

Online Submissions: http://www.wjgnet.com/1007-9327office wjg@wjgnet.com doi:10.3748/wjg.v16.i48.6155 World J Gastroenterol 2010 December 28; 16(48): 6155-6162 ISSN 1007-9327 (print) ISSN 2219-2840 (online) © 2010 Baishideng. All rights reserved.

BRIEF ARTICLE

Increasing the frequency of CIK cells adoptive immunotherapy may decrease risk of death in gastric cancer patients

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Supported by The National Natural Science Foundation of China, No. 30872176, 30950022 and 30972703; grants of Jiangsu Province and Soochow University Medical Development Foundation, No. EE126765

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 Received:
 October 12, 2010
 Revised:
 November 16, 2010

 Accepted:
 November 23, 2010
 Revised:
 November 16, 2010

Published online: December 28, 2010

Abstract

AIM: To analyze the correlation between cytokineinduced killer (CIK) cells adoptive immunotherapy and cancer-related death in gastric cancer patients.

METHODS: One hundred and fifty-six gastric cancer

patients after operation at the Third Affiliated Hospital of Soochow University were enrolled in this study. Their clinical data including demographic characteristics, operation time, tumor size, pathological type and staging, tumor metastasis, outcome of chemotherapy or CIK cells adoptive immunotherapy, survival time or time of death were collected with a standard structured questionnaire. Kaplan-Meier method was used to estimate the median survival time, and the 2- and 5- year survival rates. Hazard risk (HR) and 95% confidence interval (95% CI) of CIK cells adoptive immunotherapy for gastric cancer were calculated using the two-stage time-dependent covariates Cox model.

RESULTS: The survival time of gastric cancer patients was longer after CIK cells adoptive immunotherapy than after chemotherapy ($\chi^2 = 10.907$, P = 0.001). The median survival time of gastric cancer patients was also longer after CIK cells adoptive immunotherapy than after chemotherapy (49 mo vs 27 mo, P < 0.05). The 2- and 5-year survival rates of gastric cancer patients were significantly higher after CIK cells adoptive immunotherapy than after chemotherapy (73.5% vs 52.6%, 40.4% vs 23.9%, P < 0.05). A significant difference was observed in the survival curve for patients who received CIK cells adoptive immunotherapy (0, 1-10, 11-25, and over 25 frequencies) (χ^2 = 14.534, P = 0.002). The frequencies of CIK cells adoptive immunotherapy were significantly related with the decreasing risk of death in gastric cancer patients after adjustment for sex and age of the patients, tumor stage and relapse (HR = 0.54, 95% CI: 0.36-0.80) when the first stage Cox model was used to define the subjects who remained alive beyond 36 mo as survivors. However, no correlation was observed between the frequencies of death in CIK cells adoptive immunotherapy and the risk of gastric cancer patients (HR = 1.09, 95% CI: 0.63-0.89) when the second stage Cox model was used to define the subjects who survived for more than 36 mo as survivors.

CONCLUSION: The survival time of the gastric cancer



patients treated with chemotherapy combined with CIK cells adoptive immunotherapy is significantly longer than that of the patients treated with chemotherapy alone and increasing the frequency of CIK cells adoptive immuno-therapy seems to benefit patients more.

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Key words: Immunotherapy; Cytokine-induced killer cells; Gastric cancer; Survival analysis; Probability

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Jiang JT, Shen YP, Wu CP, Zhu YB, Wei WX, Chen LJ, Zheng X, Sun J, Lu BF, Zhang XG. Increasing the frequency of CIK cells adoptive immunotherapy may decrease risk of death in gastric cancer patients. *World J Gastroenterol* 2010; 16(48): 6155-6162 Available from: URL: http://www.wjgnet.com/1007-9327/full/v16/ i48/6155.htm DOI: http://dx.doi.org/10.3748/wjg.v16.i48.6155

INTRODUCTION

Gastric cancer is one of the most common causes of cancer-related death in China^[1]. Its incidence in Jiangsu Province of China is particularly high, and its mortality rate is much higher than the national average^[2]. The early clinical detection rate of gastric cancer is less than 15%, and about 85% cases of gastric cancer are at the advanced stage when their gastric cancer is diagnosed^[3]. Surgery is the standard treatment procedure for localized and resectable gastric cancer^[4]. However, surgery alone does not improve the 5-year survival rate of local advanced gastric cancer patients^[3]. Although standardized surgical resection and adjuvant therapeutic modalities are available for gastric cancer, the survival rate of advanced gastric cancer patients remains very low after operation^[5]. About 60% of gastric cancer patients usually experience local recurrence and metastasis to other organs^[6]. It has been demonstrated that local recurrence and distant metastasis constitute a major problem in the failure of cancer therapies^[/]. Therefore, considerable efforts are needed to improve the current therapeutic modalities and to explore new therapies. In recent years, immune therapy has become the fourth important treatment modality for malignant tumors following surgery, radiotherapy and chemotherapy^[8-10].

A number of adoptive immunotherapy with killer cells have been reported, including lymphokine-activated killer cells^[11], tumor infiltrating lymphocytes^[12], or anti-CD3 monoclonal antibody-induced killer cells^[13]. However, their therapeutic efficacy is limited due to their low antitumor activities^[14]. At present, cytokine-induced killer (CIK) cells are a new type of anti-tumor effector cells, which can proliferate rapidly *in vitro*, with a stronger antitumor activity and a broader target tumor spectrum than the reported anti-tumor effector cells^[10,15]. Moreover, CIK cells can regulate and enhance immune function^[16]. Studies have reported the level of tumor markers, change in cellular immune functions, exploration of molecule targets and a short-term efficacy in gastric cancer patients after chemotherapy plus CIK cells immunotherapy despite some side effects^[17,19]. However, the relation between the frequencies of CIK cells immunotherapy and its clinical efficacy has not been examined. In the present study, data obtained from 156 gastric cancer patients in fitting multivariate Cox model showed that more frequencies of CIK cells immunotherapy could improve the survival rate of gastric cancer patients.

