# Influence of Calcium on the Inotropic Actions of Hyperosmotic Agents, Norepinephrine, Paired Electrical Stimulation, and Treppe

JAMES T. WILLERSON, J. STANLEY CRIE, ROBERT C. ADCOCK, GORDON H. TEMPLETON, and KERN WILDENTHAL

From the Pauline and Adolph Weinberger Laboratory for Cardiopulmonary Research, Departments of Internal Medicine and Physiology, The University of Texas Southwestern Medical School, Dallas, Texas 75235

ABSTRACT To analyze the interaction of calcium ion concentration with hypertonic agents and with other inotropic interventions, isolated right ventricular cat papillary muscles were studied under isometric conditions in Krebs-Ringer bicarbonate solution. Extracellular calcium concentrations were varied between 2.5 and 11.0 mM. Maximal inotropic effects occurred between 5 and 8.0 mM calcium and further elevation to 11.0 mM was without additional influence. The effect of hyperosmotic sucrose and mannitol on papillary muscle performance was compared with that of 10-6 M norepinephrine at calcium concentrations of 2.5 and 10.0 mM and with paired electrical stimulation in 10.0 mM calcium. Both norepinephrine and the hyperosmotic agents produced significant increases in developed tension and in the maximal rate of tension rise (dT/dt) in Krebs-Ringer in 2.5 and 4.0 mM calcium. In 10 mM calcium norepinephrine increased developed tension and dT/dt, but sucrose and mannitol caused no change or small reductions in both. Paired electrical stimulation, like hyperosmolality, caused no increase in dT/dt in 10 mM calcium.

The presence of a potent pharmacological inhibitor of systolic calcium transfer across the cell membrane (D600,  $10^{-6}$  M) reduced developed tension and  $\mathrm{d}T/\mathrm{d}t$  by  $76\pm2.7$  and  $74\pm2.0\%$ , respectively, and prevented and in fact reversed the expected increase in  $\mathrm{d}T/\mathrm{d}t$  associated with an increase in rate of stimulation (treppe). However, hypertonic mannitol and paired

pacing persisted in causing marked increases in developed tension and  $\mathrm{d}T/\mathrm{d}t$  even in the presence of D600, suggesting that their inotropic effects are not dependent on increased intracellular transfer of calcium during systole through cell membrane channels in which D600 acts as a competitive inhibitor.

The results of these studies suggest that apparent functional saturation of intracellular calcium receptor sites eliminates any additional inotropic effect of hyperosmolality or paired pacing. The data are compatible with the hypothesis that the inotropic effects of hyperosmolality and of paired pacing result from an increase in calcium concentration at the myofilaments during contraction. The increase induced by hyperosmolality might occur because of an increase in the total amount of calcium released into the cytosol with each action potential and/or as a passive consequence of cellular dehydration. Norepinephrine has the capacity to increase contractility even when intracellular calcium receptor sites appear to be functionally saturated, suggesting that it may act at least in part by a mechanism that is independent of changes in net intracellular calcium concentration.

### INTRODUCTION

The net inotropic effect of many interventions can vary widely depending on the extracellular concentration of calcium (1-5). Undoubtedly, part of the reason for this is simply that the positive inotropic influence of high calcium, per se, causes improvement in muscle function to near-maximal levels, so that subsequent inotropic interventions can add little more to the muscle's performance. In addition, if a given agent acts by changing intracellular calcium concentration by one

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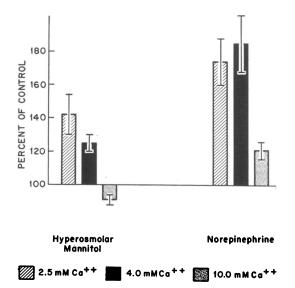


FIGURE 1 The influence of mannitol and norepinephrine on developed tension at extracellular calcium concentrations of 2.5, 4.0, and 10.0 mM is shown. The figure demonstrates that mannitol loses its inotropic effect in 10.0 mM calcium whereas that of norepinephrine is reduced but persists.

means or another, it might be supposed that prior elevation of extracellular calcium concentration above a critical level could preempt any changes the agent might otherwise produce.

