

Oxaliplatin: a review in the era of molecularly targeted therapy

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ABSTRACT

Objective

To review preclinical and clinical data for oxaliplatin in the current context of molecularly targeted therapy.

Methods of Study Selection

We searched the PubMed and PubChem databases by combining the search terms "oxaliplatin" or "platinum" or both, with "clinical trials," "pharmacokinetics," and "pharmacodynamics."

Data Extraction and Synthesis

Oxaliplatin has a complicated pharmacokinetic profile, with activity against digestive cancers in particular. It has several mechanisms of action, but cancer cells can develop resistance. Real or potential synergism has been observed when oxaliplatin is combined with other cytotoxic agents or molecularly targeted agents. Peripheral neuropathy is a prominent toxic effect.

Conclusions

Oxaliplatin lends itself to further clinical research in combination with molecularly targeted therapy.

KEY WORDS

Oxaliplatin, targeted therapy, chemotherapy, mechanism of action

1. INTRODUCTION

The advent of molecularly targeted anticancer therapy could cause a certain lack of interest in the development of novel conventional cytotoxic drugs. However, molecularly targeted agents have not shown any curative properties when administered as monotherapy. In that regard, there is hope in combining

molecularly targeted anticancer therapy with conventional cytotoxic chemotherapy. Such combinations can come about only through rational design of clinical trials, taking into account the pharmacology and clinical development of the drugs involved. It is therefore worthwhile revisiting classical chemotherapy agents, because this renewed knowledge could provide a foundation for future trials.

Oxaliplatin is the newest platinum derivative in standard chemotherapy. Here, we review oxaliplatin from the pharmacologic and drug development perspectives, and we comment on possible associations of this drug with molecularly targeted therapy.

2. CHEMICAL AND PHYSICAL PROPERTIES AND BIOTRANSFORMATION

Oxaliplatin differs from cisplatin in that the amine groups of cisplatin are replaced by diaminocyclohexane (DACH). The molecular weight of oxaliplatin is 397.3. It is slightly soluble in water, less so in methanol, and almost insoluble in ethanol and acetone 1. Its full chemical name, oxalato(trans-L-1,2-diaminocyclohexane)platinum, refers to the presence of an oxalate "leaving group" and the DACH carrier ligand, which are responsible, at least in part, for its unique properties ^{2,3}. For example, unlike cisplatin, oxaliplatin in plasma rapidly undergoes non-enzymatic transformation into reactive compounds because of displacement of the oxalate group, a process that complicates its pharmacokinetic profile. Most of the compounds appear to be pharmacologically inactive, but dichloro(DACH) platinum complexes enter the cell, where they have cytotoxic properties.

3. MECHANISMS OF ACTION

Various mechanisms of action are ascribed to oxaliplatin. Like other platinum-based compounds, oxaliplatin exerts its cytotoxic effect mostly through DNA damage. Apoptosis of cancer cells can be caused by formation of DNA lesions, arrest of DNA synthesis,

inhibition of RNA synthesis, and triggering of immunologic reactions. Oxaliplatin also exhibits synergism with other cytotoxic drugs, but the underlying mechanisms of those effects are less well understood.

3.1 DNA Lesions

At intracellular physiological concentrations of HCO₃⁻ and H₂PO₄⁻, and after aquation, dichloro(DACH)platinum compounds, once formed in plasma, enter the cell nucleus, where, with a peculiar tropism for GC-rich sites, they bind a nitrogen atom (N7) of guanine, forming DNA monoadducts, then diadducts ⁴. Although the effect of oxaliplatin is mostly on genomic DNA, adducts are also formed in nucleosomes.

Oxaliplatin can induce 3 types of crosslinks:

- DNA intra-strand crosslinks
- DNA inter-strand crosslinks
- DNA—protein crosslinks

Intra-strand crosslinks seem to be the predominant mechanism of action in the induction of DNA lesions, with binding of two Gs⁴, or less frequently, a G–A base pair.

Inter-strand crosslinks are believed to significantly contribute to the cytotoxicity of cisplatin ⁵, but seem less important in the mechanism of action of oxaliplatin. A study by Woynarowski *et al.* confirms both their presence at a low rate and their lethal properties ⁶.

As to DNA-protein crosslinks, despite their denaturating effect on enzymes and other important intracellular proteins, most studies have not proven that they cause cell death ⁵.

Monoadducts are devoid of significant cytotoxic action. Lethal DNA biadducts inhibit both DNA replication and transcription, causing apoptosis after cell cycle arrest ⁷ unless nucleotide excision repair has occurred. Formation of these DNA adducts is greater and more rapid with cisplatin than with oxaliplatin. Yet, oxaliplatin is, overall, more cytotoxic than cisplatin. The therapeutic effects of oxaliplatin therefore clearly do not depend only on the alkylating—intercalating effects of the platinum moiety ⁷.

