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NOVEL AVENUES OF DRUG DISCOVERY AND BIOMARKERS FOR DIABETES MELLITUS

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Abstract

Globally, developed nations spend a significant amount of their resources on healthcare initiatives that poorly translate into increased population life expectancy. As an example, the United States devotes sixteen percent of its gross domestic product to healthcare, the highest level in the world, but falls behind other nations that enjoy greater individual life expectancy. These observations point to the need for pioneering avenues of drug discovery to increase lifespan with controlled costs. In particular, innovative drug development for metabolic disorders such as diabetes mellitus (DM) becomes increasingly critical given that the number of diabetic individuals will increase exponentially over the next twenty years. Here we discuss the elucidation and targeting of novel cellular pathways that are intimately tied to oxidative stress in DM for new treatment strategies. Pathways that involve wingless, NAD⁺ precursors, and cytokines govern complex biological pathways that determine both cell survival and longevity during DM and its complications. Furthermore, the role of these entities as biomarkers for disease can further enhance their utility irrespective of their treatment potential. Greater understanding of the intricacies of these unique cellular mechanisms will shape future drug discovery for DM to provide focused clinical care with limited or absent long-term complications.

Keywords

biomarkers; diabetes; oxidative stress

Healthcare and metabolic disease

Compared with other nations throughout the world, the United States devotes 16% of the gross domestic product to healthcare and spending for each individual equal to \$7,290, the

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highest levels in the world¹. Total spending on pharmaceuticals is also the highest in the world with \$878 per individual. Yet, life expectancy in years in the United States equals 78.1 years and trails behind other countries such as Japan that allots 8% of the gross domestic product on healthcare, spends \$2,514 for each individual, and has a life expectancy of 82.6 years. Furthermore, the United States is ranked as having the highest level of obesity in the population at 34.3% while countries such as Japan have a 3.4% level of obesity¹. These statistics are the results of multiple factors, but also can support the arguments for not only improved preventive health measures, but also new directions to treat multiple disorders that can lead to improved lifespan while minimizing economic burden.

In particular, one can consider diabetes mellitus (DM), a metabolic disorder closely associated with increased weight gain^{2, 3}. DM reaches approximately 20 million individuals in the United States and more than 165 million individuals worldwide⁴. By 2030, DM may affect more than 360 million individuals. Additionally, a significant portion of the population has undiagnosed diabetes, illustrating the need for improved early diagnosis⁵. The incidence of impaired glucose tolerance in the young also raises further concerns⁶. Individuals with impaired glucose tolerance have a greater than twice the risk for the development of diabetic complications than individuals with normal glucose tolerance⁷.

Type 1 insulin-dependent DM is present in 5-10 percent of all diabetics, but is increasing in adolescent minority groups^{2, 8}. Furthermore, Type 1 DM leads to long-term complications throughout the body involving cardiovascular, renal, and nervous system disease⁹. Type 1 DM is associated with the presence of alleles of the Human leukocyte antigen (HLA) class II genes within the major histocompatibility complex (MHC). The disorder is considered to have autoimmune origins resulting from inflammatory infiltration of the islets of Langerhans and the selective destruction of β -cells in the pancreas that leads to insulin loss¹⁰. In Type 1 DM, activation of T-cell clones that are capable of recognizing and destroying β -cells lead eventually to severe insulin deficiency. These T-cell clones are able to escape from thymus control that yield high affinity for major histocompatibility complex (MHC) molecules with T-cell receptors but incorrect low affinity for self-peptides. Once released into the bloodstream, these T-cell clones can become activated to destroy self-antigens. In many cases, the insulin gene (INS) and the human MHC or HLA complex are believed to contain the loci with *IDDM1* and *IDDM2* to account for the susceptibility to Type 1 DM with defective antigen presentation^{11, 12}. Interestingly, a HLA class II molecule has been linked to Type 1 DM inheritance. HLA-DQ that lacks a charged aspartic acid (Asp-57) in the β -chain is believed to lead to the ineffective presentation of autoantigen peptides during thymus selection of T-cells¹³. Animal models that involve the nonobese diabetic (NOD) mice further support these findings, since these mice spontaneously develop diabetes with the human predisposing HLA-DQ corresponding molecule of H2 I-Ag. Yet, NOD mice without H2 I-Ag do not develop diabetes¹⁴.

Upon initial diagnosis, approximately ninety percent of individuals with Type 1 DM have elevated titers of autoantibodies (Type 1A DM). The remaining ten percent of Type 1 DM individuals do not have serum autoantibodies and are described as having maturity-onset diabetes of the young (MODY) that can be a result of β -cell dysfunction with autosomal-dominant inheritance (Type 1B DM)¹⁵. Other variables reported in patients with Type 1 DM include the presence of insulin resistance that is usually characteristic of Type 2 DM and can lead to neurological and vascular disease^{16, 17}. Interestingly, there is a converse overlap with Type 1 and Type 2 DM, since almost ten percent of Type 2 DM patients may have elevated serum autoantibodies¹⁸.

Monogenic inheritance does not appear to lead to Type 1 DM. Prior work demonstrates that multiple loci with possible epistatic interactions among other loci may be responsible for

genetic transmission^{19, 20}. In addition, several environmental factors may have a role with Type 1 DM such that investigations have suggested that Type 1 DM in monozygotic twins can occur with a cumulative risk of seventy percent from birth to 35 years of age^{21, 22}. Other studies indicate a concordance between monozygotic twins to be approximately fifty percent²³, suggesting that environmental factors also may lead to a predisposition for Type 1 DM. Loss of autoimmunity in Type 1 DM can be precipitated also by the exposure to infectious agents²⁴.

Type 2 noninsulin-dependent DM represents at least 80 percent of all diabetics, usually in individuals over 40 years of age, and is dramatically increasing in incidence as a result of changes in human behavior and increased body mass index^{2, 8}. Type 2 DM is characterized by a progressive deterioration of glucose tolerance with early β -cell compensation for insulin resistance (achieved by β -cell hyperplasia). This is subsequently followed by progressive decrease in β -cells mass. In contrast, gestational diabetes mellitus that represents glucose intolerance during some cases of pregnancy usually subsides after delivery.

Insulin resistance or defective insulin action occurs when physiological levels of insulin produce a subnormal physiologic response. Skeletal muscle and liver are two of the primary insulin-responsive organs responsible for maintaining normal glucose homeostasis. Insulin lowers the level of blood glucose through suppression of hepatic glucose production and stimulation of peripheral glucose uptake, but metabolic disorders can result in insulin resistance and elevated serum glucose levels. Although insulin resistance forms the basis for the development of Type 2 DM, elevated serum glucose levels also are a result of the concurrent impairment in insulin secretion. This abnormal insulin secretion may be a result of defective β -cell function, chronic exposure to free fatty acids and hyperglycemia, and the loss of inhibitory feedback through plasma glucagon levels²⁵.

Patients with DM can develop multiple complications that include immune dysfunction²⁶, sarcopenia²⁷, depression²⁸, hepatic dysfunction²⁹, renal disease³⁰, anemia and hematological disease³¹⁻³³, neurodegenerative disorders^{8, 34, 35}, and cardiovascular disease^{8, 36}. Interestingly, patients with DM are at risk for the development of cognitive disorders^{26, 37}. In a prospective population based study of 6,370 elderly individuals, patients with DM had almost twice the risk for the development of dementia³⁸. DM also has been found to increase the risk for vascular dementia in elderly subjects^{39, 40}. Although some studies have found that diabetic patients may have significantly less neuritic plaques and neurofibrillary tangles than non-diabetic patients⁴¹, other investigators report a modest adjusted relative risk of Alzheimer's disease in patients with diabetes as compared with those without diabetes to be 1.3^{42, 43}. Additional studies have described the reduced expression of genes encoding insulin in Alzheimer's patients that suggests a potential link between DM and the development of Alzheimer's disease⁴³. Alzheimer's disease can be the result of a number of etiologies^{44, 45}, such as changes in cerebral blood flow and metabolism with aging⁴⁶, sialylation and glycosylation of amyloid plaques^{47, 48}, aberrant cell cycle induction⁴⁹⁻⁵¹, amyloid toxicity⁵¹⁻⁵⁵, chemokine induction⁵⁶, exogenous toxins⁵⁷, alteration in muscarinic and nicotinic pathways^{46, 58}, and intracellular calcium changes⁵⁹. Yet, other studies point to metabolic dysfunction⁶⁰⁻⁶². For example, in animal models with brain/neuronal insulin receptor knockouts, loss of insulin signaling appears to be linked to increased phosphorylation of the microtubule-associated protein tau that occurs during Alzheimer's disease⁶³.

Oxidative stress, apoptotic injury, mitochondria, and diabetes

Many of the cellular pathways that lead to diabetic complications and insulin resistance have been linked to the generation of free radicals and oxidative stress^{2, 3, 34, 35, 62, 64, 65}. In

animal studies with Type 1 diabetic animals, oxidative stress leads to DNA damage in renal cortical cells⁶⁶. Although early effects of elevated glucose may increase the presence of potentially protective pathways⁶⁷, more prolonged exposure of elevated glucose can lead to reactive oxygen species (ROS)^{32, 68} and can be detrimental even if glucose levels are controlled⁶⁹. In addition, elevated levels of ceruloplasmin during hyperglycemia are suggestive of increased ROS⁷⁰. A number of treatment entities seek to ameliorate the effects of oxidative stress during DM^{55, 71-75}.

Oxidative stress also may promote the onset of DM by decreasing insulin sensitivity and destroying the insulin-producing cells within the pancreas. For example, ROS can penetrate through cell membranes and cause damage to β -cells of pancreas^{76, 77}. A high fat diet⁷⁸ or free fatty acids also have been shown to release ROS and contribute to mitochondrial DNA damage and impaired pancreatic β -cell function⁷⁹. Interestingly in non-diabetic rats, hyperglycemia has been shown to increase muscle protein carbonyl content and elevated levels of malondialdehyde and 4-hydroxynonenal, indicators of oxidative stress and lipid peroxidation⁸⁰. These biomarkers of oxidative stress and insulin resistance suggest that ROS contribute to the pathogenesis of hyperglycemia-induced insulin resistance^{81, 82} as well as insulin induced ROS⁷³. Hyperglycemia can lead to increased production of ROS in several cell types^{82, 83}. For example, with increased age in a rat model of nonobese Type 2 DM, increased levels of 8-OHdG and HNE-modified proteins in pancreatic beta-cells have been reported⁸⁴. Elevated glucose also has been shown to increase antioxidant enzyme levels in human endothelial cells, suggesting that elevated glucose levels may lead to a reparative process to protect cells from oxidative stress injury⁸⁵. Chronic hyperglycemia is not necessary to lead to oxidative stress injury, since even short periods of hyperglycemia, generate ROS, such as in vascular cells⁸⁶. Recent clinical correlates support these experimental studies to show that acute glucose swings in addition to chronic hyperglycemia can trigger oxidative stress mechanisms during Type 2 DM, demonstrating the importance for therapeutic interventions during acute and sustained hyperglycemic episodes⁸⁷.

At the cellular level, ROS are oxygen free radicals and other chemical entities that can lead to cell injury if left unchecked^{49, 88, 89}. Oxidative stress can result in hepatic injury^{90, 91}, pancreatitis⁹², impaired cognition^{93, 94}, neuronal injury^{44, 49, 95-99}, Parkinson's disease¹⁰⁰⁻¹⁰³, complications of epilepsy¹⁰⁴, cardiovascular disease¹⁰⁵⁻¹⁰⁷, ocular disease¹⁰⁸, age-related disorders¹⁰⁹, metal ion injury¹¹⁰, uncontrolled pain sensation¹¹¹, and promote xenobiotic toxicity^{112, 113}. ROS consist of superoxide free radicals, hydrogen peroxide, singlet oxygen, nitric oxide (NO), and peroxyxynitrite^{34, 44, 83}. ROS usually occur at low levels during normal physiological conditions and are scavenged by endogenous antioxidant systems that include superoxide dismutase (SOD), glutathione peroxidase, catalase, and vitamin D₃^{114, 115}. Additional pathways include vitamins C, E, and K^{104, 116-119}.

Oxidative stress results in cell injury through apoptotic and non-apoptotic pathways. In regards to programmed cell death (apoptosis), apoptosis can occur during DM^{2, 3, 120, 121}, anesthetic exposure¹²², tissue ischemia¹²³⁻¹²⁶, bone fatigue¹²⁷, neurodegenerative disorders^{34, 128-130} and Alzheimer's disease^{51-54, 59, 131-136}, plasticity associated with ischemic preconditioning¹³⁷, aging-related diseases^{35, 138, 139}, and toxic conditions during development^{122, 140}.

During apoptosis, the cleavage of genomic DNA into fragments occurs^{129, 141, 142} as a later event during apoptotic injury¹⁴²⁻¹⁴⁵ after the exposure of membrane phosphatidylserine (PS) residues^{146, 147}. Membrane PS exposure occurs in neurons, vascular cells, and inflammatory microglia during reduced oxygen exposure^{50, 89, 141, 148, 149}, β -amyloid (A β) exposure^{53, 55}, nitric oxide exposure¹⁵⁰⁻¹⁵⁴, and during the administration of agents that induce the production of reactive oxygen species (ROS), such as 6-hydroxydopamine¹⁰³

(Figure 1). Membrane PS externalization also occurs on platelets and has been associated with clot formation in the vascular system¹⁵⁵.

Membrane PS exposure also is involved in the activation of inflammatory cells such as microglia of the central nervous system that can dispose of injured cells^{156, 157}. For their own survival, microglia and non-neuronal cells of the brain are dependent upon several intracellular pathways, such as mTOR^{158, 159} and zinc regulation¹⁶⁰. Non-neuronal cells of the brain can be beneficial to modulate neurogenesis¹⁶¹, to function as immune surveillance for toxic products¹⁶², such as for β -amyloid⁴⁸, to block foreign organisms and viral agents from proliferating in the brain¹⁶³, to modulate vascular growth¹⁶⁴, and to allow for the repair of tissues composed of neuronal and vascular cells^{158, 165}. Yet, microglia have another side that may be detrimental to an organism. They can generate ROS^{166, 167}, may worsen events with oxidative stress injury¹⁶⁸, and activate cytokines that in some circumstances may initially lead to cell proliferation¹⁶⁹, but later can result in the demise of cells^{56, 163, 164}. For these reasons, it is important to understand the mechanisms that can activate microglia. Membrane PS exposure can become a signal for microglia to dispose of injured cells^{61, 145, 170-172}. This process can be controlled by caspase 1 and caspase 3^{89, 173, 174}. Increased expression of the phosphatidylserine receptor (PSR) on microglia also occurs to facilitate activation of microglia^{116, 175}.

Oxidative stress and apoptotic cell death during disorders such as DM are also strongly associated to cellular energy maintenance and intact mitochondrial function^{8, 26, 64, 176-178}. ROS exposure can result in the opening of the mitochondrial membrane permeability transition pore^{145, 179-181}, reduce mitochondrial NAD⁺ stores, and result in apoptotic cell injury⁴⁴. Free fatty acids also can lead to ROS release, mitochondrial DNA damage, and impaired pancreatic β -cell function⁷⁹. In patients with Type 2 DM, skeletal muscle mitochondria have been described to be smaller than those in control subjects¹⁸². A decrease in the levels of mitochondrial proteins and mitochondrial DNA in adipocytes also has been correlated with the development of type 2 DM¹⁸³.

Innovative drug discovery for DM

Multiple pathways may lead to a loss in cell survival and longevity during DM that are broad in nature and consist of several precipitating factors. Yet, oxidant-induced injury and the cellular pathways responsible for this signaling are thought to be primary mediators of the disability that ensues during DM. As a result, novel and pioneering directions for drug discovery are required for safe and effective treatments for DM. In addition, it is the understanding of the complex nature of cellular pathways and their intimate relationship that is critical for the development of successful drug platforms. Here we present novel cellular pathways that are strongly bound to oxidant pathways in DM. These pathways involve wingless genes with Wnt, NAD⁺ precursors with nicotinamide, forkhead transcription factors of the "O" class, and the cytokine and growth factor erythropoietin (EPO), each of which determine cellular development, survival and injury mechanisms, and longevity.

Novel cellular pathways and diabetes

Wnt

Proteins derived from the *Drosophila Wingless (Wg)* and the mouse *Int-1* genes are secreted cysteine-rich glycosylated proteins that play a role in a variety of cellular functions^{53, 72, 175, 184}. Wnt proteins determine multiple cellular functions that involve embryonic cell proliferation, cell differentiation, and cell survival that involve neurons, cardiomyocytes, endothelial cells, red blood cells, tumors, adipose tissue as well as several other cell types¹⁸⁵⁻¹⁹⁸. Recent work in clinical disease indicates that abnormalities in Wnt pathways, such

as with transcription factor 7-like 2 gene, may impart increased risk for type 2 DM in some populations¹⁹⁹⁻²⁰¹ and have a strong association with the development of obesity²⁰² (Table 1). The family member Wnt5b has been shown to have an elevated expression in adipose tissue, the pancreas, and the liver in patients with DM, suggesting control of metabolic pathways by Wnt²⁰³. In addition, clinical studies in patients with coronary artery disease and the combined metabolic syndrome with hypertension, hyperlipidemia, and DM have shown impaired Wnt signaling through a missense mutation in LRP-6²⁰⁴. Experimental studies in mice with hyperglycemia through a high fat diet also show increased expression of Wnt3a and Wnt7a²⁰⁵.

It also has been suggested that intact Wnt family members may offer glucose tolerance and increased insulin sensitivity²⁰⁶ as well as protect glomerular mesangial cells from elevated glucose induced apoptosis²⁰⁷. Animals that over express Wnt10b with a high-fat diet experienced a reduction in bodyweight, hyperinsulinemia, triglyceride plasma levels, and improved glucose homeostasis²⁰⁸. Cell culture studies demonstrate that the Wnt1 protein that can be controlled by the growth factor EPO is necessary and sufficient to impart cellular protection during elevated glucose exposure^{72, 209, 210}. EPO maintains the expression of Wnt1 during elevated glucose exposure and prevents loss of Wnt1 expression that would occur in the absence of EPO during elevated glucose. In addition, blockade of Wnt1 with a Wnt1 antibody can neutralize the protective capacity of EPO, illustrating that Wnt1 is a critical component in the cytoprotection of EPO during elevated glucose exposure⁷².

Wnt may foster cellular protection during DM through the novel regulation of protein kinase B (Akt) (Table 1). Activation of Akt can promote cell survival, such as during cell proliferation²¹¹, progenitor cell development¹⁶⁹, blood-brain barrier permeability²¹², inflammation^{163, 213}. Ischemic-preconditioning²¹⁴, neurodegeneration¹⁰³, hyperglycemia^{67, 215}, hypoxia²¹⁶, amyloid toxicity^{52, 53, 131, 132, 217}, excitotoxicity⁹⁵, amyloid production²¹⁸, cardiomyopathy²¹⁹, cellular aging²²⁰, and oxidative stress^{145, 171, 221}. Akt activation also can modulate microglial cell activation^{145, 171, 180}, regulate transcription factors²²², maintain mitochondrial membrane potential ($\Delta\Psi_m$), prevent cytochrome c release^{150, 180, 223}, and block caspase activity^{180, 216, 223}. However, as a “pro-survival pathway”, it should be recognized that Akt activation may be deleterious, such as during cancer resistance to chemotherapy²²⁴.

A number of observations support the dependence of Wnt on Akt activation and support the premise that Wnt can modulate DM complications through Akt^{192, 225, 226}. For example, neuronal cell differentiation requires Wnt signaling and trophic factor induction through Akt activity²²⁷ and differentiation of cardiomyocytes proceeds only with Akt activation²²⁸. Wnt also has been shown in preadipocytes to increase Akt phosphorylation²²⁹ and the Wnt-induced secreted protein in a fibroblast cell line uses Akt to block apoptotic death²³⁰. Secreted Frizzled-related proteins (sFRPs), which can modulate Wnt signaling, also employ Akt for cardiac tissue repair²³¹, reduction in tissue injury during pressure overload cardiac hypertrophy is tied to Akt activation²³², and cardiac ischemic preconditioning appear to rely upon Akt²³³. In the neuronal system, Wnt over-expression can independently increase the phosphorylation and the activation of Akt to promote neuronal protection. Inhibition of the phosphatidylinositol 3-kinase (PI 3-K) pathway or gene silencing of Akt expression prevents Wnt from blocking apoptotic injury and microglial activation⁵³.