Recent studies have demonstrated that hyperosmotic mannitol and sucrose have an inotropic effect in intact animals (6, 7) and in isolated heart muscle (8, 9). Hyperosmotic agents have also been shown to improve ventricular function, reduce the extent of myocardial injury, and improve total and collateral coronary blood flow during acute coronary insufficiency in anesthetized animals (10) and to reduce hypoxic depression in isolated cat papillary muscle (11). When osmolality rises to over 100 mosmol above control levels (>400 mosmol/kg H<sub>2</sub>O), both the contractile elements and the viscoelastic components of the myocardium are altered, but below that level hyperosmolality affects only the contractile elements of the heart (9, 12). The mechanism by which hyperosmotic agents exert their inotropic effect on contractile elements is not yet known, but Koch-Weser (8) has suggested that they may act by causing an increase in the concentration of intracellular calcium available at the myofilaments, perhaps as a passive consequence of cellular dehydration. Little and Sleator (13) have shown that hyperosmotic solutions increase the intracellular transfer of calcium from extracellular sources; however, their calculations indicated that the observed changes in calcium transfer were probably of insufficient magnitude to explain the entire inotropic effect of hyperosmolality. No other data have been available to support the contention that calcium concentration per se is linked to the inotropic effect of hyperosmotic agents.

Accordingly, the present experiments were designed to explore the interactions of the inotropic effects of hyperosmolality and increased calcium concentration. Particular attention was directed at determining whether or not prior elevation of extracellular calcium above a critical level abolishes the inotropic effect of hyperosmolality. For comparison, the interactions of calcium and paired electrical stimulation and of calcium and norepinephrine were analyzed similarly. In addition, studies were made of the ability of pharmacological blockade of systolic calcium flux to abolish the inotropic actions of hyperosmolality, paired electrical stimulation, and treppe.

#### **METHODS**

Isolated right ventricular papillary muscles were obtained from cats that had been anesthetized with intraperitoneal sodium pentobarbital (60 mg/kg). After midline thoracotomy, the heart was excised and a papillary muscle was removed from the right ventricle and transferred to a muscle bath. The bath contained Krebs-Ringer bicarbonate solution with 18 mM glucose and calcium of 2.5-10.0 mM. The medium was equilibrated with 95% O<sub>2</sub> plus 5% CO<sub>2</sub>, and the temperature was held constant by an external water bath. The muscles, which contracted isometrically, were held between spring-loaded clips one of which formed the end of a rigid pin that penetrated the bottom of the bath and attached directly to a Statham G1-4-250 force transducer (Statham Instruments, Inc., Oxnard, Calif.). The end of the muscle with chordae attached was connected to a lever mounted on a rigid Palmer stand. The muscles were stimulated with impulses (20% above threshold) through platinum electrodes placed parallel to the long axis of the muscles. Tension and its first derivative (dT/dt)

TABLE I

Effect of Increasing Extracellular Calcium Concentration
on Papillary Muscle Performance

Number of muscles	Extra- cellular calcium concen- tration	Resting tension ±1 SEM	Developed tension ±1 SEM	Maximal rate of tension rise ±1 SEM	
	mM	g/mm²	g/mm²	g/mm²/s	
8	2.5	$0.4 \pm 0.08$	$4.5 \pm 0.65$	$24 \pm 3.2$	
8	5.0	$0.3 \pm 0.05$ §	$6.5 \pm 0.83*$	$40 \pm 5.8*$	
8	8.0	$0.2 \pm 0.04 \ddagger$	$6.9 \pm 0.95 \ddagger$	$45 \pm 7.6$ §	
8	11.0	$0.2 \pm 0.02$ §	$6.8 \pm 0.96$ §	$45 \pm 7.3$ §	

The statistical comparisons are made between the calcium value for which the P value is provided and the calcium value immediately preceding it.

<sup>\*</sup> P < 0.001.

P < 0.05.

 $<sup>\</sup>S P > 0.05.$ 

were recorded on a Hewlett-Packard direct writer (Hewlett-Packard Co., Palo Alto, Calif.). Values were subsequently corrected for cross-sectional area. The mean cross-sectional area for the papillary muscles studied was 0.95 ±0.07 (SEM) mm<sup>3</sup>.