MATERIALS AND METHODS

Patients

One hundred and fifty-six primary gastric cancer patients after operation at the Third Affiliated Hospital of Soochow University (Jiangsu Province, China) were enrolled in this study. Those who did not meet the inclusion criteria, or had other tumors were excluded.

A standard questionnaire was designed to collect the data from the patients, including demographic characteristics, operation time, tumor size and location, pathological type and staging, tumor metastasis, outcome of chemotherapy or CIK cells immunotherapy. Meanwhile, time of relapse and death of the patients was recorded. Patients received 6 cycles of chemotherapy before CIK cells immunotherapy. Some patients underwent CIK cells immunotherapy due to cancer recurrence during chemotherapy. Recurrence of gastric cancer was defined when local, peritoneal or distant metastasis was detected at any site during chemotherapy^[20]. The study was conducted according to the principles of the Declaration of Helsinki. All patients gave their informed consent prior to inclusion in the study. The study was approved by the Ethics Committee of the Third Affiliated Hospital of Soochow University.

Eighty-one patients (62 males, 19 females) at the age of 59.9 \pm 10.5 years with a median age of 60.5 years who underwent chemotherapy alone served as chemotherapy group (group I), and those (60 males at the age of 62.4 \pm 10.8 years with a median age 60.5 years, 15 females at the age of 51.0 \pm 10.7 years with a median age of 50 years) who received chemotherapy plus CIK cells immunotherapy served as treatment group (group II) (Table 1).

Preparation of CIK cells and treatment

Peripheral blood mononuclear cells (PBMC) were collected with a COBE spectra blood cell separator (Gambro BCT, Inc., Lakewood, USA). Viability of PBMC was assessed by trypan blue exclusion. PBMC (2.0×10^6 /mL) were plated onto 6-well dishes (Nunc, Denmark) and cultured with medium I containing RPMI 1640 in the presence of 1.0×10^6 U/L human interferon- γ (IFN- γ , Shanghai Fosun Pharma Co., China), 5.0×10^5 U/L recombinant human interleukin-2 (IL-2, Shangdong Quangang Pharmaceutical Co., China), 10% inactivated human serum, 25 mmol/L HEPES, 2 mmol/L L-glutamine, 100 U/mL penicillin and 100 µg/mL streptomycin. The cells were incubated in a humidified atmosphere containing 5% CO₂ at 37°C. Monoclonal antibody (MAb) against CD3



Demographic and			χ²	Р				
clinical features		Group I	Group Ⅱ					
Sex								
Men	122	62 (76.5)	60 (80)					
Women	34	19 (23.5)	15 (20)	0.273	0.601			
Age (yr)		. ,	. ,					
≤ 45	14	7 (8.6)	7 (9.3)					
$45 < age \le 60$	71	36 (44.5)	35 (46.7)	0.047^{a}	0.977			
> 60	71	38 (46.9)	33 (44.0)					
Tumor site ^b								
Gastric cardia								
Yes	58	30 (37.0)	28 (37.3)	0.000	0.070			
No	98	51 (63.0)	47 (62.7)	0.002	0.970			
Gastric body								
Yes	64	35 (43.2)	29 (38.7)					
No	92	46 (56.8)	46 (61.3)	0.332	0.564			
Gastric antrum		. ,	. ,					
Yes	36	17 (21.0)	19 (25.3)					
No	120	64 (79.0)	56 (74.7)	0.414	0.520			
Tumor size ^c (cm)		()	~ /					
< 5	76	43 (59.7)	33 (55.0)	0.000	0 505			
≥ 5	56	29 (40.3)	27 (45.0)	0.299	0.585			
Histological type ^c								
Differentiated	52	28 (35.4)	24 (64.6)	0.000	0.0/2			
Poorly-differentiated	94	51 (38.5)	43 (64.2)	0.002	0.962			
Invasion ^c								
Yes	112	55 (76.4)	57 (89.1)	0.545	0.050			
No	24	17 (23.6)	7 (10.9)	3.745	0.053			
Lymph node metastas	is ^c							
Yes	100	55 (76.4)	45 (70.3)	0 (10	0.400			
No	36	17 (23.6)	19 (29.7)	0.643	0.423			
Relapse		. ,	. ,					
Yes	98	58 (28.4)	40 (46.7)	//	0.010			
No	58	23 (71.6)	35 (53.3)	5.566	0.018			
Pathological grade								
1	2	1 (1.3)	1 (1.3)					
2	27	18 (22.2)	9 (12.0)	0.07/	0.005			
3	92	44 (54.3)	48 (64.0)	2.976	0.395			
4	35	18 (22.2)	17 (22.7)					
Tumor stage								
I	14	9 (11.1)	5 (6.7)					
Ш	22	12 (14.8)	10 (13.3)	2.129 ^a	0 546			
Ш	102	49 (60.5)	53 (70.7)	2.129	0.546			
IV	18	11 (13.6)	7 (9.3)					

Table 1 Distribution of demographic and clinical characteris-tics in two groups

"cmh2 χ^2 -test; "Two cases of gastric cancer involving gastric cardia and body; "Missed cases.

(100 µg/L, Antibody Diagnostic Inc., USA) and IL-1 α (1.0 × 10⁵ U/L, Promega) were added after 24 h culture. Supernatant was aspirated and the cells were cultured in Medium II in the absence of INF- γ after another 48 h culture. The cells were transferred to Kolle flasks (Nunc, Denmark) and cultured in the same medium after 1 wk. The medium was changed every 3 d. The cytotoxic activity was assayed as previously described^[17,21]. Patients in group I were treated with oxaliplatin (120 mg/m² D1, 5-Fu 400 mg/m² CIV 24 h D1-5, CF 200 mg/m² D1-5) as previously described^[22] with the doses adjusted according to the toxicity. Patients in group II received CIK cells immunotherapy as previously described^[17] after 6 cycles of chemotherapy, with 1.0 × 10⁹ CIK cells transfused into the patients within 1 h.

