HIV/HCV Co-infection: Pathogenesis, Clinical Complications, Treatment, and New Therapeutic Technologies

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Published online: 11 January 2011

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Abstract World-wide, hepatitis C virus (HCV) accounts for approximately 130 million chronic infections, with an overall 3% prevalence. Four to 5 million persons are coinfected with HIV. It is well established that HIV has a negative impact on the natural history of HCV, including a higher rate of viral persistence, increased viral load, and more rapid progression to fibrosis, end-stage liver disease, and death. Whether HCV has a negative impact on HIV disease progression continues to be debated. However, following the introduction of effective combination antiretroviral therapy, the survival of coinfected individuals has significantly improved and HCV-associated diseases have emerged as the most important co-morbidities. In this review, we summarize the newest studies regarding the pathogenesis of HIV/HCV coinfection, including effects of coinfection on HIV disease progression, HCV-associated liver disease, the immune system, kidney and cardiovascular disease, and neurologic status; and effectiveness of current anti-HIV and HCV therapies and proposed new treatment strategies.

Keywords HIV/HCV coinfection · HIV/HCV progression · HIV/HCV treatment guidelines · HIV/HCV pathogenesis · HIV/HCV immunopathogenesis · HIV/HCV comorbidities

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Introduction

Worldwide, there are an estimated 130 million chronic hepatitis C virus (HCV) infections, with an overall prevalence of 3%. Approximately 4 to 5 million persons are co-infected with HIV. In the US and Western Europe, among HIV-infected persons, HCV prevalence is 72% to 95% among injection drug users (IDU), 1% to 12% in men who have sex with men (MSM), and 9% to 27% in heterosexuals [1]. After acute infection, 15% to 45% clear the virus, and 20% to 30% with persistent viremia develop liver fibrosis, and potentially cirrhosis, liver failure, and hepatocellular carcinoma [2]. Transmission is primarily through IDU or transfusion of infected blood or blood products, although sexual transmission has been suggested among HIV-infected women and documented among MSM [3–5].

HCV is an enveloped positive-strand RNA virus with six genotypes. Genotype 1 predominates in North America [6] and is associated with higher HCV RNA levels [7]. Replication occurs primarily in hepatocytes but also in extrahepatic reservoirs, including PBMCs, in patients with HIV [8].

Recent reviews have discussed many aspects of coinfection, including new noninvasive methods for assessing liver fibrosis [9, 10], current anti-HCV treatment regimens [11–13], novel agents and therapeutic strategies [12, 14, 15•], virologic and immunologic features in pathogenesis [16–19], and clinical outcomes [16, 20, 21]. In this review, we summarize other studies providing new information regarding HIV/HCV coinfection.

Overview

The pathogenesis of HIV and HCV disease progression, including immune system dysfunction and clinical compli-

cations in the setting of coinfection, are summarized in Fig. 1 and described below.

HIV Disease Progression in the Setting of HCV Co-infection

Although most studies demonstrate increased mortality among co-infected individuals, a recent meta-analysis of over 30 studies with over 100,000 patients found no increase in mortality in co-infected patients in the pre-HAART era. Post-HAART, co-infection increased risk of overall mortality but not of AIDS-defining conditions [22]. In contrast, an Italian cohort study found a twofold increased AIDS risk among co-infected patients [23]. The Women's Interagency HIV Study (WIHS) found an almost twofold increased AIDS risk among co-infected women without a CD4 count <200 cells/μL and for ART-naïve women [24••]. The Italian cohort showed increases in bacterial and mycotic infections and WIHS found increases

in bacterial pneumonia, HIV encephalopathy, and wasting syndrome, suggesting the need for earlier and more aggressive HIV and HCV treatment in co-infected individuals [23, 24••].

Recent studies found high levels of T-cell activation in co-infected compared to HIV monoinfected individuals even following HAART [24••, 25, 26]. Chronic immune activation may lead to immune dysfunction and cytokine production, causing enhanced HIV and HCV replication and lower T-cell counts [25]. The WIHS study showed that high levels of activated CD8 T cells are associated with incident AIDS among HCV-viremic women but not HCV-uninfected women, and CD4 activation predicted AIDS in both groups [24••, 25]. Suppression of HCV with therapy reduces activation [26]. These results again support early treatment of HIV and HCV.