Eight papillary muscles, contracting at 12/min at an initial resting tension of 1 g, were allowed to stabilize for 30-60 min at 29°C in Krebs-Ringer bicarbonate solution containing 2.5 mM calcium. After initial stabilization the pacing rate and temperature were increased (to 20/min and 37°C, respectively) to facilitate intracellular calcium transfer (14). After a 30-min period of stabilization the length of the muscle at which maximal tension development was present (Lmax) was determined. Control tensions and their maximum derivatives (max dT/dt) were then recorded, the muscle bath was rapidly emptied of its fluid content and replaced with an identical medium containing 5 mM calcium rather than the initial 2.5 mM. Another 30min stabilization period followed, and tensions and max dT/dt were again recorded. Subsequently solutions containing 7.5 and 10.0 mM calcium were evaluated in an identical manner.

Another group of eight muscles was studied in 2.5 mM calcium at Lmax and 37°C and a rate of 20/min after stabilization periods as described above. After control tensions and max dT/dt were recorded, the fluid in the bath was rapidly replaced with a new Krebs-Ringer bicarbonate solution supplemented with either mannitol (50 mosmol above control) or sucrose (90 mosmol above control). Another 30-min stabilization period was provided, and tensions and max dT/dt were again recorded. The muscles were again immersed in the control solution for 30 min and repeat control contractions were recorded. Finally nor-epinephrine-HCl (10 $^{-6}$  M) was added to the bath and tension was recorded after 3 $^{-4}$  min.

22 additional papillary muscles were tested in an identical manner except that after initial stabilization at 2.5 mM calcium, the bathing medium was switched to one containing either 4.0 or 10.0 mM calcium and the rest of the experiment was performed at that level.

Eight papillary muscles were evaluated in 10.0 mM calcium under conditions identical to those previously described. After final stabilization and recording of control values, paired pacing was instituted. The absolute refractory period was determined by gradually increasing the delay between the first and second pacing stimulus by 10-ms intervals until the first double peak contour was obtained in the developed tension tracing. Sustained paired pacing was then continued for a 2-3-min period while tensions and max dT/dt were recorded. Peak developed tension was obtained from the maximal contraction after the initiation of paired pacing. This was nearly always the first or one of the first contractions after the onset of effective paired pacing. Only those contractions that were free of after contractions were used (15).

A final group of nine muscles was treated with D600, a potent pharmacological inhibitor of systolic calcium transfer at the cell membrane (5). The muscles were allowed to stabilize in control solution at  $30^{\circ}$ C and 12 contractions/min, after which  $10^{-6}$  M of D600 was added to the bath. After 1-2 h, when contractile depression had stabilized, paired pacing was instituted as described earlier. The paired pacing was discontinued after a 2-3-min period and the muscle allowed to restabilize. Hyperosmotic mannitol was then added and tensions and dT/dt recorded when stable. In four muscles the effect of increasing the stimulation rate

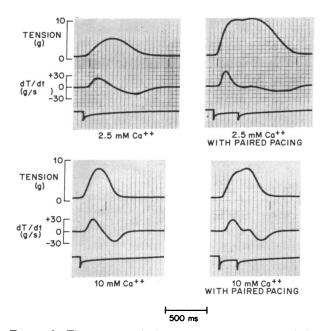


FIGURE 2 The top panel demonstrates the characteristic effect of paired pacing in increasing developed tension and dT/dt in 2.5 mM calcium at 37°C. The bottom panel demonstrates the absence of a significant positive inotropic effect of paired pacing when the calcium concentration is 10 mM.

(treppe effect) after the addition of D600 was also determined.

Statistical comparisons were made using Student's t test for paired observations. Differences were considered significant when P was less than 0.05.

## RESULTS

Calcium dose-response curve. Eight papillary muscles were studied to obtain a dose-response curve to increasing concentrations of extracellular calcium at the pacing rate of 20/min and temperature of 37°C. There were significant increases in both developed tension and max  $\mathrm{d}T/\mathrm{d}t$  as the extracellular calcium concentration was increased from 2.5 to 8.0 mM (Table I). Developed tension increased 48 and 57% in 5.0 and 7.5 mM calcium, while maximal  $\mathrm{d}T/\mathrm{d}t$  increased 67 and 88%. There was no further significant increase in either developed tension or max  $\mathrm{d}T/\mathrm{d}t$  as the extracellular calcium concentration was further increased from 8.0 to 11.0 mM.

