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Atrial Defibrillation Voltage: Falling to a New Low

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The mechanisms by which an electric shock terminates cardiac arrhythmias have been the subject of a large body of research (for review, see¹). These studies have been driven by the understanding that a significant reduction in shock energy can only be achieved by full appreciation of the mechanisms by which a shock interacts with the heart and then exploiting them to devise novel low-voltage therapeutic approaches.

The research has demonstrated that the response of the myocardium to the shock involves simultaneous occurrence of positive and negative membrane polarization^{2–4}. Detailed analysis of the etiology of this “virtual electrode polarization” (VEP) has demonstrated that tissue structure is responsible for its formation of VEP as well as for its shape, location, polarity, and intensity. The effect of tissue structure is two-fold. First, discontinuities in tissue structure (i.e. conductivities) force current to cross the membranes of neighboring cells, giving rise to VEP. Intercellular and interlaminar clefts^{5,6}, or tissue lesions⁷ are possible factors in this process. Second, continuous tissue structure such as ventricular shape and fiber architecture also give rise to VEP⁸.

Action potential duration in the myocardium can be either extended by positive VEP or shortened by negative VEP, and strong negative VEP can completely abolish the action potential, creating a new, post-shock excitable area⁹. Propagation through the post-shock excitable area has proven to directly determine the outcome of a defibrillation shock. A shock succeeds in extinguishing fibrillatory wavefronts if excitations manage to traverse the newly-created post-shock excitable area before the rest of the myocardium recovers from refractoriness. Decreasing the post-shock excitable area could thus increase the likelihood of defibrillation success and lower the defibrillation voltage¹⁰; however, this has proven difficult since the post-shock excitable areas are often hidden deep in the ventricular wall^{10,11}.

When the shock happens to affect resting cells (those that are part of the pre-shock, fibrillatory wavefront’s excitable gap), positive VEPs immediately depolarize these cells; these cells become “secondary sources” emitting new wavefronts, rapidly overwhelming any effects of negative VEP. The targeted activation of cells in the fibrillatory wavefront’s excitable gap by electric shocks thus has the potential to eliminate the reentrant circuit by rendering tissue refractory. Furthermore, since this is an excitation process (by positive VEP), the external current needs to only bring cells to the excitation threshold, requiring less energy as compared to de-excitation (by negative VEP) where an activated cell is forced into premature recovery.

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The figure illustrates this process. It presents a simulation of a diastolic excitation of the right ventricular (RV) wall of a high-resolution rabbit heart model¹², by a brief (5ms) low-voltage electric field directed across the RV wall; short- and long-axis views of the wall are shown. Positive VEP is generated at numerous locations, giving rise to subsequent activations. The endocardial deep valleys created by the trabeculations serve as “lenses”, focusing the external current, bringing the respective tissue regions to the excitation threshold, and rendering them as “secondary sources”. As simulations with this high-resolution 3D structure model¹² demonstrate, this is an important mechanism by which low-voltage shocks generate numerous new activations in the excitable regions of the heart.

The larger the number of positive VEPs resulting in excitation, the faster the activation of the entire excitable region occurs¹³. Even partial activation of the excitable gap by positive VEPs, particularly close to the crest of the wave, could result in defibrillation success since it could speed the wave by inducing a shift in the wave tip¹⁴, resulting in subsequent termination of reentry¹⁵.

The question is: how to best achieve such targeted activation of the reentrant wave’s excitable gap? A single low-voltage shock is unlikely to be effective since its ability to activate the gap depends on the timing of its delivery. Delivery of a train of several low-voltage pulses is the logical alternative, since it offers independence of the therapy outcome from the phase of the reentrant wave, as demonstrated by Ripplinger et al¹⁶ in the first low-voltage termination of ventricular tachycardia in a rabbit heart with chronic infarction. Interestingly, a train of monophasic pulses was found¹⁶ more effective in recruiting cells in the excitable gap than biphasic pulses, as the reversed phase of the biphasic pulse “undermined” the excitatory effect of the first phase, resulting in VEPs of decreased magnitude in the excitable gap..