Statistical analysis

All data were loaded into the Epidata3.0 database with double-check, and analyzed with the SAS software package (version 9.13; SAS Institute, Cary, NC, USA). The data were expressed as mean \pm SD. χ^2 -test or cmh2 χ^2 -test was used to compare the difference in balance between the concerned clinical indexes and to find the confounding factors between the two groups. Survival data were analyzed using Kaplan-Meier method and log-rank test to estimate the median survival time, 2- and 5-year survival rates, and to determine if the survival curves for the two groups were different.

When the frequency of CIK cells immunotherapy and the survival time were introduced into the Cox model, the interaction item was significantly associated with the death of gastric cancer patients (wald- $\chi^2 = 4.946$, P = 0.0261). A two-stage time-dependent Cox model was established to precisely estimate the hazard risk (HR) and 95% confidence interval (95% CI) of the association between the frequency of CIK cells immunotherapy and the death of gastric cancer patients. Because the median survival time of gastric cancer patients was about 36 mo, 36 mo was used as the optimum cutoff point.

The first stage Cox model involved 154 patients with a survival time of over 36 mo who were defined as survivors. Otherwise, the survival status was the same as the original definition.

The second stage Cox model only involved 56 patients with a survival time longer than 36 mo, and their survival status was defined as the original definition.

Pearson correlation test was performed between Schoenfeld residual of the frequencies of CIK cells immunotherapy and the survival time of gastric cancer patients to determine whether the frequency of CIK cells immunotherapy is a time-dependent variable in the two Cox models^[23].

RESULTS

Distribution of demographic and clinical characteristics in two groups

No statistical difference was found in sex and age of the patients, tumor site, histological type, invasion depth, lymph node metastasis, pathological grade, tumor size and stage between the two groups. However, the number of patients was significantly greater in group II with recurrent disease than in group I (46.7% *vs* 28.4%, $\chi^2 = 5.566$, P = 0.018) (Table 1), suggesting that more patients with relapse should receive CIK cells immunotherapy.

Demographic and clinical characteristics of patients after CIK cells immunotherapy

No statistical difference was observed in sex and age of the patients, tumor site, histological type, invasion depth, lymph node metastasis, pathological grade or tumor size after CIK cells immunotherapy (0, 1-10, 11-25, and over 25 frequencies). However, a significant difference was found in cancer recurrence and stage after CIK cells immunotherapy (Table 2).

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Demographic and clinical features	n	Free	χ²	Р			
		0	1-10	11-25	> 25		
Sex							
Men	122	62 (76.5)	39 (84.8)	13 (86.7)	8 (57.1)	5.573	0.134
Women	34	19 (23.5)	7 (15.2)	2 (13.3)	6 (42.9)		
Age (yr)							
≤ 45	14	7 (8.6)	5 (10.9)	1 (6.6)	1 (7.1)	1.153	0.97
$45 < age \le 60$	71	36 (44.4)	20 (43.5)	7 (46.7)	8 (57.1)		
> 60	71	38 (47.0)	21 (45.6)	7 (46.7)	5 (35.8)		
Tumor site ^a							
Gastric cardia							
Yes	58	30 (37.0)	17 (37.0)	5 (33.3)	6 (42.9)	0.290	0.96
No	98	51 (63.0)	29 (63.0)	10 (66.7)	8 (57.1)		
Gastric body		. ,	. ,	. ,	. ,		
Yes	64	35 (43.2)	19 (41.3)	5 (33.3)	5 (35.7)	0.691	0.87
No	92	46 (56.8)	27 (58.7)	10 (66.7)	9 (64.3)		
Gastric antrum							
Yes	36	17 (21.0)	10 (21.7)	4 (26.7)	5 (35.7)	1.614	0.65
No	120	64 (79.0)	36 (78.3)	11 (73.3)	9 (64.3)	11011	0.00
Tumor size ^c (cm)	120	01(15.0)	56 (70.5)	11 (70.0)	5 (01.0)		
< 5	76	43 (59.7)	17 (45.9)	7 (58.3)	9 (81.8)	4.834	0.18
≥5	56	29 (40.3)	20 (54.1)	5 (41.7)	2 (18.2)	1.001	0.10
≓ 5 Histological type ^c	50	29 (40.3)	20 (34.1)	5 (41.7)	2 (10.2)		
Differentiated	52	28 (25 4)	12 (20.0)	6 (40 0)	6 (E0 0)	1.760	0.62
	94	28 (35.4)	12 (30.0)	6 (40.0)	6 (50.0)	1.760	0.624
Poorly-differentiated	94	51 (64.6)	28 (70.0)	9 (60.0)	6 (50.0)		
Invasion	440		2 0 (00 5)	10 (00 0)	0 (01 0)	4.007	0.00
Yes	112	55 (76.4)	38 (90.5)	10 (90.9)	9 (81.8)	4.226	0.23
No	24	17 (23.6)	4 (9.5)	1 (9.1)	2 (18.2)		
Lymph node metastasis ^c		()		_ /	- ()		
Yes	100	55 (76.4)	31 (75.6)	7 (58.3)	7 (63.6)	2.371	0.49
No	36	17 (23.6)	10 (24.4)	5 (41.7)	4 (36.4)		
Relapse							
Yes	98	58 (71.6)	31 (67.4)	8 (53.3)	1 (7.1)	15.633	0.000
No	58	23 (28.4)	15 (32.6)	7 (46.7)	13 (92.9)		
Pathological grade							
1	2	1 (1.2)	1 (2.2)	0 (0.0)	0 (0.0)	2.976 ^b	0.395
2	27	18 (22.2)	8 (17.4)	0 (0.0)	1 (7.1)		
3	92	44 (54.3)	25 (54.4)	12 (80.0)	11 (78.6)		
4	35	18 (22.2)	12 (26.0)	3 (20.0)	2 (14.3)		
Tumor stage							
I	14	9 (11.1)	3 (6.5)	0 (0.0)	2 (14.2)	13.66 ^b	0.038
Π	22	23 (28.4)	3 (6.5)	1 (6.7)	6 (42.9)		
Ш	102	38 (46.9)	34 (73.9)	13 (86.7)	6 (42.9)		
IV	18	11 (13.6)	6 (13.1)	1 (6.6)	0 (0.0)		