There was a significant fall in resting tensions as extracellular calcium concentration increased from 2.5 to 8.0 and 11.0 mM, possibly as the result of stress relaxation (16) or possibly simply because of a slight instability of the preparation with the passage of time.

Hyperosmotic mannitol and sucrose. Hyperosmotic mannitol (50 mosmol above control) caused increases in total developed tension of 38% in the solution con-

Table II

Effect of Hypertonic Sucrose and Mannitol and 10<sup>-6</sup> M Norepinephrine on Papillary

Muscle Performance at Varying Calcium Concentrations

Number of muscles	[Ca++]	Intervention	Resting tension ±1 SEM	Developed tension ±1 SEM	Maximal rate of tension rise ±1 SEM
			g/mm²	g/mm²	g/mm²/s
4	2.5	Control	$1.0 \pm 0.41$	$5.9 \pm 0.93$	· 40±5.8
4	2.5	Sucrose	$0.7 \pm 0.32*$	$8.3 \pm 0.99*$	$63 \pm 5.3*$
4	2.5	Control 2	$0.9 \pm 0.52$	$4.9 \pm 0.69$	$33 \pm 4.4$
4	2.5	Norepinephrine	$0.8 \pm 0.46$	$8.6 \pm 0.89 \ddagger$	$74 \pm 3.7*$
6	10.0	Control	$1.1 \pm 0.19$	$6.9 \pm 1.01$	$59 \pm 10.6$
6	10.0	Sucrose	$0.9 \pm 0.16$	$6.4 \pm 0.91$	$54 \pm 9.4$
6	10.0	Control 2	$0.6 \pm 0.12$	$6.2 \pm 1.04$	$52 \pm 10.0$
6	10.0	Norepinephrine	$0.6 \pm 0.14$	$7.3 \pm 1.01*$	$62 \pm 9.2 \ddagger$
5	2.5	Control	$1.2 \pm 0.23$	$3.7 \pm 1.16$	$24 \pm 2.5$
5	2.5	Mannitol	$0.8 \pm 0.16*$	$5.1 \pm 1.40 \ddagger$	$39 \pm 11.6$ §
5	2.5	Control 2	$0.7 \pm 0.20$	$3.2 \pm 0.86$	$20 \pm 5.6$
5	2.5	Norepinephrine	$0.7 \pm 0.20$	$5.3 \pm 1.10*$	$49 \pm 8.8*$
8	10.0	Control	$1.6 \pm 0.30$	$9.7 \pm 0.76$	$82 \pm 8.1$
8	10.0	Mannitol	$1.3 \pm 0.26$ §	$9.0 \pm 0.73 \ddagger$	$75 \pm 8.4$ §
6	10.0	Control 2	$1.1 \pm 0.27$	$9.0 \pm 0.79$	$74 \pm 9.8$
6	10.0	Norepinephrine	$1.2 \pm 0.34$	$10.9 \pm 0.99$ ‡	$93 \pm 12.2*$

Statistical comparisons are made between each value and the control immediately preceding it.

taining 2.5 mM calcium and of 26% in 4.0 mM calcium. At 10 mM calcium a similar concentration of mannitol caused no change or a small decrease in total tension (mean -8%) (Fig. 1; Table II). The maximal rate of tension rise changed similarly (Table II).

As with mannitol, developed tension and  $\mathrm{d}T/\mathrm{d}t$  rose significantly when hyperosmotic sucrose (90 mosmol above control) was added to a solution containing 2.5 mM calcium but remained constant or fell in the presence of 10.0 mM calcium (Table II). With both hyperosmotic agents, resting tension tended to remain constant or to fall slightly, indicating that neither contracture nor a decrease in compliance had occurred in the muscles. The decreases in resting tension were especially marked after 30 min exposure to the hyperosmotic agents in 2.5 mM calcium, suggesting that the chronic inotropic change that had occurred had induced some stress relaxation.