The apoptotic pathway of colon cancer cells after exposure to oxaliplatin involves caspase 3 activation, translocation of Bax in the mitochondria, and release of cytochrome C in the cytosol ⁸.

Some aspects of the DNA lesions are relatively specific to oxaliplatin. For example, the conformation of oxaliplatin adducts, as compared with those of cisplatin or carboplatin adducts, makes binding with the mismatch repair (MMR) protein complex more difficult, presumably resulting in greater irreversibility of the lesions. In addition, the bulky DACH compound is postulated to more effectively prevent

DNA synthesis than does the *cis*-diamine carrier ligand of cisplatin ².

3.2 Arrest of DNA Synthesis

Experiments looking at the mechanism of synergism between oxaliplatin and 5-fluorouracil (5_{FU}) have uncovered a direct inhibitory effect of oxaliplatin on thymidylate synthase, preventing the incorporation of thymidine in nucleic acid synthesis ⁹. This antimetabolite-like effect results in arrest of the mitotic process. Because oxaliplatin is usually combined with 5_{FU}, itself a thymidylate synthase inhibitor, it is unclear whether this mechanism of action of oxaliplatin plays an important role *in vivo* of its own.

3.3 Inhibition of Messenger RNA Synthesis

Inhibition of DNA replication is not always sufficient to cause cell death. Inhibition of transcription at the initiation and elongation phases also plays a key role. Three main mechanisms of transcription inhibition are postulated for oxaliplatin ¹⁰:

- **Binding of transcription factors:** At the initiation stage, platinum—DNA adducts can serve as binding sites for transcription factors, especially when those factors have a strong chemical affinity for platinum. Thus, natural binding of the transcription factors to their promoter sites is prevented.
- *Inhibition of RNA polymerases:* This inhibition is established for cisplatin, but is presumably also true of oxaliplatin. The bases of platinum—DNA adducts are not able to enter the active site of an enzyme such as pol II.
- Role of nucleosomal DNA adducts: These adducts have the potential to block access by the RNA polymerase to the DNA template.

3.4 Immunologic Mechanisms

It has recently been discovered that oxaliplatin can cause the immunogenic death of colon cancer cells in murine and human cell lines 11. After exposure to oxaliplatin, colon cancer cells emit several immunogenic signals on their surface before undergoing apoptosis. These signals trigger the production of interferon y by T cells and also interact with the tolllike receptor 4 of dendritic cells, the whole process resulting in a sort of tumour vaccine. A particularly convincing argument of the importance of this mechanism is that humans carrying a mutant allele of the TLR4 gene resulting in loss of function were found to experience a lesser benefit from oxaliplatin chemotherapy in the metastatic setting, with a statistically significant shorter progression-free and overall survival.

3.5 Mechanism of Action in Oxaliplatin-Based Combinations

Because single-agent oxaliplatin has low activity in many tumours, it is often combined with other chemotherapeutic agents, 5_{FU} being the most common.

The exact mechanism of synergism between 5_{FU} and oxaliplatin is complex, but experimental observations suggest that oxaliplatin can downregulate or inhibit dihydropyrimidine dehydrogenase, slowing the catabolism of 5_{FU}⁹.

Oxaliplatin has been combined with other cytotoxic therapeutic agents with varying degrees of success. The mechanism of action of these combinations is less well documented and beyond the scope of this article.

Relatively few data document the potential for synergism of oxaliplatin-based combinations with molecularly targeted therapy. *In vitro* experiments show enhanced cytotoxicity to cisplatin in cells pretreated with rapamycin, after suppression of DNA repair mechanisms by the latter agent ¹². Combinations of oxaliplatin with inhibitors of the mammalian target of rapamycin should be evaluated in the clinical setting.

3.6 Resistance to Oxaliplatin

Despite initial sensitivity to oxaliplatin, most cancer cells will eventually develop resistance. Many mechanisms of resistance have been described or hypothesized because of similarity between oxaliplatin and cisplatin ^{13,14}. The intracellular fate of the drug can be affected by decreased uptake (resulting in lower intracellular concentration) or inactivation by structural or spatial changes (conjugation with glutathione or sequestration with metallothionein). However, the most important mechanisms seem to be related to DNA repair: MMR, or nucleotide excision repair (NER). Cells that overexpress ERCC1, an excision repair enzyme, are resistant to oxaliplatin ¹⁵.

The combination of oxaliplatin with other antineoplastic drugs may prevent resistance—or even reverse it. For instance, *in vitro* assays show that cetuximab reduces the expression of components of NER pathways, used by the cell to remove platinum—DNA adducts ¹⁶. A potential area of research would be the combination of oxaliplatin with inhibitors of Aurora kinases and of poly–adenosine diphosphate ribose polymerase ¹⁷.

3.7 Drug Interactions

Recent data show that oxaliplatin has little to no effect on cytochrome P450, one of the main enzymes involved in drug biotransformation ¹⁸. That finding suggests that, when necessary, oxaliplatin may be safe to use in co-administration with other commonly used drugs.