^aTumor sites in some cases were repeated; ^bBecause the theoretical value is less than 1, χ^2 was performed for patients who received cytokine-induced killer (CIK) cells immunotherapy at the frequencies of 11-25 and > 25, and for those who underwent CIK cells immunotherapy at the frequencies of 1-10, 11-25 and > 25; [°]Missed cases.

Survival time of patients in two groups

The survival time of gastric cancer patients in group II was much longer than that of those in group I (χ^2 = 10.907, P = 0.001, Figure 1). The median survival time of patients in group II was also longer than that of those in group I (49 mo vs 27 mo).

Two- and 5-year survival rates of patients in two groups

The 2- and 5-year survival rates of patients in group II were significantly higher than those of patients in group I (73.5% vs 52.6%, 40.4% vs 23.9%, P < 0.05) (Table 3).

Survival time of patients after CIK cells immunotherapy

Because the CIK cells immunotherapy seemed effective

against gastric cancer, whether the frequency of CIK cells immunotherapy affects its efficacy was determined. The survival curve was obviously higher for the patients after CIK cells immunotherapy plus chemotherapy than after chemotherapy alone. The survival time of gastric cancer patients was significantly longer after CIK cells immunotherapy than after chemotherapy ($\chi^2 = 14.534$, P = 0.002, Figure 1).

Time-dependent Cox model analysis of CIK cells immunotherapy and prognosis of patients

The frequency of CIK cells immunotherapy was a strong time-dependent variable. A significant difference was observed in the survival time and the frequency of CIK cells immunotherapy between the two models ($\chi^2 = 27.990$, P

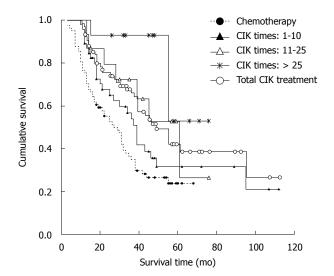


Figure 1 Survival rates of patients after cytokine-induced killer cells immunotherapy plus chemotherapy and chemotherapy alone. CIK: Cytokineinduced killer.

< 0.0001). However, it could not stratify the theoretical hypothesis of proportional hazard model. Hence, we tried to improve the analyzing results of the Cox model by dividing the patients into two stages with a relatively short survival time. Because of the relatively short interval time, the frequency of CIK cells immunotherapy may be a time-independent variable and can satisfy the assumption of proportional hazard model. The median survival time (36 mo) of the patients was used as the optimum cut-point.

In the first stage Cox model, the frequency of CIK cells immunotherapy was not a time-dependent variable, thus refusing the hypothesis of non proportional hazard model by Pearson correlation test between Schoenfeld residual of CIK cells immunotherapy frequency and survival time (r = 0.04, F = 0.10, P = 0.751). After the patients who remained alive beyond 36 mo after treatment were defined as survivors, the frequency of CIK cells immunotherapy was significantly associated with the decreasing risk of death in gastric cancer patients after adjustment for sex and age of the patients, tumor stage and relapse (HR = 0.54, 95% CI: 0.36-0.80) (Table 4).

In the second stage Cox model, the frequency of CIK cells immunotherapy was not a time-dependent variable (r = 0.307, F = 1.98, P = 0.176). When the second stage Cox model involving only 56 patients with a survival time of over 36 mo was fitted, no association was observed between the frequency of CIK cells immunotherapy and the survival time of gastric cancer patients (HR = 1.09, 95% CI: 0.63-0.89) (Table 5).

DISCUSSION

In China, gastric cancer patients are usually diagnosed at a relatively advanced stage with metastasis to other organs^[24]. A number of treatment modalities are available for gastric cancer, such as surgical resection combined with chemotherapy^[25,26], radiotherapy^[27,28], thermotherapy^[29,30] and/or traditional Chinese medicine^[31,32]. However,

Table 3 Two- and five-year survival rates of gastric cancer patients in two groups

Groups	Survival rate (%)	95% CI	u-value	Р
2-yr				
Chemotherapy	52.6	41.7-63.6	2.721	0.007
CIK treatment	73.5	63.3-83.7		
5-yr				
Chemotherapy	23.9	13.5-34.3	1.913	0.0526
CIK treatment	40.4	27.1-53.7		

CIK: Cytokine-induced killer; 95% CI: 95% confidence interval.

the 5-year survival rate of advanced gastric cancer patients is still very poor^[33]. It has been shown that cellular immunotherapy can promote host anti-cancer immunity, thus prolonging the survival time of gastric cancer patients^[34]. Treatment of gastric cancer with autologous CIK cells is one of the promising cellular immunotherapies^[35].

The results of the present study demonstrate that CIK cells immunotherapy plus chemotherapy for gastric cancer has more potential benefits than chemotherapy alone. Therefore, the effect of adjuvant cellular therapies for gastric cancer has drawn more attention of oncologists. It has been shown that adjuvant radiotherapy and chemotherapy for gastric cancer after curative resection can improve the disease-free and overall survival time of gastric cancer patients^[25,36]. Residual tumor cells after chemotherapy can be removed by the host immune system^[37]. However, several cycles of chemotherapy can decrease the immune functions of gastric cancer patients^[38], and have been suspected to be one of the reasons for the high relapse rate of gastric cancer after postoperative systemic chemotherapy. Immunotherapy can directly kill cancer cells and boost the immune responses against the tumor^[39]. Therefore, immunotherapy should be beneficial for gastric cancer patients, and not conducive to the growth of tumor cells.