Nicotinamide

As the amide form of vitamin B₃ (niacin), nicotinamide or nicotinic acid which is the water soluble form vitamin B₃, is obtained through synthesis or as a dietary source and supplement²³⁴. The principal form of niacin in dietary plant sources is nicotinic acid that is rapidly absorbed through the gastrointestinal epithelium²³⁵. Nicotinamide is subsequently

generated through the conversion of nicotinic acid in the liver or through the hydrolysis of NAD^+ ⁶². After nicotinamide is obtained in the body, it functions as the precursor for the coenzyme β -nicotinamide adenine dinucleotide (NAD^+)^{61, 236} and also is essential for the synthesis of nicotinamide adenine dinucleotide phosphate (NADP^+)²³⁷. Initially, nicotinamide is changed to its mononucleotide form (NMN) with the enzyme nicotinic acid/nicotinamide adenyltransferase yielding the dinucleotides NAAD^+ and NAD^+ . NAAD^+ also yields NAD^+ through NAD^+ synthase²³⁸ or NAD^+ can be synthesized through nicotinamide riboside kinase that phosphorylates nicotinamide riboside to NMN^{239, 240}.

The cellular pathways of nicotinamide are essential for energy metabolism and may directly impact normal physiology as well as disease progression²⁴¹⁻²⁴⁴. Nicotinamide through NAD^+ has a critical physiological role in cellular metabolism and can be directly utilized by cells to synthesize NAD^+ ^{34, 61, 116}. Nicotinamide also participates in energy metabolism through the tricarboxylic acid cycle by utilizing NAD^+ in the mitochondrial respiratory electron transport chain for the production of ATP, DNA synthesis, and DNA repair²⁴⁵⁻²⁴⁷. In addition, nicotinamide can significantly increase NAD^+ levels in vulnerable regions of the ischemic brain, suggesting that nicotinamide may prevent cell injury through the maintenance of NAD^+ levels²⁴⁸. During axonal degeneration, nicotinamide also may promote protection through NAD^+ -dependent mechanisms²⁴⁹. Nicotinamide also appears to function directly at the level of mitochondrial membrane pore formation^{61, 250, 251} to prevent the release of cytochrome c²⁵². Nicotinamide can prevent mitochondrial membrane depolarization during exposure to either *tert*-butylhydroperoxide or atractyloside²⁵³. There are other mechanisms that nicotinamide may use to maintain cellular metabolic homeostasis through the maintenance of mitochondrial membrane potential^{181, 250}. Nicotinamide can phosphorylate Bad²⁵² to prevent mitochondrial membrane depolarization and subsequent cytochrome c release. Nicotinamide also may inhibit the assembly of the mitochondrial permeability transition pore complex similar to the action of cyclosporin A²⁵⁴ as well as stabilize cellular energy metabolism through ATP pathways²⁵⁵.

In addition to its role in metabolism, nicotinamide can be essential for cellular differentiation, such as for human embryonic stem cells²⁵⁶. Nicotinamide has protean endocrine effects^{257, 258}, can scavenge ROS, and offers cellular protection for both neuronal^{253, 259, 260} and vascular cells^{34, 61, 116, 236}. In neuronal cell populations, nicotinamide protects against free radical injury¹⁸¹, anoxia²⁶¹, excitotoxicity²⁶², homocysteine toxicity²⁶³, ethanol-induced neuronal injury²⁶⁴, and oxygen-glucose deprivation^{253, 265}. In cortical neurons, nicotinamide blocks cell injury during ROS generating toxins such as tertiary butylhydroperoxide²⁶⁶. Nicotinamide also can protect both rod and cone photoreceptor cells against N-methyl-N-nitrosourea toxicity^{267, 268} as well as against glycation end products in all layers of the retina²⁶⁹. In animal studies, nicotinamide improves cognitive function, cell survival, and reduces edema following cortical trauma²⁷⁰⁻²⁷⁵, limits axonal degeneration²⁴⁹, reduces cerebral ischemia²⁷⁶⁻²⁷⁸ sometimes more effectively in models that were absent of comorbidities²⁷⁹, prevents spinal cord injury^{280, 281}, and lessens disability in models of Parkinson's disease^{100, 282, 283}.

In regards to the vascular system^{253, 259, 260}, nicotinamide promotes vascular integrity^{61, 116, 236} which may be crucial for tissue growth and repair²⁸⁴. Nicotinamide can protect the function of the blood brain barrier^{270, 271}, influence arteriolar dilatation and blood flow²⁸⁵, increase skin vascular permeability²⁸⁶, inhibit atherosclerotic plaque formation through inhibition of poly(ADP-ribose) polymerase²⁸⁷, and foster platelet production through megakaryocyte maturation²⁸⁸. Nicotinamide can enhance endothelial cell viability during ROS exposure^{181, 250, 261, 289}. Nicotinamide also may reverse a previously sustained early apoptotic injury^{61, 181, 250, 252, 253, 261}, suggesting that apoptosis prior to reaching genomic DNA degradation is dynamic and reversible in nature^{61, 181, 251, 261}. Yet, some studies in

mice suggest that nicotinamide may either prevent or contribute to atherosclerotic plaques over a three to six month progression²⁹⁰. Although the mechanisms are not clear, it is conceivable that these events may occur during oxidative stress and the production of acidosis-induced cellular toxicity²⁹¹⁻²⁹³. Nicotinamide cannot prevent cellular injury during intracellular acidification paradigms¹⁸¹.

Since nicotinamide is closely aligned with cellular energy management, it may play a significant role during DM and the complications of this disorder (Table 1). For example, nicotinamide appears to have a close relationship with metabolic pathways that may lead to clinical cognitive changes²⁹⁴. Nicotinamide also has been shown to maintain normal fasting blood glucose with streptozotocin-induced DM in animal models^{295, 296}. Nicotinamide can limit peripheral nerve injury during elevated glucose²⁹⁷, reverse Type 1 DM in mice with acetyl-lcarnitine²⁹⁸, and block oxidant stress^{242, 250, 252, 264, 299}. Nicotinamide also affects levels of O-N-acetylglucosamin(O-GlcNAc)ylated proteins³⁰⁰ and can significantly improve glucose utilization, prevent excessive lactate production in ischemic animal models³⁰¹. In clinical conditions, oral nicotinamide administration, nicotinamide (1200mg/m²/day) protects β -cell function and prevents clinical disease in islet-cell antibody-positive first-degree relatives of Type 1 DM³⁰². Nicotinamide administration (25mg/kg) has been shown in patients with recent onset Type 1 DM combined with intensive insulin therapy for up to two years after diagnosis to reduce HbA_{1c} levels³⁰³. Also relevant to patients with DM and renal insufficiency, nicotinamide can reduce intestinal absorption of phosphate and prevent the development of hyperphosphatemia³⁰⁴. As a caveat for caution, some studies have reported that prolonged exposure to nicotinamide may lead to impaired β -cell function and reduction in cell growth^{305, 306} as well as elevated nicotinamide levels may foster DM³⁰⁷. Furthermore, nicotinamide also may inhibit P450 and hepatic metabolism³⁰⁸ and play a role in the progression of other disorders such as Parkinson's disease²⁸³.

One novel pathway that may control some of the beneficial effects of nicotinamide during DM involves the forkhead transcription factors of the "O" class (FoxOs)^{309, 310} (Table 1). These transcription factors either inhibit or activate target gene expression by binding to DNA through the forkhead domain that relies upon fourteen protein-DNA contacts^{309, 311-314}. The term for these transcription factors is derived in part from imaging studies. On X-ray crystallography³¹⁵ or nuclear magnetic resonance imaging³¹⁶, the forkhead domain is described as a "winged helix" as a result of a butterfly-like appearance. The original nomenclature for these proteins, such as forkhead in rhabdomyosarcoma (*FKHR*), the Drosophila gene fork head (*fkh*), and Forkhead RElated ACTivator (FREAC)-1 and -2, has been replaced³¹⁷. The current nomenclature for human Fox proteins places all letters in uppercase, otherwise only the initial letter is listed as uppercase for the mouse, and for all other chordates the initial and subclass letters are in uppercase³¹⁸. Members of this family that include FoxO1, FoxO3, FoxO4, and FoxO6 are found throughout the body^{82, 191, 317}. These proteins are expressed in tissues of the reproductive system of males and females, skeletal muscle, the cardiovascular system, lung, liver, pancreas, spleen, thymus, and the nervous system^{157, 313, 314, 319}. Modulation of FoxOs is a viable therapeutic target for systems that involve metabotropic glutamate receptors⁹⁶, neurotrophins³²⁰, cancer^{157, 313, 321}, and cytokines²²² to foster intended cell survival.

Interestingly, FoxO proteins can modulate cell cycle progression to prevent tumor growth^{157, 313, 322}. For example, administration of the Bcr-Abl tyrosine kinase inhibitor imatinib in chronic myelogenous leukemia cell lines blocks cell proliferation and promotes apoptotic cell death through FoxO3a and increased TRAIL production³²³. The transcription factor E2F-1 that oversees cell cycle progression increases expression of FoxO1 and FoxO3a to lead to cell cycle arrest³²⁴. Other work indicates that FoxO proteins utilize the p53

upstream regulator p19(Arf) through Myc to block cell cycle induction and lymphoma progression³²⁵.

Since attempted initiation of the cell cycle such as in neurons may be detrimental and can lead to cell death^{49, 50, 326, 327}, one may consider the ability of FoxO proteins to block cell cycle progression to be beneficial in these circumstances. In regards to cell metabolism and DM, FoxO proteins may be cytoprotective. Interferon-gamma driven expression of tryptophan catabolism by cytotoxic T lymphocyte antigen 4 may activate Foxo3a to protect dendritic cells from injury in nonobese diabetic mice³²⁸. In addition, adipose tissue-specific expression of Foxo1 in mice improves glucose tolerance and sensitivity to insulin during an elevated fat diet³²⁹. FoxO proteins also may protect against diminished mitochondrial energy levels known to occur during insulin resistance such as in the elderly populations^{2, 3, 8}. In caloric restricted mice that have decreased energy reserves, Foxo1, Foxo3a, and Foxo4 mRNA levels were noted to progressively increase over two years³³⁰. These observations complement studies in *Drosophila* and mammalian cells that demonstrate an increase in insulin signaling to regulate cellular metabolism during the up-regulation of FoxO1 expression³³¹.

However, the role of FoxO proteins in different cell systems can be variable and do not consistently point to a beneficial effect of FoxO proteins. FoxO3a controls early activation and subsequent apoptotic injury in microglia through caspase action of caspase 3, 8, and 9^{55, 75}, illustrating that targeting FoxO3a activity may limit apoptotic caspase activity and promote cell survival (Figure 1). In clinical conditions, analysis of the genetic variance in *FOXO1a* and *FOXO3a* on metabolic profiles, age-related diseases, fertility, fecundity, and mortality in patients have observed higher HbA_{1c} levels and increased mortality risk associated with specific haplotypes of *FOXO1a*³³². These clinical observations may indicate that elevated glucose levels can reduce post-translational phosphorylation of FOXO1, FOXO3a, and FOXO4 and initiate cellular apoptosis³³³. In addition, mice with a constitutively active Foxo1 transgene have increased microsomal triglyceride transfer protein and high plasma triglyceride levels³³⁴. Increased transcriptional activity of FoxO1, such as by the Sirt1 activator resveratrol, also can decrease insulin mediated glucose uptake and result in insulin resistance³³⁵. Overexpression of Foxo1 in skeletal muscles of mice can lead to reduced skeletal muscle mass and poor glycemic control³³⁶. Other studies that block the expression of Foxo1 in normal and cachectic mice³³⁷ or reduce FoxO3 expression³³⁸ demonstrate positive effects with an increase in skeletal muscle mass or resistance to muscle atrophy.

As the pathways with cellular metabolism and FoxOs begin to unravel, nicotinamide becomes an attractive agent to consider for DM^{61, 62, 116, 236}. Nicotinamide inhibits FoxO protein activity through phosphorylation²⁵³ and may be protective through two separate mechanisms of post-translational modification of FoxO3a^{35, 157, 191, 314, 317}. Nicotinamide not only can maintain phosphorylation of FoxO3a and inhibit its activity to potentially block caspase 3 activity²⁵³, but also can preserve the integrity of the FoxO3a protein to block FoxO3a proteolysis that can yield pro-apoptotic amino-terminal fragments²⁵³. During oxidative stress, an initial inhibitory phosphorylation of FoxO3a at the regulatory phosphorylation sites (Thr³² and Ser²⁵³) occurs^{253, 339}. Yet, loss of phosphorylated FoxO3a expression appears to subsequently result over twelve hours, possibly by caspase degradation, which can raise the vulnerability of neurons to apoptotic injury²⁵³. The loss of both FoxO3a phosphorylation and the integrity of this transcription factor may then lead to apoptosis. FoxO3a proteolysis occurs during cell injury yielding an amino-terminal (Nt) fragment that can become biologically active and lead to cellular injury³⁴⁰. Nicotinamide, through the phosphorylation of FoxO3a blocks apoptotic cell injury and prevents caspase 3 activity²⁵³.

Nicotinamide is closely linked to cell longevity pathways that involve not only FoxOs, but also sirtuins^{116, 341, 342}. FoxO proteins are deacetylated by histone deacetylases. These include the sirtuin Sirt1, a NAD⁺-dependent deacetylase and the mammalian ortholog of the silent information regulator 2 (Sir2) protein³¹⁰, that can control multiple processes such as cell injury, lifespan, and metabolism^{343, 344}. FoxO proteins and sirtuins have been associated with cell longevity and aging as shown by early studies linking DAF-16 in *Caenorhabditis elegans*^{157, 310, 344-346}. Furthermore, sirtuins are tied to cellular metabolism^{343, 347} and increased cell survival^{344, 345, 348-350}. Yet, the relationship among nicotinamide, FoxO transcription factors, and sirtuins is not entirely clear (Table 1). For example, some studies suggest that stimulation of Sirt1 during starvation is dependent upon FoxO3a activity as well as p53³⁵¹. During exercise, an up-regulation of FoxO3a and Sirt1 activity is observed in the heart of rats³⁵², suggesting that physical activity may be beneficial for the cardiovascular system through FoxO proteins. Other work has shown that Sirt1 may repress the activity of FoxO1, FoxO3a, and FoxO4, illustrating that cellular longevity may benefit from reduction in FoxO protein generated apoptosis³⁵³.

However, nicotinamide prevents oxidant-induced apoptotic injury usually in a specific concentration range. Administration of nicotinamide in a range of 5.0 - 25.0 mmol/L significantly protects cells during oxidative stress injuries. This concentration range is similar to other injury paradigms in both animal models²⁶⁸ and in cell culture models^{61, 181, 250}. In contrast to these cytoprotective concentrations of nicotinamide that also can modulate offers gene regulation³⁵⁴, a reduction in nicotinamide levels during nicotinamidase expression supports increased cellular survival and longevity^{348, 350}. Nicotinamide can block cellular Sir2 by intercepting an ADP-ribosyl-enzyme-acetyl peptide intermediate with the regeneration of NAD⁺ (transglycosidation)³⁵⁵. Physiological concentrations of nicotinamide noncompetitively inhibit Sir2, suggesting that nicotinamide is a physiologically relevant regulator of Sir2 enzymes³⁵⁶. Interestingly, nicotinamidase expression which reduces nicotinamide concentrations prevents both apoptotic late DNA degradation and early PS exposure that appears to depend upon increased Sirt1 activity and may serve to modulate inflammatory cell activation^{348, 350}. In addition, inhibition of sirtuin (Sirt1) activity either by pharmacological methods or siRNA gene silencing is detrimental to cell survival during oxidative stress and blocks nicotinamidase protection, further supporting that Sirt1 activity may be necessary for nicotinamidase protection during oxidative stress. As a result, in relation to cell longevity, it is the lower concentrations of nicotinamide that can function as an inhibitor of sirtuins that are necessary for the promotion of increased lifespan and cellular survival^{250, 252, 253, 261, 348, 350, 357}, at least in yeast and metazoans^{116, 341, 342}. Sirtuins also may prevent nicotinamide from assisting with DNA repair by altering the accessibility of DNA damaged sites for repair enzymes³⁵⁸. Furthermore, sirtuin activators, at least at the experimental animal level, may promote glucose homeostasis and insulin sensitivity^{62, 343, 344, 350, 359} while also reducing the risk of obesity³⁶⁰.

Erythropoietin

The growth factor and cytokine EPO is approved by the Food and Drug Administration for the treatment of anemia, but continued new work has identified this agent for the potential treatment of multiple disorders^{209, 361}. Clinical considerations include treatment for depression³⁶², Alzheimer's disease^{52, 363, 364}, Parkinson's disease³⁶⁵, immune system dysfunction^{150, 222, 223, 366, 367}, neurodegeneration^{52, 71, 150, 223, 368-371}, cardiovascular disorders^{180, 216, 222, 372-379}, spinal cord injury^{380, 381}, brain edema³⁸², fertility³⁸³, trauma³⁸⁴⁻³⁸⁶, shock³⁸⁷⁻³⁸⁹, infection³⁹⁰⁻³⁹², pulmonary disease³⁹³⁻³⁹⁵, renal disease^{68, 396-398}, gastrointestinal disorders³⁹⁹⁻⁴⁰¹, ocular disease⁴⁰²⁻⁴⁰⁴, and metabolic disorders^{2, 33, 34, 71, 72, 405}. New studies further support the use of intravitreal EPO injections in patients⁴⁰⁶.

EPO is required for erythropoiesis⁴⁰⁷⁻⁴⁰⁹, but also functions in other organs and tissues, such as the brain, heart, and vascular system^{216, 222, 223, 410-412}. EPO production is believed to occur throughout the body^{83, 361, 413} and can be detected in the breath of healthy individuals⁴¹⁴. The principal organs of EPO production and secretion are the kidney, liver, brain, and uterus^{210, 339, 415}.

In regards to EPO during DM, plasma EPO is often low in diabetic patients with anemia⁴¹⁶ or without anemia⁴¹⁷. The inability of these individuals to produce EPO in response to a declining hemoglobin levels suggests an impaired EPO response during DM⁴¹⁸ (Table 1). Yet, increased EPO secretion during diabetic pregnancies may represent the body's attempt at endogenous protection against the complications of DM^{419, 420}. This potential cytoprotective capacity of EPO may be important during complications of DM, such as those that involve cognitive impairment. For example, EPO may improve cognitive ability. EPO may reduce in animal models apoptotic pathways during periods of hyperoxia in the developing brain^{421, 422}. Furthermore, clinical disorders may have periods of hyperoxia followed by cerebral hypoperfusion and hypoxia that can lead to cerebral injury with associated oxidative stress⁴²³. EPO under these conditions also may be protective since it can promote neurite outgrowth⁴²⁴ and also may regulate hemoglobin levels that have recently been associated with cognitive decline⁴²⁵. Elevated EPO concentrations during infant maturation have been correlated with increased Mental Development Index scores⁴²⁶ and EPO may prevent toxic effects of agents used to control cognitive function such as haloperidol⁴²⁷.

In relation to clinical relevance, EPO in diabetic as well as non-diabetic patients with severe, resistant congestive heart failure can decrease fatigue, increase left ventricular ejection fraction, and significantly decrease hospitalization stay⁴²⁸ (Table 1). In addition, EPO can serve to reverse the complications of anemia during DM³³. Experimental work during elevated glucose also has demonstrated that EPO can significantly improve vascular cell survival in a 1.0 ng/ml range⁷². EPO administration in patients also can significantly increase plasma levels of EPO well above this range of 1.0 ng/ml that has been associated with potential EPO cellular protection in patients with cardiac or renal disease^{429, 430}, illustrating that the effects of EPO observed during *in vitro* studies may parallel the cellular processes altered by EPO in patients with DM⁴²⁶.