4. TOXICITY PROFILE

Oxaliplatin causes adverse reactions that narrow its therapeutic index. The target organs are mainly the hematopoietic system, the peripheral nerves, and the gastrointestinal (GI) system.

4.1 The Hematopoietic System

Oxaliplatin is moderately myelotoxic, more so than cisplatin. The severity of myelotoxicity is proportional to the dose, typically 85–135 mg/m² intravenously. Grades 3 and 4 neutropenia are common, but with only a 4% incidence of neutropenic fever ¹⁹. Anemia and thrombocytopenia are usually not severe.

Like many other cytotoxic drugs, oxaliplatin presumably affects progenitor cells in the bone marrow. It also enters peripheral blood cells: DNA adducts are present in leukocytes after oxaliplatin administration ²⁰. Whether this action contributes to hematologic toxicity is uncertain, but the number of platinum—DNA adducts in the blood cells of patients treated with cisplatin correlates with the degree of leucopenia and thrombocytopenia ²¹.

Other, less frequent, mechanisms of hematologic toxicity have been described. For instance, hypersensitivity reactions after repeated infusions of oxaliplatin can cause hemolytic anemia and secondary immune thrombocytopenia ²². In addition, rare cases of secondary acute leukemia have also been reported, as with other alkylating agents ²³.

4.2 The Peripheral Nerves

Peripheral neuropathy is extremely common in oxaliplatin-treated subjects. It exists in an acute and a chronic form, believed to result from distinct, but overlapping, pathophysiologic mechanisms.

Acute peripheral neuropathy is characterized by paresthesia, dysethesia, or allodynia affecting the extremities, the lips, and the oropharyngolaryngeal area during or shortly after oxaliplatin infusion. It is often triggered by exposure to cold. It usually subsides within a few hours or days ²⁴. Experimental data suggest that oxaliplatin affects voltage-gated sodium channels in complex pathways involving calcium. Calcium itself is chelated by oxalate, a metabolite of oxaliplatin ²⁵.

Chronic oxaliplatin-induced peripheral neuropathy results from cumulative exposure to the drug. The incidence of grades 3 and 4 neuropathy is about 15% ²⁶ in patients who have received a cumulative dose of about 800 mg/m². It essentially involves the extremities. Although described initially as a degenerative process of the axons, in which the previously mentioned sodium ion channels play a role, it is now thought as well to be a state secondary to accumulation of platinum compounds in the dorsal root ganglia cells, causing atrophy and mitochondrial

dysfunction ²⁷. It is irreversible in fewer than 5% of cases. Most of the time, peripheral neuropathy manifests itself as decreased distal sensations and proprioception. As with the acute form, involvement of motor fibres is rare.

An understanding of the mechanism of peripheral neuropathy induced by oxaliplatin is crucial for the prevention and treatment of this phenomenon $^{28,\bar{29}}$. Many drugs for that purpose (for example, xaliproden, gabapentin) have been unsuccessfully tested. However, despite a previous controversy, the results of a retrospective study 30 and of an incompletely accrued randomized controlled trial 31 indicate a benefit with infusions of calcium gluconate and magnesium sulphate before and after oxaliplatin administration. The benefit consists of a significant reduction in the incidence of chronic peripheral neuropathy symptoms secondary to oxaliplatin. Although initially described, the improvement of acute neurotoxicity by this intervention has not been confirmed. There is no evidence to suggest a decrease in the anticancer effects of oxaliplatin when calcium and magnesium infusions are administered

An additional research question is whether oxaliplatin can safely be combined with anti-neoplastics that have a different mechanism of neurotoxicity. Few studies in that regard have been performed, but uncontrolled trials suggest no increase in the incidence of severe peripheral neuropathy when oxaliplatin is associated with vinca alkaloids ³², taxanes ³³, and proteasome inhibitors ³⁴.

4.3 The GI System

The GI side effects attributed to oxaliplatin consist mainly of nausea, vomiting, and diarrhea ³⁵. Usually mild to moderate in intensity, they are considered to be nonspecific toxic effects of the drug on the rapidly dividing cells of the GI tract.

5. PHARMACOKINETICS

5.1 Generalities

Oxaliplatin is often administered concomitantly with 5_{FU}. Phase I trials have shown no alteration of oxaliplatin pharmacokinetics when 5_{FU} is administered concomitantly ³⁶. In addition, most papers do not address the pharmacokinetics of oxaliplatin *per se*, but of the platinum content. In fact, shortly after infusion, oxaliplatin forms many different platinum compounds that bind to blood or cell proteins. These molecules are thought to be of no pharmacologic interest ³⁷. Therefore, in most experiments, only the ultrafilterable platinum component is measured, which complicates interpretation of the pharmacologic data. Platinum derived from oxaliplatin is described as having a "tri-exponential" pattern of elimination, the

half-lives being successively 0.28 hour, 16.3 hours, and 273 hours.

