Peyman A, Knolle J, Baron R, Teti A, Horton MA. An antisense oligonucleotide targeting the alphaV integrin gene inhibits adhesion and induces apoptosis in breast cancer cells. *Eur J Cancer* 2000;36:397–409. [PubMed: 10708943]
56. Gillespie MT, Thomas RJ, Pu ZY, Zhou H, Martin TJ, Findlay DM. Calcitonin receptors, bone sialoprotein and osteopontin are expressed in primary breast cancers. *Int J Cancer* 1997;73:812–5. [PubMed: 9399657]
57. Van der Velde-Zimmermann D, Verdaasdonk MA, Rademakers LH, De Weger RA, Van den Tweel JG, Joling P. Fibronectin distribution in human bone marrow stroma: matrix assembly and

- tumor cell adhesion via alpha5 beta1 integrin. *Exp Cell Res* 1997;230:111–20. [PubMed: 9013713]
58. Takayama S, Ishii S, Ikeda T, Masamura S, Doi M, Kitajima M. The relationship between bone metastasis from human breast cancer and integrin alpha(v)beta3 expression. *Anticancer Res* 2005;25:79–83. [PubMed: 15816522]
 59. Nakamura I, Duong le T, Rodan SB, Rodan GA. Involvement of alpha(v)beta3 integrins in osteoclast function. *J Bone Miner Metab* 2007;25:337–44. [PubMed: 17968485]
 60. Pecheur I, Peyruchaud O, Serre CM, Guglielmi J, Voland C, Bourre F, Margue C, Cohen-Solal M, Buffet A, Kieffer N, Clezardin P. Integrin alpha(v)beta3 expression confers on tumor cells a greater propensity to metastasize to bone. *FASEB J* 2002;16:1266–8. [PubMed: 12153995]
 61. Sloan EK, Pouliot N, Stanley KL, Chia J, Moseley JM, Hards DK, Anderson RL. Tumor-specific expression of alphavbeta3 integrin promotes spontaneous metastasis of breast cancer to bone. *Breast Cancer Res* 2006;8:R20. [PubMed: 16608535]
 62. Zhao Y, Bachelier R, Treilleux I, Pujuguet P, Peyruchaud O, Baron R, Clement-Lacroix P, Clezardin P. Tumor alphavbeta3 integrin is a therapeutic target for breast cancer bone metastases. *Cancer Res* 2007;67:5821–30. [PubMed: 17575150]
 63. Martin-Thouvenin V, Gendron MC, Hogervorst F, Figdor CG, Lanotte M. Phorbol ester-induced promyelocytic leukemia cell adhesion to marrow stromal cells involves fibronectin specific alpha 5 beta 1 integrin receptors. *J Cell Physiol* 1992;153:95–102. [PubMed: 1387876]
 64. Korah R, Boots M, Wieder R. Integrin alpha5beta1 promotes survival of growth-arrested breast cancer cells: an in vitro paradigm for breast cancer dormancy in bone marrow. *Cancer Res* 2004;64:4514–22. [PubMed: 15231661]
 65. Liesveld JL, Dipersio JF, Abboud CN. Integrins and adhesive receptors in normal and leukemic CD34+ progenitor cells: potential regulatory checkpoints for cellular traffic. *Leuk Lymphoma* 1994;14:19–28. [PubMed: 7522718]
 66. Lang SH, Clarke NW, George NJ, Testa NG. Primary prostatic epithelial cell binding to human bone marrow stroma and the role of alpha2beta1 integrin. *Clin Exp Metastasis* 1997;15:218–27. [PubMed: 9174123]
 67. Hall CL, Dai J, van Golen KL, Keller ET, Long MW. Type I collagen receptor (alpha 2 beta 1) signaling promotes the growth of human prostate cancer cells within the bone. *Cancer Res* 2006;66:8648–54. [PubMed: 16951179]
 68. Hall CL, DUBYK CW, Riesenberger TA, Shein D, Keller ET, van Golen KL. Type I collagen receptor (alpha2beta1) signaling promotes prostate cancer invasion through RhoC GTPase. *Neoplasia* 2008;10:797–803. [PubMed: 18670640]
 69. Mori Y, Shimizu N, Dallas M, Niewolna M, Story B, Williams PJ, Mundy GR, Yoneda T. Anti-alpha4 integrin antibody suppresses the development of multiple myeloma and associated osteoclastic osteolysis. *Blood* 2004;104:2149–54. [PubMed: 15138161]
 70. Michigami T, Shimizu N, Williams PJ, Niewolna M, Dallas SL, Mundy GR, Yoneda T. Cell-cell contact between marrow stromal cells and myeloma cells via VCAM-1 and alpha(4)beta(1)-integrin enhances production of osteoclast-stimulating activity. *Blood* 2000;96:1953–60. [PubMed: 10961900]
 71. Matsuura N, Puzon-McLaughlin W, Irie A, Morikawa Y, Kakudo K, Takada Y. Induction of experimental bone metastasis in mice by transfection of integrin alpha 4 beta 1 into tumor cells. *Am J Pathol* 1996;148:55–61. [PubMed: 8546226]
 72. Teitelbaum SL, Ross FP. Genetic regulation of osteoclast development and function. *Nat Rev Genet* 2003;4:638–49. [PubMed: 12897775]
 73. Ross FP, Teitelbaum SL. alphavbeta3 and macrophage colony-stimulating factor: partners in osteoclast biology. *Immunol Rev* 2005;208:88–105. [PubMed: 16313343]
 74. Novack DV, Teitelbaum SL. The osteoclast: friend or foe? *Annu Rev Pathol* 2008;3:457–84. [PubMed: 18039135]
 75. Ross FP, Chappel J, Alvarez JI, Sander D, Butler WT, Farach-Carson MC, Mintz KA, Robey PG, Teitelbaum SL, Cheresch DA. Interactions between the bone matrix proteins osteopontin and bone sialoprotein and the osteoclast integrin alpha v beta 3 potentiate bone resorption. *J Biol Chem* 1993;268:9901–7. [PubMed: 8486670]