Protection in the hematological and vascular system by EPO may rely upon modulation of FoxOs and Wnt signaling^{209, 361} (Table 1). EPO fosters erythroid progenitor cell development through the regulation of FoxO activity^{361, 413, 431} and may require regulation of specific gene expression through an EPO-FoxO3a association to promote erythropoiesis in cultured cells⁴³². In addition, EPO exerts cellular protection through Wnt signaling. Cell culture studies demonstrate that the Wnt1 protein is necessary and sufficient to impart cellular protection during elevated glucose exposure⁷². EPO maintains the expression of Wnt1 during elevated glucose exposure and prevents loss of Wnt1 expression that would occur in the absence of EPO during elevated glucose. In addition, blockade of Wnt1 with a Wnt1 antibody can neutralize the protective capacity of EPO, illustrating that Wnt1 is a critical component in the cytoprotection of EPO during elevated glucose exposure⁷². Furthermore, EPO during elevated glucose and similar to other models of oxidative stress can block neuronal degeneration⁷¹, prevents renal cell apoptosis⁴⁰⁵, apoptotic DNA degradation, and degeneration in cardiac and vascular cell models^{180, 216, 222, 411, 433}. Protection by EPO also is related to the maintenance of mitochondrial membrane potential, since loss of mitochondrial membrane potential is known to lead to apoptotic cell injury^{166, 434}. EPO has the capacity to prevent the depolarization of the mitochondrial membrane that also affects the release of cytochrome c^{150, 216, 435} (Figure 2).

Biomarkers and future considerations

Primary cellular pathways that may be critical for the development of new drug therapies also may possess great utility to function as biomarkers that can predict the diagnosis, onset, or progression of disease. Biomarkers can be used for the determination of specific genes, proteins, or products of cellular and biological processes. In addition, biomarkers can represent the response of cells or tissues to therapeutic strategies⁸¹. Many of the biological outcomes described for Wnt, nicotinamide, and EPO encompass necessary cellular pathways for development, growth, and survival. Yet, these entities also hold the potential to further disease progression or be set in motion during disease onset as an endogenous protective response. In regards to Wnt, signaling for this glycoprotein may indicate the onset of early tissue injury during conditions such as elevated glucose⁷², amyloid toxicity⁵³, or cardiac ischemia^{233, 436} and alert the need for rapid responsive treatments to limit cell injury. Yet, increased Wnt expression also may suggest disease progression as well as a poor prognostic response to prior therapies^{34, 177, 192}. Aberrant Wnt signaling has been associated with advanced prostate cancer and bone metastases⁴³⁷, the genesis of cancer stem cells¹⁸⁵, the reliance of FoxO pathways to promote unregulated cell proliferation in cancer⁴³⁸, and the vulnerability of patients with Type 2 DM to develop colorectal tumors⁴³⁹.

Independent from Wnt, FoxOs and EPO may serve as essential biomarkers. For example, the absence of FoxO proteins can suggest tumor progression⁴⁴⁰. On the converse side, the up-regulation of FoxO proteins can indicate a positive response to chemotherapy^{321, 441, 442} and also be an indicator and responsive agent to ischemic tissue⁴⁴³. In some scenarios, expression of FoxO proteins may suggest tissue protection and recovery, since FoxO3a activity may enhance vascular smooth muscle antioxidant properties in aged animals and be beneficial to the cardiovascular system during physical exertion³⁵². In a similar manner, EPO also may be a positive indicator of cytoprotective responses^{209, 210, 339}. Although EPO has not been shown to correlate with Psychomotor Development Index or an overall incidence of neurodevelopmental impairment, in clinical studies infants with elevated EPO possessed higher Mental Development Index scores than infants with lower EPO concentrations, suggesting that the presence of EPO may correlate with a positive developmental course. Advanced cognitive function also may rely upon appropriate levels of EPO that can be followed since both low and high levels of EPO in the elderly can be associated with diminished cognitive function⁴²⁵. Yet, the presence of EPO may affect a number of other systems in addition to the brain⁴⁴⁴. Recent work has shown that the presence of a truncated form of EPO receptor can function as a dominant negative regulator of EPO signaling and lead to hypertension, suggesting that monitoring of this biomarker may identify individuals susceptible to hypertension⁴⁴⁵. Although EPO has recently been reported to prevent drug-induced fibrosis and possible endothelial damage during chemotherapy⁴⁴⁶, it is important to note that EPO also may foster tumor progression^{447, 448}. EPO and its receptor are present in tumor specimens, may block tumor cell apoptosis through Akt⁴⁴⁹, enhance metastatic disease,⁴⁵⁰ and complicate radiotherapy by assisting with tumor angiogenesis⁴⁵¹.

Drug development for any disorder becomes a complex enterprise, especially for disorders such as DM with the multiple complications of this disease that can ensue through oxidant stress pathways. Nevertheless, elucidation of novel pathways and their biological role that have an intimate relationship with agents such as Wnt, nicotinamide, and EPO are vital for the successful treatment of clinical disease. In addition, defining the ability of these agents to function as potential biomarkers for disease can further enhance the utility of new entities irrespective of their treatment potential. Given that the present percentage of the gross domestic product for U.S. healthcare spending is the highest in the world, life expectancy in the U.S. trails behind other countries, and that the number of individuals affected by DM globally is expected to climb exponentially over the years, it becomes critical to understand

both the clinical efficacy of novel treatments as well as the potential complications of these agents especially in a variety of circumstances that may not only involve essential cellular repair but also undesirable cellular proliferation.

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References

1. Development OfEC-0a. OECD Health Data 2009. 2009. www.ecosante.org/oeed.htm
2. Maiese K, Chong ZZ, Shang YC. Mechanistic insights into diabetes mellitus and oxidative stress. *Curr Med Chem* 2007;14(16):1729–1738. [PubMed: 17627510]
3. Maiese K, Morhan SD, Chong ZZ. Oxidative stress biology and cell injury during type 1 and type 2 diabetes mellitus. *Curr Neurovasc Res* Feb;2007 4(1):63–71. [PubMed: 17311546]
4. Chan JC, Malik V, Jia W, et al. Diabetes in Asia: epidemiology, risk factors, and pathophysiology. *Jama* May 27;2009 301(20):2129–2140. [PubMed: 19470990]
5. Rebecchi KR, Wenke JL, Go EP, Desaire H. Label-free quantitation: a new glycoproteomics approach. *J Am Soc Mass Spectrom* Jun;2009 20(6):1048–1059. [PubMed: 19278867]
6. Dabelea D, Bell RA, D'Agostino RB Jr. et al. Incidence of diabetes in youth in the United States. *JAMA* Jun 27;2007 297(24):2716–2724. [PubMed: 17595272]
7. Harris MI, Eastman RC. Early detection of undiagnosed diabetes mellitus: a US perspective. *Diabetes Metab Res Rev* Jul-Aug;2000 16(4):230–236. [PubMed: 10934451]
8. Maiese K. Diabetic stress: new triumphs and challenges to maintain vascular longevity. *Expert Rev Cardiovasc Ther* Mar;2008 6(3):281–284. [PubMed: 18327989]
9. Daneman D. Type 1 diabetes. *Lancet* Mar 11;2006 367(9513):847–858. [PubMed: 16530579]
10. Bonner-Weir S. Life and death of the pancreatic beta cells. *Trends Endocrinol Metab* Nov;2000 11(9):375–378. [PubMed: 11042468]
11. Awata T, Kurihara S, Kikuchi C, et al. Evidence for association between the class I subset of the insulin gene minisatellite (IDDM2 locus) and IDDM in the Japanese population. *Diabetes* Oct; 1997 46(10):1637–1642. [PubMed: 9313762]
12. Baisch JM, Weeks T, Giles R, Hoover M, Stastny P, Capra JD. Analysis of HLA-DQ genotypes and susceptibility in insulin-dependent diabetes mellitus. *N Engl J Med* Jun 28;1990 322(26): 1836–1841. [PubMed: 2348836]
13. Todd JA, Bell JI, McDevitt HO. HLA-DQ beta gene contributes to susceptibility and resistance to insulin-dependent diabetes mellitus. *Nature* Oct 15-21;1987 329(6140):599–604. [PubMed: 3309680]
14. Lund T, O'Reilly L, Hutchings P, et al. Prevention of insulin-dependent diabetes mellitus in non-obese diabetic mice by transgenes encoding modified I-A beta-chain or normal I-E alpha-chain. *Nature* Jun 21;1990 345(6277):727–729. [PubMed: 2163026]
15. Permutt MA, Wasson J, Cox N. Genetic epidemiology of diabetes. *J Clin Invest* Jun;2005 115(6): 1431–1439. [PubMed: 15931378]
16. Kernan WN, Inzucchi SE, Viscoli CM, Brass LM, Bravata DM, Horwitz RI. Insulin resistance and risk for stroke. *Neurology* Sep 24;2002 59(6):809–815. [PubMed: 12349850]
17. Orchard TJ, Olson JC, Erbey JR, et al. Insulin resistance-related factors, but not glycemia, predict coronary artery disease in type 1 diabetes: 10-year follow-up data from the Pittsburgh Epidemiology of Diabetes Complications Study. *Diabetes Care* May;2003 26(5):1374–1379. [PubMed: 12716791]
18. Pietropaolo M, Barinas-Mitchell E, Pietropaolo SL, Kuller LH, Trucco M. Evidence of islet cell autoimmunity in elderly patients with type 2 diabetes. *Diabetes* Jan;2000 49(1):32–38. [PubMed: 10615947]

19. Bottino R, Trucco M. Multifaceted therapeutic approaches for a multigenic disease. *Diabetes Dec*; 2005 54(Suppl 2):S79–86. [PubMed: 16306345]
20. Davies JL, Kawaguchi Y, Bennett ST, et al. A genome-wide search for human type 1 diabetes susceptibility genes. *Nature Sep 8;1994 371(6493):130–136*. [PubMed: 8072542]
21. Kyvik KO, Green A, Beck-Nielsen H. Concordance rates of insulin dependent diabetes mellitus: a population based study of young Danish twins. *Bmj Oct 7;1995 311(7010):913–917*. [PubMed: 7580548]
22. Melanitou E. The autoimmune contrivance: genetics in the mouse model. *Clin Immunol Dec;2005 117(3):195–206*. [PubMed: 16188504]
23. Redondo MJ, Yu L, Hawa M, et al. Heterogeneity of type I diabetes: analysis of monozygotic twins in Great Britain and the United States. *Diabetologia Mar;2001 44(3):354–362*. [PubMed: 11317668]
24. Luppi P, Rossiello MR, Faas S, Trucco M. Genetic background and environment contribute synergistically to the onset of autoimmune diseases. *J Mol Med Aug;1995 73(8):381–393*. [PubMed: 8528740]
25. Del Prato S, Marchetti P. Beta- and alpha-cell dysfunction in type 2 diabetes. *Horm Metab Res Nov-Dec;2004 36(11-12):775–781*. [PubMed: 15655708]
26. Hao J, Shen W, Tian C, et al. Mitochondrial nutrients improve immune dysfunction in the type 2 diabetic Goto-Kakizaki rats. *J Cell Mol Med Apr;2009 13(4):701–711*. [PubMed: 18410524]
27. Morley JE. Diabetes, sarcopenia, and frailty. *Clin Geriatr Med Aug;2008 24(3):455–469*. vi. [PubMed: 18672182]
28. McIntyre RS, Rasgon NL, Kemp DE, et al. Metabolic syndrome and major depressive disorder: co-occurrence and pathophysiologic overlap. *Curr Diab Rep Feb;2009 9(1):51–59*. [PubMed: 19192425]
29. Wu SY, Wang GF, Liu ZQ, et al. Effect of geniposide, a hypoglycemic glucoside, on hepatic regulating enzymes in diabetic mice induced by a high-fat diet and streptozotocin. *Acta Pharmacol Sin Feb;2009 30(2):202–208*. [PubMed: 19122671]
30. Guarnieri G, Zanetti M, Vinci P, Cattin MR, Barazzoni R. Insulin resistance in chronic uremia. *J Ren Nutr Jan;2009 19(1):20–24*. [PubMed: 19121765]
31. Aso Y, Suganuma R, Wakabayashi S, et al. Anemia is associated with an elevated serum level of high-molecular-weight adiponectin in patients with type 2 diabetes independently of renal dysfunction. *Transl Res Oct;2009 154(4):175–182*. [PubMed: 19766961]
32. Gossai D, Lau-Cam CA. The effects of taurine, taurine homologs and hypotaurine on cell and membrane antioxidative system alterations caused by type 2 diabetes in rat erythrocytes. *Adv Exp Med Biol 2009;643:359–368*. [PubMed: 19239167]
33. Singh DK, Winocour P, Farrington K. Erythropoietic stress and anemia in diabetes mellitus. *Nat Rev Endocrinol Apr;2009 5(4):204–210*. [PubMed: 19352318]
34. Maiese K. Triple play: Promoting neurovascular longevity with nicotinamide, WNT, and erythropoietin in diabetes mellitus. *Biomed Pharmacother April - May;2008 62(4):218–232*. [PubMed: 18342481]
35. Maiese, K.; Chong, Z.; Li, F. Reducing oxidative stress and enhancing neurovascular longevity during diabetes mellitus.. In: Maiese, K., editor. *Neurovascular Medicine: Pursuing Cellular Longevity for Healthy Aging*. Oxford University Press; New York, NY: 2009. p. 540-564. ISBN13: 978-0-19-532669-7, ISBN10: 0-19-532669-5
36. Donahoe SM, Stewart GC, McCabe CH, et al. Diabetes and mortality following acute coronary syndromes. *JAMA Aug 15;2007 298(7):765–775*. [PubMed: 17699010]
37. Kuhad A, Bishnoi M, Tiwari V, Chopra K. Suppression of NF-kappabeta signaling pathway by tocotrienol can prevent diabetes associated cognitive deficits. *Pharmacol Biochem Behav Apr; 2009 92(2):251–259*. [PubMed: 19138703]
38. Ott A, Stolk RP, van Harskamp F, Pols HA, Hofman A, Breteler MM. Diabetes mellitus and the risk of dementia: The Rotterdam Study. *Neurology Dec 10;1999 53(9):1937–1942*. [PubMed: 10599761]
39. Schnaider Beeri M, Goldbourt U, Silverman JM, et al. Diabetes mellitus in midlife and the risk of dementia three decades later. *Neurology Nov 23;2004 63(10):1902–1907*. [PubMed: 15557509]

40. Xu WL, Qiu CX, Wahlin A, Winblad B, Fratiglioni L. Diabetes mellitus and risk of dementia in the Kungsholmen project: a 6-year follow-up study. *Neurology* Oct 12;2004 63(7):1181–1186. [PubMed: 15477535]
41. Beeri MS, Silverman JM, Davis KL, et al. Type 2 diabetes is negatively associated with Alzheimer's disease neuropathology. *J Gerontol A Biol Sci Med Sci* Apr;2005 60(4):471–475. [PubMed: 15933386]
42. Maiese K, Chong ZZ, Hou J, Shang YC. New strategies for Alzheimer's disease and cognitive impairment. *Oxid Med Cell Longev* 2009;2(5):279–290. [PubMed: 20716915]
43. Steen E, Terry BM, Rivera EJ, et al. Impaired insulin and insulin-like growth factor expression and signaling mechanisms in Alzheimer's disease--is this type 3 diabetes? *J Alzheimers Dis* Feb;2005 7(1):63–80. [PubMed: 15750215]
44. Chong ZZ, Li F, Maiese K. Oxidative stress in the brain: Novel cellular targets that govern survival during neurodegenerative disease. *Prog Neurobiol* Feb;2005 75(3):207–246. [PubMed: 15882775]
45. Chong ZZ, Li F, Maiese K. Stress in the brain: novel cellular mechanisms of injury linked to Alzheimer's disease. *Brain Res Brain Res Rev* Jul;2005 49(1):1–21. [PubMed: 15960984]
46. Maiese K, Holloway HH, Larson DM, Soncrant TT. Effect of acute and chronic arecoline treatment on cerebral metabolism and blood flow in the conscious rat. *Brain Res* 1994;641(1):65–75. [PubMed: 8019853]
47. Kim HS, Lim JY, Sul D, et al. Neuroprotective effects of the new diterpene, CBNU06 against beta-amyloid-induced toxicity through the inhibition of NF-kappaB signaling pathway in PC12 cells. *Eur J Pharmacol* Nov 10;2009 622(1-3):25–31. [PubMed: 19765580]
48. Salminen A, Kaamiranta K. Siglec receptors and hiding plaques in Alzheimer's disease. *J Mol Med* Jul;2009 87(7):697–701. [PubMed: 19390836]
49. Chong ZZ, Li F, Maiese K. Attempted Cell Cycle Induction in Post-Mitotic Neurons Occurs in Early and Late Apoptotic Programs Through Rb, E2F1, and Caspase 3. *Curr Neurovasc Res* Feb; 2006 3(1):25–39. [PubMed: 16472123]
50. Lin SH, Chong ZZ, Maiese K. Cell cycle induction in post-mitotic neurons proceeds in concert with the initial phase of programmed cell death in rat. *Neurosci Lett* Sep 14;2001 310(2-3):173–177. [PubMed: 11585595]
51. Majd S, Zarifkar A, Rastegar K, Takhshid MA. Different fibrillar Abeta 1-42 concentrations induce adult hippocampal neurons to reenter various phases of the cell cycle. *Brain Res* Jul 7;2008 1218:224–229. [PubMed: 18533137]
52. Chong ZZ, Li F, Maiese K. Erythropoietin requires NF-kappaB and its nuclear translocation to prevent early and late apoptotic neuronal injury during beta-amyloid toxicity. *Curr Neurovasc Res* Dec;2005 2(5):387–399. [PubMed: 16375720]
53. Chong ZZ, Li F, Maiese K. Cellular demise and inflammatory microglial activation during beta-amyloid toxicity are governed by Wnt1 and canonical signaling pathways. *Cell Signal* Jun;2007 19(6):1150–1162. [PubMed: 17289346]
54. Majd S, Rastegar K, Zarifkar A, Takhshid MA. Fibrillar beta-amyloid (Abeta) (1-42) elevates extracellular Abeta in cultured hippocampal neurons of adult rats. *Brain Res* Dec 14;2007 1185:321–327. [PubMed: 17961521]
55. Shang YC, Chong ZZ, Hou J, Maiese K. The forkhead transcription factor FoxO3a controls microglial inflammatory activation and eventual apoptotic injury through caspase 3. *Curr Neurovasc Res* Feb;2009 6(1):20–31. [PubMed: 19355923]
56. Bakshi P, Margenthaler E, Laporte V, Crawford F, Mullan M. Novel role of CXCR2 in regulation of gamma-secretase activity. *ACS Chem Biol* Dec 19;2008 3(12):777–789. [PubMed: 19067586]
57. Lu J, Wu DM, Zheng YL, et al. Trace amounts of copper exacerbate beta amyloid-induced neurotoxicity in the cholesterol-fed mice through TNF-mediated inflammatory pathway. *Brain Behav Immun* Feb;2009 23(2):193–203. [PubMed: 18835350]
58. Bitner RS, Nikkel AL, Markosyan S, Otte S, Puttfarcken P, Gopalakrishnan M. Selective alpha7 nicotinic acetylcholine receptor activation regulates glycogen synthase kinase3beta and decreases tau phosphorylation in vivo. *Brain Res* Apr 10;2009 1265:65–74. [PubMed: 19230830]