One paper reported the pharmacokinetics of oxaliplatin itself rather than of platinum after oxaliplatin infusion 38 . The half-life $(t_{1/2})$ of oxaliplatin is 14.1 minutes. Another half-life of 45 minutes is related to *in vivo* degradation in blood rather than to elimination. Also, "a significant correlation between the clearance and the *in vivo* degradation rate constants" was found, suggesting that there could be a physiologic link between those two processes.

5.2 Impaired Kidney Function

In a study examining the pharmacokinetics of oxaliplatin in the setting of renal function impairment ³⁹, 34 patients were stratified according to creatinine clearance and received oxaliplatin at various dose levels.

The area under the curve (AUC) increased with lower creatinine clearance, supporting the understanding that the clearance of oxaliplatin occurs largely through renal mechanisms. However, no increased toxicity was observed, even with an increased AUC secondary to renal dysfunction.

5.3 Impaired Liver Function

Similarly, 60 cancer patients with liver dysfunction were stratified according to the results of liver function tests (total bilirubin, aspartate aminotransferase, and alkaline phosphatase) or their status as liver transplant recipients ⁴⁰. They received oxaliplatin 60–130 mg/m² according to a dose-escalation protocol.

Unlike renal insufficiency, liver dysfunction does not seem to affect oxaliplatin clearance and AUC, except in the group with the most severe abnormalities. No increased side effects were seen in the patients tested.

5.4 Long-Term Retention of Oxaliplatin Derivatives

Given that the 3rd half-life of oxaliplatin is in the order of hundreds of hours, accumulation of the drug in tissues may presumably be expected. In this regard, a study examined long-term retention of platinum 8–75 months after treatment with cisplatin and oxaliplatin ⁴¹.

The results showed that the plasma concentration of platinum in individuals previously exposed to oxaliplatin or cisplatin is larger by a factor of 30 than that in unexposed controls. The metal is found in both whole and ultrafilterable plasma. Risk factors for persistent high levels are decreased glomerular function and high cumulative dose. The authors demonstrated that the platinum found is still reactive, capable of forming platinum—DNA adducts *in vitro*. Although the physiologic significance of this reactivity is unknown, the findings are of concern with regard to long-term toxic effects such as secondary malignancies.

6. CLINICAL DEVELOPMENT OF OXALIPLATIN

6.1 Early Studies

The development of oxaliplatin was born of the need to find an alternative to cisplatin, an effective agent in various cancers, but substantially toxic. A recognized limitation of cisplatin was also its lack of activity against colorectal cancer, one of the most common human malignancies.

The phase I studies evaluated activity and safety for a range of doses. Unlike the usual classical studies, in which patient cohorts are given progressively higher doses of the studied drug, Mathé *et al.* used a different design: Doses were escalated in each study patient until the maximally efficient dose range, defined as between 45 mg/m² and 67 mg/m² administered intravenously, was reached ⁴². An absence of nephrotoxicity, setting oxaliplatin apart from cisplatin, was observed. Hints of activity against lung cancer, breast cancer, melanoma, and hepatoma were noted.

In a more conventional phase I trial ⁴³, dose escalation reached 200 mg/m² delivered intravenously. At that dose level, the characteristic peripheral neuropathy was recognized, leading to the recommendation, now accepted, that the maximum dose to be used in clinic be 135 mg/m² administered intravenously. Activity was also seen in various tumours, including some that had been pretreated with cisplatin.

Pharmacokinetic studies in that early period were also conducted. Although synergism between oxaliplatin and 5FU was rapidly recognized, relatively few pharmacodynamic studies were performed, possibly because of an assumption that oxaliplatin and cisplatin shared the same mechanism of action. However, interest in unmasking specific aspects of oxaliplatin arose when it was discovered that oxaliplatin and cisplatin are not cross-resistant.

6.2 Oxaliplatin in Colorectal Cancer

The modest activity (10%) of single-agent oxaliplatin in 5FU-refractory colorectal cancer was recognized early ⁴⁴. This activity rate is about 20% in untreated cases, according to a paper published in 1998 ⁴⁵. In both articles, the authors concluded that oxaliplatin combinations should be explored to improve outcomes. In fact, an uncontrolled study had already suggested synergy of 5FU and oxaliplatin, with reported response rates as high as 58% in a heterogeneous cohort of untreated and previously treated colorectal cancer patients ⁴⁶.

The high activity of 5FU-leucovorin-oxaliplatin in 5FU-refractory colorectal carcinoma was confirmed in later trials, and the combination was given the acronym FOLFOX ⁴⁷. Several versions of this regimen have been developed (from FOLFOX1 to FOLFOX7) to improve 5FU-related toxicity and patient convenience. There is

no strong evidence that any of the FOLFOX variations is superior to the others in terms of efficacy.