Several pathways for active HCV infection impacting HIV infection have been proposed (Table 1). HCV co-infection may increase immune activation, leading to CD4 T-cell apoptosis in HIV-untreated patients and more rapid

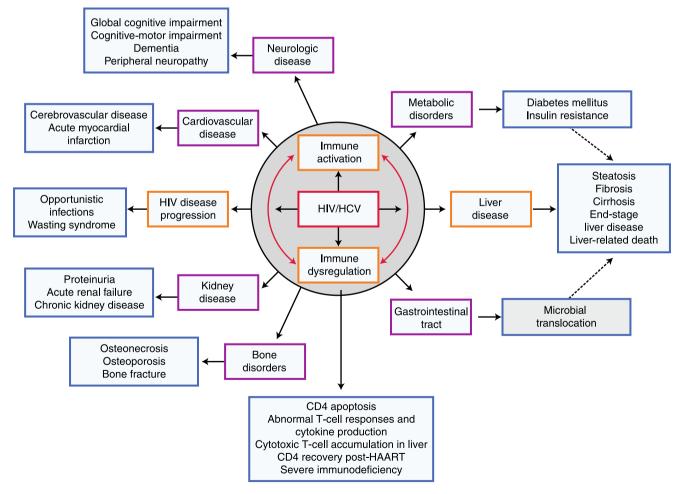


Fig. 1 Pathogenesis of HIV/HCV co-infection: Immune activation and dysregulation, effects on HIV and HCV disease progression, and complications in multiple organ systems



Table 1 Pathogenesis of HIV/HCV-associated diseases and complications

Affected organ/ system	Complication	Proposed mechanism of action	References
Immune	↑ HIV disease progression	↑ CD4 and CD8 T-cell activation, leading to ↑ immune dysfunction and cytokine production, enhanced HIV and HCV production, and ↓ T-cell counts	[24••, 25, 26]
		HCV-induced ↑ CD4 T-cell apoptosis, leading to severe immunodeficiency	[27]
		HCV-induced ↓ CD4 recovery post-HAART	[28]
	↑Cryoglobulinemia	↑ Cryoglobulin production by activated B cells	[53]
	↓ CRP production in liver	Unknown	[54]
Liver	↑ Steatosis	HIV-related mitochondrial translocation (†LPS)-induced immune activation	[16, 18, 26, 42]
	↑ Fibrosis	↑ Activation of CD4 and CD8 T cells	[26]
	↑ Cirrhosis	\uparrow Levels of cytokines (eg, IL-4, IL-5, IL-13, TGF- β , IFN- γ and TNF- α) and chemokines (eg, IP-10 and IL-8)	[17, 18]
	↑ ESLD	HIV-induced ↓ HCV-specific CD4 and CD8 T-cell responses	[16, 17]
	↑ HCC	↑ Levels of cytotoxic CD8 T cells in liver	[39, 40]
		↑ Activation of HSC	[18, 19, 43]
		↑ Apoptosis of hepatocytes and HSC, mediated by TRAIL	[16–19, 44]
		↑ Collagen synthesis by HIV-infected HSC	[43]
		↑ HSC production of collagen by HIV-infected Kupffer cells	[16]
		Insulin resistance–associated hyperinsulinemia and hyperglycemia stimulate HSC to \(\cap \) connective tissue growth factor and extracellular matrix	[45, 46]
Liver/metabolic	Diabetes mellitus	Unknown	[68]
	Insulin resistance	Unknown	[46]
Cardiovascular		HIV- and HCV-associated chronic inflammation leads to endothelial dysfunction, causing ↑ sCAMS	[59, 60]
Hematologic	Thrombocytopenia	Sequestration of platelets in cirrhosis and portal hypertension	[55]
		↓ Production of thrombopoietin in advanced liver disease	[55]
Kidney	Proteinuria	Unknown	[56]
	Acute interstitial nephritis Acute tubular necrosis	Unknown	[58]
	MPGN Acute renal failure	Stimulation of B cells to \(\tau \)cryoglobulin production and deposits in renal vessels	[53, 56–58]
	Chronic kidney disease		
Gastrointestinal	Mitochondrial translocation	HIV-inducted †gut permeability and depletion of CD4 cells from gut-associated lymphoid tissue	[16]
Central/peripheral nervous	Neurocognitive syndromes	HIV and HCV replication in brain	[61]
	Peripheral neuropathy	HCV core protein activates glia and ↑HIV-associated	[62]
	syndromes	neurotoxicity LPS-induced monocyte activation and ↑HIV-associated dementia	[65]
Bone	Osteoporosis Osteonecrosis	↓ Bone mass in chronic liver disease	[70]

CRP C-reactive protein; ESLD end-stage liver disease; HCC hepatocellular carcinoma; HSC hepatic stellate cells; LEE liver enzyme elevations; LPS lipopolysaccharides; MPGN membranoproliferative glomerulonephritis; sCAMS, soluble cellular adhesion molecules; TRAIL, tumor necrosis factor—related apoptosis-inducing ligand

progression to severe immunodeficiency [27]. However, the impact of HCV infection on CD4 cell recovery following HAART is conflicting; some reports note a poorer CD4 response in co-infected compared to monoinfected patients [28], and others do not [27, 29–31].