Norepinephrine. The addition of  $10^{-6}$  M norepinephrine significantly increased developed tension and max  $\mathrm{d}T/\mathrm{d}t$  in 2.5, 4.0, and 10.0 mM calcium environments (Fig. 1; Table II). Increases in developed tension and max  $\mathrm{d}T/\mathrm{d}t$  were less in high calcium, but

consistent increases persisted even at the highest concentration. Thus, developed tension rose by 21% and max dT/dt by 26% after the addition of  $10^{-6}$  M norepinephrine to a solution containing 10.0 mM calcium. There was no significant change in resting tension.

Paired pacing in the presence of 10.0 mM calcium. To determine whether 10.0 mM extracellular calcium concentrations provided maximal functional intracellular calcium concentrations, eight additional papillary muscles were studied (Fig. 2; Table III). Maximal dT/dt was not significantly changed by the use of paired pacing in the presence of 10.0 mM calcium at 37°C and a pacing rate of 20/min. Developed tension increased slightly in five of eight muscles studied. The time to peak tension also increased in these same five muscles. As noted earlier, the maximal effect of paired pacing was usually noted during the first few beats. Thereafter, sustained paired pacing produced a progressive decrease in developed tension and max dT/dt often coinciding with the appearance of after contractions.

Influence of D600 on inotropic effect of hyperosmotic mannitol, paired pacing, and the treppe effect. D600

<sup>\*</sup> P < 0.001.

P < 0.01.

<sup>§</sup> P < 0.05.

<sup>||</sup> Two of the papillary muscles developed arrhythmias when norepinephrine was added.

reduced developed tension and dT/dt by  $76\pm2.7\%$  (P < 0.001) and  $74\pm2.0\%$  (P < 0.001), respectively (Fig. 3). Paired pacing and hyperosmotic mannitol produced significant increases in developed tension and dT/dt even in the presence of D600 (Fig. 3). In contrast the positive treppe effect which is characterized by an increase in dT/dt with increasing rate of stimulation was prevented and in fact reversed by D600 (Fig. 4). The influence of D600 on increases in developed tension as a consequence of treppe was directionally similar but not as dramatic.

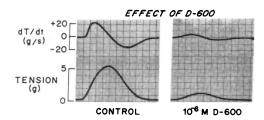
#### DISCUSSION

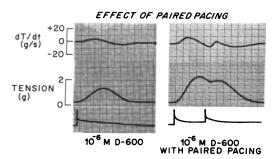
Intracellular calcium levels rise in direct relation to increases in extracellular calcium, as a consequence of increased binding to sarcoplasmic reticulum, sarcolemma, mitochondria, and other cellular storage sites (17, 18). Correspondingly, an increased amount of calcium is released into the cytoplasm during excitation, resulting in greater binding of calcium to troponin, increased formation of actin and myosin cross bridges, and increased contractile strength (19). The maximal

TABLE III

Effect of 10 mM Calcium and 10 mM Calcium Combined
with Paired Pacing on Papillary Muscle Performance

	Resting tension	Developed tension	Maximal rate of tension rise	Time to peak tension
	g/mm²	g/mm²	$g/mm^2/s$	ms
10 mM	calcium			
1	0.2	5.3	46	220
2	0.5	8.5	90	190
3	0.8	7.2	70	220
4	0.4	5.1	33	300
5	0.5	3.7	35	210
6	0.7	3.5	35	240
7	0.4	4.9	41	200
8	0.4	3.9	39	180
Mean	0.5	5.3	49	220
SE	0.07	0.62	7.3	13.2
Paired p	acing and 10	mM calcium		
1	0.2	6.2	46	270
2	0.5	8.5	90	200
3	0.8	7.2	70	220
4	0.4	5.5	33	400
5	0.5	3.9	34	290
6	0.7	3.5	35	230
7	0.4	5.3	41	300
8	0.4	4.1	39	230
Mean	0.5	5.5	49	268
SE	0.07	0.61	7.3	22.7
	NS	P < 0.05	NS	P < 0.02