In the present study, CIK cells were obtained upon culturing PBMC in the presence of IFN-y, IL-2, anti-CD-3MAb, and IL-1 $\alpha^{[13]}$. This method allows us to generate a large number of CIK cells. In addition, the anti-tumor cell activity of CIK cells is stronger than that of anti-tumor effector cells^[16]. The effector cells in our culture are believed to be CD3⁺CD56⁺. CIK cells have a higher survival rate, proliferation capacity, and killing activity than their target cells^[40,41], and can secrete a variety of cytokines, which further enhance the cytotoxicity of immune effector cells^[42] and change the tumor microenvironment to favor cancer eradication. In addition, CIK cells can kill both autologous and allogeneic tumor cells^[43,44], as well as multi-drug resistance cells and FasL-positive cells^[45,46]. Accordingly, CIK cells immunotherapy combined with chemotherapy may have a synergistic effect.

Several studies^[17,47-49] showed that CIK cells immunotherapy can significantly improve the immune functions of cancer patients, such as an increase in CD3⁺CD56⁺ level. However, the clinical data are not enough to demonstrate the effectiveness of CIK cells immunotherapy. The results of this retrospective study, based on the follow-up

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Table 4 First stage time-dependent multivariate Cox model analysis of cytokine-induced killer cells immunotherapy at different fre- quencies and prognosis of the patients ^{1,2} ($n = 154^3$)						
Variables ⁴	β	S β	Wald- χ^2	<i>P</i> -value	HR	95% CI
No. of CIK infusion	-0.620	0.200	9.592	0.002	0.54	0.36-0.80
Sex	0.294	0.298	0.969	0.325	1.34	0.75-2.41
Age	0.508	0.200	6.492	0.011	1.66	1.12-2.46
Tumor stages	0.739	0.202	13.377	0.0003	2.10	1.40-3.11
Relapse (yes or no)	3.363	0.719	21.848	< 0.0001	28.87	7.05-118.25

¹All patients with survival time of longer than 36 mo (median value for total patients) were defined as the survivors; ²Pearson correlation test between Schoen-feld residual of cytokine-induced killer (CIK) cells immunotherapy frequency and survival time (r = 0.01, F = 0.01, P = 0.936); ³Missed patients; ⁴Variable value definition in the Cox model (frequency of CIK cells immunotherapy: 0 = 0 time, 1 = 1-10 times, 2 = 11-25 times, 3 = more than 25 times; Sex: 1 = man, 2 = wom-an), age ($0 = \le 45$ yr, 1 = about 60 yr, 2 = over 60 yr), tumor stage (0 = stage I, 1 = stage II, 2 = stage II, 3 = stage IV), relapse (1 = yes, 0 = no). HR: Hazard risk; 95% CI: 95% confidence interval.

Table 5 Second stage time-dependent multivariate Cox model analysis of cytokine-induced killer cells immunotherapy at different frequencies and prognosis of patients^{1,2} (n = 56)

Variables ³	β	S β	Wald- χ^2	<i>P</i> -value	HR	95% CI
Number of CIK infusion	0.089	0.280	0.102	0.750	1.09	0.63-1.89
Sex	0.676	0.619	1.191	0.275	1.97	0.58-6.62
Age	-0.318	0.442	0.518	0.472	0.73	0.31-1.73
Tumor stages	-0.471	0.341	1.909	0.167	0.62	0.32-1.22
Relapse (yes or no)	2.203	0.558	15.582	< 0.0001	9.05	3.03-27.02

¹This model only involving the patients with a survival time longer than 36 mo. The survival rate of each patient was similar; ²Pearson correlation test between Schoenfeld residual of cytokine-induced killer (CIK) cells immunotherapy frequency and survival time (r = 0.307, F = 1.98, P = 0.176); ³Variable value definition in the Cox model (frequency of CIK cells immunotherapy: 0 = 0 time, 1 = 1-10 times, 2 = 11-25 times, 3 = more than 25 times; Sex: 1 = man, 2 = woman), age ($0 = \le 45$ yr, 1 = about 60 yr, 2 = over 60 yr), tumor stage (0 = stage I, 1 = stage II, 2 = stage II, 3 = stage IV), relapse (1 = yes, 0 = no). HR: Hazard risk; 95% CI: 95% confidence interval.

of 156 gastric cancer patients, show that the frequency of CIK cells immunotherapy can significantly prolong the survival time of gastric cancer patients.

However, the frequency of CIK cells immunotherapy may significantly decrease the risk of cancer-related death in gastric cancer patients. The common Cox model was not preferred in our analysis because the balance test showed that tumor relapse and stage were different in patients of the two groups. There are two reasons that support our conclusion. First, except for tumor relapse and stage, the balance test showed the following factors, including age and sex of the patients, tumor size, site and invasiveness, lymph node metastasis and pathological grade did not affect us to assign the gastric cancer patients into groups I and II. Second, due to the fact that the frequency of CIK cells immunotherapy was a timedependent variable in the Cox model, a two-stage timedependent Cox model adjustment was made for some confounding factors (tumor relapse and stage), thus the false results were avoided when the common Cox model was used to make our analysis more reliable.