59. Vaisid T, Barnoy S, Kosower NS. Calpastatin overexpression attenuates amyloid-beta-peptide toxicity in differentiated PC12 cells. *Neuroscience* Oct 28;2008 156(4):921–931. [PubMed: 18786620]
60. Erol A. Unraveling the Molecular Mechanisms Behind the Metabolic Basis of Sporadic Alzheimer's Disease. *J Alzheimers Dis.* Feb 16;2009
61. Maiese K, Chong ZZ. Nicotinamide: necessary nutrient emerges as a novel cytoprotectant for the brain. *Trends Pharmacol Sci* May;2003 24(5):228–232. [PubMed: 12767721]
62. Maiese K, Chong ZZ, Hou J, Shang YC. The vitamin nicotinamide: translating nutrition into clinical care. *Molecules* 2009;14(9):3446–3485. [PubMed: 19783937]
63. Schubert M, Gautam D, Surjo D, et al. Role for neuronal insulin resistance in neurodegenerative diseases. *Proc Natl Acad Sci U S A* Mar 2;2004 101(9):3100–3105. [PubMed: 14981233]
64. Newsholme P, Haber EP, Hirabara SM, et al. Diabetes associated cell stress and dysfunction: role of mitochondrial and non-mitochondrial ROS production and activity. *J Physiol* Aug 15;2007 583(Pt 1):9–24. [PubMed: 17584843]
65. Szabo C. Role of nitrosative stress in the pathogenesis of diabetic vascular dysfunction. *Br J Pharmacol* Mar;2009 156(5):713–727. [PubMed: 19210748]
66. Simone S, Gorin Y, Velagapudi C, Abboud HE, Habib SL. Mechanism of oxidative DNA damage in diabetes: tuberin inactivation and downregulation of DNA repair enzyme 8-oxo-7,8-dihydro-2'-deoxyguanosine-DNA glycosylase. *Diabetes* Oct;2008 57(10):2626–2636. [PubMed: 18599524]
67. Gaddini L, Villa M, Matteucci A, et al. Early effects of high glucose in retinal tissue cultures Renin-Angiotensin system-dependent and -independent signaling. *Neurobiol Dis* Aug;2009 35(2): 278–285. [PubMed: 19481149]
68. Singh DK, Winocour P, Farrington K. Mechanisms of disease: the hypoxic tubular hypothesis of diabetic nephropathy. *Nat Clin Pract Nephrol* Apr;2008 4(4):216–226. [PubMed: 18268525]
69. Barbosa NB, Oliveira C, Araldi D, Folmer V, Rocha JB, Nogueira CW. Acute diphenyl diselenide treatment reduces hyperglycemia but does not change delta-aminolevulinic acid dehydratase activity in alloxan-induced diabetes in rats. *Biol Pharm Bull* Dec;2008 31(12):2200–2204. [PubMed: 19043199]
70. Memisogullari R, Bakan E. Levels of ceruloplasmin, transferrin, and lipid peroxidation in the serum of patients with Type 2 diabetes mellitus. *J Diabetes Complications* Jul-Aug;2004 18(4): 193–197. [PubMed: 15207835]
71. Chattopadhyay M, Walter C, Mata M, Fink DJ. Neuroprotective effect of herpes simplex virus-mediated gene transfer of erythropoietin in hyperglycemic dorsal root ganglion neurons. *Brain* Apr;2009 132(Pt 4):879–888. [PubMed: 19244253]
72. Chong ZZ, Shang YC, Maiese K. Vascular injury during elevated glucose can be mitigated by erythropoietin and Wnt signaling. *Curr Neurovasc Res* Aug;2007 4(3):194–204. [PubMed: 17691973]
73. Lin J, Zheng S, Chen A. Curcumin attenuates the effects of insulin on stimulating hepatic stellate cell activation by interrupting insulin signaling and attenuating oxidative stress. *Lab Invest* Dec; 2009 89(12):1397–1409. [PubMed: 19841616]
74. Liu W, Liu P, Tao S, et al. Berberine inhibits aldose reductase and oxidative stress in rat mesangial cells cultured under high glucose. *Arch Biochem Biophys* Jul 15;2008 475(2):128–134. [PubMed: 18471986]
75. Shang YC, Chong ZZ, Hou J, Maiese K. FoxO3a Governs Early Microglial Proliferation and Employs Mitochondrial Depolarization with Caspase 3, 8, and 9 Cleavage During Oxidant Induced Apoptosis. *Curr Neurovasc Res* Nov 1;2009 6(4):223–238. [PubMed: 19807657]
76. Chen H, Li X, Epstein PN. MnSOD and catalase transgenes demonstrate that protection of islets from oxidative stress does not alter cytokine toxicity. *Diabetes* May;2005 54(5):1437–1446. [PubMed: 15855331]
77. Lepore DA, Shinkel TA, Fiscaro N, et al. Enhanced expression of glutathione peroxidase protects islet beta cells from hypoxia-reoxygenation. *Xenotransplantation* Jan;2004 11(1):53–59. [PubMed: 14962293]

78. Ribeiro MC, Barbosa NB, de Almeida TM, et al. High-fat diet and hydrochlorothiazide increase oxidative stress in brain of rats. *Cell Biochem Funct* Oct;2009 27(7):473–478. [PubMed: 19784960]
79. Rachek LI, Thornley NP, Grishko VI, LeDoux SP, Wilson GL. Protection of INS-1 cells from free fatty acid-induced apoptosis by targeting hOGG1 to mitochondria. *Diabetes* Apr;2006 55(4):1022–1028. [PubMed: 16567524]
80. Haber CA, Lam TK, Yu Z, et al. N-acetylcysteine and taurine prevent hyperglycemia-induced insulin resistance in vivo: possible role of oxidative stress. *Am J Physiol Endocrinol Metab* Oct; 2003 285(4):E744–753. [PubMed: 12799318]
81. Maiese K. Marking the onset of oxidative stress: Biomarkers and novel strategies. *Oxid Med Cell Longev* 2009;2(1):1. [PubMed: 20046637]
82. Maiese K, Hou J, Chong ZZ, Shang YC. Erythropoietin, forkhead proteins, and oxidative injury: biomarkers and biology. *ScientificWorldJournal* 2009;9:1072–1104. [PubMed: 19802503]
83. Maiese K, Chong ZZ, Hou J, Shang YC. Erythropoietin and oxidative stress. *Curr Neurovasc Res* 2008;5(2):125–142. [PubMed: 18473829]
84. Ihara Y, Toyokuni S, Uchida K, et al. Hyperglycemia causes oxidative stress in pancreatic beta-cells of GK rats, a model of type 2 diabetes. *Diabetes* Apr;1999 48(4):927–932. [PubMed: 10102716]
85. Ceriello A, dello Russo P, Amstad P, Cerutti P. High glucose induces antioxidant enzymes in human endothelial cells in culture. Evidence linking hyperglycemia and oxidative stress. *Diabetes* Apr;1996 45(4):471–477. [PubMed: 8603769]
86. Yano M, Hasegawa G, Ishii M, et al. Short-term exposure of high glucose concentration induces generation of reactive oxygen species in endothelial cells: implication for the oxidative stress associated with postprandial hyperglycemia. *Redox Rep* 2004;9(2):111–116. [PubMed: 15231066]
87. Monnier L, Mas E, Ginet C, et al. Activation of oxidative stress by acute glucose fluctuations compared with sustained chronic hyperglycemia in patients with type 2 diabetes. *JAMA* Apr 12;2006 295(14):1681–1687. [PubMed: 16609090]
88. De Felice FG, Velasco PT, Lambert MP, et al. Aβ oligomers induce neuronal oxidative stress through an N-methyl-D-aspartate receptor-dependent mechanism that is blocked by the Alzheimer drug memantine. *J Biol Chem* Apr 13;2007 282(15):11590–11601. [PubMed: 17308309]
89. Lin SH, Maiese K. The metabotropic glutamate receptor system protects against ischemic free radical programmed cell death in rat brain endothelial cells. *J Cereb Blood Flow Metab* 2001;21(3):262–275. [PubMed: 11295881]
90. Nomoto M, Miyata M, Yin S, et al. Bile acid-induced elevated oxidative stress in the absence of farnesoid X receptor. *Biol Pharm Bull* Feb;2009 32(2):172–178. [PubMed: 19182371]
91. Walsh KB, Toledo AH, Rivera-Chavez FA, Lopez-Neblina F, Toledo-Pereyra LH. Inflammatory mediators of liver ischemia-reperfusion injury. *Exp Clin Transplant* Jun;2009 7(2):78–93. [PubMed: 19715511]
92. Escobar J, Pereda J, Arduini A, et al. Cross-talk between oxidative stress and pro-inflammatory cytokines in acute pancreatitis: a key role for protein phosphatases. *Curr Pharm Des* 2009;15(26): 3027–3042. [PubMed: 19754377]
93. Hammoud DA, Hoffman JM, Pomper MG. Molecular neuroimaging: from conventional to emerging techniques. *Radiology* Oct;2007 245(1):21–42. [PubMed: 17885179]
94. Swan GE, Lessov-Schlaggar CN. The effects of tobacco smoke and nicotine on cognition and the brain. *Neuropsychol Rev* Sep;2007 17(3):259–273. [PubMed: 17690985]
95. Campos-Esparza MR, Sanchez-Gomez MV, Matute C. Molecular mechanisms of neuroprotection by two natural antioxidant polyphenols. *Cell Calcium* Apr;2009 45(4):358–368. [PubMed: 19201465]
96. Chong ZZ, Li F, Maiese K. Group I Metabotropic Receptor Neuroprotection Requires Akt and Its Substrates that Govern FOXO3a, Bim, and beta-Catenin During Oxidative Stress. *Curr Neurovasc Res* May;2006 3(2):107–117. [PubMed: 16719794]
97. He Z, Lu Q, Xu X, Huang L, Chen J, Guo L. DDPH ameliorated oxygen and glucose deprivation-induced injury in rat hippocampal neurons via interrupting Ca²⁺ overload and glutamate release. *Eur J Pharmacol* Jan 28;2009 603(1-3):50–55. [PubMed: 19105952]

98. Lehtinen MK, Tegelberg S, Schipper H, et al. Cystatin B deficiency sensitizes neurons to oxidative stress in progressive myoclonus epilepsy, EPM1. *J Neurosci* May 6;2009 29(18):5910–5915. [PubMed: 19420257]
99. Ye J, Han Y, Wang C, Yu W. Cytoprotective effect of polypeptide from *Chlamys farreri* on neuroblastoma (SH-SY5Y) cells following HO exposure involves scavenging ROS and inhibition JNK phosphorylation. *J Neurochem* Oct;2009 111(2):441–451. [PubMed: 19682211]
100. Anderson DW, Bradbury KA, Schneider JS. Broad neuroprotective profile of nicotinamide in different mouse models of MPTP-induced parkinsonism. *Eur J Neurosci* Aug;2008 28(3):610–617. [PubMed: 18702732]
101. Morissette M, Al Sweidi S, Callier S, Di Paolo T. Estrogen and SERM neuroprotection in animal models of Parkinson's disease. *Mol Cell Endocrinol* Aug 13;2008 290(1-2):60–69. [PubMed: 18515001]
102. Morissette M, Le Saux M, D'Astous M, et al. Contribution of estrogen receptors alpha and beta to the effects of estradiol in the brain. *J Steroid Biochem Mol Biol* Feb;2008 108(3-5):327–338. [PubMed: 17936613]
103. Rodriguez-Blanco J, Martin V, Herrera F, Garcia-Santos G, Antolin I, Rodriguez C. Intracellular signaling pathways involved in post-mitotic dopaminergic PC12 cell death induced by 6-hydroxydopamine. *J Neurochem* Oct;2008 107(1):127–140. [PubMed: 18665912]
104. Sales Santos I, da Rocha Tomé A, Saldanha G, Ferreira P, Militão G, de Freitas R. Oxidative stress in the hippocampus during experimental seizures can be ameliorated with the antioxidant ascorbic acid. *Oxid Med Cell Longev* 2009;2(4):23–30.
105. Maiese K, Chong ZZ, Shang YC, Hou J. Therapeutic promise and principles: Metabotropic glutamate receptors. *Oxid Med Cell Longev* Jul 1;2008 1(1):1–14. [PubMed: 19750024]
106. Probst-Hensch NM, Imboden M, Felber Dietrich D, et al. Glutathione S-transferase polymorphisms, passive smoking, obesity, and heart rate variability in nonsmokers. *Environ Health Perspect* Nov;2008 116(11):1494–1499. [PubMed: 19057702]
107. Yemisli M, Gursoy-Ozdemir Y, Vural A, Can A, Topalkara K, Dalkara T. Pericyte contraction induced by oxidative-nitrative stress impairs capillary reflow despite successful opening of an occluded cerebral artery. *Nat Med* Sep;2009 15(9):1031–1037. [PubMed: 19718040]
108. Fatma N, Kubo E, Toris CB, Stamer WD, Camras CB, Singh DP. PRDX6 attenuates oxidative stress- and TGFbeta-induced abnormalities of human trabecular meshwork cells. *Free Radic Res* Sep;2009 43(9):783–795. [PubMed: 19572226]
109. Gomes P, Simão S, Silva E, et al. Aging increases oxidative stress and renal expression of oxidant and antioxidant enzymes that are associated with an increased trend in systolic blood pressure. *Oxid Med Cell Longev* 2009;2(3):19–26. [PubMed: 20046641]
110. Pedersen MO, Jensen R, Pedersen DS, et al. Metallothionein-I+II in neuroprotection. *Biofactors* Jul-Aug;2009 35(4):315–325. [PubMed: 19655389]
111. Chuang HH, Lin S. Oxidative challenges sensitize the capsaicin receptor by covalent cysteine modification. *Proc Natl Acad Sci U S A* Nov 24;2009 106(47):20097–20102. [PubMed: 19897733]
112. Astiz M, de Alaniz MJ, Marra CA. Effect of pesticides on cell survival in liver and brain rat tissues. *Ecotoxicol Environ Saf* Oct;2009 72(7):2025–2032. [PubMed: 19493570]
113. Slotkin TA, Seidler FJ. Protein kinase C is a target for diverse developmental neurotoxicants: transcriptional responses to chlorpyrifos, diazinon, dieldrin and divalent nickel in PC12 cells. *Brain Res* Mar 31;2009 1263:23–32. [PubMed: 19368821]
114. Hamden K, Carreau S, Jamoussi K, et al. 1Alpha,25 dihydroxyvitamin D3: therapeutic and preventive effects against oxidative stress, hepatic, pancreatic and renal injury in alloxan-induced diabetes in rats. *J Nutr Sci Vitaminol (Tokyo)* Jun;2009 55(3):215–222. [PubMed: 19602829]
115. Regulska M, Leskiewicz M, Budziszewska B, et al. Inhibitory effects of 1,25-dihydroxyvitamin D(3) and its low-calcemic analogues on staurosporine-induced apoptosis. *Pharmacol Rep* Jul-Aug;2007 59(4):393–401. [PubMed: 17901567]
116. Li F, Chong ZZ, Maiese K. Cell Life Versus Cell Longevity: The Mysteries Surrounding the NAD(+) Precursor Nicotinamide. *Curr Med Chem* 2006;13(8):883–895. [PubMed: 16611073]

117. Li J, Wang H, Rosenberg PA. Vitamin K prevents oxidative cell death by inhibiting activation of 12-lipoxygenase in developing oligodendrocytes. *J Neurosci Res* Jul;2009 87(9):1997–2005. [PubMed: 19235890]
118. Ozsoy N, Candoken E, Akev N. Implications for degenerative disorders: Antioxidative activity, total phenols, flavonoids, ascorbic acid, β -carotene, α -tocopherol in Aloe vera. *Oxid Med Cell Longev* 2009;2(2):99–106. [PubMed: 20357932]
119. Then SM, Mazlan M, Mat Top G, Wan Ngah WZ. Is vitamin E toxic to neuron cells? *Cell Mol Neurobiol* Jun;2009 29(4):485–496. [PubMed: 19172392]
120. El-Mir MY, Detaille D, G RV, et al. Neuroprotective role of antidiabetic drug metformin against apoptotic cell death in primary cortical neurons. *J Mol Neurosci* 2008;34(1):77–87. [PubMed: 18040888]
121. Kui L, Weiwei Z, Ling L, et al. Ghrelin inhibits apoptosis induced by high glucose and sodium palmitate in adult rat cardiomyocytes through the PI3K-Akt signaling pathway. *Regul Pept* Jun 5;2009 155(1-3):62–69. [PubMed: 19289146]
122. Head BP, Patel HH, Niesman IR, Drummond JC, Roth DM, Patel PM. Inhibition of p75 neurotrophin receptor attenuates isoflurane-mediated neuronal apoptosis in the neonatal central nervous system. *Anesthesiology* Apr;2009 110(4):813–825. [PubMed: 19293698]
123. Han Z, Xiao MJ, Shao B, Zheng RY, Yang GY, Jin K. Attenuation of ischemia-induced rat brain injury by 2-(2-benzofuranyl)-2-imidazoline, a high selectivity ligand for imidazoline I(2) receptors. *Neurol Res* May;2009 31(4):390–395. [PubMed: 19508825]
124. Maiese K. From the Bench to the Bedside: The Molecular Management of Cerebral Ischemia. *Clinical Neuropharm* 1998;21(1):1–7.
125. Maiese K, Pek L, Berger SB, Reis DJ. Reduction in focal cerebral ischemia by agents acting at imidazole receptors. *J Cereb Blood Flow Metab* 1992;12(1):53–63. [PubMed: 1345758]
126. Nakka VP, Gusain A, Mehta SL, Raghbir R. Molecular mechanisms of apoptosis in cerebral ischemia: multiple neuroprotective opportunities. *Mol Neurobiol* Feb;2008 37(1):7–38. [PubMed: 18066503]
127. Cardoso L, Herman BC, Verborgt O, Laudier D, Majeska RJ, Schaffler MB. Osteocyte apoptosis controls activation of intracortical resorption in response to bone fatigue. *J Bone Miner Res* Apr; 2009 24(4):597–605. [PubMed: 19049324]
128. Arboleda G, Morales LC, Benitez B, Arboleda H. Regulation of ceramide-induced neuronal death: cell metabolism meets neurodegeneration. *Brain Res Rev* Mar;2009 59(2):333–346. [PubMed: 18996148]
129. Maiese K, Vincent AM. Critical temporal modulation of neuronal programmed cell injury. *Cell Mol Neurobiol* 2000;20(3):383–400. [PubMed: 10789835]
130. Zhong J, Zheng W, Huang L, et al. PrP106-126 amide causes the semi-penetrated poration in the supported lipid bilayers. *Biochim Biophys Acta* Jun;2007 1768(6):1420–1429. [PubMed: 17451641]
131. Burgos-Ramos E, Puebla-Jimenez L, Arilla-Ferreiro E. Minocycline provides protection against beta-amyloid(25-35)-induced alterations of the somatostatin signaling pathway in the rat temporal cortex. *Neuroscience* Jul 17;2008 154(4):1458–1466. [PubMed: 18555616]
132. Burgos-Ramos E, Puebla-Jimenez L, Arilla-Ferreiro E. Minocycline prevents A β (25-35)-induced reduction of somatostatin and neprilysin content in rat temporal cortex. *Life Sci* Feb 13;2009 84(7-8):205–210. [PubMed: 19101571]
133. Casoli T, Di Stefano G, Giorgetti B, et al. Release of beta-amyloid from high-density platelets: implications for Alzheimer's disease pathology. *Ann N Y Acad Sci* Jan;2007 1096:170–178. [PubMed: 17405928]
134. Kelley BJ, Knopman DS. Alternative medicine and Alzheimer disease. *Neurologist* Sep;2008 14(5):299–306. [PubMed: 18784599]
135. Liang WS, Dunckley T, Beach TG, et al. Altered neuronal gene expression in brain regions differentially affected by Alzheimer's disease: a reference data set. *Physiol Genomics* Apr 22;2008 33(2):240–256. [PubMed: 18270320]