76. McHugh KP. Mice lacking b3 integrins are osteosclerotic because of dysfunctional osteoclasts. *Journal of Clinical Investigation* 2000;105:433–440. [PubMed: 10683372]
77. Bakewell SJ, Nestor P, Prasad S, Tomasson MH, Dowland N, Mehrotra M, Scarborough R, Kanter J, Abe K, Phillips D, Weilbaecher KN. Platelet and osteoclast beta3 integrins are critical for bone metastasis. *Proc Natl Acad Sci U S A* 2003;100:14205–10. [PubMed: 14612570]
78. Zambonin Zallone A, Teti A, Gaboli M, Marchisio PC. Beta 3 subunit of vitronectin receptor is present in osteoclast adhesion structures and not in other monocyte-macrophage derived cells. *Connect Tissue Res* 1989;20:143–9. [PubMed: 2482152]
79. Crippes BA, Engleman VW, Settle SL, Delarco J, Ornberg RL, Helfrich MH, Horton MA, Nickols GA. Antibody to beta3 integrin inhibits osteoclast-mediated bone resorption in the thyroparathyroidectomized rat. *Endocrinology* 1996;137:918–24. [PubMed: 8603604]
80. Chellaiah MA. Regulation of podosomes by integrin alphavbeta3 and Rho GTPase-facilitated phosphoinositide signaling. *Eur J Cell Biol* 2006;85:311–7. [PubMed: 16460838]
81. Faccio R, Takeshita S, Zallone A, Ross FP, Teitelbaum SL. c-Fms and the alphavbeta3 integrin collaborate during osteoclast differentiation. *J Clin Invest* 2003;111:749–58. [PubMed: 12618529]
82. Faccio R, Grano M, Colucci S, Zallone AZ, Quaranta V, Pelletier AJ. Activation of alphav beta3 integrin on human osteoclast-like cells stimulates adhesion and migration in response to osteopontin. *Biochem Biophys Res Commun* 1998;249:522–5. [PubMed: 9712729]
83. Miyachi A, Alvarez J, Greenfield EM, Teti A, Grano M, Colucci S, Zambonin-Zallone A, Ross FP, Teitelbaum SL, Cheresch D, et al. Recognition of osteopontin and related peptides by an alpha v beta 3 integrin stimulates immediate cell signals in osteoclasts. *J Biol Chem* 1991;266:20369–74. [PubMed: 1939092]
84. Rucci N, DiGiacinto C, Orru L, Millimaggi D, Baron R, Teti A. A novel protein kinase C alpha-dependent signal to ERK1/2 activated by alphaVbeta3 integrin in osteoclasts and in Chinese hamster ovary (CHO) cells. *J Cell Sci* 2005;118:3263–75. [PubMed: 16014375]
85. Zhang Z, Baron R, Horne WC. Integrin engagement, the actin cytoskeleton, and c-Src are required for the calcitonin-induced tyrosine phosphorylation of paxillin and HEF1, but not for calcitonin-induced Erk1/2 phosphorylation. *J Biol Chem* 2000;275:37219–23. [PubMed: 10954702]
86. Clohisy DR, Ramnaraine ML. Osteoclasts are required for bone tumors to grow and destroy bone. *J Orthop Res* 1998;16:660–6. [PubMed: 9877389]
87. Honore P, Luger NM, Sabino MA, Schwei MJ, Rogers SD, Mach DB, O'Keefe PF, Ramnaraine ML, Clohisy DR, Mantyh PW. Osteoprotegerin blocks bone cancer-induced skeletal destruction, skeletal pain and pain-related neurochemical reorganization of the spinal cord. *Nat Med* 2000;6:521–8. [PubMed: 10802707]
88. Carlinfante G, Vassiliou D, Svensson O, Wendel M, Heinegard D, Andersson G. Differential expression of osteopontin and bone sialoprotein in bone metastasis of breast and prostate carcinoma. *Clin Exp Metastasis* 2003;20:437–44. [PubMed: 14524533]
89. Heinegard D, Andersson G, Reinholt FP. Roles of osteopontin in bone remodeling. *Ann N Y Acad Sci* 1995;760:213–22. [PubMed: 7540374]
90. Katayama Y, House CM, Udagawa N, Kazama JJ, McFarland RJ, Martin TJ, Findlay DM. Casein kinase 2 phosphorylation of recombinant rat osteopontin enhances adhesion of osteoclasts but not osteoblasts. *J Cell Physiol* 1998;176:179–87. [PubMed: 9618157]
91. Hayashi C, Rittling S, Hayata T, Amagasa T, Denhardt D, Ezura Y, Nakashima K, Noda M. Serum osteopontin, an enhancer of tumor metastasis to bone, promotes B16 melanoma cell migration. *J Cell Biochem* 2007;101:979–86. [PubMed: 17390343]
92. Denhardt DT, Chambers AF. Overcoming obstacles to metastasis --defenses against host defenses: osteopontin (OPN) as a shield against attack by cytotoxic host cells. *J Cell Biochem* 1994;56:48–51. [PubMed: 7528752]
93. Nemoto H, Rittling SR, Yoshitake H, Furuya K, Amagasa T, Tsuji K, Nifuji A, Denhardt DT, Noda M. Osteopontin deficiency reduces experimental tumor cell metastasis to bone and soft tissues. *J Bone Miner Res* 2001;16:652–9. [PubMed: 11315992]
94. Ohyama Y, Nemoto H, Rittling S, Tsuji K, Amagasa T, Denhardt DT, Nifuji A, Noda M. Osteopontin-deficiency suppresses growth of B16 melanoma cells implanted in bone and osteoclastogenesis in co-cultures. *J Bone Miner Res* 2004;19:1706–11. [PubMed: 15355566]

95. Desai B, Rogers MJ, Chellaiah MA. Mechanisms of osteopontin and CD44 as metastatic principles in prostate cancer cells. *Mol Cancer* 2007;6:18. [PubMed: 17343740]
96. Hullinger TG, Taichman RS, Linseman DA, Somerman MJ. Secretory products from PC-3 and MCF-7 tumor cell lines upregulate osteopontin in MC3T3-E1 cells. *J Cell Biochem* 2000;78:607–16. [PubMed: 10861858]
97. Abe M, Hiura K, Wilde J, Shioyasono A, Moriyama K, Hashimoto T, Kido S, Oshima T, Shibata H, Ozaki S, Inoue D, Matsumoto T. Osteoclasts enhance myeloma cell growth and survival via cell-cell contact: a vicious cycle between bone destruction and myeloma expansion. *Blood* 2004;104:2484–91. [PubMed: 15187021]
98. Standal T, Borset M, Sundan A. Role of osteopontin in adhesion, migration, cell survival and bone remodeling. *Exp Oncol* 2004;26:179–84. [PubMed: 15494684]
99. Chen YJ, Wei YY, Chen HT, Fong YC, Hsu CJ, Tsai CH, Hsu HC, Liu SH, Tang CH. Osteopontin increases migration and MMP-9 up-regulation via alphavbeta3 integrin, FAK, ERK, and NF-kappaB-dependent pathway in human chondrosarcoma cells. *J Cell Physiol* 2009;221:98–108. [PubMed: 19475568]
100. Lundberg P, Koskinen C, Baldock PA, Lothgren H, Stenberg A, Lerner UH, Oldenborg PA. Osteoclast formation is strongly reduced both in vivo and in vitro in the absence of CD47/SIRPalpha-interaction. *Biochem Biophys Res Commun* 2007;352:444–8. [PubMed: 17126807]
101. Uluckan O, Becker SN, Deng H, Zou W, Prior JL, Piwnica-Worms D, Frazier WA, Weilbaecher KN. CD47 regulates bone mass and tumor metastasis to bone. *Cancer Res* 2009;69:3196–204. [PubMed: 19276363]
102. Han X, Sterling H, Chen Y, Saginario C, Brown EJ, Frazier WA, Lindberg FP, Vignery A. CD47, a ligand for the macrophage fusion receptor, participates in macrophage multinucleation. *J Biol Chem* 2000;275:37984–92. [PubMed: 10964914]
103. Rachkovsky M, Sodi S, Chakraborty A, Avissar Y, Bolognia J, McNiff JM, Platt J, Bermudes D, Pawelek J. Melanoma × macrophage hybrids with enhanced metastatic potential. *Clin Exp Metastasis* 1998;16:299–312. [PubMed: 9626809]
104. Vignery A. Macrophage fusion: are somatic and cancer cells possible partners? *Trends Cell Biol* 2005;15:188–93. [PubMed: 15817374]
105. Hanahan D, Weinberg RA. The hallmarks of cancer. *Cell* 2000;100:57–70. [PubMed: 10647931]
106. Stupack DG, Cheresh DA. Integrins and angiogenesis. *Curr Top Dev Biol* 2004;64:207–38. [PubMed: 15563949]
107. Hood JD, Cheresh DA. Role of integrins in cell invasion and migration. *Nat Rev Cancer* 2002;2:91–100. [PubMed: 12635172]
108. Hodivala-Dilke K. alphavbeta3 integrin and angiogenesis: a moody integrin in a changing environment. *Curr Opin Cell Biol* 2008;20:514–9. [PubMed: 18638550]
109. Silva R, D'Amico G, Hodivala-Dilke KM, Reynolds LE. Integrins: the keys to unlocking angiogenesis. *Arterioscler Thromb Vasc Biol* 2008;28:1703–13. [PubMed: 18658045]
110. Stockmann C, Doedens A, Weidemann A, Zhang N, Takeda N, Greenberg JI, Cheresh DA, Johnson RS. Deletion of vascular endothelial growth factor in myeloid cells accelerates tumorigenesis. *Nature* 2008;456:814–8. [PubMed: 18997773]
111. Morgan EA, Schneider J, TB, Uluckan O, Heller EA, Hurchla MA, Deng H, Floyd DH, Berdy A, Prior JL, Piwnica-Worms D, Teitelbaum SL, FPR, Weilbaecher K. Dissection of platelet and myeloid cell defects by conditional targeting of the $\beta 3$ integrin subunit. *FASEB*. 2009 in press.
112. Cackowski FC, Anderson JL, Patrene KD, Choksi RJ, Shapiro SD, Windle JJ, Blair HC, Roodman GD. Osteoclasts are important for bone angiogenesis. *Blood* 115:140–9. [PubMed: 19887675]
113. Capoccia BJ, Shepherd RM, Link DC. G-CSF and AMD3100 mobilize monocytes into the blood that stimulate angiogenesis in vivo through a paracrine mechanism. *Blood* 2006;108:2438–45. [PubMed: 16735597]
114. De Palma M, Naldini L. Role of haematopoietic cells and endothelial progenitors in tumour angiogenesis. *Biochim Biophys Acta* 2006;1766:159–66. [PubMed: 16857321]