After the postoperative adjuvant chemotherapy, most residual tumor cells sensitive to chemotherapy were removed. Chemotherapy may suppress the immune function and therefore immunotherapy is necessary for boosting immunity. It was reported that the number of $\rm CD3^+$ cells and the $\rm CD4^+/\rm CD8^+$ ratio are significantly lower in most gastric cancer patients than in healthy controls after che-

motherapy^[17]. In the present study, the number of CD3⁺ and CD4⁺ cells was significantly increased, while the number of CD8⁺ cells was declined and the CD4⁺/CD8⁺ ratio was increased after CIK cells immunotherapy, suggesting that CIK cells also have an immune modulating function in addition to their anti-tumor function^[50]. Single CIK cells immunotherapy has an *in vivo* effect against gastric cancer for about one month^[51]. In contrast, the number of CD3⁺ cells and the CD4⁺/CD8⁺ ratio maintain at a high level after three cycles of CIK cells immunotherapy^[17].

Our study has a few limitations. First, it was a retrospective cohort/observational study rather than a strictlydesigned randomized trial. Since the patients were not assigned into CIK cells immunotherapy group and chemotherapy group, imbalance of certain clinical factors between the two groups could not be avoided. Second, there were some unknown potential reasons for choice of treatment regimens, which might affect our conclusion, despite the fact that the balance test was performed and some confounding factors were adjusted in the Cox model. Therefore, a randomized clinical trial is necessary to justify the benefit of CIK cells immunotherapy for gastric cancer. The benefit of radiochemotherapy and S1 chemotherapy for gastric cancers, established in a recent clinical trial^[52], is important to determine whether CIK cells immunotherapy provides additional benefit when it is used in combination with radiotherapy and chemotherapy.



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In conclusion, more frequencies of CIK cells are necessary for gastric cancer. The survival time of gastric cancer patients is significantly longer after chemotherapy plus CIK cells immunotherapy than after chemotherapy alone.

COMMENTS

Background

Gastric cancer is one of the most common causes of cancer-related death in China. Although standardized surgical resection and numerous adjuvant therapeutic modalities are available for gastric cancer, the postoperative survival rate of advanced stage cancer patients remains very low. In recent years, immune therapy for malignant tumors has become the fourth important tumor treatment modality following surgery, radiotherapy and chemotherapy. Cellular immunotherapy can promote host anti-cancer immunity, thus prolonging the survival time of gastric cancer patients. Treatment with autologous cytokine-induced killer (CIK) cells is one of the promising cellular immunotherapies.

Research frontiers

A number of adoptive cells immunotherapy have been reported, including using lymphokine activated killer cells, tumor infiltrating lymphocytes, and anti-CD3 monoclonal antibody-induced killer cells. However, their therapeutic efficacy is limited due to their low anti-tumor activities. CIK cells, a new type of anti-tumor effector cells, can proliferate rapidly *in vitro*, with a stronger anti-tumor activity and a broader spectrum of tumor targets than the reported anti-tumor effector cells. Moreover, CIK cells can regulate and enhance immune function.

Innovations and breakthroughs

CIK cells immunotherapy can decrease levels of tumor markers, change immune functions, and achieve a short-term efficacy against gastric cancer. However, the relation between the frequency of CIK cells immunotherapy and its clinical efficacy has not been examined. In the present study, data obtained from 156 gastric cancer patients were used in fitting multivariate Cox model, showing that more frequencies of CIK cells immunotherapy improve the survival rate of gastric cancer patients.

Applications

The survival time of gastric cancer patients was significantly longer after chemotherapy plus CIK cells immunotherapy than after chemotherapy alone, and more frequencies of CIK cells immunotherapy benefited gastric cancer patients more. This strategy can be applied in treatment of gastric cancer.

Terminology

CIK cells are cytokine-induced killer cells and a new type of anti-tumor effector cells, which can proliferate rapidly *in vitro* with a stronger anti-tumor activity and a broader spectrum of tumor targets than the reported anti-tumor effector cells. Moreover, CIK cells can regulate and enhance immune function.

Peer review

This study showed beneficial effect of CIK cells immunotherapy on gastric cancer, thus improving the 2- and 5-year survival rates of gastric cancer patients. The study is well designed and the data are believable.

REFERENCES

- Shang J, Pena AS. Multidisciplinary approach to understand the pathogenesis of gastric cancer. *World J Gastroenterol* 2005; 11: 4131-4139
- 2 Shen X, Zhang J, Yan Y, Yang Y, Fu G, Pu Y. Analysis and estimates of the attributable risk for environmental and genetic risk factors in gastric cancer in a Chinese population. J *Toxicol Environ Health A* 2009; 72: 759-766
- 3 Varadhachary G, Ajani JA. Gastric cancer. *Clin Adv Hematol Oncol* 2005; **3**: 118-124
- 4 Valentini V, Cellini F, Minsky BD, Mattiucci GC, Balducci M, D'Agostino G, D'Angelo E, Dinapoli N, Nicolotti N, Valentini C, La Torre G. Survival after radiotherapy in gastric cancer: systematic review and meta-analysis. *Radiother Oncol* 2009; 92: 176-183
- 5 **Pisani P**, Parkin DM, Bray F, Ferlay J. Estimates of the worldwide mortality from 25 cancers in 1990. *Int J Cancer* 1999; **83**: 18-29