The real value of oxaliplatin in metastatic colorectal cancer has been demonstrated in randomized trials. For example, a phase III study conducted by de Gramont ⁴⁸ comparing 5FU-leucovorin with FOLFOX4 in previously untreated metastatic colorectal cancer, showed a significant improvement in response rate (50.7% vs. 22.3%) and progression-free survival (9 months vs. 6.2 months) in favour of FOLFOX. Similar results have been reported in trials in which capecitabine was substituted for 5FU and leucovorin, forming the XELOX or CAPOX regimen ⁴⁹. In addition, oxaliplatin in combination with fluoropyrimidines remains useful, even after progression on 5FU-irinotecan, another standard regimen for metastatic colorectal cancer ⁵⁰.

Given the effectiveness of oxaliplatin-based chemotherapy in advanced colorectal carcinoma, there was obvious interest for studies in the adjuvant setting. In patients who have undergone surgery for stage III colon cancer, 5FU or capecitabine (compared with observation) significantly improves the cure rate. Those results are furthered by the addition of oxaliplatin to the 5FU backbone. A recent publication confirms that, as compared with 5FU, FOLFOX improves overall survival in stage III patients ⁵¹.

Molecularly targeted cancer therapy is also added to oxaliplatin-based chemotherapy for the purpose of synergism. For now, the agents that are routinely used in that context are monoclonal antibodies. Clinical trials of bevacizumab, the antibody against vascular endothelial growth factor, in combination with FOLFOX or XELOX for metastatic colorectal cancer have shown positive results in terms of response rate, progressionfree survival, or overall survival, depending on the particular clinical setting 52,53. Improved response rates and progression-free survival are seen when an antibody (cetuximab or panitumumab) against the epidermal growth factor receptor (EGFR) is added to FOLFOX or XELOX, although the benefit of those antibodies is limited to patients harbouring an unmutated KRAS gene in their colorectal tumour 54,55.

Disappointingly, adding both bevacizumab and an anti-EGFR antibody to FOLFOX or XELOX is ineffective and even deleterious ⁵⁶. In addition, neither bevacizumab nor cetuximab improves overall survival or disease-free survival when given in combination with oxaliplatin-based chemotherapy in the adjuvant setting ^{57,58}. Overall, these results show how little is understood about the interaction of oxaliplatin with antibodies, and how useful pharmacodynamic studies would be in that regard.

Even fewer data are available regarding potential combinations of oxaliplatin with anti-angiogenic small molecules and multikinase inhibitors. The occasional antagonistic effects encountered in some *in vitro* experiments ⁵⁹ highlight the need for careful research.

6.3 Other Cancers

Given its success in colorectal cancer, oxaliplatin has been tested in other digestive cancers. Because of its less toxic profile, many research protocols substitute it for cisplatin. The present review considers only gastroesophageal and pancreatic cancers.

In a phase III trial, Cunningham *et al.* ⁶⁰ randomized patients with advanced gastric or esophageal cancer to either epirubicin—cisplatin—5_{FU} or epirubicin—oxaliplatin—capecitabine. Response rate and progression-free survival were similar in both groups, suggesting equivalency between cisplatin and oxaliplatin, and between 5_{FU} and capecitabine respectively.

Oxaliplatin in combination with 5FU or gemcitabine shows promising results as a salvage regimen for metastatic pancreatic cancer after failure of gemcitabine ⁶¹, but no results of phase III trials have yet been published in definitive form.

7. FUTURE PERSPECTIVES

Although it belongs to the same class of drugs as cisplatin and carboplatin, oxaliplatin shows marked differences in its pharmacokinetic and pharmacodynamic profiles, as well as in its spectrum of antitumour activity and its toxicity.

The pharmacology of oxaliplatin has uncovered one of the most appealing characteristics of the drug: its lack of cross-resistance with cisplatin and carboplatin, which complements its manageable toxicity profile. Pharmacodynamic characterization of oxaliplatin has also helped in achieving an understanding of the pathogenesis of the accompanying peripheral neuropathy, which should result in more effective management of that complication.

The discovery of multiple mechanisms of action of oxaliplatin, coupled with increased knowledge of cellular mechanisms of resistance, should facilitate design of clinical trials with novel combinations. An obvious goal would be to improve the results of current chemotherapy regimens without excessive toxicity. In particular, future studies should focus on combinations of oxaliplatin with molecularly targeted agents, because those combinations present potential therapeutic complementarity, with little overlap in pharmacologic characteristics.