115. Papaspyridonos M, Lyden D. Chapter 11. The role of bone marrow-derived cells in tumor angiogenesis and metastatic progression. *Methods Enzymol* 2008;444:255–69. [PubMed: 19007668]
116. Ramjaun AR, Hodivala-Dilke K. The role of cell adhesion pathways in angiogenesis. *Int J Biochem Cell Biol* 2009;41:521–30. [PubMed: 18762270]
117. Brooks PC, Clark RA, Cheresh DA. Requirement of vascular integrin alpha v beta 3 for angiogenesis. *Science* 1994;264:569–71. [PubMed: 7512751]
118. Mahabeshwar GH, Byzova TV. Vascular integrin signaling. *Methods Enzymol* 2008;443:199–226. [PubMed: 18772018]
119. Brooks PC, Montgomery AM, Rosenfeld M, Reisfeld RA, Hu T, Klier G, Cheresh DA. Integrin alpha v beta 3 antagonists promote tumor regression by inducing apoptosis of angiogenic blood vessels. *Cell* 1994;79:1157–64. [PubMed: 7528107]
120. Brooks PC, Silletti S, von Schalscha TL, Friedlander M, Cheresh DA. Disruption of angiogenesis by PEX, a noncatalytic metalloproteinase fragment with integrin binding activity. *Cell* 1998;92:391–400. [PubMed: 9476898]
121. Silletti S, Kessler T, Goldberg J, Boger DL, Cheresh DA. Disruption of matrix metalloproteinase 2 binding to integrin alpha v beta 3 by an organic molecule inhibits angiogenesis and tumor growth in vivo. *Proc Natl Acad Sci U S A* 2001;98:119–24. [PubMed: 11134507]
122. Maeshima Y, Yerramalla UL, Dhanabal M, Holthaus KA, Barbashov S, Kharbanda S, Reimer C, Manfredi M, Dickerson WM, Kalluri R. Extracellular matrix-derived peptide binds to alpha(v)beta(3) integrin and inhibits angiogenesis. *J Biol Chem* 2001;276:31959–68. [PubMed: 11399763]
123. Nemeth JA, Cher ML, Zhou Z, Mullins C, Bhagat S, Trikha M. Inhibition of alpha(v)beta3 integrin reduces angiogenesis, bone turnover, and tumor cell proliferation in experimental prostate cancer bone metastases. *Clin Exp Metastasis* 2003;20:413–20. [PubMed: 14524530]
124. Reynolds LE, Wyder L, Lively JC, Taverna D, Robinson SD, Huang X, Sheppard D, Hynes RO, Hodivala-Dilke KM. Enhanced pathological angiogenesis in mice lacking beta3 integrin or beta3 and beta5 integrins. *Nat Med* 2002;8:27–34. [PubMed: 11786903]
125. Reynolds AR, Reynolds LE, Nagel TE, Lively JC, Robinson SD, Hicklin DJ, Bodary SC, Hodivala-Dilke KM. Elevated Flk1 (vascular endothelial growth factor receptor 2) signaling mediates enhanced angiogenesis in beta3-integrin-deficient mice. *Cancer Res* 2004;64:8643–50. [PubMed: 15574772]
126. Carmeliet P. Integrin indecision. *Nat Med* 2002;8:14–6. [PubMed: 11786895]
127. Martin KH, Slack JK, Boerner SA, Martin CC, Parsons JT. Integrin connections map: to infinity and beyond. *Science* 2002;296:1652–3. [PubMed: 12040184]
128. Wang XQ, Sun P, Paller AS. Inhibition of integrin-linked kinase/protein kinase B/Akt signaling: mechanism for ganglioside-induced apoptosis. *J Biol Chem* 2001;276:44504–11. [PubMed: 11577096]
129. Zhao H, Ross FP, Teitelbaum SL. Unoccupied alpha(v)beta3 integrin regulates osteoclast apoptosis by transmitting a positive death signal. *Mol Endocrinol* 2005;19:771–80. [PubMed: 15591537]
130. Reynolds AR, Hart IR, Watson AR, Welti JC, Silva RG, Robinson SD, Da Violante G, Gourlaouen M, Salih M, Jones MC, Jones DT, Saunders G, Kostourou V, Perron-Sierra F, Norman JC, Tucker GC, Hodivala-Dilke KM. Stimulation of tumor growth and angiogenesis by low concentrations of RGD-mimetic integrin inhibitors. *Nat Med* 2009;15:392–400. [PubMed: 19305413]
131. Friedlander M, Brooks PC, Shaffer RW, Kincaid CM, Varner JA, Cheresh DA. Definition of two angiogenic pathways by distinct alpha v integrins. *Science* 1995;270:1500–2. [PubMed: 7491498]
132. Bellahcene A, Chaplet M, Bonjean K, Castronovo V. Zoledronate inhibits alphavbeta3 and alphavbeta5 integrin cell surface expression in endothelial cells. *Endothelium* 2007;14:123–30. [PubMed: 17497369]
133. Hirbe AC, Roelofs AJ, Floyd DH, Deng H, Becker SN, Lanigan LG, Apicelli AJ, Xu Z, Prior JL, Eagleton MC, Pivnicka-Worms D, Rogers MJ, Weilbaecher K. The bisphosphonate zoledronic

- acid decreases tumor growth in bone in mice with defective osteoclasts. *Bone* 2009;44:908–16. [PubMed: 19442620]
134. Ottewell PD, Monkkonen H, Jones M, Lefley DV, Coleman RE, Holen I. Antitumor effects of doxorubicin followed by zoledronic acid in a mouse model of breast cancer. *J Natl Cancer Inst* 2008;100:1167–78. [PubMed: 18695136]
 135. Gao L, Deng H, Zhao H, Hirbe A, Harding J, Ratner L, Weilbaecher K. HTLV-1 Tax transgenic mice develop spontaneous osteolytic bone metastases prevented by osteoclast inhibition. *Blood* 2005;106:4294–302. [PubMed: 16118323]
 136. Coleman RE, Guise TA, Lipton A, Roodman GD, Berenson JR, Body JJ, Boyce BF, Calvi LM, Hadji P, McCloskey EV, Saad F, Smith MR, Suva LJ, Taichman RS, Vessella RL, Weilbaecher KN. Advancing treatment for metastatic bone cancer: consensus recommendations from the Second Cambridge Conference. *Clin Cancer Res* 2008;14:6387–95. [PubMed: 18927277]
 137. Gnant M, Mlineritsch B, Schippinger W, Luschin-Ebengreuth G, Postlberger S, Menzel C, Jakesz R, Seifert M, Hubalek M, Bjelic-Radisic V, Samonigg H, Tausch C, Eidtmann H, Steger G, Kwasny W, Dubsy P, Fridrik M, Fitzal F, Stierer M, Rucklinger E, Greil R, Marth C. Endocrine therapy plus zoledronic acid in premenopausal breast cancer. *N Engl J Med* 2009;360:679–91. [PubMed: 19213681]
 138. Aft R, Naughton M, Trinkaus K, Watson M, Ylagan L, Chavez-MacGregor M, Zhai J, Kuo S, Shannon W, Diemer K, Herrmann V, Dietz J, Ali A, Ellis M, Weiss P, Eberlein T, Ma C, Fracasso PM, Zoberi I, Taylor M, Gillanders W, Pluard T, Mortimer J, Weilbaecher K. Effect of zoledronic acid on disseminated tumour cells in women with locally advanced breast cancer: an open label, randomised, phase 2 trial. *Lancet Oncol* 11:421–8. [PubMed: 20362507]
 139. Gnant M. The evolving role of zoledronic acid in early breast cancer. *Onco Targets Ther* 2009;2:95–104. [PubMed: 20616898]
 140. Tanjore H, Zeisberg EM, Gerami-Naini B, Kalluri R. Beta1 integrin expression on endothelial cells is required for angiogenesis but not for vasculogenesis. *Dev Dyn* 2008;237:75–82. [PubMed: 18058911]
 141. Senger DR, Claffey KP, Benes JE, Perruzzi CA, Sergiou AP, Detmar M. Angiogenesis promoted by vascular endothelial growth factor: regulation through alpha1beta1 and alpha2beta1 integrins. *Proc Natl Acad Sci U S A* 1997;94:13612–7. [PubMed: 9391074]
 142. Pozzi A, Moberg PE, Miles LA, Wagner S, Soloway P, Gardner HA. Elevated matrix metalloproteinase and angiostatin levels in integrin alpha 1 knockout mice cause reduced tumor vascularization. *Proc Natl Acad Sci U S A* 2000;97:2202–7. [PubMed: 10681423]
 143. Kim S, Bell K, Mousa SA, Varner JA. Regulation of angiogenesis in vivo by ligation of integrin alpha5beta1 with the central cell-binding domain of fibronectin. *Am J Pathol* 2000;156:1345–62. [PubMed: 10751360]
 144. Boudreau NJ, Varner JA. The homeobox transcription factor Hox D3 promotes integrin alpha5beta1 expression and function during angiogenesis. *J Biol Chem* 2004;279:4862–8. [PubMed: 14610084]
 145. Garmy-Susini B, Jin H, Zhu Y, Sung RJ, Hwang R, Varner J. Integrin alpha4beta1-VCAM-1-mediated adhesion between endothelial and mural cells is required for blood vessel maturation. *J Clin Invest* 2005;115:1542–51. [PubMed: 15902308]
 146. Nikolopoulos SN, Blaikie P, Yoshioka T, Guo W, Giancotti FG. Integrin beta4 signaling promotes tumor angiogenesis. *Cancer Cell* 2004;6:471–83. [PubMed: 15542431]
 147. Chung J, Bachelder RE, Lipscomb EA, Shaw LM, Mercurio AM. Integrin (alpha 6 beta 4) regulation of eIF-4E activity and VEGF translation: a survival mechanism for carcinoma cells. *J Cell Biol* 2002;158:165–74. [PubMed: 12105188]
 148. Rodriguez-Manzaneque JC, Lane TF, Ortega MA, Hynes RO, Lawler J, Iruela-Arispe ML. Thrombospondin-1 suppresses spontaneous tumor growth and inhibits activation of matrix metalloproteinase-9 and mobilization of vascular endothelial growth factor. *Proc Natl Acad Sci U S A* 2001;98:12485–90. [PubMed: 11606713]
 149. John AS, Rothman VL, Tuszynski GP. Thrombospondin-1 (TSP-1) Stimulates Expression of Integrin alpha6 in Human Breast Carcinoma Cells: A Downstream Modulator of TSP-1-Induced Cellular Adhesion. *J Oncol* 2010;2010:645376. [PubMed: 20631908]