HCV-Associated Liver Disease Progression in the Setting of HIV Co-infection

Despite reductions of morbidity and mortality in HIV-infected individuals on HAART, liver-related deaths now



represent a leading cause of death in this population. primarily due to HCV co-infection. HCV-associated liver disease, including fibrosis, cirrhosis, and end-stage liver disease (ESLD), is accelerated in HIV-infected individuals. Progression to cirrhosis is threefold higher in co-infected than monoinfected patients, and approximately 33% progress to cirrhosis in less than 20 years [32...]. A recent metaanalysis of 17 studies found cirrhosis in 21% of 3567 coinfected individuals after 20 years of infection and 49% after 30 years [33]. Another meta-analysis of 27 studies among 7666 individuals found that co-infected individuals had a twofold increased rate of cirrhosis compared to monoinfected [33]. Severe liver fibrosis and cirrhosis were also found in 10% to 25% of co-infected HCV viremic patients with normal alanine aminotransferase (ALT) levels [34, 35]. A 20-year prospective study found increased risk of hepatitis/liver-related deaths despite HAART among coinfected drug users (DUs) compared to HCV-monoinfected DUs, providing further support that HIV accelerates liver disease in the HAART era [36].

Hepatic steatosis (HS), a common (40%–75%) complication of HCV monoinfection and HCV/HIV co-infection, is associated with rapid fibrosis progression [37], although a recent meta-analysis found that it is not necessarily more common in co-infected than HCV monoinfected patients [38]. HS is associated with increased body mass index, diabetes, elevated ALT levels, HCV genotype 3, necroinflammation, and fibrosis [37, 38].

Pathogenesis of Liver Disease in Co-infection

Mechanisms to explain accelerated liver disease in coinfected patients are not well understood but may include direct viral effects and immunologic alterations such as immune activation, apoptosis, and diminished HCVspecific T-cell responses (Table 1) [16-18]. Immune activation by HIV induces cytokine changes (eg, IL-4, IL-5, and IL-13, TGF-β) that increase liver inflammation and fibrosis [17, 18]. Coinfection increases apoptosis of hepatocytes through a Fas/FasL pathway that could account for accelerated liver disease [17, 18]. Accumulation of cytotoxic CD8 T cells in the liver that increases inflammatory mediators in co-infected compared to HCVmonoinfected patients may also lead to increased tissue damage in co-infected patients [39, 40]. New evidence shows HIV-specific CD8 T cells accumulate in the liver in co-infection and produce TNF- α , which is associated with liver fibrosis [18, 41]. Recently, HIV-related microbial translocation that causes systemic activation has been linked with severity of HCV-related liver disease [42].

Increasing evidence finds HIV replication in hepatocytes and hepatic stellate cells (HSC) [18, 19, 43]. HIV infection of activated HSC promotes collagen expression and secretion of

proinflammatory cytokines [43]. In addition to infection, HIV proteins induce hepatocytes to apoptosis and release of inflammatory chemokines and cytokines that promote fibrosis [18, 19]. Further, HIV and HCV co-infection may increase tumor necrosis factor—related apoptosis—inducing ligand (TRAIL)-mediated apoptosis of hepatocytes [17, 18, 44].

Insulin resistance (IR), also common in chronic HCV, appears to be critical in liver steatosis and liver disease progression. A recent study found IR associated with liver fibrosis and steatosis in HCV-monoinfected but not co-infected patients [45], but another study found IR associated with liver fibrosis in co-infected patients [46]. The mechanism for IR in liver disease among HCV-infected patients is unknown, but hyperinsulinemia and hyperglycemia stimulate HSC, leading to increased connective tissue growth factor and accumulation of extracellular matrix [46].

New Technologies for Diagnosis and Staging of Liver Disease

Although liver biopsy is the gold standard for staging liver disease, its invasive nature, serious complications, sampling error, and inherent heterogeneity have prompted research to identify noninvasive methods [10, 21, 47]. Promising methodologies include transient elastography/elastometry (TE) and serologic biomarkers.