8. CONFLICT OF INTEREST DISCLOSURES

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9. REFERENCES

- Ibrahim A, Hirschfeld S, Cohen MH, Griebel DJ, Williams GA, Pazdur R. FDA drug approval summaries: oxaliplatin. *Oncologist* 2004;9:8–12.
- Raymond E, Chaney SG, Taamma A, Cvitkovic E. Oxaliplatin: a review of preclinical and clinical studies. *Ann Oncol* 1998:9:1053-71.
- United States, National Institutes of Health, National Cancer Institute (NCI). Home > NCI Drug Dictionary > O > Oxaliplatin [Web resource]. Bethesda, MD: NCI; n.d. [Available at: www.cancer.gov/drugdictionary/?CdrID=42374; cited April 1, 2010]
- Faivre S, Chan D, Salinas R, Woynarowska B, Woynarowski JM. DNA strand breaks and apoptosis induced by oxaliplatin in cancer cells. *Biochem Pharmacol* 2003;66:225–37.
- Zwelling LA, Anderson T, Kohn KW. DNA-protein and DNA interstrand cross-linking by *cis*- and *trans*-platinum(II) diamminedichloride in L1210 mouse leukemia cells and relation to cytotoxicity. *Cancer Res* 1979;39:365–9.
- Woynarowski JM, Faivre S, Herzig MC, et al. Oxaliplatininduced damage of cellular DNA. Mol Pharmacol 2000;58:920– 7
- Di Francesco A, Ruggiero A, Riccardi R. Cellular and molecular aspects for drugs of the future: oxaliplatin. *Cell Mol Life Sci* 2002;59:1914–27.
- Arango D, Wilson AJ, Shi Q, et al. Molecular mechanisms of action and prediction of response to oxaliplatin in colorectal cancer cells. Br J Cancer 2004;91:1931–46.
- Fischel JL, Formento P, Ciccolini J, et al. Impact of the oxaliplatin–5 fluorouracil–folinic acid combination on respective intracellular determinants of drug activity. Br J Cancer 2002;86:1162–8.
- Todd RC, Lippard SJ. Inhibition of transcription by platinum antitumor compounds. *Metallomics* 2009;1:280–91.
- Tesniere A, Schlemmer F, Boige V, et al. Immunogenic death of colon cancer cells treated with oxaliplatin. Oncogene 2010;29:482–91.
- Beuvink I, Boulay A, Fumagalli S, et al. The mtor inhibitor RAD001 sensitizes tumor cells to DNA-damaged induced apoptosis through inhibition of p21 translation. Cell 2005;120:747–59
- 13. Reardon JT, Vaisman A, Chaney SG, Sancar A. Efficient nucleotide excision repair of cisplatin, oxaliplatin, and bis-aceto-ammine-dichloro-cyclohexylamine-platinum(IV) (JM216) platinum intrastrand DNA diadducts. *Cancer Res* 1999;59:3968–71.
- 14. El-Akawi Z, Abu-Hadid M, Perez R, *et al.* Altered glutathione metabolism in oxaliplatin resistant ovarian carcinoma cells. *Cancer Lett* 1996;105:5–14.
- Arnould S, Hennebelle I, Canal P, Bugat R, Guichard S. Cellular determinants of oxaliplatin sensitivity in colon cancer cell lines. *Eur J Cancer* 2003;39:112–19.
- 16. Prewett M, Deevi DS, Bassi R, *et al.* Tumors established with cell lines selected for oxaliplatin resistance respond to oxaliplatin if combined with cetuximab. *Clin Cancer Res* 2007;13:7432–40.
- Martin LP, Hamilton TC, Schilder RJ. Platinum resistance: the role of DNA repair pathways. Clin Cancer Res 2008;14:1291–5.