150. Eliceiri BP, Cheresh DA. Adhesion events in angiogenesis. *Curr Opin Cell Biol* 2001;13:563–8. [PubMed: 11544024]
151. Calvi LM, Adams GB, Weibrecht KW, Weber JM, Olson DP, Knight MC, Martin RP, Schipani E, Divieti P, Bringhurst FR, Milner LA, Kronenberg HM, Scadden DT. Osteoblastic cells regulate the haematopoietic stem cell niche. *Nature* 2003;425:841–6. [PubMed: 14574413]
152. Brenner S, Whiting-Theobald N, Kawai T, Linton GF, Rudikoff AG, Choi U, Ryser MF, Murphy PM, Sechler JM, Malech HL. CXCR4-transgene expression significantly improves marrow engraftment of cultured hematopoietic stem cells. *Stem Cells* 2004;22:1128–33. [PubMed: 15579633]
153. Kahn J, Byk T, Jansson-Sjostrand L, Petit I, Shivtiel S, Nagler A, Hardan I, Deutsch V, Gazit Z, Gazit D, Karlsson S, Lapidot T. Overexpression of CXCR4 on human CD34+ progenitors increases their proliferation, migration, and NOD/SCID repopulation. *Blood* 2004;103:2942–9. [PubMed: 15070669]
154. Zhang J, Niu C, Ye L, Huang H, He X, Tong WG, Ross J, Haug J, Johnson T, Feng JQ, Harris S, Wiedemann LM, Mishina Y, Li L. Identification of the haematopoietic stem cell niche and control of the niche size. *Nature* 2003;425:836–41. [PubMed: 14574412]
155. Papayannopoulou T. Mechanisms of stem-/progenitor-cell mobilization: the anti-VLA-4 paradigm. *Semin Hematol* 2000;37:11–8. [PubMed: 10718154]
156. Hidalgo A, Peired AJ, Weiss LA, Katayama Y, Frenette PS. The integrin alphaMbeta2 anchors hematopoietic progenitors in the bone marrow during enforced mobilization. *Blood* 2004;104:993–1001. [PubMed: 15100152]
157. Stier S, Ko Y, Forkert R, Lutz C, Neuhaus T, Grunewald E, Cheng T, Dombkowski D, Calvi LM, Rittling SR, Scadden DT. Osteopontin is a hematopoietic stem cell niche component that negatively regulates stem cell pool size. *J Exp Med* 2005;201:1781–91. [PubMed: 15928197]
158. Christopher MJ, Liu F, Hilton MJ, Long F, Link DC. Suppression of CXCL12 production by bone marrow osteoblasts is a common and critical pathway for cytokine-induced mobilization. *Blood* 2009;114:1331–9. [PubMed: 19141863]
159. Adams GB, Martin RP, Alley IR, Chabner KT, Cohen KS, Calvi LM, Kronenberg HM, Scadden DT. Therapeutic targeting of a stem cell niche. *Nat Biotechnol* 2007;25:238–43. [PubMed: 17237769]
160. Mendez-Ferrer S, Frenette PS. Hematopoietic stem cell trafficking: regulated adhesion and attraction to bone marrow microenvironment. *Ann N Y Acad Sci* 2007;1116:392–413. [PubMed: 18083941]
161. Eash KJ, Means JM, White DW, Link DC. CXCR4 is a key regulator of neutrophil release from the bone marrow under basal and stress granulopoiesis conditions. *Blood* 2009;113:4711–9. [PubMed: 19264920]
162. Kollet O, Dar A, Shivtiel S, Kalinkovich A, Lapid K, Sztainberg Y, Tesio M, Samstein RM, Goichberg P, Spiegel A, Elson A, Lapidot T. Osteoclasts degrade endosteal components and promote mobilization of hematopoietic progenitor cells. *Nat Med* 2006;12:657–64. [PubMed: 16715089]
163. Bungartz G, Stiller S, Bauer M, Muller W, Schippers A, Wagner N, Fassler R, Brakebusch C. Adult murine hematopoiesis can proceed without beta1 and beta7 integrins. *Blood* 2006;108:1857–64. [PubMed: 16735603]
164. Jiang Y, Prosper F, Verfaillie CM. Opposing effects of engagement of integrins and stimulation of cytokine receptors on cell cycle progression of normal human hematopoietic progenitors. *Blood* 2000;95:846–54. [PubMed: 10648395]
165. Voura EB, Billia F, Iscove NN, Hawley RG. Expression mapping of adhesion receptor genes during differentiation of individual hematopoietic precursors. *Exp Hematol* 1997;25:1172–9. [PubMed: 9328454]
166. Hemler ME, Lobb RR. The leukocyte beta 1 integrins. *Curr Opin Hematol* 1995;2:61–7. [PubMed: 9371973]
167. Coulombel L, Auffray I, Gaugler MH, Roseblatt M. Expression and function of integrins on hematopoietic progenitor cells. *Acta Haematol* 1997;97:13–21. [PubMed: 8980606]