TE, which uses ultrasound readings to measure liver elasticity or stiffness, is well tolerated and can accurately stage fibrosis and cirrhosis [35, 47], although discriminating power may be low [48]. TE may also be useful in patients with normal aminotransferase levels [34, 35].

Serological markers correlated with liver fibrosis stage can be classified into three groups: 1) indices from routine blood tests of liver function [eg, APRI (aspartate aminotransferase [AST]-to-platelet ratio index) and Fib-4 (age, AST, platelets, and ALT level)]; 2) markers of extracellular matrix metabolism (eg, hyaluronic acid); and 3) indices combining markers of both types [49]. These are valid markers of liver fibrosis and predictive of HCV-related liver mortality in monoinfected and co-infected individuals [49, 50].

Other promising noninvasive technologies include DNA microarray analysis to identify novel biomarkers to assess fibrosis [51] and MRI to measure steatosis and predict adipose tissue and metabolic factors associated with steatosis in co-infected patients [52].

Other Complications of HIV/HCV Co-infection

Other Immune Dysregulation and Hematologic Disorders

Although both HCV and HIV activate B cells, their interaction in the lymphoid system needs study [16]. Both



viruses induce B lymphocytes to synthesize cryoglobulins, but a recent study found only a marginal influence of HIV infection on cryoglobulin production in HCV-infected patients [53]. A large retrospective study found HCV co-infected patients had lower C-reactive protein (CRP) levels than HIV monoinfected patients, suggesting that HCV decreases the liver's ability to secrete CRP [54]. HIV-associated thrombocytopenia remains an important problem in the HAART era and is associated with cirrhosis but also HCV infection without serious liver disease [55]. This may be an important consideration in staging liver disease using APRI, which is calculated using platelet count.

Kidney Disease

A meta-analysis of 24 observational studies and clinical trials found that HCV co-infection is associated with increased HIV-related kidney disease risk, including proteinuria and acute renal failure, compared to HIV monoinfection [56]. A large retrospective study of HIV-infected veterans also found that HCV co-infection was associated with higher chronic kidney disease rates; HCV prevalence increased with worsening estimated glomerular filtration rate (eGFR) [57]. The largest study of biopsy-proven renal disease found that HIV/HCV-associated nephropathies reduced survival compared to HCV monoinfection [58].

Cardiovascular Disease

HCV co-infection is associated with a higher prevalence of cardiovascular disease (CVD) than HIV monoinfection. HCV co-infection is associated with increased risk of cerebrovascular disease and a trend toward increased risk of acute myocardial infarction among HIV-infected patients [59]. HIV-infected patients have an increased CVD risk because HAART and HIV-associated chronic inflammation can cause endothelial dysfunction [60]. Circulating soluble cellular adhesion molecules (CAMS) released by the vascular endothelium, including soluble intercellular adhesion molecule-1 (sICAM-1) and vascular adhesion molecule-1 (sVCAM-1), are higher in coinfected patients stably treated with HAART than healthy controls [60]. HAART can lower sICAM-1 and sVCAM-1, suggesting that HAART can improve endothelial cell function and decrease CVD risk by decreasing plasma HIV-RNA levels, increasing T-cell number and function, and decreasing immune activation. Patients with advanced HCV infection have higher levels of sICAM-1 and sVCAM-1, suggesting that HCV infection also causes endothelial dysfunction, and response to HCV treatment might reduce CVD risk [60].



Both HIV and HCV replicate in the brain and cerebrospinal fluid (CSF) and are implicated in neurocognitive and peripheral neuropathy syndromes [61-64]. Co-infected subjects show significant cognitive-motor impairment compared to HIV-monoinfected patients [63] and higher rates of global cognitive impairment, especially in learning and memory [64]. Recently, HCV RNA and antigens were found in brains of co-infected patients [61]. HCV core protein activates human glia and contributes to HIVassociated neurotoxicity [62]. Plasma lipopolysaccharides (LPS), indicators of microbial translocation from the gut, induce monocyte activation in HIV infection and may contribute to HIV-associated dementia (HAD) by increased trafficking of activated monocytes into the brain [65]. That LPS levels are higher in HCV co-infected patients suggests that HCV may influence HAD pathogenesis [65].

In contrast to findings of increased neurocognitive impairment in co-infected patients, a retrospective study from the AIDS Clinical Trials Group Longitudinal Linked Randomized Trials (ALLRT) found that active HCV infection did not exacerbate clinically significant neurocognitive dysfunction or peripheral neuropathy in individuals with controlled HIV infection [66].