- Masek V, Anzenbacherova E, Machova M, Brabec V, Anzenbacher P. Interaction of antitumour platinum complexes with human liver microsomal cytochromes P450. *Anticancer Drugs* 2009;20:305–11.
- 19. Tournigand C, André T, Achille E, *et al.* FOLFIRI followed by FOLFOX6 or the reverse sequence in advanced colorectal cancer: a randomized GERCOR study. *J Clin Oncol* 2004;22:229–37.
- Pieck AC, Drescher A, Wiesmann KG, et al. Oxaliplatin–DNA adduct formation in white blood cells of cancer patients. Br J Cancer 2008;98:1959–65.
- Veal GJ, Dias C, Price L, et al. Influence of cellular factors and pharmacokinetics on the formation of platinum—DNA adducts in leukocytes of children receiving cisplatin therapy. Clin Cancer Res 2001;7:2205–12.
- 22. Koutras AK, Makatsoris T, Paliogianni F, *et al.* Oxaliplatin-induced acute-onset thrombocytopenia, hemorrhage and hemolysis. *Oncology* 2004;67:179–82.
- Carneiro BA, Kaminer L, Eldibany M, Sreekantaiah C, Kaul K, Locker GY. Oxaliplatin-related acute myelogenous leukemia. Oncologist 2006;11:261–2.
- Argyriou AA, Polychronopoulos P, Iconomou G, Chroni E, Kalofonos HP. A review on oxaliplatin-induced peripheral nerve damage. *Cancer Treat Rev* 2008;34:368–77.
- Grolleau F, Gamelin L, Boisdron–Celle M, Lapied B, Pelhate M, Gamelin E. A possible explanation for a neurotoxic effect of the anticancer agent oxaliplatin on neuronal voltage-gated sodium channels. *J Neurophysiol* 2001;85:2293–7.
- 26. Maindrault–Goebel F, Louvet C, André T, *et al.* Oxaliplatin added to the simplified bimonthly leucovorin and 5-fluorouracil regimen as second-line therapy for metastatic colorectal cancer (FOLFOX6). *Eur J Cancer* 1999;35:1338–42.
- 27. Cavaletti G, Tredici G, Petruccioli MG, *et al*. Effects of different schedules of oxaliplatin treatment on the peripheral nervous system of the rat. *Eur J Cancer* 2001;37:2457–63.
- Krishnan AV, Goldstein D, Friedlander M, Kiernan MC. Oxaliplatin and axonal Na+ channel function in vivo. Clin Cancer Res 2006;12:4481–4.
- 29. Park S, Krishnan A, Lin CY, Goldstein D, Friedlander M, Kiernan M. Mechanisms underlying chemotherapy-induced neurotoxicity and the potential for neuroprotective strategies. *Curr Med Chem* 2008;15:3081–94.
- Gamelin L, Boisdron–Celle M, Delva R, et al. Prevention of oxaliplatin-related neurotoxicity by calcium and magnesium infusions. Clin Cancer Res 2004;10:4055–61.
- 31. Grothey A, Nikcevich DA, Sloan JA, *et al.* Evaluation of the effect of intravenous calcium and magnesium (CaMg) on chronic and acute neurotoxicity associated with oxaliplatin: results from a placebo-controlled phase III trial [abstract 4009]. *J Clin Oncol* 2008;26:. [Available online at www.asco.org/ ASCOv2/Meetings/Abstracts?&vmview=abst_detail_view&confID=55&abstractID=32445; cited December 17, 2010]
- 32. Mir O, Alexandre J, Ropert S, *et al.* Vinorelbine and oxaliplatin in stage IV nonsmall cell lung cancer patients unfit for cisplatin: a single-center experience. *Anticancer Drugs* 2009;20:105–8.
- Kuo DYS, Blank SV, Christos PJ, et al. Paclitaxel plus oxaliplatin for recurrent or metastatic cervical cancer: a New York Cancer Consortium Study. Gynecol Oncol 2010;116:442–6.
- 34. Cohen SJ, Engstrom PF, Lewis NL, *et al.* Phase 1 study of capecitabine and oxaliplatin in combination with the proteasome

- inhibitor bortezomib in patients with advanced solid tumors. *Am J Clin Oncol* 2008;31:1–5.
- Zafar S, Marcello J, Wheeler J, et al. Treatment-related toxicity and supportive care in metastatic colorectal cancer. J Support Oncol 2010;8:15–20.
- Graham MA, Lockwood GF, Greenslade D, Brienza S, Bayssas M, Gamelin E. Clinical pharmacokinetics of oxaliplatin: a critical review. *Clin Cancer Res* 2000;6:1205–18.
- 37. Kweekel DM, Gelderblom H, Guchelaar J. Pharmacology of oxaliplatin and the use of pharmacogenomics to individualize therapy. *Cancer Treat Rev* 2005;31:90–105.
- Ehrsson H, Wallin I, Yachnin J. Pharmacokinetics of oxaliplatin in humans. *Med Oncol* 2002;19:261–5.
- Takimoto CH, Graham MA, Lockwood G, et al. Oxaliplatin pharmacokinetics and pharmacodynamics in adult cancer patients with impaired renal function. Clin Cancer Res 2007;13:4832–9.
- Synold TW, Takimoto CH, Doroshow JH, et al. Dose-escalating and pharmacologic study of oxaliplatin in adult cancer patients with impaired hepatic function: a National Cancer Institute Organ Dysfunction Working Group study. Clin Cancer Res 2007;13:3660–6.
- Brouwers E, Huitema A, Beijnen J, Schellens J. Long-term platinum retention after treatment with cisplatin and oxaliplatin. BMC Clin Pharmacol 2008;8:7.
- 42. Mathé G, Kidani Y, Triana K, et al. A phase I trial of trans-1-diaminocyclohexane oxalato-platinum (L-OHP). Biomed Pharmacother 1986;40:372–6.
- 43. Extra JM, Espie M, Calvo F, Ferme C, Mignot L, Marty M. Phase I study of oxaliplatin in patients with advanced cancer. *Cancer Chemother Pharmacol* 1990;25:299–303.
- 44. Machover D, Diaz–Rubio E, de Gramont A, *et al.* Two consecutive phase II studies of oxaliplatin (L-OHP) for treatment of patients with advanced colorectal carcinoma who were resistant to previous treatment with fluoropyrimidines. *Ann Oncol* 1996;7:95–8.
- 45. Diaz–Rubio E, Sastre J, Zaniboni A, *et al.* Oxaliplatin as single agent in previously untreated colorectal carcinoma patients: a phase π multicentric study. *Ann Oncol* 1998;9:105–8.
- 46. Lévi F, Misset JL, Brienza S, Adam R, et al. A chronopharmacologic phase π clinical trial with 5-fluorouracil, folinic acid, and oxaliplatin using an ambulatory multichannel programmable pump. High antitumour effectiveness against metastatic colorectal cancer. Cancer 1992;69:893–900.
- 47. de Gramont A, Tournigand C, Louvet C, *et al.* Oxaliplatin, folinic acid and 5-fluorouracil (FOLFOX) in pretreated patients with metastatic advanced cancer. The GERCOD [French]. *Rev Med Interne* 1997;18:769–75.
- de Gramont A, Figer A, Seymour M, et al. Leucovorin and fluorouracil with or without oxaliplatin as first-line treatment in advanced colorectal cancer. J Clin Oncol 2000:18:2938–47.
- 49. Cassidy J, Tabernero J, Twelves C, et al. XELOX (capecitabine plus oxaliplatin): active first-line therapy for patients with metastatic colorectal cancer. J Clin Oncol 2004;22:2084–91.
- Culy CR, Clemett D, Wiseman LR. Oxaliplatin: a review of its pharmacological properties and clinical efficacy in metastatic colorectal cancer and its potential in other malignancies. *Drugs* 2000;60:895–924.