168. Verfaillie CM, McCarthy JB, McClave PB. Differentiation of primitive human multipotent hematopoietic progenitors into single lineage clonogenic progenitors is accompanied by alterations in their interaction with fibronectin. *J Exp Med* 1991;174:693–703. [PubMed: 1875168]
169. Oostendorp RA, Reisbach G, Spitzer E, Thalmeier K, Dienemann H, Mergenthaler HG, Dormer P. VLA-4 and VCAM-1 are the principal adhesion molecules involved in the interaction between blast colony-forming cells and bone marrow stromal cells. *Br J Haematol* 1995;91:275–84. [PubMed: 8547062]
170. Liesveld JL, Winslow JM, Frediani KE, Ryan DH, Abboud CN. Expression of integrins and examination of their adhesive function in normal and leukemic hematopoietic cells. *Blood* 1993;81:112–21. [PubMed: 7678062]
171. Qian H, Tryggvason K, Jacobsen SE, Ekblom M. Contribution of alpha6 integrins to hematopoietic stem and progenitor cell homing to bone marrow and collaboration with alpha4 integrins. *Blood* 2006;107:3503–10. [PubMed: 16439681]
172. Schreiber TD, Steidl C, Essl M, Abele H, Geiger K, Muller CA, Aicher WK, Klein G. The integrin alpha9beta1 on hematopoietic stem and progenitor cells: involvement in cell adhesion, proliferation and differentiation. *Haematologica* 2009;94:1493–501. [PubMed: 19608669]
173. Yin T, Li L. The stem cell niches in bone. *J Clin Invest* 2006;116:1195–201. [PubMed: 16670760]
174. Kaplan RN, Riba RD, Zacharoulis S, Bramley AH, Vincent L, Costa C, MacDonald DD, Jin DK, Shido K, Kerns SA, Zhu Z, Hicklin D, Wu Y, Port JL, Altorki N, Port ER, Ruggero D, Shmelkov SV, Jensen KK, Rafii S, Lyden D. VEGFR1-positive haematopoietic bone marrow progenitors initiate the pre-metastatic niche. *Nature* 2005;438:820–7. [PubMed: 16341007]
175. Chavakis E, Aicher A, Heeschen C, Sasaki K, Kaiser R, El Makhfi N, Urbich C, Peters T, Scharffetter-Kochanek K, Zeiher AM, Chavakis T, Dimmeler S. Role of beta2-integrins for homing and neovascularization capacity of endothelial progenitor cells. *J Exp Med* 2005;201:63–72. [PubMed: 15623573]
176. McAllister SS, Gifford AM, Greiner AL, Kelleher SP, Saelzler MP, Ince TA, Reinhardt F, Harris LN, Hylander BL, Repasky EA, Weinberg RA. Systemic endocrine instigation of indolent tumor growth requires osteopontin. *Cell* 2008;133:994–1005. [PubMed: 18555776]
177. Lyden D, Hattori K, Dias S, Costa C, Blaikie P, Butros L, Chadburn A, Heissig B, Marks W, Witte L, Wu Y, Hicklin D, Zhu Z, Hackett NR, Crystal RG, Moore MA, Hajar KA, Manova K, Benezra R, Rafii S. Impaired recruitment of bone-marrow-derived endothelial and hematopoietic precursor cells blocks tumor angiogenesis and growth. *Nat Med* 2001;7:1194–201. [PubMed: 11689883]
178. Pazolli E, Luo X, Brehm S, Carbery K, Chung JJ, Prior JL, Doherty J, Demehri S, Salavaggione L, Piwnica-Worms D, Stewart SA. Senescent stromal-derived osteopontin promotes preneoplastic cell growth. *Cancer Res* 2009;69:1230–9. [PubMed: 19155301]
179. Wei SC, Tsao PN, Yu SC, Shun CT, Tsai-Wu JJ, Wu CH, Su YN, Hsieh FJ, Wong JM. Placenta growth factor expression is correlated with survival of patients with colorectal cancer. *Gut* 2005;54:666–72. [PubMed: 15831913]
180. Marcellini M, De Luca N, Riccioni T, Ciucci A, Orecchia A, Lacal PM, Ruffini F, Pesce M, Cianfarani F, Zambruno G, Orlandi A, Failla CM. Increased melanoma growth and metastasis spreading in mice overexpressing placenta growth factor. *Am J Pathol* 2006;169:643–54. [PubMed: 16877362]
181. Kang Y, Siegel PM, Shu W, Drobnjak M, Kakonen SM, Cordon-Cardo C, Guise TA, Massague J. A multigenic program mediating breast cancer metastasis to bone. *Cancer Cell* 2003;3:537–49. [PubMed: 12842083]
182. Muller A, Homey B, Soto H, Ge N, Catron D, Buchanan ME, McClanahan T, Murphy E, Yuan W, Wagner SN, Barrera JL, Mohar A, Verastegui E, Zlotnik A. Involvement of chemokine receptors in breast cancer metastasis. *Nature* 2001;410:50–6. [PubMed: 11242036]
183. Sun YX, Fang M, Wang J, Cooper CR, Pienta KJ, Taichman RS. Expression and activation of alpha(v)beta(3) integrins by SDF-1/CXC12 increases the aggressiveness of prostate cancer cells. *Prostate* 2007;67:61–73. [PubMed: 17034033]

184. Sun YX, Wang J, Shelburne CE, Lopatin DE, Chinnaiyan AM, Rubin MA, Pienta KJ, Taichman RS. Expression of CXCR4 and CXCL12 (SDF-1) in human prostate cancers (PCa) in vivo. *J Cell Biochem* 2003;89:462–73. [PubMed: 12761880]
185. Sun YX, Schneider A, Jung Y, Wang J, Dai J, Cook K, Osman NI, Koh-Paige AJ, Shim H, Pienta KJ, Keller ET, McCauley LK, Taichman RS. Skeletal localization and neutralization of the SDF-1(CXCL12)/CXCR4 axis blocks prostate cancer metastasis and growth in osseous sites in vivo. *J Bone Miner Res* 2005;20:318–29. [PubMed: 15647826]
186. Smith MC, Luker KE, Garbow JR, Prior JL, Jackson E, Piwnica-Worms D, Luker GD. CXCR4 regulates growth of both primary and metastatic breast cancer. *Cancer Res* 2004;64:8604–12. [PubMed: 15574767]
187. Pardo-Cabanas M, Bartolome RA, Wright N, Hidalgo A, Drager AM, Teixido J. Integrin alpha4beta1 involvement in stromal cell-derived factor-1alpha-promoted myeloma cell transendothelial migration and adhesion: role of cAMP and the actin cytoskeleton in adhesion. *Exp Cell Res* 2004;294:571–80. [PubMed: 15023543]
188. Mahabeleshwar GH, Feng W, Phillips DR, Byzova TV. Integrin signaling is critical for pathological angiogenesis. *J Exp Med* 2006;203:2495–507. [PubMed: 17030947]
189. Feng W, McCabe NP, Mahabeleshwar GH, Somanath PR, Phillips DR, Byzova TV. The angiogenic response is dictated by beta3 integrin on bone marrow-derived cells. *J Cell Biol* 2008;183:1145–57.
190. Gabrilovich DI, Nagaraj S. Myeloid-derived suppressor cells as regulators of the immune system. *Nat Rev Immunol* 2009;9:162–74. [PubMed: 19197294]
191. Yang L, Edwards CM, Mundy GR. Gr-1+CD11b+ myeloid-derived suppressor cells: formidable partners in tumor metastasis. *J Bone Miner Res* 25:1701–6. [PubMed: 20572008]
192. Bingle L, Brown NJ, Lewis CE. The role of tumour-associated macrophages in tumour progression: implications for new anticancer therapies. *J Pathol* 2002;196:254–65. [PubMed: 11857487]
193. Dirx AE, Oude Egbrink MG, Wagstaff J, Griffioen AW. Monocyte/macrophage infiltration in tumors: modulators of angiogenesis. *J Leukoc Biol* 2006;80:1183–96. [PubMed: 16997855]
194. Jin H, Su J, Garmy-Susini B, Kleeman J, Varner J. Integrin alpha4beta1 promotes monocyte trafficking and angiogenesis in tumors. *Cancer Res* 2006;66:2146–52. [PubMed: 16489015]
195. Hume DA. The mononuclear phagocyte system. *Curr Opin Immunol* 2006;18:49–53. [PubMed: 16338128]
196. Sweet MJ, Hume DA. CSF-1 as a regulator of macrophage activation and immune responses. *Arch Immunol Ther Exp (Warsz)* 2003;51:169–77. [PubMed: 12894871]
197. Sica A, Allavena P, Mantovani A. Cancer related inflammation: the macrophage connection. *Cancer Lett* 2008;267:204–15. [PubMed: 18448242]
198. Gordon S, Taylor PR. Monocyte and macrophage heterogeneity. *Nat Rev Immunol* 2005;5:953–64. [PubMed: 16322748]
199. Martinez FO, Sica A, Mantovani A, Locati M. Macrophage activation and polarization. *Front Biosci* 2008;13:453–61. [PubMed: 17981560]
200. Mantovani A, Sozzani S, Locati M, Allavena P, Sica A. Macrophage polarization: tumor-associated macrophages as a paradigm for polarized M2 mononuclear phagocytes. *Trends Immunol* 2002;23:549–55. [PubMed: 12401408]
201. Chen ZG, Bottazzi B, Wang JM, Mantovani A. Tumor-associated macrophages in metastasizing tumors. *Adv Exp Med Biol* 1988;233:61–71. [PubMed: 3066158]
202. Chambers SK, Kacinski BM, Ivins CM, Carcangiu ML. Overexpression of epithelial macrophage colony-stimulating factor (CSF-1) and CSF-1 receptor: a poor prognostic factor in epithelial ovarian cancer, contrasted with a protective effect of stromal CSF-1. *Clin Cancer Res* 1997;3:999–1007. [PubMed: 9815777]
203. Canioni D, Salles G, Mounier N, Brousse N, Keuppens M, Morchhauser F, Lamy T, Sonet A, Rousselet MC, Foussard C, Xerri L. High numbers of tumor-associated macrophages have an adverse prognostic value that can be circumvented by rituximab in patients with follicular lymphoma enrolled onto the GELA-GOELAMS FL-2000 trial. *J Clin Oncol* 2008;26:440–6. [PubMed: 18086798]