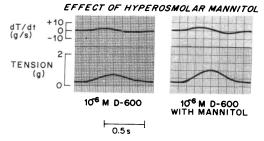
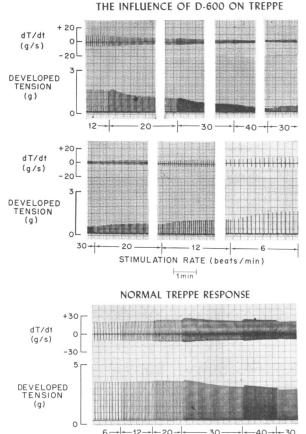


FIGURE 3 The top panel shows the depressant effect of a 1-h exposure to D600. The middle panel demonstrates that paired pacing still produces a significant increase in developed tension and  $\mathrm{d}T/\mathrm{d}t$ , even in the presence of D600. The bottom panel demonstrates that mannitol also continued to exert its inotropic effect even in the presence of D600, as evidenced by the increases in both tension and  $\mathrm{d}T/\mathrm{d}t$  after mannitol.

inotropic effect produced by increasing extracellular calcium concentrations in the present study, as in others (4, 14), was observed to occur between 5.0 and 8.0 mM; there was no further increase as the extracellular calcium concentration was increased to 11.0 mM. This suggests that intracellular calcium storage sites become functionally saturated with calcium as a consequence of the elevations of extracellular calcium, even though the total amount of bound calcium may continue to increase as extracellular concentrations rise still higher (18).

Like increased extracellular calcium concentration, paired electrical stimulation is thought to increase contractility by increasing the amount of calcium available at the myofilaments (1, 2, 20). As has been reported previously under other experimental conditions (1), paired pacing provided no additional increase in the



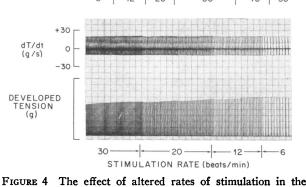


FIGURE 4 The effect of altered rates of stimulation in the presence (top panel) and absence (bottom panel) of D600 on isometrically contracting cat papillary muscles. The characteristic immediate increases in  $\mathrm{d}T/\mathrm{d}t$  that accompany increased stimulation rate are prevented and reversed in the presence of D600. The effect of D600 on immediate changes in developed tension with increased stimulation rate is directionally similar but less marked. Note that the immediate increases in developed tension with increased stimulation rate are not sustained at the higher pacing rates even in the absence of D600.

rate of tension development in muscles exposed to maximal calcium in this study. The small increase in total tension development that occurred with paired pacing was a function of a prolongation in the time to peak tension rather than an increase in  $\mathrm{d}T/\mathrm{d}t$ . Thus, it appears that only the duration of the active state was changed, and not its intensity. It is the latter that is best correlated with the amount of calcium present at the myofilaments during contraction (21). These results suggest that the myofilament receptor complexes are functionally saturated when extracellular calcium levels exceed 8.0 mM, when a temperature of 37°C and pacing rate of 20/min are present.

On the other hand, 10<sup>-6</sup> M norepinephrine did produce a modest but significant increase in developed tension and in the maximal rate of tension rise over and above that resulting from maximal inotropic concentrations of extracellular calcium. This finding is not in agreement with some previous analyses of calcium-norepinephrine interaction (4). The reason for this discrepancy is not readily apparent, though it may be relevant to note that temperature, pacing rate, and glucose concentrations were all higher in our experiments. Whatever the reason, it appears from our results that under some conditions, norepinephrine retains the capacity to increase contractility in the face of maximal external calcium concentrations at a time when paired pacing is ineffective. At present it is not clear whether this reflects an ability of norepinephrine to increase some critical fraction of intracellular calcium above that possible by increasing extracellular concentrations of calcium or by paired pacing, or whether it implies that at least a portion of norepinephrine's inotropic potential is not calcium dependent. Additional investigations will be needed to settle this question.

The inotropic effect of hyperosmolality is critically influenced by extracellular calcium concentration. Like paired pacing, hyperosmolality failed to increase contractility when extracellular calcium was 10.0 mM. On the other hand, inhibition of intracellular systolic transfer of calcium with D600, a potent pharmacological inhibitor (5), failed to prevent the inotropic effect either of hyperosmotic mannitol or of paired pacing, though it did reduce and reverse the expected increase in dT/dt resulting from increasing rate of stimulation (treppe effect) just as has been reported previously for verapamil, another inhibitor of intracellular transfer of calcium at the cell membrane (22).