Because sensory neuropathy (SN) is a common complication of HIV infection and certain HIV treatments and is associated with HCV infection, there are concerns about possible synergistic effects of these viruses on the peripheral nervous system [67]. However, a survey among HIV-infected patients in six international sites found HCV seropositivity was not associated with increased SN risk [67].

Diabetes Mellitus

One of the largest prospective studies of HIV-infected persons and uninfected controls found that while HIV infection itself is not associated with increased diabetes mellitus (DM) risk, HCV coinfection is associated with higher risk, with a similar trend in the HIV-uninfected group. Risk was not altered by presence of liver damage as measured by ALT and AST levels [68]. A large prospective study of HIV-infected and uninfected women also found that HCV infection and higher HCV RNA levels were associated with incident diabetes, suggesting that HCV-infected patients should be screened to diabetes [69].

Bone Complications

Among the emerging comorbidities in the HAART era are bone complications, including osteonecrosis or osteoporosis, but the role of HCV and HAART is controversial [70].



In a national cohort of patients on combination ART, the 10-year incidence of bone fractures was 3.6-fold higher in HCV co-infected patients perhaps because patients with chronic liver disease have decreased bone mass [70]. Active monitoring of co-infected patients is recommended for prevention of bone fractures.

Treatment of HIV in Co-infected Patients

The timing of initiation of HAART in relation to anti-HCV therapy in co-infected patients poses challenges for clinicians. HAART may slow liver disease progression and might therefore be initiated earlier in co-infected than HIV monoinfected patients [11, 71]. On the other hand, HAART might increase fibrosis in co-infected patients through cumulative hepatotoxicity [11, 13, 72]. Recent guidelines recommend that HAART generally be initiated first to slow liver disease progression and increase CD4 count, but certain drugs should be avoided (eg, ZDV, didanosine, stavudine, and abacavir) and others should be monitored for hepatoxicity [32., 73.]. Although ART interruption is deleterious to the HIV-infected population, the first randomized study of ART interruptions in HCV co-infected persons found that interruption was particularly unsafe in co-infected persons because of elevated nonopportunistic disease death, though not liver disease death [74].

Treatment of HCV in Co-infected Patients

Treatment of chronic HCV in co-infected individuals is a priority because of their more rapid progression to ESLD, poor tolerance of ART, and greater risk of hepatoxicity [75]. Clearance is associated with regression of liver fibrosis and reduced risk of ART hepatoxicity [75, 76]. However, anti-HCV treatment is less effective in co-infected patients [75].

HCV treatment guidelines for co-infected patients are published [77], but there is a lack of consensus regarding key factors that might inform initiation and duration of therapy, including stage of HIV and HCV disease and viral load, HCV genotype, degree of hepatic fibrosis, and patient's readiness to tolerate and adhere to treatment. These have clinical importance because they influence safety, tolerability, and success of therapy. Investigations continue to identify better predictors of treatment response that could guide the pretreatment evaluation process and permit earlier termination of ineffective treatment, reducing additional cost and adverse effects of ineffective therapy. There are no guidelines for the clinical management and treatment of co-infected children, and the limited experience in their management and

lack of evidence base to guide policy is a barrier to achieving optimal care [78].

Current Standard Therapeutic Regimen

Pegylated interferon plus ribavirin (pegIFN + RBV) is the standard HCV treatment in both monoinfected and co-infected patients [77, 79]. Although guidelines recommend a fixed course of 48 weeks to optimize HCV treatment in co-infected patients [77], a recent study explored response-guided therapy, with duration based on virologic response at treatment weeks 4, 12, and 24 [80]. The results were encouraging, with 55% achieving a sustained viral response (SVR). Among patients who failed a prior suboptimal treatment regimen, retreatment with pegIFN + RBV for 12 months achieved an SVR in nearly one third, an encouraging outcome in light of earlier studies with lower retreatment response rates [75].

Predictors of Treatment Response

The best predictors of treatment outcome are virologic response kinetics, including rapid viral response (RVR), defined as HCV viral load below the level of detectability 4 weeks after treatment initiation, early viral response (EVR), defined as undetectable HCV load or a 2 log drop from baseline 12 weeks after therapy initiation, and SVR, defined as undetectable HCV load 24 weeks after therapy completion [81]. While undetectable HCV RNA at week 4 is the best predictor of SVR in co-infected patients, baseline serum HCV RNA is an independent predictor of SVR in HCV genotype 1 patients [82]. Although these predictors use absence of serum HCV RNA as the marker of treatment success, negative-strand HCV RNA in PBMCs in the absence of plasma HCV RNA has been reported [8].