136. Verdaguer E, Susana Gde A, Clemens A, Pallas M, Camins A. Implication of the transcription factor E2F-1 in the modulation of neuronal apoptosis. *Biomed Pharmacother* Aug;2007 61(7): 390–399. [PubMed: 17178208]
137. Sommer C. Neuronal plasticity after ischemic preconditioning and TIA-like preconditioning ischemic periods. *Acta Neuropathol* May;2009 117(5):511–523. [PubMed: 19084975]
138. Braga M, Sinha Hikim AP, Datta S, et al. Involvement of oxidative stress and caspase 2-mediated intrinsic pathway signaling in age-related increase in muscle cell apoptosis in mice. *Apoptosis* Jun;2008 13(6):822–832. [PubMed: 18461459]
139. Maiese, K.; Chong, ZZ.; Kang, J. Transformation into treatment: Novel therapeutics that begin within the cell.. In: Maiese, K., editor. *Neuronal and Vascular Plasticity: Elucidating Basic Cellular Mechanisms for Future Therapeutic Discovery*. Kluwer Academic Publishers; Norwell, MA: 2003. p. 1-26.
140. Gross J, Machulik A, Amarjargal N, et al. Expression of apoptosis-related genes in the organ of Corti, modiolus and stria vascularis of newborn rats. *Brain Res* Aug 8;2007 1162:56–68. [PubMed: 17612509]
141. Maiese K, Ahmad I, TenBroeke M, Gallant J. Metabotropic glutamate receptor subtypes independently modulate neuronal intracellular calcium. *J Neurosci Res* 1999;55:472–485. [PubMed: 10723057]
142. Maiese K, Vincent AM. Membrane asymmetry and DNA degradation: functionally distinct determinants of neuronal programmed cell death. *J Neurosci Res* 2000;59(4):568–580. [PubMed: 10679797]
143. Dombroski D, Balasubramanian K, Schroit AJ. Phosphatidylserine expression on cell surfaces promotes antibody- dependent aggregation and thrombosis in beta2-glycoprotein I-immune mice. *J Autoimmun* 2000;14(3):221–229. [PubMed: 10756084]
144. Jessel R, Haertel S, Socaciu C, Tykhonova S, Diehl HA. Kinetics of apoptotic markers in exogenously induced apoptosis of EL4 cells. *J Cell Mol Med* 2002;6(1):82–92. [PubMed: 12003671]
145. Kang JQ, Chong ZZ, Maiese K. Critical role for Akt1 in the modulation of apoptotic phosphatidylserine exposure and microglial activation. *Mol Pharmacol* Sep;2003 64(3):557–569. [PubMed: 12920191]
146. Maiese K, Vincent A, Lin SH, Shaw T. Group I and Group III metabotropic glutamate receptor subtypes provide enhanced neuroprotection. *J Neurosci Res* 2000;62(2):257–272. [PubMed: 11020218]
147. Mari C, Karabiyikoglu M, Goris ML, Tait JF, Yenari MA, Blankenberg FG. Detection of focal hypoxic-ischemic injury and neuronal stress in a rodent model of unilateral MCA occlusion/ reperfusion using radiolabeled annexin V. *Eur J Nucl Med Mol Imaging* May;2004 31(5):733–739. [PubMed: 14985868]
148. Maiese K. The dynamics of cellular injury: transformation into neuronal and vascular protection. *Histol Histopathol* 2001;16(2):633–644. [PubMed: 11332719]
149. Vincent AM, Maiese K. Direct temporal analysis of apoptosis induction in living adherent neurons. *J Histochem Cytochem* 1999;47(5):661–672. [PubMed: 10219058]
150. Chong ZZ, Lin SH, Kang JQ, Maiese K. Erythropoietin prevents early and late neuronal demise through modulation of Akt1 and induction of caspase 1, 3, and 8. *J Neurosci Res* Mar 1;2003 71(5):659–669. [PubMed: 12584724]
151. Chong ZZ, Lin SH, Kang JQ, Maiese K. The tyrosine phosphatase SHP2 modulates MAP kinase p38 and caspase 1 and 3 to foster neuronal survival. *Cell Mol Neurobiol* Oct;2003 23(4-5):561–578. [PubMed: 14514016]
152. Maiese K, Boccone L. Neuroprotection by peptide growth factors against anoxia and nitric oxide toxicity requires modulation of protein kinase C. *J Cereb Blood Flow Metab* 1995;15(3):440–449. [PubMed: 7714002]
153. Maiese K, Boniece IR, Skurat K, Wagner JA. Protein kinases modulate the sensitivity of hippocampal neurons to nitric oxide toxicity and anoxia. *J Neurosci Res* 1993;36(1):77–87. [PubMed: 8230323]

154. Maiese K, TenBroeke M, Kue I. Neuroprotection of lubeluzole is mediated through the signal transduction pathways of nitric oxide. *J Neurochem* 1997;68(2):710–714. [PubMed: 9003060]
155. Leytin V, Allen DJ, Mykhaylov S, Lyubimov E, Freedman J. Thrombin-triggered platelet apoptosis. *J Thromb Haemost* Dec;2006 4(12):2656–2663. [PubMed: 16961585]
156. Gilfillan AM, Rivera J. The tyrosine kinase network regulating mast cell activation. *Immunol Rev* Mar;2009 228(1):149–169. [PubMed: 19290926]
157. Maiese K, Chong ZZ, Shang YC, Hou J. A “FOXO” in sight: targeting Foxo proteins from conception to cancer. *Med Res Rev* May;2009 29(3):395–418. [PubMed: 18985696]
158. Chong ZZ, Li F, Maiese K. The pro-survival pathways of mTOR and protein kinase B target glycogen synthase kinase-3beta and nuclear factor-kappaB to foster endogenous microglial cell protection. *Int J Mol Med* Feb;2007 19(2):263–272. [PubMed: 17203200]
159. Dello Russo C, Lisi L, Tringali G, Navarra P. Involvement of mTOR kinase in cytokine-dependent microglial activation and cell proliferation. *Biochem Pharmacol* Nov 1;2009 78(9):1242–1251. [PubMed: 19576187]
160. Lee SJ, Cho KS, Koh JY. Oxidative injury triggers autophagy in astrocytes: the role of endogenous zinc. *Glia* Sep;2009 57(12):1351–1361. [PubMed: 19229997]
161. Martin I, Andres CR, Vedrine S, et al. Effect of the oligodendrocyte myelin glycoprotein (OMgp) on the expansion and neuronal differentiation of rat neural stem cells. *Brain Res* Aug 11;2009 1284:22–30. [PubMed: 19501059]
162. Geijtenbeek TB, Gringhuis SI. Signalling through C-type lectin receptors: shaping immune responses. *Nat Rev Immunol* Jul;2009 9(7):465–479. [PubMed: 19521399]
163. Williams R, Dhillon NK, Hegde ST, et al. Proinflammatory cytokines and HIV-1 synergistically enhance CXCL10 expression in human astrocytes. *Glia* May;2009 57(7):734–743. [PubMed: 18985732]
164. Zhao L, Ma W, Fariss RN, Wong WT. Retinal vascular repair and neovascularization are not dependent on CX3CR1 signaling in a model of ischemic retinopathy. *Exp Eye Res* Jun;2009 88(6):1004–1013. [PubMed: 19176215]
165. Dringen R. Oxidative and antioxidative potential of brain microglial cells. *Antioxid Redox Signal* Sep-Oct;2005 7(9-10):1223–1233. [PubMed: 16115027]
166. Maiese K, Chong ZZ. Insights into oxidative stress and potential novel therapeutic targets for Alzheimer disease. *Restor Neurol Neurosci* 2004;22(2):87–104. [PubMed: 15272144]
167. Sankarapandi S, Zweier JL, Mukherjee G, Quinn MT, Huso DL. Measurement and characterization of superoxide generation in microglial cells: evidence for an NADPH oxidase-dependent pathway. *Arch Biochem Biophys* 1998;353(2):312–321. [PubMed: 9606965]
168. Denes A, Ferenczi S, Halasz J, Kornyei Z, Kovacs KJ. Role of CX3CR1 (fractalkine receptor) in brain damage and inflammation induced by focal cerebral ischemia in mouse. *J Cereb Blood Flow Metab* Oct;2008 28(10):1707–1721. [PubMed: 18575457]
169. Wu Y, Peng H, Cui M, Whitney NP, Huang Y, Zheng JC. CXCL12 increases human neural progenitor cell proliferation through Akt-1/FOXO3a signaling pathway. *J Neurochem* May;2009 109(4):1157–1167. [PubMed: 19302476]
170. Chong ZZ, Kang JQ, Maiese K. Metabotropic glutamate receptors promote neuronal and vascular plasticity through novel intracellular pathways. *Histol Histopathol* Jan;2003 18(1):173–189. [PubMed: 12507297]
171. Kang JQ, Chong ZZ, Maiese K. Akt1 protects against inflammatory microglial activation through maintenance of membrane asymmetry and modulation of cysteine protease activity. *J Neurosci Res* Oct 1;2003 74(1):37–51. [PubMed: 13130504]
172. Mallat M, Marin-Teva JL, Cheret C. Phagocytosis in the developing CNS: more than clearing the corpses. *Curr Opin Neurobiol* Feb;2005 15(1):101–107. [PubMed: 15721751]
173. Chong ZZ, Kang J, Li F, Maiese K. mGluRI Targets Microglial Activation and Selectively Prevents Neuronal Cell Engulfment Through Akt and Caspase Dependent Pathways. *Curr Neurovasc Res* Jul;2005 2(3):197–211. [PubMed: 16181114]
174. Li F, Chong ZZ, Maiese K. Microglial integrity is maintained by erythropoietin through integration of Akt and its substrates of glycogen synthase kinase-3beta, beta-catenin, and nuclear factor-kappaB. *Curr Neurovasc Res* Aug;2006 3(3):187–201. [PubMed: 16918383]

175. Li F, Chong ZZ, Maiese K. Winding through the WNT pathway during cellular development and demise. *Histol Histopathol* Jan;2006 21(1):103–124. [PubMed: 16267791]
176. He XL, Wang YH, Gao M, Li XX, Zhang TT, Du GH. Baicalein protects rat brain mitochondria against chronic cerebral hypoperfusion-induced oxidative damage. *Brain Res* Jan 16;2009 1249:212–221. [PubMed: 18977207]
177. Maiese K, Chong ZZ, Shang YC, Hou J. Rogue proliferation versus restorative protection: where do we draw the line for Wnt and forkhead signaling? *Expert Opin Ther Targets* Jul;2008 12(7): 905–916. [PubMed: 18554157]
178. Plecita-Hlavata L, Lessard M, Santorova J, Bewersdorf J, Jezek P. Mitochondrial oxidative phosphorylation and energetic status are reflected by morphology of mitochondrial network in INS-1E and HEP-G2 cells viewed by 4Pi microscopy. *Biochim Biophys Acta* Jul-Aug;2008 1777(7-8):834–846. [PubMed: 18452700]
179. Di Lisa F, Menabo R, Canton M, Barile M, Bernardi P. Opening of the mitochondrial permeability transition pore causes depletion of mitochondrial and cytosolic NAD⁺ and is a causative event in the death of myocytes in postischemic reperfusion of the heart. *J Biol Chem* Jan 26;2001 276(4):2571–2575. [PubMed: 11073947]
180. Chong ZZ, Kang JQ, Maiese K. Apaf-1, Bcl-xL, Cytochrome c, and Caspase-9 Form the Critical Elements for Cerebral Vascular Protection by Erythropoietin. *J Cereb Blood Flow Metab* Mar; 2003 23(3):320–330. [PubMed: 12621307]
181. Lin SH, Vincent A, Shaw T, Maynard KI, Maiese K. Prevention of nitric oxide-induced neuronal injury through the modulation of independent pathways of programmed cell death. *J Cereb Blood Flow Metab* 2000;20(9):1380–1391. [PubMed: 10994860]
182. Kelley DE, He J, Menshikova EV, Ritov VB. Dysfunction of mitochondria in human skeletal muscle in type 2 diabetes. *Diabetes* Oct;2002 51(10):2944–2950. [PubMed: 12351431]
183. Choo HJ, Kim JH, Kwon OB, et al. Mitochondria are impaired in the adipocytes of type 2 diabetic mice. *Diabetologia* Apr;2006 49(4):784–791. [PubMed: 16501941]
184. Speese SD, Budnik V. Wnts: up-and-coming at the synapse. *Trends Neurosci* Jun;2007 30(6): 268–275. [PubMed: 17467065]
185. Espada J, Calvo MB, Diaz-Prado S, Medina V. Wnt signalling and cancer stem cells. *Clin Transl Oncol* Jul;2009 11(7):411–427. [PubMed: 19574199]
186. Jozwiak J, Kotulska K, Grajkowska W, et al. Upregulation of the WNT pathway in tuberous sclerosis-associated subependymal giant cell astrocytomas. *Brain Dev* Jun;2007 29(5):273–280. [PubMed: 17071037]
187. Jozwiak J, Wlodarski P. Hamartin and tuberlin modulate gene transcription via beta-catenin. *J Neurooncol* Sep;2006 79(3):229–234. [PubMed: 16552619]
188. Kikuchi A, Yamamoto H, Sato A. Selective activation mechanisms of Wnt signaling pathways. *Trends Cell Biol* Mar;2009 19(3):119–129. [PubMed: 19208479]
189. Lee JH, Lee EO, Kang JL, Chong YH. Concomitant degradation of beta-catenin and GSK-3 beta potently contributes to glutamate-induced neurotoxicity in rat hippocampal slice cultures. *J Neurochem* Aug;2008 106(3):1066–1077. [PubMed: 18445133]
190. Luo JM, Dai CF, Lin SY, Huang PQ. Asymmetric syntheses and Wnt signal inhibitory activity of melleumin A and four analogues of melleumins A and B. *Chem Asian J* Feb 2;2009 4(2):328–335. [PubMed: 19072738]
191. Maiese K, Hou J, Chong ZZ, Shang YC. A fork in the path: Developing therapeutic inroads with FoxO proteins. *Oxid Med Cell Longev* 2009;2(3):119–126. [PubMed: 20592766]
192. Maiese K, Li F, Chong ZZ, Shang YC. The Wnt signaling pathway: Aging gracefully as a protectionist? *Pharmacol Ther* Apr;2008 118(1):58–81. [PubMed: 18313758]
193. Mercado-Gomez O, Hernandez-Fonseca K, Villavicencio-Queijeiro A, Massieu L, Chimal-Monroy J, Arias C. Inhibition of Wnt and PI3K signaling modulates GSK-3beta activity and induces morphological changes in cortical neurons: role of tau phosphorylation. *Neurochem Res* Aug;2008 33(8):1599–1609. [PubMed: 18461448]
194. Muruganandan S, Roman AA, Sinal CJ. Adipocyte differentiation of bone marrow-derived mesenchymal stem cells: cross talk with the osteoblastogenic program. *Cell Mol Life Sci* Jan; 2009 66(2):236–253. [PubMed: 18854943]

195. Sutton LP, Honardoust D, Mouyal J, Rajakumar N, Rushlow WJ. Activation of the canonical Wnt pathway by the antipsychotics haloperidol and clozapine involves dishevelled-3. *J Neurochem* Jul;2007 102(1):153–169. [PubMed: 17472703]
196. Vettor R, Milan G, Franzin C, et al. THE ORIGIN OF INTERMUSCULAR ADIPOSE TISSUE AND ITS PATHOPHYSIOLOGICAL IMPLICATIONS. *Am J Physiol Endocrinol Metab*. Sep 8;2009
197. Wang Z, Havasi A, Gall JM, Mao H, Schwartz JH, Borkan SC. Beta-catenin promotes survival of renal epithelial cells by inhibiting Bax. *J Am Soc Nephrol* Sep;2009 20(9):1919–1928. [PubMed: 19696224]
198. Wilusz M, Majka M. Role of the Wnt/beta-catenin network in regulating hematopoiesis. *Arch Immunol Ther Exp (Warsz)* Jul-Aug;2008 56(4):257–266. [PubMed: 18726147]
199. Grant SF, Thorleifsson G, Reynisdottir I, et al. Variant of transcription factor 7-like 2 (TCF7L2) gene confers risk of type 2 diabetes. *Nat Genet* Mar;2006 38(3):320–323. [PubMed: 16415884]
200. Lehman DM, Hunt KJ, Leach RJ, et al. Haplotypes of Transcription Factor 7-Like 2 (TCF7L2) Gene and Its Upstream Region Are Associated With Type 2 Diabetes and Age of Onset in Mexican Americans. *Diabetes* Feb;2007 56(2):389–393. [PubMed: 17259383]
201. Scott LJ, Bonnycastle LL, Willer CJ, et al. Association of transcription factor 7-like 2 (TCF7L2) variants with type 2 diabetes in a Finnish sample. *Diabetes* Sep;2006 55(9):2649–2653. [PubMed: 16936217]
202. Guo YF, Xiong DH, Shen H, et al. Polymorphisms of the low-density lipoprotein receptor-related protein 5 (LRP5) gene are associated with obesity phenotypes in a large family-based association study. *J Med Genet* Oct;2006 43(10):798–803. [PubMed: 16723389]
203. Kanazawa A, Tsukada S, Sekine A, et al. Association of the gene encoding wingless-type mammary tumor virus integration-site family member 5B (WNT5B) with type 2 diabetes. *Am J Hum Genet* Nov;2004 75(5):832–843. [PubMed: 15386214]
204. Mani A, Radhakrishnan J, Wang H, et al. LRP6 mutation in a family with early coronary disease and metabolic risk factors. *Science* Mar 2;2007 315(5816):1278–1282. [PubMed: 17332414]
205. Al-Aly Z, Shao JS, Lai CF, et al. Aortic Msx2-Wnt calcification cascade is regulated by TNF-alpha-dependent signals in diabetic Ldlr^{-/-} mice. *Arterioscler Thromb Vasc Biol* Dec;2007 27(12):2589–2596. [PubMed: 17932314]
206. Wright WS, Longo KA, Dolinsky VW, et al. Wnt10b Inhibits Obesity in ob/ob and Agouti Mice. *Diabetes* Feb;2007 56(2):295–303. [PubMed: 17259372]
207. Lin CL, Wang JY, Huang YT, Kuo YH, Surendran K, Wang FS. Wnt/beta-catenin signaling modulates survival of high glucose-stressed mesangial cells. *J Am Soc Nephrol* Oct;2006 17(10):2812–2820. [PubMed: 16943306]
208. Aslanidi G, Kroutov V, Philipsberg G, et al. Ectopic expression of Wnt10b decreases adiposity and improves glucose homeostasis in obese rats. *Am J Physiol Endocrinol Metab* Sep;2007 293(3):E726–736. [PubMed: 17578883]
209. Maiese K, Chong ZZ, Li F, Shang YC. Erythropoietin: Elucidating new cellular targets that broaden therapeutic strategies. *Prog Neurobiol* 2008;85:194–213. [PubMed: 18396368]
210. Maiese K, Chong ZZ, Shang YC. Raves and risks for erythropoietin. *Cytokine Growth Factor Rev* Apr;2008 19(2):145–155. [PubMed: 18299246]
211. Gayer CP, Chaturvedi LS, Wang S, Craig DH, Flanigan T, Basson MD. Strain-induced proliferation requires the phosphatidylinositol 3-kinase/AKT/glycogen synthase kinase pathway. *J Biol Chem* Jan 23;2009 284(4):2001–2011. [PubMed: 19047055]
212. An J, Zhang C, Polavarapu R, Zhang X, Yepes M. Tissue-type plasminogen activator and the low-density lipoprotein receptor-related protein induce Akt phosphorylation in the ischemic brain. *Blood* Oct 1;2008 112(7):2787–2794. [PubMed: 18628488]
213. Slaets H, Dumont D, Vanderlocht J, et al. Leukemia inhibitory factor induces an antiapoptotic response in oligodendrocytes through Akt-phosphorylation and up-regulation of 14-3-3. *Proteomics* Mar;2008 8(6):1237–1247. [PubMed: 18338825]
214. Dreixler JC, Hemmert JW, Shenoy SK, et al. The role of Akt/protein kinase B subtypes in retinal ischemic preconditioning. *Exp Eye Res* Mar;2009 88(3):512–521. [PubMed: 19084003]

215. Anitha M, Gondha C, Sutliff R, et al. GDNF rescues hyperglycemia-induced diabetic enteric neuropathy through activation of the PI3K/Akt pathway. *J Clin Invest* Feb;2006 116(2):344–356. [PubMed: 16453021]
216. Chong ZZ, Kang JQ, Maiese K. Erythropoietin is a novel vascular protectant through activation of Akt1 and mitochondrial modulation of cysteine proteases. *Circulation* Dec 3;2002 106(23):2973–2979. [PubMed: 12460881]
217. Burgos-Ramos E, Martos-Moreno GA, Lopez MG, et al. The N-terminal tripeptide of insulin-like growth factor-I protects against beta-amyloid-induced somatostatin depletion by calcium and glycogen synthase kinase 3 beta modulation. *J Neurochem* Apr;2009 109(2):360–370. [PubMed: 19220704]
218. Tamagno E, Guglielmotto M, Giliberto L, et al. JNK and ERK1/2 pathways have a dual opposite effect on the expression of BACE1. *Neurobiol Aging* Oct;2009 30(10):1563–1573. [PubMed: 18255190]
219. Kim KH, Oudit GY, Backx PH. Erythropoietin protects against doxorubicin-induced cardiomyopathy via a phosphatidylinositol 3-kinase-dependent pathway. *J Pharmacol Exp Ther* Jan;2008 324(1):160–169. [PubMed: 17928571]
220. Tajés M, Yeste-Velasco M, Zhu X, et al. Activation of Akt by lithium: Pro-survival pathways in aging. *Mech Ageing Dev* Jan 3;2009 130(4):253–261. [PubMed: 19162061]
221. Chong ZZ, Kang JQ, Maiese K. Akt1 drives endothelial cell membrane asymmetry and microglial activation through Bcl-x(L) and caspase 1, 3, and 9. *Exp Cell Res* Jun 10;2004 296(2):196–207. [PubMed: 15149850]
222. Chong ZZ, Maiese K. Erythropoietin involves the phosphatidylinositol 3-kinase pathway, 14-3-3 protein and FOXO3a nuclear trafficking to preserve endothelial cell integrity. *Br J Pharmacol* Apr;2007 150(7):839–850. [PubMed: 17339844]
223. Chong ZZ, Kang JQ, Maiese K. Erythropoietin fosters both intrinsic and extrinsic neuronal protection through modulation of microglia, Akt1, Bad, and caspase-mediated pathways. *Br J Pharmacol* Mar;2003 138(6):1107–1118. [PubMed: 12684267]
224. Fujiwara M, Izuishi K, Sano T, et al. Modulating effect of the PI3-kinase inhibitor LY294002 on cisplatin in human pancreatic cancer cells. *J Exp Clin Cancer Res* 2008;27:76. [PubMed: 19032736]
225. Chong ZZ, Li F, Maiese K. Activating Akt and the brain's resources to drive cellular survival and prevent inflammatory injury. *Histol Histopathol* Jan;2005 20(1):299–315. [PubMed: 15578447]
226. Chong ZZ, Maiese K. Targeting WNT, protein kinase B, and mitochondrial membrane integrity to foster cellular survival in the nervous system. *Histol Histopathol* Apr;2004 19(2):495–504. [PubMed: 15024710]
227. Fukumoto S, Hsieh CM, Maemura K, et al. Akt participation in the Wnt signaling pathway through Dishevelled. *J Biol Chem* 2001;276(20):17479–17483. [PubMed: 11278246]
228. Naito AT, Akazawa H, Takano H, et al. Phosphatidylinositol 3-kinase-Akt pathway plays a critical role in early cardiomyogenesis by regulating canonical Wnt signaling. *Circ Res* Jul 22;2005 97(2):144–151. [PubMed: 15994435]
229. Longo KA, Kennell JA, Ochocinska MJ, Ross SE, Wright WS, MacDougald OA. Wnt signaling protects 3T3-L1 preadipocytes from apoptosis through induction of insulin-like growth factors. *J Biol Chem* 2002;277(41):38239–38244. [PubMed: 12154096]
230. Su F, Overholtzer M, Besser D, Levine AJ. WISP-1 attenuates p53-mediated apoptosis in response to DNA damage through activation of the Akt kinase. *Genes Dev* 2002;16(1):46–57. [PubMed: 11782444]
231. Mirotsov M, Zhang Z, Deb A, et al. Secreted frizzled related protein 2 (Sfrp2) is the key Akt-mesenchymal stem cell-released paracrine factor mediating myocardial survival and repair. *Proc Natl Acad Sci U S A* Jan 30;2007 104(5):1643–1648. [PubMed: 17251350]
232. van de Schans VA, van den Borne SW, Strzelecka AE, et al. Interruption of Wnt signaling attenuates the onset of pressure overload-induced cardiac hypertrophy. *Hypertension* Mar;2007 49(3):473–480. [PubMed: 17210832]