- 6 Ajani JA. Evolving chemotherapy for advanced gastric cancer. *Oncologist* 2005; **10** Suppl 3: 49-58
- 7 Catalano V, Labianca R, Beretta GD, Gatta G, de Braud F, Van Cutsem E. Gastric cancer. Crit Rev Oncol Hematol 2005; 54: 209-241
- 8 **Dougan M**, Dranoff G. Immune therapy for cancer. *Annu Rev Immunol* 2009; **27**: 83-117
- 9 Stroncek D, Berlyne D, Fox B, Gee A, Heimfeld S, Lindblad R, Loper K, McKenna D Jr, Rooney C, Sabatino M, Wagner E, Whiteside T, Wood D, Heath-Mondoro T. Developments in clinical cell therapy. *Cytotherapy* 2010; **12**: 425-428
- 10 Hontscha C, Borck Y, Zhou H, Messmer D, Schmidt-Wolf IG. Clinical trials on CIK cells: first report of the international registry on CIK cells (IRCC). J Cancer Res Clin Oncol 2010; Epub ahead of print
- Rosenberg S. Lymphokine-activated killer cells: a new approach to immunotherapy of cancer. J Natl Cancer Inst 1985; 75: 595-603
- 12 **Rosenberg SA**, Spiess P, Lafreniere R. A new approach to the adoptive immunotherapy of cancer with tumor-infiltrating lymphocytes. *Science* 1986; **233**: 1318-1321
- 13 Yun YS, Hargrove ME, Ting CC. In vivo antitumor activity of anti-CD3-induced activated killer cells. *Cancer Res* 1989; 49: 4770-4774
- 14 Takahashi H, Nakada T, Puisieux I. Inhibition of human colon cancer growth by antibody-directed human LAK cells in SCID mice. *Science* 1993; 259: 1460-1463
- 15 Schmidt-Wolf IG, Negrin RS, Kiem HP, Blume KG, Weissman IL. Use of a SCID mouse/human lymphoma model to evaluate cytokine-induced killer cells with potent antitumor cell activity. *J Exp Med* 1991; **174**: 139-149
- 16 Schmidt-Wolf IG, Lefterova P, Mehta BA, Fernandez LP, Huhn D, Blume KG, Weissman IL, Negrin RS. Phenotypic characterization and identification of effector cells involved in tumor cell recognition of cytokine-induced killer cells. *Exp Hematol* 1993; 21: 1673-1679
- 17 Jiang J, Xu N, Wu C, Deng H, Lu M, Li M, Xu B, Wu J, Wang R, Xu J, Nilsson-Ehle P. Treatment of advanced gastric cancer by chemotherapy combined with autologous cytokineinduced killer cells. *Anticancer Res* 2006; 26: 2237-2242
- 18 Jiang JT, Wu CP, Shi LR, Xu N, Deng HF, Lu MY, Ji M, Zhu YB, Zhang XG. Side effects during treatment of advanced gastric carcinoma by chemotherapy combined with CIK-cell transfusion in elderly people. *Chin J Clin Oncol* 2008; 5: 79-82
- 19 Jiang J, Zhu Y, Wu C, Shen Y, Wei W, Chen L, Zheng X, Sun J, Lu B, Zhang X. Tumor expression of B7-H4 predicts poor survival of patients suffering from gastric cancer. *Cancer Immunol Immunother* 2010; 59: 1707-1714
- 20 Yoo CH, Noh SH, Shin DW, Choi SH, Min JS. Recurrence following curative resection for gastric carcinoma. *Br J Surg* 2000; 87: 236-242
- 21 Jiang JT, Wu CP, Deng HF, Lu MY, Li M, Wang H, Wu J. Compare cytotoxicity of cytokine-induced killer cells with tumor infiltrating lymphocytes in vitro. *Jianyan Yixue* 2005; 20: 322-324
- 22 De Vita F, Orditura M, Matano E, Bianco R, Carlomagno C, Infusino S, Damiano V, Simeone E, Diadema MR, Lieto E, Castellano P, Pepe S, De Placido S, Galizia G, Di Martino N, Ciardiello F, Catalano G, Bianco AR. A phase II study of biweekly oxaliplatin plus infusional 5-fluorouracil and folinic acid (FOLFOX-4) as first-line treatment of advanced gastric cancer patients. *Br J Cancer* 2005; **92**: 1644-1649
- 23 **Schoenfeld DA**, Richter JR. Nomograms for calculating the number of patients needed for a clinical trial with survival as an endpoint. *Biometrics* 1982; **38**: 163-170
- 24 **Chen L**, Tian H, Chen J, He ZG, Tao SF, Lokesh G, Peng SY. Surgical management of gastric stump cancer: a report of 37 cases. J Zhejiang Univ Sci B 2005; **6**: 38-42
- 25 **Kollmannsberger C**, Budach W, Stahl M, Schleucher N, Hehr T, Wilke H, Schleicher J, Vanhoefer U, Jehle EC, Oechsle K, Trarbach T, Boehlke I, Kanz L, Hartmann JT,



Bokemeyer C. Adjuvant chemoradiation using 5-fluorouracil/folinic acid/cisplatin with or without paclitaxel and radiation in patients with completely resected high-risk gastric cancer: two cooperative phase II studies of the AIO/ARO/ ACO. *Ann Oncol* 2005; **16**: 1326-1333