- 51. André T, Boni C, Navarro M, *et al.* Improved overall survival with oxaliplatin, fluorouracil, and leucovorin as adjuvant treatment in stage II or III colon cancer in the MOSAIC trial. *J Clin Oncol* 2009;27:3109–16.
- Giantonio BJ, Catalano PJ, Meropol NJ, et al. Bevacizumab in combination with oxaliplatin, fluorouracil, and leucovorin (FOLFOX4) for previously treated metastatic colorectal cancer: results from the Eastern Cooperative Oncology Group Study E3200. J Clin Oncol 2007;25:1539–44.
- 53. Saltz LB, Clarke S, Diaz–Rubio E, *et al.* Bevacizumab in combination with oxaliplatin-based chemotherapy as first-line therapy in metastatic colorectal cancer: a randomized phase III study. *J Clin Oncol* 2008;26:2013–19.
- 54. Amado RG, Wolf M, Peeters M, *et al.* Wild-type *KRAS* is required for panitumumab efficacy in patients with metastatic colorectal cancer. *J Clin Oncol* 2008;26:1626–34.
- 55. Douillard J, Siena S, Cassidy J, *et al.* Randomized phase 3 study of panitumumab with FOLFOX4 compared to FOLFOX4 alone as 1st-line treatment (tx) for metastatic colorectal cancer (mcRc): the PRIME trial [abstract 10LBA]. *Eur J Cancer* 2009;7(suppl):6. [Available online at: download.journals.elsevierhealth.com/pdfs/journals/1359-6349/PIIS1359634909720397.pdf; cited December 24, 2010]
- Tol J, Koopman M, Cats A, et al. Chemotherapy, bevacizumab, and cetuximab in metastatic colorectal cancer. N Engl J Med 2009;360:563–72.
- 57. Alberts SR, Sargent DJ, Smyrk TC, et al. Adjuvant mfolfox6 with or without cetuxiumab (cmab) in KRAS wild-type (wt) patients (pts) with resected stage III colon cancer (cc): results from NCCTG Intergroup phase III trial N0147 [abstract CRA3507].

 J Clin Oncol 2010;28: [Available online at www.asco.org/ascov2/Meetings/Abstracts?&vmview=abst_detail_view&confID=74&abstractID=41265; cited December 17, 2010]

- 58. Wolmark N, Yothers G, O'Connell M, et al. A phase III trial comparing mfolfox6 to mfolfox6 plus bevacizumab in stage II or III carcinoma of the colon: results of NSABP protocol C-08 [abstract LBA4]. *J Clin Oncol* 2009;27:. [Available online at www.asco. org/ascov2/Meetings/Abstracts?&vmview=abst_detail_view&confID=65&abstractID=31978; cited December 17, 2010]
- 59. Heim M, Scharifi M, Zisowsky J, *et al.* The Raf kinase inhibitor BAY 43-9006 reduces cellular uptake of platinum compounds and cytotoxicity in human colorectal carcinoma cell lines. *Anticancer Drugs* 2005;16:129–36.
- Cunningham D, Starling N, Rao S, et al. Capecitabine and oxaliplatin for advanced esophagogastric cancer. N Engl J Med 2008;358:36-46.
- 61. Oettle H, Pelzer U, Stieler J, et al. Oxaliplatin/folinic acid/5-fluorouracil [24h] (OFF) plus best supportive care versus best supportive care alone (BSC) in second-line therapy of gemcitabine-refractory advanced pancreatic cancer (CONKO 003) [abstract 4031]. *J Clin Oncol* 2005;23:. [Available online at www.asco.org/ASCOv2/Meetings/Abstracts?&vmview=abst_detail_view&confID=34&abstractID=32322; cited December 17, 2010]

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