233. Barandon L, Dufourcq P, Costet P, et al. Involvement of FrzA/sFRP-1 and the Wnt/frizzled pathway in ischemic preconditioning. *Circ Res* Jun 24;2005 96(12):1299–1306. [PubMed: 15920021]
234. DiPalma JR, Thayer WS. Use of niacin as a drug. *Annu Rev Nutr* 1991;11:169–187. [PubMed: 1832551]
235. Rex A, Fink H. Pharmacokinetic aspects of reduced nicotinamide adenine dinucleotide (NADH) in rats. *Front Biosci* 2008;13:3735–3741. [PubMed: 18508468]
236. Li F, Chong ZZ, Maiese K. Navigating novel mechanisms of cellular plasticity with the NAD⁺ precursor and nutrient nicotinamide. *Front Biosci* 2004;9:2500–2520. [PubMed: 15353303]
237. Jackson TM, Rawling JM, Roebuck BD, Kirkland JB. Large supplements of nicotinic acid and nicotinamide increase tissue NAD⁺ and poly(ADP-ribose) levels but do not affect diethylnitrosamine-induced altered hepatic foci in Fischer-344 rats. *J Nutr* Jun;1995 125(6):1455–1461. [PubMed: 7782898]
238. Wojcik M, Seidle HF, Bieganowski P, Brenner C. Glutamine-dependent NAD⁺ synthetase. How a two-domain, three-substrate enzyme avoids waste. *J Biol Chem* Nov 3;2006 281(44):33395–33402. [PubMed: 16954203]
239. Khan JA, Forouhar F, Tao X, Tong L. Nicotinamide adenine dinucleotide metabolism as an attractive target for drug discovery. *Expert Opin Ther Targets* May;2007 11(5):695–705. [PubMed: 17465726]
240. Khan JA, Xiang S, Tong L. Crystal structure of human nicotinamide riboside kinase. *Structure* Aug;2007 15(8):1005–1013. [PubMed: 17698003]
241. Depeint F, Bruce WR, Shangari N, Mehta R, O'Brien PJ. Mitochondrial function and toxicity: role of the B vitamin family on mitochondrial energy metabolism. *Chem Biol Interact* Oct 27;2006 163(1-2):94–112. [PubMed: 16765926]
242. Hara N, Yamada K, Shibata T, Osago H, Hashimoto T, Tsuchiya M. Elevation of cellular NAD levels by nicotinic acid and involvement of nicotinic acid phosphoribosyltransferase in human cells. *J Biol Chem* Aug 24;2007 282(34):24574–24582. [PubMed: 17604275]
243. Williams AC, Ramsden DB. Pellagra: A clue as to why energy failure causes diseases? *Med Hypotheses* 2007;69(3):618–628. [PubMed: 17349750]
244. Williams AC, Ramsden DB. Hydrogen symbioses in evolution and disease. *Qjm* Jul;2007 100(7):451–459. [PubMed: 17566009]
245. Magni G, Amici A, Emanuelli M, Orsomando G, Raffaelli N, Ruggieri S. Enzymology of NAD⁺ homeostasis in man. *Cell Mol Life Sci* Jan;2004 61(1):19–34. [PubMed: 14704851]
246. Lin SJ, Guarente L. Nicotinamide adenine dinucleotide, a metabolic regulator of transcription, longevity and disease. *Curr Opin Cell Biol* Apr;2003 15(2):241–246. [PubMed: 12648681]
247. Hageman GJ, Stierum RH. Niacin, poly(ADP-ribose) polymerase-1 and genomic stability. *Mutat Res* Apr 18;2001 475(1-2):45–56. [PubMed: 11295153]
248. Sadanaga-Akiyoshi F, Yao H, Tanuma S, et al. Nicotinamide attenuates focal ischemic brain injury in rats: with special reference to changes in nicotinamide and NAD⁺ levels in ischemic core and penumbra. *Neurochem Res* Aug;2003 28(8):1227–1234. [PubMed: 12834263]
249. Wang J, Zhai Q, Chen Y, et al. A local mechanism mediates NAD-dependent protection of axon degeneration. *J Cell Biol* Aug 1;2005 170(3):349–355. [PubMed: 16043516]
250. Chong ZZ, Lin SH, Maiese K. Nicotinamide Modulates Mitochondrial Membrane Potential and Cysteine Protease Activity during Cerebral Vascular Endothelial Cell Injury. *J Vasc Res* 2002;39(2):131–147. [PubMed: 12011585]
251. Maiese K, Lin S, Chong ZZ. Elucidating neuronal and vascular injury through the cytoprotective agent nicotinamide. *Curr Med Chem-Imm, Endoc. & Metab. Agents* 2001;1(3):257–267.
252. Chong ZZ, Lin SH, Li F, Maiese K. The sirtuin inhibitor nicotinamide enhances neuronal cell survival during acute anoxic injury through Akt, Bad, PARP, and mitochondrial associated “anti-apoptotic” pathways. *Curr Neurovasc Res* Oct;2005 2(4):271–285. [PubMed: 16181120]
253. Chong ZZ, Lin SH, Maiese K. The NAD⁺ precursor nicotinamide governs neuronal survival during oxidative stress through protein kinase B coupled to FOXO3a and mitochondrial membrane potential. *J Cereb Blood Flow Metab* Jul;2004 24(7):728–743. [PubMed: 15241181]

254. Halestrap AP, Woodfield KY, Connern CP. Oxidative stress, thiol reagents, and membrane potential modulate the mitochondrial permeability transition by affecting nucleotide binding to the adenine nucleotide translocase. *J Biol Chem* Feb 7;1997 272(6):3346–3354. [PubMed: 9013575]
255. La Piana G, Marzulli D, Consalvo MI, Lofrumento NE. Cytochrome c-induced cytosolic nicotinamide adenine dinucleotide oxidation, mitochondrial permeability transition, and apoptosis. *Arch Biochem Biophys* Feb 15;2003 410(2):201–211. [PubMed: 12573279]
256. Idelson M, Alper R, Obolensky A, et al. Directed differentiation of human embryonic stem cells into functional retinal pigment epithelium cells. *Cell Stem Cell* Oct 2;2009 5(4):396–408. [PubMed: 19796620]
257. Aoyagi S, Archer TK. Nicotinamide uncouples hormone-dependent chromatin remodeling from transcription complex assembly. *Mol Cell Biol* Jan;2008 28(1):30–39. [PubMed: 17954562]
258. Bruno V, Battaglia G, Copani A, et al. Activation of class II or III metabotropic glutamate receptors protects cultured cortical neurons against excitotoxic degeneration. *Eur J Neurosci* 1995;7(9):1906–1913. [PubMed: 8528465]
259. Anderson DW, Bradbury KA, Schneider JS. Neuroprotection in Parkinson models varies with toxin administration protocol. *Eur J Neurosci* Dec;2006 24(11):3174–3182. [PubMed: 17156378]
260. Feng Y, Paul IA, LeBlanc MH. Nicotinamide reduces hypoxic ischemic brain injury in the newborn rat. *Brain Res Bull* Mar 31;2006 69(2):117–122. [PubMed: 16533659]
261. Lin SH, Chong ZZ, Maiese K. Nicotinamide: A Nutritional Supplement that Provides Protection Against Neuronal and Vascular Injury. *J Med Food* 2001;4(1):27–38. Spring. [PubMed: 12639285]
262. Slomka M, Zieminska E, Salinska E, Lazarewicz JW. Neuroprotective effects of nicotinamide and 1-methylnicotinamide in acute excitotoxicity in vitro. *Folia Neuropathol* 2008;46(1):69–80. [PubMed: 18368629]
263. Slomka M, Zieminska E, Lazarewicz J. Nicotinamide and 1-methylnicotinamide reduce homocysteine neurotoxicity in primary cultures of rat cerebellar granule cells. *Acta Neurobiol Exp (Wars)* 2008;68(1):1–9. [PubMed: 18389009]
264. Ieraci A, Herrera DG. Nicotinamide Protects against Ethanol-Induced Apoptotic Neurodegeneration in the Developing Mouse Brain. *PLoS Med* Feb 21;2006 3(4):e101. [PubMed: 16478293]
265. Shen CC, Huang HM, Ou HC, Chen HL, Chen WC, Jeng KC. Protective effect of nicotinamide on neuronal cells under oxygen and glucose deprivation and hypoxia/reoxygenation. *J Biomed Sci* Jul-Aug;2004 11(4):472–481. [PubMed: 15153782]
266. Sonee M, Martens JR, Evers MR, Mukherjee SK. The effect of tertiary butylhydroperoxide and nicotinamide on human cortical neurons. *Neurotoxicology* Jun;2003 24(3):443–448. [PubMed: 12782109]
267. Kiuchi K, Kondo M, Ueno S, et al. Functional rescue of N-methyl-N-nitrosourea-induced retinopathy by nicotinamide in Sprague-Dawley rats. *Curr Eye Res* Jun;2003 26(6):355–362. [PubMed: 12868016]
268. Kiuchi K, Yoshizawa K, Shikata N, Matsumura M, Tsubura A. Nicotinamide prevents N-methyl-N-nitrosourea-induced photoreceptor cell apoptosis in Sprague-Dawley rats and C57BL mice. *Exp Eye Res* Mar;2002 74(3):383–392. [PubMed: 12014919]
269. Reber F, Geffarth R, Kasper M, et al. Graded sensitiveness of the various retinal neuron populations on the glyoxal-mediated formation of advanced glycation end products and ways of protection. *Graefes Arch Clin Exp Ophthalmol* Mar;2003 241(3):213–225. [PubMed: 12644946]
270. Hoane MR, Gilbert DR, Holland MA, Pierce JL. Nicotinamide reduces acute cortical neuronal death and edema in the traumatically injured brain. *Neurosci Lett* Nov 6;2006 408(1):35–39. [PubMed: 16987607]
271. Hoane MR, Kaplan SA, Ellis AL. The effects of nicotinamide on apoptosis and blood-brain barrier breakdown following traumatic brain injury. *Brain Res* Dec 13;2006 1125(1):185–193. [PubMed: 17109832]

272. Hoane MR, Pierce JL, Holland MA, Anderson GD. Nicotinamide treatment induces behavioral recovery when administered up to 4 hours following cortical contusion injury in the rat. *Neuroscience* Jun 26;2008 154(3):861–868. [PubMed: 18514428]
273. Hoane MR, Pierce JL, Kaufman NA, Beare JE. Variation in chronic nicotinamide treatment after traumatic brain injury can alter components of functional recovery independent of histological damage. *Oxid Med Cell Longev* 2008;1(1):45–52.
274. Holland MA, Tan AA, Smith DC, Hoane MR. Nicotinamide treatment provides acute neuroprotection and GFAP regulation following fluid percussion injury. *J Neurotrauma* Feb;2008 25(2):140–152. [PubMed: 18260797]
275. Wallis RA, Panizzon KL, Girard JM. Traumatic neuroprotection with inhibitors of nitric oxide and ADP- ribosylation. *Brain Res* 1996;710(1-2):169–177. [PubMed: 8963656]
276. Yang J, Klaidman L, Chang M, et al. Nicotinamide therapy protects against both necrosis and apoptosis in a stroke model. *Pharmacol Biochem Behav* 2002;73(4):901–910. [PubMed: 12213537]
277. Gupta S, Kaul CL, Sharma SS. Neuroprotective effect of combination of poly (ADP-ribose) polymerase inhibitor and antioxidant in middle cerebral artery occlusion induced focal ischemia in rats. *Neurol Res* Jan;2004 26(1):103–107. [PubMed: 14977067]
278. Sakakibara Y, Mitha AP, Ayoub IA, Ogilvy CS, Maynard KI. Delayed treatment with nicotinamide (vitamin B3) reduces the infarct volume following focal cerebral ischemia in spontaneously hypertensive rats, diabetic and non-diabetic Fischer 344 rats. *Brain Res Mar 22;2002* 931(1):68–73. [PubMed: 11897090]
279. Macleod MR, O'Collins T, Howells DW, Donnan GA. Pooling of animal experimental data reveals influence of study design and publication bias. *Stroke* May;2004 35(5):1203–1208. [PubMed: 15060322]
280. Brewer KL, Hardin JS. Neuroprotective effects of nicotinamide after experimental spinal cord injury. *Acad Emerg Med* Feb;2004 11(2):125–130. [PubMed: 14759952]
281. Isbir CS, Ak K, Kurtkaya O, et al. Ischemic preconditioning and nicotinamide in spinal cord protection in an experimental model of transient aortic occlusion. *Eur J Cardiothorac Surg* Jun; 2003 23(6):1028–1033. [PubMed: 12829083]
282. Williams A, Ramsden D. Nicotinamide: a double edged sword. *Parkinsonism Relat Disord* Nov; 2005 11(7):413–420. [PubMed: 16183323]
283. Williams AC, Cartwright LS, Ramsden DB. Parkinson's disease: the first common neurological disease due to auto-intoxication? *Qjm* Mar;2005 98(3):215–226. [PubMed: 15728403]
284. Beck H, Plate KH. Angiogenesis after cerebral ischemia. *Acta Neuropathol* May;2009 117(5): 481–496. [PubMed: 19142647]
285. Giulumian AD, Meszaros LG, Fuchs LC. Endothelin-1-induced contraction of mesenteric small arteries is mediated by ryanodine receptor Ca²⁺ channels and cyclic ADP-ribose. *J Cardiovasc Pharmacol* Dec;2000 36(6):758–763. [PubMed: 11117376]
286. Pietrzak L, Mogielnicki A, Buczko W. Nicotinamide and its metabolite N-methylnicotinamide increase skin vascular permeability in rats. *Clin Exp Dermatol* Apr;2009 34(3):380–384. [PubMed: 19175785]
287. Oumouna-Benachour K, Hans CP, Suzuki Y, et al. Poly(ADP-ribose) polymerase inhibition reduces atherosclerotic plaque size and promotes factors of plaque stability in apolipoprotein E-deficient mice: effects on macrophage recruitment, nuclear factor-kappaB nuclear translocation, and foam cell death. *Circulation* May 8;2007 115(18):2442–2450. [PubMed: 17438151]
288. Giammona LM, Fuhrken PG, Papoutsakis ET, Miller WM. Nicotinamide (vitamin B3) increases the polyploidisation and proplatelet formation of cultured primary human megakaryocytes. *Br J Haematol* Nov;2006 135(4):554–566. [PubMed: 17054670]
289. Slominska EM, Yuen A, Osman L, Gebicki J, Yacoub MH, Smolenski RT. Cytoprotective effects of nicotinamide derivatives in endothelial cells. *Nucleosides Nucleotides Nucleic Acids* Jun;2008 27(6):863–866. [PubMed: 18600553]
290. Mateuszuk L, Khomich TI, Slominska E, et al. Activation of nicotinamide N-methyltransferase and increased formation of 1-methylnicotinamide (MNA) in atherosclerosis. *Pharmacol Rep* Jan-Feb; 2009 61(1):76–85. [PubMed: 19307695]

291. Vincent AM, Maiese K. Nitric oxide induction of neuronal endonuclease activity in programmed cell death. *Exp Cell Res* 1999;246(2):290–300. [PubMed: 9925743]
292. Vincent AM, TenBroeke M, Maiese K. Metabotropic glutamate receptors prevent programmed cell death through the modulation of neuronal endonuclease activity and intracellular pH. *Exp Neurol* 1999;155(1):79–94. [PubMed: 9918707]
293. Vincent AM, TenBroeke M, Maiese K. Neuronal intracellular pH directly mediates nitric oxide-induced programmed cell death. *J Neurobiol* 1999;40(2):171–184. [PubMed: 10413448]
294. Young GS, Jacobson EL, Kirkland JB. Water maze performance in young male Long-Evans rats is inversely affected by dietary intakes of niacin and may be linked to levels of the NAD⁺ metabolite cADPR. *J Nutr* Apr;2007 137(4):1050–1057. [PubMed: 17374675]
295. Reddy S, Bibby NJ, Wu D, Swinney C, Barrow G, Elliott RB. A combined casein-free-nicotinamide diet prevents diabetes in the NOD mouse with minimum insulinitis. *Diabetes Res Clin Pract* Aug;1995 29(2):83–92. [PubMed: 8591703]
296. Hu Y, Wang Y, Wang L, et al. Effects of nicotinamide on prevention and treatment of streptozotocin-induced diabetes mellitus in rats. *Chin Med J (Engl)* Nov;1996 109(11):819–822. [PubMed: 9275363]
297. Stevens MJ, Li F, Drel VR, et al. Nicotinamide reverses neurological and neurovascular deficits in streptozotocin diabetic rats. *J Pharmacol Exp Ther* Jan;2007 320(1):458–464. [PubMed: 17021258]
298. Cresto JC, Fabiano de Bruno LE, Cao GF, et al. The association of acetyl-L-carnitine and nicotinamide remits the experimental diabetes in mice by multiple low-dose streptozotocin. *Pancreas* Nov;2006 33(4):403–411. [PubMed: 17079947]
299. Chlopicki S, Swies J, Mogielnicki A, et al. 1-Methylnicotinamide (MNA), a primary metabolite of nicotinamide, exerts anti-thrombotic activity mediated by a cyclooxygenase-2/prostacyclin pathway. *Br J Pharmacol Sep;2007 152(2):230–239*. 300. [PubMed: 17641676]
300. Lee HI, Cho HJ, Han JA, et al. Transient downregulation of protein O-N-acetylglucosaminylation by treatment of high-dose nicotinamide in human cells. *Exp Mol Med* Apr 30;2008 40(2):246–253. [PubMed: 18446063]
301. Tam D, Tam M, Maynard KI. Nicotinamide modulates energy utilization and improves functional recovery from ischemia in the in vitro rabbit retina. *Ann N Y Acad Sci* Aug;2005 1053:258–268. [PubMed: 16179531]
302. Olmos PR, Hodgson MI, Maiz A, et al. Nicotinamide protected first-phase insulin response (FPIR) and prevented clinical disease in first-degree relatives of type-1 diabetics. *Diabetes Res Clin Pract* Mar;2006 71(3):320–333. [PubMed: 16233932]
303. Crino A, Schiaffini R, Ciampalini P, et al. A two year observational study of nicotinamide and intensive insulin therapy in patients with recent onset type 1 diabetes mellitus. *J Pediatr Endocrinol Metab* Aug;2005 18(8):749–754. [PubMed: 16200840]
304. Eto N, Miyata Y, Ohno H, Yamashita T. Nicotinamide prevents the development of hyperphosphataemia by suppressing intestinal sodium-dependent phosphate transporter in rats with adenine-induced renal failure. *Nephrol Dial Transplant* Jul;2005 20(7):1378–1384. [PubMed: 15870221]
305. Liu HK, Green BD, Flatt PR, McClenaghan NH, McCluskey JT. Effects of long-term exposure to nicotinamide and sodium butyrate on growth, viability, and the function of clonal insulin secreting cells. *Endocr Res* Feb;2004 30(1):61–68. [PubMed: 15098920]
306. Reddy S, Salari-Lak N, Sandler S. Long-term effects of nicotinamide-induced inhibition of poly(adenosine diphosphate-ribose) polymerase activity in rat pancreatic islets exposed to interleukin-1 beta. *Endocrinology* 1995;136(5):1907–1912. [PubMed: 7720637]
307. Zhou SS, Li D, Sun WP, et al. Nicotinamide overload may play a role in the development of type 2 diabetes. *World J Gastroenterol* Dec 7;2009 15(45):5674–5684. [PubMed: 19960564]
308. Gaudineau C, Auclair K. Inhibition of human P450 enzymes by nicotinic acid and nicotinamide. *Biochem Biophys Res Commun* May 7;2004 317(3):950–956. [PubMed: 15081432]
309. Maiese K, Chong ZZ, Shang YC. “Sly as a FOXO”: New paths with Forkhead signaling in the brain. *Curr Neurovasc Res* Nov;2007 4(4):295–302. [PubMed: 18045156]