Together, these observations suggest that the inotropic effect of hyperosmotic mannitol and of paired pacing, is dependent on changes in intracellular calcium but is not dependent on increases in the systolic transfer of calcium through the membrane channels that carry the slow inward current.

It remains possible, however, that calcium transfer across the cell membrane might be increased through channels that are not influenced by D600, or that hyper-

osmolality might cause an increase in the amount of calcium released from intracellular storage sites during each action potential, as has been suggested to occur with paired stimulation. Also, augmentation of contraction by some mechanism that is not mediated by calcium remains a potential explanation for the persistent inotropic effect of mannitol after D600 blockade. Another and more obvious explanation is that, as Koch-Weser has implied, hyperosmolality may affect cardiac performance at least in part by passively increasing calcium availability to the myofilaments during contraction, as a consequence of cellular dehydration and a reduction in cell size (8). Cellular dehydration after exposure to severely hyperosmotic solutions (2-3× isotonicity) is manifested by gross swelling of the Ttubular system and a marked shrinkage in cell size (23). Others have shown that lesser degrees of hyperosmolality, such as were used in the present study, result in a loss of tissue water from beating heart tissues of approximately 10% (24). Koch-Weser (8) has shown that such a change occurs gradually over a period of 30 min, by which time a new steady state has developed. This loss of tissue water would result passively in a corresponding increase in the concentration of cytoplasmic calcium for any given number of ions that are released into the cytoplasm (25). Nayler (26) has suggested a similar mechanism to explain the interaction of calcium and severe hypertonicity (two to three times normal) in determining the severity of contracture in toad myocardium.

The failure of hyperosmotic mannitol and sucrose to produce a significant increase in developed tension and dT/dt in 10.0 mM calcium is not simply a reflection of the muscles already contracting maximally, inasmuch as increases in performance were observed in response to norepinephrine in the same muscles under the same conditions. It might theoretically be possible that substrate limitation at the relatively high pacing rate (20/min) and temperature (37°C) could inhibit responsiveness to hyperosmolality but not to a glycogenolytic agent such as norepinephrine. Such a possibility was considered in these studies, and additional experiments were performed in three papillary muscles at a pacing rate of 12/min and a temperature of 29°C. These are conditions in which there would be much less chance that substrate availability might be a limiting factor. In each of these muscles developed tension and max dT/dt remained constant or declined after the addition of hypertonic mannitol to a solution containing 10.0 mM calcium but rose after norepinephrine. In addition, the high glucose concentration (18 mM) used in these experiments argues against the possibility that substrate limitation could be an explanation

for the differences in the response to hyperosmotic agents and norepinephrine.

We were also concerned by the possibility that the observed unresponsiveness to hyperosmolality at extracellular calcium concentration of 10.0 mM might have been a consequence of general deterioration of the preparation with time. Accordingly, two additional experiments were made in which fresh muscles were placed into 10.0 mM calcium after a brief equilibration period in 2.5 mM calcium and subsequently exposed to mannitol after a very short period in vitro. Results in these muscles were identical to those described earlier: mannitol had no inotropic effect.

In summary, the present study provides further evidence that hyperosmolality, paired electrical stimulation, and norepinephrine exert a potent inotropic influence on isolated cardiac muscle at normal calcium concentrations. In addition, the results show that the effectiveness of hyperosmolality, like that of paired electrical stimulation, is critically influenced by extracellular calcium concentration, but that the inotropic effect of norepinephrine, while reduced, is not prevented by maximal functional extracellular calcium concentrations. Moreover, inhibition of systolic calcium flux by a potent pharmacological inhibitor, D600, does not prevent the inotropic response to hypertonic mannitol. Finally, it is of special interest that D600 was found to reduce and in fact reverse the expected increase in dT/dt with treppe but not that of paired electrical stimulation. Thus, the drug's inhibition of the cell's ability to transport calcium through channels that carry the slow inward current during systole (5, 22) seems to disclose a fundamental difference in the mechanism of action of the two interventions. Further definition of this difference should provide a fruitful area for future study.

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