204. Reyes-Reyes M, Mora N, Gonzalez G, Rosales C. beta1 and beta2 integrins activate different signalling pathways in monocytes. *Biochem J* 2002;363:273–80. [PubMed: 11931654]
205. Jin H, Aiyer A, Su J, Borgstrom P, Stupack D, Friedlander M, Varner J. A homing mechanism for bone marrow-derived progenitor cell recruitment to the neovasculature. *J Clin Invest* 2006;116:652–62. [PubMed: 16498499]
206. Sato T, Nakai T, Tamura N, Okamoto S, Matsuoka K, Sakuraba A, Fukushima T, Uede T, Hibi T. Osteopontin/Eta-1 upregulated in Crohn's disease regulates the Th1 immune response. *Gut* 2005;54:1254–62. [PubMed: 16099792]
207. Schneider JG, Zhu Y, Coleman T, Semenovich CF. Macrophage beta3 integrin suppresses hyperlipidemia-induced inflammation by modulating TNFalpha expression. *Arterioscler Thromb Vasc Biol* 2007;27:2699–706. [PubMed: 17951320]
208. Savill J, Dransfield I, Hogg N, Haslett C. Vitronectin receptor-mediated phagocytosis of cells undergoing apoptosis. *Nature* 1990;343:170–3. [PubMed: 1688647]
209. Savill J, Hogg N, Ren Y, Haslett C. Thrombospondin cooperates with CD36 and the vitronectin receptor in macrophage recognition of neutrophils undergoing apoptosis. *J Clin Invest* 1992;90:1513–22. [PubMed: 1383273]
210. Law DA, DeGuzman FR, Heiser P, Ministri-Madrid K, Killeen N, Phillips DR. Integrin cytoplasmic tyrosine motif is required for outside-in alphaIIb beta3 signalling and platelet function. *Nature* 1999;401:808–11. [PubMed: 10548108]
211. Taverna D, Moher H, Crowley D, Borsig L, Varki A, Hynes RO. Increased primary tumor growth in mice null for beta3- or beta3/beta5-integrins or selectins. *Proc Natl Acad Sci U S A* 2004;101:763–8. [PubMed: 14718670]
212. Kanamori M, Kawaguchi T, Berger MS, Pieper RO. Intracranial microenvironment reveals independent opposing functions of host alphaVbeta3 expression on glioma growth and angiogenesis. *J Biol Chem* 2006;281:37256–64. [PubMed: 17028191]
213. Galarneau H, Villeneuve J, Gowing G, Julien JP, Vallieres L. Increased glioma growth in mice depleted of macrophages. *Cancer Res* 2007;67:8874–81. [PubMed: 17875729]
214. Gabrilovich DI, Bronte V, Chen SH, Colombo MP, Ochoa A, Ostrand-Rosenberg S, Schreiber H. The terminology issue for myeloid-derived suppressor cells. *Cancer Res* 2007;67:425. author reply 426. [PubMed: 17210725]
215. Sinha P, Clements VK, Bunt SK, Albelda SM, Ostrand-Rosenberg S. Cross-talk between myeloid-derived suppressor cells and macrophages subverts tumor immunity toward a type 2 response. *J Immunol* 2007;179:977–83. [PubMed: 17617589]
216. Honn KV, Tang DG, Crissman JD. Platelets and cancer metastasis: a causal relationship? *Cancer Metastasis Rev* 1992;11:325–51. [PubMed: 1423821]
217. Billroth T. Pathology and therapeutics, in fifty lectures. *Clin Orthop* 2003;1871:4–11. [PubMed: 12616036]
218. Gouin-Thibault I, Achkar A, Samama MM. The thrombophilic state in cancer patients. *Acta Haematol* 2001;106:33–42. [PubMed: 11549775]
219. Boucharaba A, Serre CM, Gres S, Saulnier-Blache JS, Bordet JC, Guglielmi J, Clezardin P, Peyruchaud O. Platelet-derived lysophosphatidic acid supports the progression of osteolytic bone metastases in breast cancer. *J Clin Invest* 2004;114:1714–25. [PubMed: 15599396]
220. Phillips DR, Charo IF, Scarborough RM. GPIIb-IIIa: the responsive integrin. *Cell* 1991;65:359–62. [PubMed: 2018971]
221. Smyth SS, Reis ED, Vaananen H, Zhang W, Collier BS. Variable protection of beta 3-integrin-deficient mice from thrombosis initiated by different mechanisms. *Blood* 2001;98:1055–62. [PubMed: 11493451]
222. Hodivala-Dilke KM, McHugh KP, Tsakiris DA, Rayburn H, Crowley D, Ullman-Cullere M, Ross FP, Collier BS, Teitelbaum S, Hynes RO. Beta3-integrin-deficient mice are a model for Glanzmann thrombasthenia showing placental defects and reduced survival. *J Clin Invest* 1999;103:229–38. [PubMed: 9916135]
223. Jurasz P, Stewart MW, Radomski A, Khadour F, Duszyk M, Radomski MW. Role of von Willebrand factor in tumour cell-induced platelet aggregation: differential regulation by NO and prostacyclin. *Br J Pharmacol* 2001;134:1104–12. [PubMed: 11682459]

224. Karpatkin S. Role of thrombin in tumor angiogenesis, implantation, and metastasis. *Pathophysiol Haemost Thromb* 2003;33 (Suppl 1):54–5. [PubMed: 12955005]
225. Nierodzik ML, Karpatkin S. Thrombin induces tumor growth, metastasis, and angiogenesis: Evidence for a thrombin-regulated dormant tumor phenotype. *Cancer Cell* 2006;10:355–62. [PubMed: 17097558]
226. Nierodzik ML, Plotkin A, Kajumo F, Karpatkin S. Thrombin stimulates tumor-platelet adhesion in vitro and metastasis in vivo. *J Clin Invest* 1991;87:229–36. [PubMed: 1845869]
227. Gasic GJ, Gasic TB, Stewart CC. Antimetastatic effects associated with platelet reduction. *Proceedings of National Academy of Sciences, USA* 1968;61:46–52.
228. Izumi Y, Taniuchi Y, Tsuji T, Smith CW, Nakamori S, Fidler IJ, Irimura T. Characterization of human colon carcinoma variant cells selected for sialyl Lex carbohydrate antigen: liver colonization and adhesion to vascular endothelial cells. *Exp Cell Res* 1995;216:215–21. [PubMed: 7529188]
229. Kim YJ, Borsig L, Varki NM, Varki A. P-selectin deficiency attenuates tumor growth and metastasis. *Proc Natl Acad Sci U S A* 1998;95:9325–30. [PubMed: 9689079]
230. Karpatkin S, Pearlstein E, Ambrogio C, Collier BS. Role of adhesive proteins in platelet tumor interaction in vitro and metastasis formation in vivo. *J Clin Invest* 1988;81:1012–9. [PubMed: 3280598]
231. Lerner WA, Pearlstein E, Ambrogio C, Karpatkin S. A new mechanism for tumor induced platelet aggregation. Comparison with mechanisms shared by other tumor with possible pharmacologic strategy toward prevention of metastases. *Int J Cancer* 1983;31:463–9. [PubMed: 6299977]
232. Nierodzik ML, Bain RM, Liu LX, Shivji M, Takeshita K, Karpatkin S. Presence of the seven transmembrane thrombin receptor on human tumour cells: effect of activation on tumour adhesion to platelets and tumor tyrosine phosphorylation. *Br J Haematol* 1996;92:452–7. [PubMed: 8603016]
233. Mannori G, Crottet P, Cecconi O, Hanasaki K, Aruffo A, Nelson RM, Varki A, Bevilacqua MP. Differential colon cancer cell adhesion to E-, P-, and L-selectin: role of mucin-type glycoproteins. *Cancer Res* 1995;55:4425–31. [PubMed: 7545541]
234. Bromberg ME, Bailly MA, Konigsberg WH. Role of protease-activated receptor 1 in tumor metastasis promoted by tissue factor. *Thromb Haemost* 2001;86:1210–4. [PubMed: 11816709]
235. Camerer E, Qazi AA, Duong DN, Cornelissen I, Advincula R, Coughlin SR. Platelets, protease-activated receptors, and fibrinogen in hematogenous metastasis. *Blood* 2004;104:397–401. [PubMed: 15031212]
236. Francis JL, Amirhosravi A. Effect of antihemostatic agents on experimental tumor dissemination. *Semin Thromb Hemost* 2002;28:29–38. [PubMed: 11885023]
237. Hu L, Lee M, Campbell W, Perez-Solar R, Karpatkin S. Role of Endogenous Thrombin in Tumor Implantation, Seeding and Spontaneous Metastasis. *Blood*. 2004
238. Palumbo JS, Talmage KE, Massari JV, La Jeunesse CM, Flick MJ, Kombrinck KW, Jirouskova M, Degen JL. Platelets and fibrin(ogen) increase metastatic potential by impeding natural killer cell-mediated elimination of tumor cells. *Blood* 2005;105:178–85. [PubMed: 15367435]
239. Pearlstein E, Ambrogio C, Karpatkin S. Effect of antiplatelet antibody on the development of pulmonary metastases following injection of CT26 colon adenocarcinoma, Lewis lung carcinoma, and B16 amelanotic melanoma tumor cells into mice. *Cancer Res* 1984;44:3884–7. [PubMed: 6744304]
240. Rickles FR, Patierno S, Fernandez PM. Tissue factor, thrombin, and cancer. *Chest* 2003;124:58S–68S. [PubMed: 12970125]
241. Savage B, Almus-Jacobs F, Ruggeri ZM. Specific synergy of multiple substrate-receptor interactions in platelet thrombus formation under flow. *Cell* 1998;94:657–66. [PubMed: 9741630]
242. Shi X, Gangadharan B, Brass LF, Ruf W, Mueller BM. Protease-activated receptors (PAR1 and PAR2) contribute to tumor cell motility and metastasis. *Mol Cancer Res* 2004;2:395–402. [PubMed: 15280447]