310. Maiese K, Chong ZZ, Shang YC. OutFOXing disease and disability: the therapeutic potential of targeting FoxO proteins. *Trends Mol Med* Apr 8;2008 14(5):219–227. [PubMed: 18403263]
311. Coffey PJ. When less is more: the PI3K pathway as a determinant of tumor response to dietary restriction. *Cell Res* Jul;2009 19(7):797–799. [PubMed: 19581877]
312. Jacobsen EA, Ananieva O, Brown ML, Chang Y. Growth, differentiation, and malignant transformation of pre-B cells mediated by inducible activation of v-Abl oncogene. *J Immunol* Jun 1;2006 176(11):6831–6838. [PubMed: 16709843]
313. Maiese K, Chong ZZ, Shang YC, Hou J. Clever cancer strategies with FoxO transcription factors. *Cell Cycle* Dec 15;2008 7(24):3829–3839. [PubMed: 19066462]
314. Maiese K, Chong ZZ, Shang YC, Hou J. FoxO proteins: cunning concepts and considerations for the cardiovascular system. *Clin Sci (Lond)* Feb;2009 116(3):191–203. [PubMed: 19118491]
315. Clark KL, Halay ED, Lai E, Burley SK. Co-crystal structure of the HNF-3/fork head DNA-recognition motif resembles histone H5. *Nature* Jul 29;1993 364(6436):412–420. [PubMed: 8332212]
316. Jin C, Marsden I, Chen X, Liao X. Sequence specific collective motions in a winged helix DNA binding domain detected by 15N relaxation NMR. *Biochemistry* Apr 28;1998 37(17):6179–6187. [PubMed: 9558357]
317. Maiese K, Chong Z, Hou J, Shang Y. The “O” Class: Crafting clinical care with FoxO transcription factors. *Adv Exp Med Biol* 2009;665:242–260. [PubMed: 20429429]
318. Kaestner KH, Knochel W, Martinez DE. Unified nomenclature for the winged helix/forkhead transcription factors. *Genes Dev* Jan 15;2000 14(2):142–146. [PubMed: 10702024]
319. Lappas M, Lim R, Riley C, Rice GE, Permezel M. Localisation and expression of FoxO1 proteins in human gestational tissues. *Placenta* Mar;2009 30(3):256–262. [PubMed: 19150739]
320. Zheng WH, Kar S, Quirion R. FKHRL1 and its homologs are new targets of nerve growth factor Trk receptor signaling. *J Neurochem* Mar;2002 80(6):1049–1061. [PubMed: 11953455]
321. Fei M, Lu M, Wang Y, et al. Arsenic trioxide-induced growth arrest of human hepatocellular carcinoma cells involving FOXO3a expression and localization. *Med Oncol* 2009;26(2):178–185. [PubMed: 18937079]
322. Lei H, Quelle FW. FOXO transcription factors enforce cell cycle checkpoints and promote survival of hematopoietic cells after DNA damage. *Mol Cancer Res* Aug;2009 7(8):1294–1303. [PubMed: 19671690]
323. Kikuchi S, Nagai T, Kunitama M, Kirito K, Ozawa K, Komatsu N. Active FKHRL1 overcomes imatinib resistance in chronic myelogenous leukemia-derived cell lines via the production of tumor necrosis factor-related apoptosis-inducing ligand. *Cancer Sci* Dec;2007 98(12):1949–1958. [PubMed: 17900262]
324. Nowak K, Killmer K, Gessner C, Lutz W. E2F-1 regulates expression of FOXO1 and FOXO3a. *Biochim Biophys Acta* Apr;2007 1769(4):244–252. [PubMed: 17482685]
325. Bouchard C, Lee S, Paulus-Hock V, Loddenkemper C, Eilers M, Schmitt CA. FoxO transcription factors suppress Myc-driven lymphomagenesis via direct activation of Arf. *Genes Dev* Nov 1;2007 21(21):2775–2787. [PubMed: 17974917]
326. Bethea CL, Reddy AP, Tokuyama Y, Henderson JA, Lima FB. Protective actions of ovarian hormones in the serotonin system of macaques. *Front Neuroendocrinol* Jul;2009 30(2):212–238. [PubMed: 19394356]
327. Chong ZZ, Kang JQ, Maiese K. Essential cellular regulatory elements of oxidative stress in early and late phases of apoptosis in the central nervous system. *Antioxid Redox Signal* Apr;2004 6(2):277–287. [PubMed: 15025929]
328. Fallarino F, Bianchi R, Orabona C, et al. CTLA-4-Ig activates forkhead transcription factors and protects dendritic cells from oxidative stress in nonobese diabetic mice. *J Exp Med* Oct 18;2004 200(8):1051–1062. [PubMed: 15492127]
329. Nakae J, Cao Y, Oki M, et al. Forkhead transcription factor FoxO1 in adipose tissue regulates energy storage and expenditure. *Diabetes* Mar;2008 57(3):563–576. [PubMed: 18162510]
330. Furuyama T, Yamashita H, Kitayama K, Higami Y, Shimokawa I, Mori N. Effects of aging and caloric restriction on the gene expression of Foxo1, 3, and 4 (FKHR, FKHRL1, and AFX) in the rat skeletal muscles. *Microsc Res Tech* Nov 15;2002 59(4):331–334. [PubMed: 12424797]

331. Puig O, Tjian R. Transcriptional feedback control of insulin receptor by dFOXO/FOXO1. *Genes Dev* Oct 15;2005 19(20):2435–2446. [PubMed: 16230533]
332. Kim JR, Jung HS, Bae SW, et al. Polymorphisms in FOXO gene family and association analysis with BMI. *Obesity (Silver Spring)* Feb;2006 14(2):188–193. [PubMed: 16571842]
333. Marchetti V, Menghini R, Rizza S, et al. Benfotiamine counteracts glucose toxicity effects on endothelial progenitor cell differentiation via Akt/FoxO signaling. *Diabetes* Aug;2006 55(8): 2231–2237. [PubMed: 16873685]
334. Kamagate A, Dong HH. Foxo1 integrates insulin signaling to VLDL production. *Cell Cycle* 2008;7(20):3162–3170. [PubMed: 18927507]
335. Ni YG, Wang N, Cao DJ, et al. FoxO transcription factors activate Akt and attenuate insulin signaling in heart by inhibiting protein phosphatases. *Proc Natl Acad Sci U S A* Dec 18;2007 104(51):20517–20522. [PubMed: 18077353]
336. Kamei Y, Miura S, Suzuki M, et al. Skeletal muscle FOXO1 (FKHR) transgenic mice have less skeletal muscle mass, down-regulated Type I (slow twitch/red muscle) fiber genes, and impaired glycemic control. *J Biol Chem* Sep 24;2004 279(39):41114–41123. [PubMed: 15272020]
337. Liu CM, Yang Z, Liu CW, et al. Effect of RNA oligonucleotide targeting Foxo-1 on muscle growth in normal and cancer cachexia mice. *Cancer Gene Ther* Dec;2007 14(12):945–952. [PubMed: 17885675]
338. Sandri M, Lin J, Handschin C, et al. PGC-1 α protects skeletal muscle from atrophy by suppressing FoxO3 action and atrophy-specific gene transcription. *Proc Natl Acad Sci U S A* Oct 31;2006 103(44):16260–16265. [PubMed: 17053067]
339. Maiese K, Li F, Chong ZZ. Erythropoietin in the brain: can the promise to protect be fulfilled? *Trends Pharmacol Sci* 2004;25(11):577–583. [PubMed: 15491780]
340. Charvet C, Alberti I, Luciano F, et al. Proteolytic regulation of Forkhead transcription factor FOXO3a by caspase-3-like proteases. *Oncogene* Jul 17;2003 22(29):4557–4568. [PubMed: 12881712]
341. Porcu M, Chiarugi A. The emerging therapeutic potential of sirtuin-interacting drugs: from cell death to lifespan extension. *Trends Pharmacol Sci* Feb;2005 26(2):94–103. [PubMed: 15681027]
342. Saunders LR, Verdin E. Sirtuins: critical regulators at the crossroads between cancer and aging. *Oncogene* Aug 13;2007 26(37):5489–5504. [PubMed: 17694089]
343. Taylor DM, Maxwell MM, Luthi-Carter R, Kazantsev AG. Biological and potential therapeutic roles of sirtuin deacetylases. *Cell Mol Life Sci* Dec;2008 65(24):4000–4018. [PubMed: 18820996]
344. Zschoernig B, Mahlknecht U. SIRTUIN 1: regulating the regulator. *Biochem Biophys Res Commun* Nov 14;2008 376(2):251–255. [PubMed: 18774777]
345. Tang BL, Chua CE. SIRT1 and neuronal diseases. *Mol Aspects Med* Jun;2008 29(3):187–200. [PubMed: 17397914]
346. Zwaal RF, Schroit AJ. Pathophysiologic implications of membrane phospholipid asymmetry in blood cells. *Blood* 1997;89(4):1121–1132. [PubMed: 9028933]
347. Canto C, Auwerx J. Caloric restriction, SIRT1 and longevity. *Trends Endocrinol Metab* Sep;2009 20(7):325–331. [PubMed: 19713122]
348. Balan V, Miller GS, Kaplun L, et al. Life span extension and neuronal cell protection by *Drosophila* nicotinamidase. *J Biol Chem* Oct 10;2008 283(41):27810–27819. [PubMed: 18678867]
349. Bureau G, Longpre F, Martinoli MG. Resveratrol and quercetin, two natural polyphenols, reduce apoptotic neuronal cell death induced by neuroinflammation. *J Neurosci Res* Feb 1;2008 86(2): 403–410. [PubMed: 17929310]
350. Chong ZZ, Maiese K. Enhanced Tolerance against Early and Late Apoptotic Oxidative Stress in Mammalian Neurons through Nicotinamidase and Sirtuin Mediated Pathways. *Curr Neurovasc Res* Aug;2008 5(3):159–170. [PubMed: 18691073]
351. Nemoto S, Fergusson MM, Finkel T. Nutrient availability regulates SIRT1 through a forkhead-dependent pathway. *Science* Dec 17;2004 306(5704):2105–2108. [PubMed: 15604409]
352. Ferrara N, Rinaldi B, Corbi G, et al. Exercise Training Promotes SIRT1 Activity in Aged Rats. *Rejuvenation Res* Dec 10;2008 11(1):139–150. [PubMed: 18069916]

353. Motta MC, Divecha N, Lemieux M, et al. Mammalian SIRT1 represses forkhead transcription factors. *Cell* Feb 20;2004 116(4):551–563. [PubMed: 14980222]
354. Lee HI, Jang SY, Kang HT, Hwang ES. p53-, SIRT1-, and PARP-1-independent downregulation of p21WAF1 expression in nicotinamide-treated cells. *Biochem Biophys Res Commun* Apr 4;2008 368(2):298–304. [PubMed: 18230337]
355. Jackson MD, Schmidt MT, Oppenheimer NJ, Denu JM. Mechanism of nicotinamide inhibition and transglycosidation by Sir2 histone/protein deacetylases. *J Biol Chem* Dec 19;2003 278(51):50985–50998. [PubMed: 14522996]
356. Bitterman KJ, Anderson RM, Cohen HY, Latorre-Esteves M, Sinclair DA. Inhibition of silencing and accelerated aging by nicotinamide, a putative negative regulator of yeast sir2 and human SIRT1. *J Biol Chem* Nov 22;2002 277(47):45099–45107. [PubMed: 12297502]
357. Cai AL, Zipfel GJ, Sheline CT. Zinc neurotoxicity is dependent on intracellular NAD levels and the sirtuin pathway. *Eur J Neurosci* Oct;2006 24(8):2169–2176. [PubMed: 17042794]
358. Kruszewski M, Szumiel I. Sirtuins (histone deacetylases III) in the cellular response to DNA damage--facts and hypotheses. *DNA Repair (Amst)* Nov 21;2005 4(11):1306–1313. [PubMed: 16084131]
359. Yoshizaki T, Milne JC, Imamura T, et al. SIRT1 exerts anti-inflammatory effects and improves insulin sensitivity in adipocytes. *Mol Cell Biol* Mar;2009 29(5):1363–1374. [PubMed: 19103747]
360. Zillikens MC, van Meurs JB, Rivadeneira F, et al. SIRT1 genetic variation is related to BMI and risk of obesity. *Diabetes* Dec;2009 58(12):2828–2834. [PubMed: 19741164]
361. Maiese K, Li F, Chong ZZ. New avenues of exploration for erythropoietin. *Jama* Jan 5;2005 293(1):90–95. [PubMed: 15632341]
362. Girgenti MJ, Hunsberger J, Duman CH, Sathyanesan M, Terwilliger R, Newton SS. Erythropoietin induction by electroconvulsive seizure, gene regulation, and antidepressant-like behavioral effects. *Biol Psychiatry* Aug 1;2009 66(3):267–274. [PubMed: 19185286]
363. Ma R, Xiong N, Huang C, et al. Erythropoietin protects PC12 cells from beta-amyloid(25-35)-induced apoptosis via PI3K/Akt signaling pathway. *Neuropharmacology* May-Jun;2009 56(6-7):1027–1034. [PubMed: 19268480]
364. Sun ZK, Yang HQ, Pan J, et al. Protective effects of erythropoietin on tau phosphorylation induced by beta-amyloid. *J Neurosci Res* Oct;2008 86(13):3018–3027. [PubMed: 18512763]
365. McLeod M, Hong M, Mukhida K, Sadi D, Ulalia R, Mendez I. Erythropoietin and GDNF enhance ventral mesencephalic fiber outgrowth and capillary proliferation following neural transplantation in a rodent model of Parkinson's disease. *Eur J Neurosci* Jul;2006 24(2):361–370. [PubMed: 16903847]
366. Cuzzocrea S, Mazzon E, di Paola R, et al. Erythropoietin reduces the degree of arthritis caused by type II collagen in the mouse. *Arthritis Rheum* Mar;2005 52(3):940–950. [PubMed: 15751086]
367. Thorne M, Moore CS, Robertson GS. Lack of TIMP-1 increases severity of experimental autoimmune encephalomyelitis: Effects of darbepoetin alfa on TIMP-1 null and wild-type mice. *J Neuroimmunol* Jun 25;2009 211(1-2):92–100. [PubMed: 19428125]
368. Sanchez PE, Fares RP, Risso JJ, et al. Optimal neuroprotection by erythropoietin requires elevated expression of its receptor in neurons. *Proc Natl Acad Sci U S A* Jun 16;2009 106(24):9848–9853. [PubMed: 19497871]
369. Sanchez PE, Navarro FP, Fares RP, et al. Erythropoietin receptor expression is concordant with erythropoietin but not with common beta chain expression in the rat brain throughout the life span. *J Comp Neurol* Jun 1;2009 514(4):403–414. [PubMed: 19330822]
370. Slotkin TA, Seidler FJ, Fumagalli F. Targeting of neurotrophic factors, their receptors, and signaling pathways in the developmental neurotoxicity of organophosphates in vivo and in vitro. *Brain Res Bull* Jul 1;2008 76(4):424–438. [PubMed: 18502319]
371. van der Kooij MA, Groenendaal F, Kavelaars A, Heijnen CJ, van Bel F. Neuroprotective properties and mechanisms of erythropoietin in in vitro and in vivo experimental models for hypoxia/ischemia. *Brain Res Rev* Nov;2008 59(1):22–33. [PubMed: 18514916]
372. Andreotti F, Agati L, Conti E, et al. Update on phase II studies of erythropoietin in acute myocardial infarction. Rationale and design of Exogenous erythroPoietin in Acute Myocardial

- Infarction: New Outlook and Dose Association Study (EPAMINONDAS). *J Thromb Thrombolysis* Nov;2009 28(4):489–495. [PubMed: 19533304]
373. Brunner S, Winogradow J, Huber BC, et al. Erythropoietin administration after myocardial infarction in mice attenuates ischemic cardiomyopathy associated with enhanced homing of bone marrow-derived progenitor cells via the CXCR-4/SDF-1 axis. *Faseb J* Feb;2009 23(2):351–361. [PubMed: 18827024]
374. Incagnoli P, Ramond A, Joyeux-Faure M, Pepin JL, Levy P, Ribuot C. Erythropoietin improved initial resuscitation and increased survival after cardiac arrest in rats. *Resuscitation* Jun;2009 80(6):696–700. [PubMed: 19406554]
375. Li CL, Jiang J, Fan YQ, Fu GS, Wang JA, Fan WM. Knockout of the tumor necrosis factor a receptor 1 gene can up-regulate erythropoietin receptor during myocardial ischemia-reperfusion injury in mice. *Chin Med J (Engl)* Mar 5;2009 122(5):566–570. [PubMed: 19323909]
376. Schlecht-Bauer D, Antier D, Machet MC, Hyvelin JM. Short- and Long-Term Cardioprotective Effect of Darbepoetin-alpha: Role of Bcl-2 Family Proteins. *J Cardiovasc Pharmacol.* Jul 10;2009
377. Smith K, Semple D, Bhandari S, Seymour AM. Cellular basis of uraemic cardiomyopathy: a role for erythropoietin? *Eur J Heart Fail* Aug;2009 11(8):732–738. [PubMed: 19633100]
378. Timmer SA, De Boer K, Knaapen P, Gotte MJ, Van Rossum AC. The potential role of erythropoietin in chronic heart failure: from the correction of anemia to improved perfusion and reduced apoptosis? *J Card Fail* May;2009 15(4):353–361. [PubMed: 19398085]
379. Uitterdijk A, Groenendijk BC, van Der Giessen WJ. Stem cell therapy for chronic heart failure. *Hellenic J Cardiol* Mar-Apr;2009 50(2):127–132. [PubMed: 19329414]
380. Matis GK, Birbilis TA. Erythropoietin in spinal cord injury. *Eur Spine J* Mar;2009 18(3):314–323. [PubMed: 19030901]
381. Yoo JY, Won YJ, Lee JH, et al. Neuroprotective effects of erythropoietin posttreatment against kainate-induced excitotoxicity in mixed spinal cultures. *J Neurosci Res* Jan;2009 87(1):150–163. [PubMed: 18711747]
382. Okutan O, Turkoglu OF, Gok HB, Beskonakli E. Neuroprotective effect of erythropoietin after experimental cold injury-induced vasogenic brain edema in rats. *Surg Neurol* Nov;2008 70(5):498–502. [PubMed: 18291472]
383. Karaca M, Odabasoglu F, Kumtepe Y, Albayrak A, Cadirci E, Keles ON. Protective effects of erythropoietin on ischemia/reperfusion injury of rat ovary. *Eur J Obstet Gynecol Reprod Biol* Jun;2009 144(2):157–162. [PubMed: 19375213]
384. Harder Y, Amon M, Schramm R, et al. Erythropoietin reduces necrosis in critically ischemic myocutaneous tissue by protecting nutritive perfusion in a dose-dependent manner. *Surgery* Apr; 2009 145(4):372–383. [PubMed: 19303985]
385. Rotter R, Menshykova M, Winkler T, et al. Erythropoietin improves functional and histological recovery of traumatized skeletal muscle tissue. *J Orthop Res* Dec;2008 26(12):1618–1626. [PubMed: 18634017]
386. Zhu L, Wang HD, Yu XG, et al. Erythropoietin prevents zinc accumulation and neuronal death after traumatic brain injury in rat hippocampus: in vitro and in vivo studies. *Brain Res* Sep 15;2009 1289:96–105. [PubMed: 19615349]
387. Aoshiba K, Onizawa S, Tsuji T, Nagai A. Therapeutic effects of erythropoietin in murine models of endotoxin shock. *Crit Care Med* Mar;2009 37(3):889–898. [PubMed: 19237893]
388. Contaldo C, Elsherbiny A, Lindenblatt N, et al. Erythropoietin enhances oxygenation in critically perfused tissue through modulation of nitric oxide synthase. *Shock* Jun;2009 31(6):599–606. [PubMed: 18838945]
389. Simon F, Calzia E, Radermacher P, Schelzig H. Beneficial effects of erythropoietin in models of shock and organ failure-nothing is simple and easy. *Shock* Feb;2009 31(2):220–221. [PubMed: 19145197]
390. Casals-Pascual C, Idro R, Picot S, Roberts DJ, Newton CR. Can erythropoietin be used to prevent brain damage in cerebral malaria? *Trends Parasitol* Jan;2009 25(1):30–36. [PubMed: 19008152]