What should the frequency of the low-voltage pulses be to achieve maximum efficacy in activating cells in the reentrant wave excitable gap? Fenton et al¹⁷ applied this concept to the termination of atrial reentrant arrhythmias; they used a train of monophasic pulses with a cycle length 5–10ms below the dominant cycle length of the arrhythmia. The choice of cycle length was possibly based on the practice of anti-tachycardia pacing, which delivers a train of stimuli coordinated with the arrhythmia cycle length in an attempt to disrupt the reentrant circuit by invading, with an outside wavefront, the excitable gap of the reentry. Fenton et al achieved atrial arrhythmia termination (flutter and fibrillation were not reported separately) at a low-voltage strength (0.9–1.4 V/cm) with a success rate of 93%.

The new study by Ambrosi et al¹⁸ in this issue of the Journal took low-voltage termination of atrial arrhythmias one step further towards an effective bioelectric therapy. Multiple monophasic pulses were applied within one or two cycle lengths of the arrhythmia. The benefit of the multiple pulses being delivered within one or two arrhythmia cycles is that at least one (or more) of the pulses will have a favorable timing for activating cells in the reentry’s excitable gap via VEP, rather than “falling on top” of the propagating wave. Ambrosi et al offered another important benefit of their approach: such a delivery of the pulse train would avoid the coincidental delivery of the atrial defibrillation/cardioversion therapy on the T-wave, which could result in ventricular arrhythmia. The thresholds for atrial arrhythmia termination with the approach by Ambrosi et al were 0.86 (for shocks within one cycle length of the arrhythmia) and 0.28 V/cm (within two cycle lengths) for atrial flutter, and 3.46 V/cm (within one cycle) for atrial fibrillation, all for 100% success of termination. For atrial flutter, this is 2.5- and 7.6-fold decrease in shock strength compared to a single shock, respectively; for atrial fibrillation, the decrease is 2-fold. The difference in termination thresholds for atrial flutter and fibrillation is most likely due to the smaller excitable gap in fibrillation as well as the fact that the constant meander of reentrant

wavefronts leads to dynamic changes in the locations of the excitable gaps, making excitation of the cells in these gaps by the same pulse train less likely to occur.

The new studies in low-voltage defibrillation, such as that by Ambrosi et al¹⁸, demonstrate that the quest to achieve termination of arrhythmias with electrical therapy, the strength of which is below the pain threshold, is in full swing. We thus anxiously await the new lows in defibrillation voltage.

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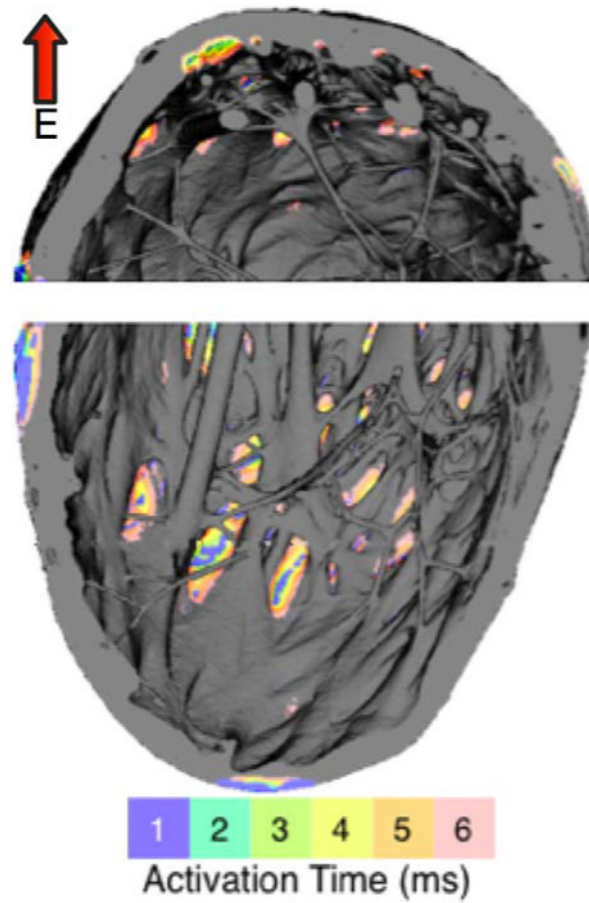


Figure.
Simulated activation pattern on the endocardial surface of the rabbit heart RV wall resulting from the application of a brief uniform electric field (E) in direction perpendicular to the middle of the wall (red arrow).