- 26 Lin WL, Li DG, Chen Q, Lu HM. Clinical and experimental study of oxaliplatin in treating human gastric carcinoma. *World J Gastroenterol* 2004; **10**: 2911-2915
- 27 **Abe M**, Nishimura Y, Shibamoto Y. Intraoperative radiation therapy for gastric cancer. *World J Surg* 1995; **19**: 544-547
- 28 Klautke G, Foitzik T, Ludwig K, Ketterer P, Klar E, Fietkau R. Neoadjuvant radiochemotherapy in locally advanced gastric carcinoma. *Strahlenther Onkol* 2004; 180: 695-700
- 29 Zuo Y, Xu M, Shen D, Lu WD, Lu JF. [Postoperative intraperitioneal hyperthermic chemoperfusion combined with intravenous chemotherapy for 82 advanced gastric cancer patients] *Zhonghua Zhongliu Zazhi* 2004; 26: 247-249
- 30 Bozzetti F, Vaglini M, Deraco M. Intraperitoneal hyperthermic chemotherapy in gastric cancer: rationale for a new approach. *Tumori* 1998; 84: 483-488
- 31 Yu QS, Zhang FZ, Tang XR. [Clinical study on early use of Chinese medicinal herbs and chemotherapy after operation of gastric cancer] Zhongguo Zhongxiyi Jiehe Zazhi 1995; 15: 459-461
- 32 Wang GT. [Treatment of operated late gastric carcinoma with prescription of strengthening the patient's resistance and dispelling the invading evil in combination with chemotherapy: follow-up study of 158 patients and experimental study in animals] *Zhongxiyi Jiehe Zazhi* 1990; **10**: 712-716, 707
- 33 Yamaguchi K, Shimamura T, Hyodo I, Koizumi W, Doi T, Narahara H, Komatsu Y, Kato T, Saitoh S, Akiya T, Munakata M, Miyata Y, Maeda Y, Takiuchi H, Nakano S, Esaki T, Kinjo F, Sakata Y. Phase I/II study of docetaxel and S-1 in patients with advanced gastric cancer. Br J Cancer 2006; 94: 1803-1808
- 34 Herberman RB. Cancer therapy by biological response modifiers. *Clin Physiol Biochem* 1987; **5**: 238-248
- 35 Margolin KA, Negrin RS, Wong KK, Chatterjee S, Wright C, Forman SJ. Cellular immunotherapy and autologous transplantation for hematologic malignancy. *Immunol Rev* 1997; 157: 231-240
- 36 Macdonald JS, Smalley SR, Benedetti J, Hundahl SA, Estes NC, Stemmermann GN, Haller DG, Ajani JA, Gunderson LL, Jessup JM, Martenson JA. Chemoradiotherapy after surgery compared with surgery alone for adenocarcinoma of the stomach or gastroesophageal junction. N Engl J Med 2001; 345: 725-730
- 37 Haynes NM, van der Most RG, Lake RA, Smyth MJ. Immunogenic anti-cancer chemotherapy as an emerging concept. *Curr Opin Immunol* 2008; 20: 545-557
- 38 Wu K, Nie Y, Guo C, Chen Y, Ding J, Fan D. Molecular basis of therapeutic approaches to gastric cancer. J Gastroenterol Hepatol 2009; 24: 37-41
- 39 Larmonier N, Fraszczak J, Lakomy D, Bonnotte B, Katsanis E. Killer dendritic cells and their potential for cancer immunotherapy. *Cancer Immunol Immunother* 2010; 59: 1-11
- 40 Hoffman DM, Gitlitz BJ, Belldegrun A, Figlin RA. Adoptive

cellular therapy. Semin Oncol 2000; 27: 221-233

- 41 Lu PH, Negrin RS. A novel population of expanded human CD3+CD56+ cells derived from T cells with potent in vivo antitumor activity in mice with severe combined immunodeficiency. J Immunol 1994; **153**: 1687-1696
- 42 Gritzapis AD, Dimitroulopoulos D, Paraskevas E, Baxevanis CN, Papamichail M. Large-scale expansion of CD3(+)CD56(+) lymphocytes capable of lysing autologous tumor cells with cytokine-rich supernatants. *Cancer Immunol Immunother* 2002; 51: 440-448
- 43 Johnson BE, Bridges JD, Sobczeck M, Gray J, Linnoila RI, Gazdar AF, Hankins L, Steinberg SM, Edison M, Frame JN, Pass H, Nesbitt J, Holden D, Mulshine JL, Glatstein E, Ihde DC. Patients with limited-stage small-cell lung cancer treated with concurrent twice-daily chest radiotherapy and etoposide/cisplatin followed by cyclophosphamide, doxorubicin, and vincristine. J Clin Oncol 1996; 14: 806-813
- 44 Scheffold C, Brandt K, Johnston V, Lefterova P, Degen B, Schöntube M, Huhn D, Neubauer A, Schmidt-Wolf IG. Potential of autologous immunologic effector cells for bone marrow purging in patients with chronic myeloid leukemia. *Bone Marrow Transplant* 1995; **15**: 33-39
- 45 Schmidt-Wolf IG, Lefterova P, Johnston V, Scheffold C, Csipai M, Mehta BA, Tsuruo T, Huhn D, Negrin RS. Sensitivity of multidrug-resistant tumor cell lines to immunologic effector cells. *Cell Immunol* 1996; 169: 85-90
- 46 Verneris MR, Kornacker M, Mailänder V, Negrin RS. Resistance of ex vivo expanded CD3+CD56+ T cells to Fas-mediated apoptosis. *Cancer Immunol Immunother* 2000; 49: 335-345
- 47 Chen FX, Liu JQ, Zhang NZ, Gong XJ, Zhang GL, Xu YM, Zhou ZH, Wang T, Huang J. [Clinical observation on adoptive immunotherapy with autologous cytokine-induced killer cells for advanced malignant tumor] *Ai Zheng* 2002; 21: 797-801
- 48 Wu C, Jiang J, Shi L, Xu N. Prospective study of chemotherapy in combination with cytokine-induced killer cells in patients suffering from advanced non-small cell lung cancer. *Anticancer Res* 2008; 28: 3997-4002
- 49 Olioso P, Giancola R, Di Riti M, Contento A, Accorsi P, Iacone A. Immunotherapy with cytokine induced killer cells in solid and hematopoietic tumours: a pilot clinical trial. *Hema*tol Oncol 2009; 27: 130-139
- 50 Kim HM, Lim J, Yoon YD, Ahn JM, Kang JS, Lee K, Park SK, Jeong YJ, Kim JM, Han G, Yang KH, Kim YJ, Kim Y, Han SB. Anti-tumor activity of ex vivo expanded cytokine-induced killer cells against human hepatocellular carcinoma. *Int Immunopharmacol* 2007; 7: 1793-1801
- 51 Cho MY, Joh YG, Kim NR, Jung SI, Bae JW, Kim YC, Koo BH, Whang CW, Suh SO. T-lymphocyte subsets in patients with AJCC stage III gastric cancer during postoperative adjuvant chemotherapy. American Joint Committee on Cancer. *Scand J Surg* 2002; **91**: 172-177
- 52 Catalano V, Labianca R, Beretta GD, Gatta G, de Braud F, Van Cutsem E. Gastric cancer. Crit Rev Oncol Hematol 2009; 71: 127-164

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