391. Mihaila RG, Rezi EC, Boitan M, Zaharie AV, Olteanu A, Deac M. Erythropoietin and the pro-inflammatory cytokines in chronic C hepatitis. *Hepatogastroenterology* May-Jun;2009 56(91-92): 751–755. [PubMed: 19621696]
392. Picot S, Bienvenu AL, Konate S, et al. Safety of epoetin beta-quinine drug combination in children with cerebral malaria in Mali. *Malar J* 2009;8:169. [PubMed: 19630971]
393. MacRedmond R, Singhera GK, Dorscheid DR. Erythropoietin inhibits respiratory epithelial cell apoptosis in a model of acute lung injury. *Eur Respir J* Jun;2009 33(6):1403–1414. [PubMed: 19164355]
394. Tascilar O, Cakmak GK, Tekin IO, et al. Protective effects of erythropoietin against acute lung injury in a rat model of acute necrotizing pancreatitis. *World J Gastroenterol* Dec 14;2007 13(46):6172–6182. [PubMed: 18069756]
395. Wu H, Dong G, Liu H, Xu B, Li D, Jing H. Erythropoietin attenuates ischemiareperfusion induced lung injury by inhibiting tumor necrosis factor-alpha and matrix metalloproteinase-9 expression. *Eur J Pharmacol* Jan 14;2009 602(2-3):406–412. [PubMed: 19061883]
396. Chang YK, Choi DE, Na KR, et al. Erythropoietin attenuates renal injury in an experimental model of rat unilateral ureteral obstruction via anti-inflammatory and anti-apoptotic effects. *J Urol* Mar;2009 181(3):1434–1443. [PubMed: 19157461]
397. Chen HH, Tarng DC, Lee KF, Wu CY, Chen YC. Epoetin alfa and darbepoetin alfa: effects on ventricular hypertrophy in patients with chronic kidney disease. *J Nephrol* Jul-Aug;2008 21(4): 543–549. [PubMed: 18651544]
398. Song YR, Lee T, You SJ, et al. Prevention of acute kidney injury by erythropoietin in patients undergoing coronary artery bypass grafting: a pilot study. *Am J Nephrol* 2009;30(3):253–260. [PubMed: 19494484]
399. Luo YH, Li ZD, Liu LX, Dong GH. Pretreatment with erythropoietin reduces hepatic ischemia-reperfusion injury. *Hepatobiliary Pancreat Dis Int* Jun;2009 8(3):294–299. [PubMed: 19502171]
400. Schmeding M, Hunold G, Ariyakhagorn V, et al. Erythropoietin reduces ischemiareperfusion injury after liver transplantation in rats. *Transpl Int*. Mar 20;2009
401. Ucan BH, Irkorucu O, Cakmak GK, et al. Erythropoietin: a possible cytoprotective cytokine in acute necrotizing pancreatitis. *J Hepatobiliary Pancreat Surg* 2009;16(4):530–537. [PubMed: 19333535]
402. Tsai JC, Song BJ, Wu L, Forbes M. Erythropoietin: a candidate neuroprotective agent in the treatment of glaucoma. *J Glaucoma* Sep;2007 16(6):567–571. [PubMed: 17873720]
403. Wang ZY, Shen LJ, Tu L, et al. Erythropoietin protects retinal pigment epithelial cells from oxidative damage. *Free Radic Biol Med* Apr 15;2009 46(8):1032–1041. [PubMed: 19136057]
404. Zhong YS, Liu XH, Cheng Y, Min YJ. Erythropoietin with retrobulbar administration protects retinal ganglion cells from acute elevated intraocular pressure in rats. *J Ocul Pharmacol Ther* Oct;2008 24(5):453–459. [PubMed: 18788995]
405. Toba H, Sawai N, Morishita M, et al. Chronic treatment with recombinant human erythropoietin exerts renoprotective effects beyond hematopoiesis in streptozotocin-induced diabetic rat. *Eur J Pharmacol* Jun 10;2009 612(1-3):106–114. [PubMed: 19356735]
406. Lagreze WA, Feltgen N, Bach M, Jehle T. Feasibility of intravitreal erythropoietin injections in humans. *Br J Ophthalmol* Dec;2009 93(12):1667–1671. [PubMed: 19692373]
407. Di Giacomo V, Sancilio S, Caravatta L, Rana RA, Di Pietro R, Cataldi A. Regulation of CREB activation by P38 mitogen activated protein kinase during human primary erythroblast differentiation. *Int J Immunopathol Pharmacol* Jul-Sep;2009 22(3):679–688. [PubMed: 19822084]
408. Koh SH, Noh MY, Cho GW, Kim KS, Kim SH. Erythropoietin increases the motility of human bone marrow-multipotent stromal cells (hBM-MSCs) and enhances the production of neurotrophic factors from hBM-MSCs. *Stem Cells Dev* Apr;2009 18(3):411–421. [PubMed: 18590375]
409. Tilling L, Chowieńczyk P, Clapp B. Progenitors in motion: mechanisms of mobilization of endothelial progenitor cells. *Br J Clin Pharmacol* Oct;2009 68(4):484–492. [PubMed: 19843051]

410. Mikati MA, Hokayem JA, Sabban ME. Effects of a single dose of erythropoietin on subsequent seizure susceptibility in rats exposed to acute hypoxia at p10. *Epilepsia* Jan;2007 48(1):175–181. [PubMed: 17241225]
411. Moon C, Krawczyk M, Paik D, et al. Erythropoietin, modified to not stimulate red blood cell production, retains its cardioprotective properties. *J Pharmacol Exp Ther* Mar;2006 316(3):999–1005. [PubMed: 16306273]
412. Um M, Gross AW, Lodish HF. A “classical” homodimeric erythropoietin receptor is essential for the antiapoptotic effects of erythropoietin on differentiated neuroblastoma SH-SY5Y and pheochromocytoma PC-12 cells. *Cell Signal* Mar;2007 19(3):634–645. [PubMed: 17045782]
413. Arcasoy MO. The non-haematopoietic biological effects of erythropoietin. *Br J Haematol* Apr; 2008 141(1):14–31. [PubMed: 18324962]
414. Schumann C, Triantafilou K, Krueger S, et al. Detection of erythropoietin in exhaled breath condensate of nonhypoxic subjects using a multiplex bead array. *Mediators Inflamm* 2006;2006(5):18061. [PubMed: 17392570]
415. Li F, Chong ZZ, Maiese K. Erythropoietin on a Tightrope: Balancing Neuronal and Vascular Protection between Intrinsic and Extrinsic Pathways. *Neurosignals* Nov-Dec;2004 13(6):265–289. [PubMed: 15627815]
416. Mojiminiyi OA, Abdella NA, Zaki MY, El Gebely SA, Mohamedi HM, Aldhahi WA. Prevalence and associations of low plasma erythropoietin in patients with Type 2 diabetes mellitus. *Diabet Med* Aug;2006 23(8):839–844. [PubMed: 16911620]
417. Symeonidis A, Kouraklis-Symeonidis A, Psiroyiannis A, et al. Inappropriately low erythropoietin response for the degree of anemia in patients with noninsulin-dependent diabetes mellitus. *Ann Hematol* Feb;2006 85(2):79–85. [PubMed: 16132904]
418. Thomas MC, Cooper ME, Tsalamandris C, MacIsaac R, Jerums G. Anemia with impaired erythropoietin response in diabetic patients. *Arch Intern Med* Feb 28;2005 165(4):466–469. [PubMed: 15738380]
419. Teramo K, Kari MA, Eronen M, Markkanen H, Hiilesmaa V. High amniotic fluid erythropoietin levels are associated with an increased frequency of fetal and neonatal morbidity in type 1 diabetic pregnancies. *Diabetologia* Oct;2004 47(10):1695–1703. [PubMed: 15502930]
420. Teramo KA, Widness JA. Increased fetal plasma and amniotic fluid erythropoietin concentrations: markers of intrauterine hypoxia. *Neonatology* 2009;95(2):105–116. [PubMed: 18776724]
421. Kaindl AM, Sifringer M, Koppelstaetter A, et al. Erythropoietin protects the developing brain from hyperoxia-induced cell death and proteome changes. *Ann Neurol* Nov;2008 64(5):523–534. [PubMed: 19067366]
422. Yis U, Kurul SH, Kumral A, et al. Effect of erythropoietin on oxygen-induced brain injury in the newborn rat. *Neurosci Lett* Dec 31;2008 448(3):245–249. [PubMed: 18973793]
423. He Z, Huang L, Wu Y, Wang J, Wang H, Guo L. DDPH: improving cognitive deficits beyond its alpha 1-adrenoceptor antagonism in chronic cerebral hypoperfused rats. *Eur J Pharmacol* Jul 7;2008 588(2-3):178–188. [PubMed: 18502414]
424. Berkingali N, Warnecke A, Gomes P, et al. Neurite outgrowth on cultured spiral ganglion neurons induced by erythropoietin. *Hear Res* Sep;2008 243(1-2):121–126. [PubMed: 18672044]
425. Shah RC, Wilson RS, Tang Y, Dong X, Murray A, Bennett DA. Relation of hemoglobin to level of cognitive function in older persons. *Neuroepidemiology* 2009;32(1):40–46. [PubMed: 19001795]
426. Bierer R, Peceny MC, Hartenberger CH, Ohls RK. Erythropoietin concentrations and neurodevelopmental outcome in preterm infants. *Pediatrics* Sep;2006 118(3):e635–640. [PubMed: 16908620]
427. Pillai A, Dhandapani KM, Pillai BA, Terry AV Jr, Mahadik SP. Erythropoietin prevents haloperidol treatment-induced neuronal apoptosis through regulation of BDNF. *Neuropsychopharmacology* Jul;2008 33(8):1942–1951. [PubMed: 17805306]
428. Silverberg DS, Wexler D, Iaina A, Schwartz D. The interaction between heart failure and other heart diseases, renal failure, and anemia. *Semin Nephrol* Jul;2006 26(4):296–306. [PubMed: 16949468]

429. Mason-Garcia M, Beckman BS, Brookins JW, et al. Development of a new radioimmunoassay for erythropoietin using recombinant erythropoietin. *Kidney Int Nov*;1990 38(5):969–975. [PubMed: 2266682]
430. Namiuchi S, Kagaya Y, Ohta J, et al. High serum erythropoietin level is associated with smaller infarct size in patients with acute myocardial infarction who undergo successful primary percutaneous coronary intervention. *J Am Coll Cardiol May 3*;2005 45(9):1406–1412. [PubMed: 15862410]
431. Cariou A, Claessens YE, Pene F, et al. Early high-dose erythropoietin therapy and hypothermia after out-of-hospital cardiac arrest: a matched control study. *Resuscitation Mar*;2008 76(3):397–404. [PubMed: 18037223]
432. Bakker WJ, van Dijk TB, Parren-van Amelsvoort M, et al. Differential regulation of Foxo3a target genes in erythropoiesis. *Mol Cell Biol May*;2007 27(10):3839–3854. [PubMed: 17353275]
433. Avasarala JR, Konduru SS. Recombinant erythropoietin down-regulates IL-6 and CXCR4 genes in TNF-alpha-treated primary cultures of human microvascular endothelial cells: implications for multiple sclerosis. *J Mol Neurosci 2005*;25(2):183–189. [PubMed: 15784966]
434. Leuner K, Hauptmann S, Abdel-Kader R, et al. Mitochondrial dysfunction: the first domino in brain aging and Alzheimer's disease? *Antioxid Redox Signal Oct*;2007 9(10):1659–1675. [PubMed: 17867931]
435. Miki T, Miura T, Yano T, et al. Alteration in erythropoietin-induced cardioprotective signaling by postinfarct ventricular remodeling. *J Pharmacol Exp Ther Apr*;2006 317(1):68–75. [PubMed: 16377761]
436. Barandon L, Couffignal T, Ezan J, et al. Reduction of infarct size and prevention of cardiac rupture in transgenic mice overexpressing FrzA. *Circulation Nov 4*;2003 108(18):2282–2289. [PubMed: 14581414]
437. Emami KH, Corey E. When prostate cancer meets bone: control by wnts. *Cancer Lett Aug 18*;2007 253(2):170–179. [PubMed: 17462819]
438. Hoogeboom D, Essers MA, Polderman PE, Voets E, Smits LM, Burgering BM. Interaction of FOXO with {beta}-Catenin Inhibits {beta}-Catenin/T Cell Factor Activity. *J Biol Chem Apr 4*;2008 283(14):9224–9230. [PubMed: 18250171]
439. Sun J, Jin T. Both Wnt and mTOR signaling pathways are involved in insulin-stimulated proto-oncogene expression in intestinal cells. *Cell Signal Jan*;2008 20(1):219–229. [PubMed: 17993259]
440. Lynch RL, Konicek BW, McNulty AM, et al. The progression of LNCaP human prostate cancer cells to androgen independence involves decreased FOXO3a expression and reduced p27KIP1 promoter transactivation. *Mol Cancer Res Mar*;2005 3(3):163–169. [PubMed: 15798096]
441. Hegde PS, Rusnak D, Bertiaux M, et al. Delineation of molecular mechanisms of sensitivity to lapatinib in breast cancer cell lines using global gene expression profiles. *Mol Cancer Ther May*; 2007 6(5):1629–1640. [PubMed: 17513611]
442. Hoekstra AV, Ward EC, Hardt JL, et al. Chemosensitization of endometrial cancer cells through AKT inhibition involves FOXO1. *Gynecol Oncol Mar*;2008 108(3):609–618. [PubMed: 18234299]
443. Bakker WJ, Harris IS, Mak TW. FOXO3a is activated in response to hypoxic stress and inhibits HIF1-induced apoptosis via regulation of CITED2. *Mol Cell Dec 28*;2007 28(6):941–953. [PubMed: 18158893]
444. Dharmarajan TS, Widjaja D. Erythropoiesis-stimulating agents in anemia: use and misuse. *J Am Med Dir Assoc Nov*;2009 10(9):607–616. [PubMed: 19883882]
445. Ioka T, Tsuruoka S, Ito C, et al. Hypertension induced by erythropoietin has a correlation with truncated erythropoietin receptor mRNA in endothelial progenitor cells of hemodialysis patients. *Clin Pharmacol Ther Aug*;2009 86(2):154–159. [PubMed: 19458615]
446. Sigounas G, Salleng KJ, Mehlhop PD, Sigounas DG. Erythropoietin ameliorates chemotherapy-induced fibrosis of the lungs in a preclinical murine model. *Int J Cancer Jun 15*;2008 122(12): 2851–2857. [PubMed: 18350568]

447. Kokhaei P, Abdalla AO, Hansson L, et al. Expression of erythropoietin receptor and in vitro functional effects of epoetins in B-cell malignancies. *Clin Cancer Res* Jun 15;2007 13(12):3536–3544. [PubMed: 17575216]
448. Maiese K, Li F, Chong ZZ. Erythropoietin and cancer. *JAMA* 2005;293(15):1858–1859. [PubMed: 15840858]
449. Hardee ME, Rabbani ZN, Arcasoy MO, et al. Erythropoietin inhibits apoptosis in breast cancer cells via an Akt-dependent pathway without modulating in vivo chemosensitivity. *Mol Cancer Ther* Feb;2006 5(2):356–361. [PubMed: 16505109]
450. Lai SY, Grandis JR. Understanding the presence and function of erythropoietin receptors on cancer cells. *J Clin Oncol* Oct 10;2006 24(29):4675–4676. [PubMed: 17028292]
451. Ceelen W, Boterberg T, Smeets P, et al. Recombinant human erythropoietin alpha modulates the effects of radiotherapy on colorectal cancer microvessels. *Br J Cancer* Mar 12;2007 96(5):692–700. [PubMed: 17299396]

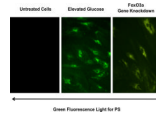


Figure 1. Transfection of FoxO3a siRNA in endothelial cells prevents apoptotic phosphatidylserine (PS) exposure during elevated D-glucose

Representative images illustrate that gene knockdown of *FoxO3a* with FoxO3a siRNA (siRNA) significantly blocks endothelial cell membrane PS externalization assessed by annexin V phycoerythrin (green fluorescence). FoxO3a siRNA alone was not toxic and non-specific scrambled siRNA did not reduce PS exposure during elevated D-glucose.

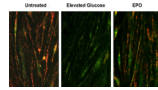


Figure 2. Erythropoietin (EPO) blocks mitochondrial depolarization during elevated D-glucose
Elevated D-glucose (20 mM) resulted in a significant decrease in the red/green fluorescence intensity ratio of mitochondria using a cationic membrane potential indicator JC-1 within 48 hours when compared with untreated endothelial cells, illustrating that elevated D-glucose leads to mitochondrial membrane depolarization. In contrast, pre-treatment with EPO (10 ng/ml) during elevated D-glucose significantly increased the red/green fluorescence intensity of mitochondria in endothelial cells, indicating that mitochondrial membrane potential was restored by EPO.

Table 1

Summary of Clinical Outcomes and Signaling Pathways with Wnt, Nicotinamide, and Erythropoietin

Therapeutic Presentation and Potential During Diabetes Mellitus (DM)	Clinical Outcomes	Signaling Pathways
<i>Wnt</i>	Wnt pathways, such as with transcription factor 7-like 2 gene, may impart increased risk for type 2 DM Wnt may have an association with the development of obesity Wnt has an elevated expression in adipose tissue, the pancreas, and the liver in patients with DM Impaired Wnt signaling through a missense mutation in LRP-6 during metabolic syndrome	Vascular/renal cell early and late apoptotic programs decreased by Wnt Wnt utilizes EPO for protection against elevated glucose Protection by Wnt through Akt and Secreted Frizzled-related protein pathways
<i>Nicotinamide</i>	Nicotinamide can maintain normal fasting blood glucose and improve glucose utilization in animal models of DM Nicotinamide can limit peripheral nerve injury during elevated glucose Nicotinamide protects β -cell function in islet-cell antibody-positive first-degree relatives of Type 1 DM Nicotinamide combined with intensive insulin therapy reduces HbA _{1c} levels Nicotinamide can reduce intestinal absorption of phosphate and prevent the development of hyperphosphatemia	Nicotinamide functions through transcription factors of the forkhead family and caspases Nicotinamide has an inverse relationship with sirtuins that can alter cell survival and cell longevity
<i>Erythropoietin (EPO)</i>	EPO is often low in DM, suggesting an impaired EPO response EPO in diabetic patients with severe, resistant congestive heart failure can decrease fatigue, increase left ventricular ejection fraction, and significantly decrease hospitalization stay EPO can serve to reverse the complications of anemia during DM EPO can protect vascular cells during DM	EPO protection in the hematological and vascular systems relies upon modulation of FoxO and Wnt EPO fosters erythroid progenitor cell development through FoxO activity EPO during elevated glucose and similar to other models of oxidative stress can block cell degeneration through Wnt Protection by EPO is governed by the maintenance of mitochondrial membrane potential