243. Amirkhosravi A, Amaya M, Siddiqui FA. Blockade of GpIIb/IIIa inhibits the release of vascular endothelial growth factor (VEGF) from tumor cell-activated platelets and experimental metastasis. *Platelets* 1999;10:285–292. [PubMed: 16801104]
244. Amirkhosravi A, Meyer T, Chang JY, Amaya M, Siddiqui F, Desai H, Francis JL. Tissue factor pathway inhibitor reduces experimental lung metastasis of B16 melanoma. *Thromb Haemost* 2002;87:930–6. [PubMed: 12083498]
245. Rafii DC, Psaila B, Butler J, Jin DK, Lyden D. Regulation of vasculogenesis by platelet-mediated recruitment of bone marrow-derived cells. *Arterioscler Thromb Vasc Biol* 2008;28:217–22. [PubMed: 18096826]
246. Li X, Koh AJ, Wang Z, Soki FN, Park SI, Pienta KJ, McCauley LK. Inhibitory effects of megakaryocytic cells in prostate cancer skeletal metastasis. *J Bone Miner Res*.
247. Beeton CA, Bord S, Ireland D, Compston JE. Osteoclast formation and bone resorption are inhibited by megakaryocytes. *Bone* 2006;39:985–90. [PubMed: 16870519]
248. Grundt A, Grafe IA, Liegibel U, Sommer U, Nawroth P, Kasperk C. Direct effects of osteoprotegerin on human bone cell metabolism. *Biochem Biophys Res Commun* 2009;389:550–5. [PubMed: 19748486]
249. Chollet ME, Brouland JP, Bal dit Sollier C, Bauduer F, Drouet L, Bellucci S. Evidence of a colocalisation of osteoprotegerin (OPG) with von Willebrand factor (VWF) in platelets and megakaryocytes alpha granules. Studies from normal and grey platelets. *Br J Haematol* 148:805–7. [PubMed: 19958354]
250. Leven RM, Tablin F. Extracellular matrix stimulation of guinea pig megakaryocyte proplatelet formation in vitro is mediated through the vitronectin receptor. *Exp Hematol* 1992;20:1316–22. [PubMed: 1283596]
251. Schmitz B, Thiele J, Otto F, Farahmand P, Henze F, Frimpong S, Wickenhauser C, Fischer R. Evidence for integrin receptor involvement in megakaryocyte-fibroblast interaction: a possible pathomechanism for the evolution of myelofibrosis. *J Cell Physiol* 1998;176:445–55. [PubMed: 9699497]
252. Lemieux JM, Horowitz MC, Kacena MA. Involvement of integrins alpha(3)beta(1) and alpha(5)beta(1) and glycoprotein IIb in megakaryocyte-induced osteoblast proliferation. *J Cell Biochem* 109:927–32. [PubMed: 20052668]
253. Escudero ND, Lacave M, Ubios AM, Mandalunis PM. Effect of monosodium olpadronate on osteoclasts and megakaryocytes: an in vivo study. *J Musculoskelet Neuronal Interact* 2009;9:109–20. [PubMed: 19516086]
254. Bernasconi S, Matteucci C, Sironi M, Conni M, Colotta F, Mosca M, Colombo N, Bonazzi C, Landoni F, Corbetta G, et al. Effects of granulocyte-monocyte colony-stimulating factor (GM-CSF) on expression of adhesion molecules and production of cytokines in blood monocytes and ovarian cancer-associated macrophages. *Int J Cancer* 1995;60:300–7. [PubMed: 7829234]
255. Zaslavsky A, Baek KH, Lynch RC, Short S, Grillo J, Folkman J, Italiano JE Jr, Ryeom S. Platelet-derived thrombospondin-1 is a critical negative regulator and potential biomarker of angiogenesis. *Blood* 115:4605–13. [PubMed: 20086246]
256. Mizejewski GJ. Role of integrins in cancer: survey of expression patterns. *Proc Soc Exp Biol Med* 1999;222:124–38. [PubMed: 10564536]
257. Khalili P, Arakelian A, Chen G, Plunkett ML, Beck I, Parry GC, Donate F, Shaw DE, Mazar AP, Rabbani SA. A non-RGD-based integrin binding peptide (ATN-161) blocks breast cancer growth and metastasis in vivo. *Mol Cancer Ther* 2006;5:2271–80. [PubMed: 16985061]
258. Trikha M, De Clerck YA, Markland FS. Contortrostatin, a snake venom disintegrin, inhibits beta 1 integrin-mediated human metastatic melanoma cell adhesion and blocks experimental metastasis. *Cancer Res* 1994;54:4993–8. [PubMed: 7520832]
259. Ramos OH, Kauskot A, Cominetti MR, Bechyne I, Salla Pontes CL, Chareyre F, Manent J, Vassy R, Giovannini M, Legrand C, Selistre-de-Araujo HS, Crepin M, Bonnefoy A. A novel alpha(v)beta (3)-blocking disintegrin containing the RGD motive, DisBa-01, inhibits bFGF-induced angiogenesis and melanoma metastasis. *Clin Exp Metastasis* 2008;25:53–64. [PubMed: 17952617]

260. Avraamides CJ, Garmy-Susini B, Varner JA. Integrins in angiogenesis and lymphangiogenesis. *Nat Rev Cancer* 2008;8:604–17. [PubMed: 18497750]
261. Fujii H, Komazawa H, Mori H, Kojima M, Itoh I, Murata J, Azuma I, Saiki I. Antimetastatic activities of synthetic Arg-Gly-Asp-Ser (RGDS) and Arg-Leu-Asp-Ser (RLDS) peptide analogues and their inhibitory mechanisms. *Biol Pharm Bull* 1995;18:1681–8. [PubMed: 8787788]
262. Brooks PC, Stromblad S, Klemke R, Visscher D, Sarkar FH, Cheresch DA. Antiintegrin alpha v beta 3 blocks human breast cancer growth and angiogenesis in human skin. *J Clin Invest* 1995;96:1815–22. [PubMed: 7560073]
263. Vaillant F, Asselin-Labat ML, Shackleton M, Forrest NC, Lindeman GJ, Visvader JE. The mammary progenitor marker CD61/beta3 integrin identifies cancer stem cells in mouse models of mammary tumorigenesis. *Cancer Res* 2008;68:7711–7. [PubMed: 18829523]
264. Feng X, Novack DV, Faccio R, Ory DS, Aya K, Boyer MI, McHugh KP, Ross FP, Teitelbaum SL. A Glanzmann's mutation in beta 3 integrin specifically impairs osteoclast function. *J Clin Invest* 2001;107:1137–44. [PubMed: 11342577]
265. Putz E, Witter K, Offner S, Stosiek P, Zippelius A, Johnson J, Zahn R, Riethmuller G, Pantel K. Phenotypic characteristics of cell lines derived from disseminated cancer cells in bone marrow of patients with solid epithelial tumors: establishment of working models for human micrometastases. *Cancer Res* 1999;59:241–8. [PubMed: 9892213]
266. Felding-Habermann B, O'Toole TE, Smith JW, Fransvea E, Ruggeri ZM, Ginsberg MH, Hughes PE, Pampori N, Shattil SJ, Saven A, Mueller BM. Integrin activation controls metastasis in human breast cancer. *Proc Natl Acad Sci U S A* 2001;98:1853–8. [PubMed: 11172040]
267. Teti A, Migliaccio S, Baron R. The role of the alphaVbeta3 integrin in the development of osteolytic bone metastases: a pharmacological target for alternative therapy? *Calcif Tissue Int* 2002;71:293–9. [PubMed: 12154391]
268. Engelman VW. A peptidomimetic antagonists of the avb3 integrin inhibits bone resorption in vitro and prevents osteoporosis in vivo. *Journal of Clinical Investigation* 1997;99:2284–2292. [PubMed: 9151803]
269. Harms JF, Welch DR, Samant RS, Shevde LA, Miele ME, Babu GR, Goldberg SF, Gilman VR, Sosnowski DM, Campo DA, Gay CV, Budgeon LR, Mercer R, Jewell J, Mastro AM, Donahue HJ, Erin N, Debies MT, Meehan WJ, Jones AL, Mbalaviele G, Nickols A, Christensen ND, Melly R, Beck LN, Kent J, Rader RK, Kotyk JJ, Pagel MD, Westlin WF, Griggs DW. A small molecule antagonist of the alpha(v)beta3 integrin suppresses MDA-MB-435 skeletal metastasis. *Clin Exp Metastasis* 2004;21:119–28. [PubMed: 15168729]
270. Nemeth JA, Nakada MT, Trikha M, Lang Z, Gordon MS, Jayson GC, Corringham R, Prabhakar U, Davis HM, Beckman RA. Alpha-v integrins as therapeutic targets in oncology. *Cancer Invest* 2007;25:632–46. [PubMed: 18027153]
271. Murphy MG, Cerchio K, Stoch SA, Gottesdiener K, Wu M, Recker R. Effect of L-000845704, an alphaVbeta3 integrin antagonist, on markers of bone turnover and bone mineral density in postmenopausal osteoporotic women. *J Clin Endocrinol Metab* 2005;90:2022–8. [PubMed: 15687321]
272. Smith JW, Ruggeri ZM, Kunicki TJ, Cheresch DA. Interaction of integrins alpha v beta 3 and glycoprotein IIb-IIIa with fibrinogen. Differential peptide recognition accounts for distinct binding sites. *J Biol Chem* 1990;265:12267–71. [PubMed: 2373693]
273. Gramoun A, Shorey S, Bashutski JD, Dixon SJ, Sims SM, Heersche JN, Manolson MF. Effects of Vitaxin, a novel therapeutic in trial for metastatic bone tumors, on osteoclast functions in vitro. *J Cell Biochem* 2007;102:341–52. [PubMed: 17390341]
274. Hariharan S, Gustafson D, Holden S, McConkey D, Davis D, Morrow M, Basche M, Gore L, Zang C, O'Bryant CL, Baron A, Galleman D, Colevas D, Eckhardt SG. Assessment of the biological and pharmacological effects of the alpha nu beta3 and alpha nu beta5 integrin receptor antagonist, cilengitide (EMD 121974), in patients with advanced solid tumors. *Ann Oncol* 2007;18:1400–7. [PubMed: 17693653]
275. Oliveira-Ferrer L, Hauschild J, Fiedler W, Bokemeyer C, Nippgen J, Celik I, Schuch G. Cilengitide induces cellular detachment and apoptosis in endothelial and glioma cells mediated

- by inhibition of FAK/src/AKT pathway. *J Exp Clin Cancer Res* 2008;27:86. [PubMed: 19114005]
276. Legler DF, Wiedle G, Ross FP, Imhof BA. Superactivation of integrin alphavbeta3 by low antagonist concentrations. *J Cell Sci* 2001;114:1545–53. [PubMed: 11282030]
277. Alghisi GC, Ponsonnet L, Ruegg C. The integrin antagonist cilengitide activates alphaVbeta3, disrupts VE-cadherin localization at cell junctions and enhances permeability in endothelial cells. *PLoS One* 2009;4:e4449. [PubMed: 19212436]
278. Guo W, Giancotti FG. Integrin signalling during tumour progression. *Nat Rev Mol Cell Biol* 2004;5:816–26. [PubMed: 15459662]
279. Sterling JA, Edwards JR, Martin TJ, Mundy GR. Advances in the biology of bone metastasis: How the skeleton affects tumor behavior. *Bone*.
280. Bisanz K, Yu J, Edlund M, Spohn B, Hung MC, Chung LW, Hsieh CL. Targeting ECM-integrin interaction with liposome-encapsulated small interfering RNAs inhibits the growth of human prostate cancer in a bone xenograft imaging model. *Mol Ther* 2005;12:634–43. [PubMed: 16039164]
281. Danen EH, Marcinkiewicz C, Cornelissen IM, van Kraats AA, Pachter JA, Ruitter DJ, Niewiarowski S, van Muijen GN. The disintegrin eristostatin interferes with integrin alpha 4 beta 1 function and with experimental metastasis of human melanoma cells. *Exp Cell Res* 1998;238:188–96. [PubMed: 9457071]
282. Nervi B, Ramirez P, Rettig MP, Uy GL, Holt MS, Ritchey JK, Prior JL, Piwnica-Worms D, Bridger G, Ley TJ, DiPersio JF. Chemosensitization of acute myeloid leukemia (AML) following mobilization by the CXCR4 antagonist AMD3100. *Blood* 2009;113:6206–14. [PubMed: 19050309]
283. Lane SW, Scadden DT, Gilliland DG. The leukemic stem cell niche: current concepts and therapeutic opportunities. *Blood* 2009;114:1150–7. [PubMed: 19401558]
284. Azab AK, Runnels JM, Pitsillides C, Moreau AS, Azab F, Leleu X, Jia X, Wright R, Ospina B, Carlson AL, Alt C, Burwick N, Roccaro AM, Ngo HT, Farag M, Melhem MR, Sacco A, Munshi NC, Hideshima T, Rollins BJ, Anderson KC, Kung AL, Lin CP, Ghobrial IM. CXCR4 inhibitor AMD3100 disrupts the interaction of multiple myeloma cells with the bone marrow microenvironment and enhances their sensitivity to therapy. *Blood* 2009;113:4341–51. [PubMed: 19139079]

Integrin Expression During Bone Metastasis

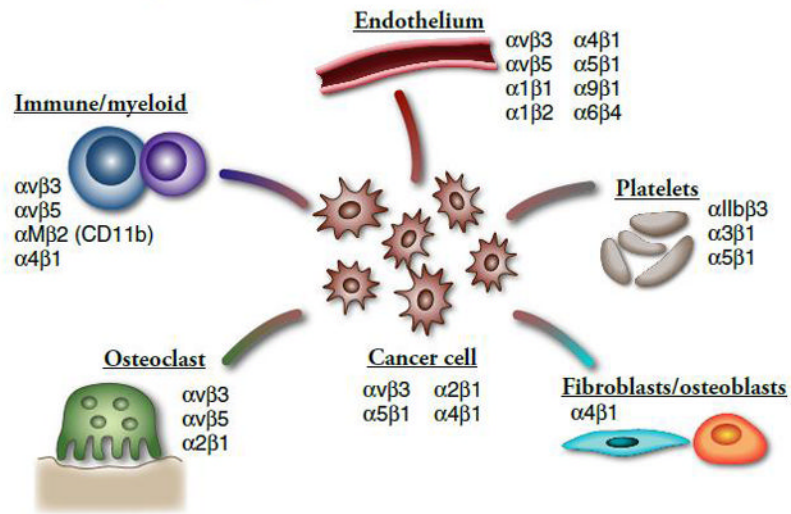


Fig. 1.
Integrin Expression During Bone Metastasis

Table 1

Extracellular matrix proteins and the main integrins that participate in bone metastasis and tumor growth in bone

Integrin	ECM Ligands
$\alpha v\beta 3$	vitronectin, osteopontin, bone sialoprotein, fibronectin, TSP-1
$\alpha 2\beta 1$	collagen I, laminin
$\alpha 4\beta 1$ (VLA-4)	VCAM-1 Fibronectin, osteopontin
$\alpha v\beta 1$	fibronectin, vitronectin,
$\alpha v\beta 5$	vitronectin, osteonectin, bone sialoprotein, fibronectin
$\alpha IIb\beta 3$	fibrinogen,
$\beta 2$	VCAM-1, ICAM-1, fibrinogen
$\alpha 1\beta 1$	collagen
$\alpha 5\beta 1$	fibronectin
$\alpha 9\beta 1$	TSP1
$\alpha 6\beta 4$	laminin, TSP-1