Results of investigations of impact of baseline CD4 count on viral response kinetics are mixed. In a large randomized study of pegIFN + RBV in co-infected patients that included a small number of patients with CD4 counts <200 cells/ μ L, SVR rates tended to increase with higher CD4 counts in genotype 1, but were independent of baseline CD4 counts for genotypes 2/3 [83]. Another large randomized study of co-infected patients found the efficacy of pegIFN + RBV was not different in patients with and without severe immunodeficiency, suggesting that advanced immunosuppression is not a major factor in predicting SVR [84].

In the AIDS Clinical Trials Group (ACTG), HCV quasispecies complexity was an important predictor of treatment outcomes, with lower baseline complexity associated with EVR and a decrease in complexity by 4 weeks associated with RVR [81]. Extrahepatic replication in B,



CD4, CD8, and NK cells at the end of 48 weeks of treatment has predicted viral relapse and, although the assays are cumbersome, they might identify patients whose treatment should be extended to 72 weeks [85].

Adverse Events Associated with Anti-HCV Therapy

IFN-related adverse events (AEs), specifically CD4 cell declines and psychiatric effects, are reported to be more common in co-infected virologic responders than non-responders [86]. Also with successful IFN therapy, alterations in cytokine pools necessary to improve immune function may have negative effects in the brain by traversing the blood–brain barrier [86].

The first meta-analysis of sex difference in AEs in co-infected individuals showed that women were more likely to develop AEs requiring treatment discontinuation or dose modification and to develop them earlier, but the types of AEs were similar [87]. Women on NNRTIs were more likely to discontinue therapy, and women on AZT were more likely to experience AEs, suggesting that in women antiviral regimen is an important predictor of treatment discontinuation and modification.

HAART as HCV Therapeutic Agent

Table 2 summarizes the therapeutic and adverse effects of HAART on HCV disease, alone or in combination with pegIFN + RBV. Early HAART may protect co-infected patients from liver fibrosis progression [71]. HAART can significantly decrease liver HCV necroinflammatory activity in co-infected patients with relatively preserved immune status, possibly by inhibiting HIV replication in the liver or decreasing level of proinflammatory cytokines [71]. In vitro, the HIV protease inhibitor nelfinavir inhibits HCV replication at concentrations showing no cytotoxicity and acts synergistically with IFN against HCV, suggesting that nelfinavir could improve the antiviral effects of IFN in co-infected patients [88].

Some ART medications, especially abacavir, may compromise the response to anti-HCV therapies, perhaps by competing intracellularly with RBV [89]. However, a large cohort study did not find an association of abacavir or other ARTs with reduced EVR or SVR [90].

The combination of HAART and pegIFN + RBN may also increase the frequency of AEs [13]. HCV therapy with zidovudine (ZDV) has been associated with higher anemia rates [13]. RBV with NRTIs such a didanosine has been

Table 2 Therapeutic and adverse effects on HCV disease of antiretroviral agents alone and in combination with anti-HCV therapy in co-infected individuals

Therapy	Therapeutic effect	Reference	Adverse event	Reference
HAART	↓ HIV replication in liver	[71]	↑ Liver enzymes (LEE) at HAART initiation (IRD)	[93]
	↓ Proinflammatory cytokines	[39, 71]	↑ Hepatoxicity	[13, 72]
	↓ Hepatic necroinflammatory activity	[71]		
	↓ Liver necrosis and inflammation	[11]		
	↓ Liver disease progression	[11, 71]	↑ Hepatocellular necrosis and steatosis	[13]
	↓ Liver mortality	[13]		
Protease inhibitors	↓ HCV replication in vitro↓ Hepatotoxicity after SVR	[88] [76]		
NRTI	· · · · · · · · · · · · · · · · · · ·	L	↑ Steatosis	[11]
			↑ Hepatotoxicity after SVR	[76]
Efavirenz	↓ Hepatotoxicity after SVR to IFN	[76]		. ,
HAART + PegIFN + RBV				
ZDV + PegIFN + RBV			↑ Anemia	[13, 32••]
PI + pegIFN + RBV	↓ Hepatotoxicity	[76]		
	Synergistic ↓ HCV replication in vitro	[88]		
NNRTI + pegIFN + RBV	↓ Hepatotoxicity	[76]		
NRTI + pegIFN + RBV			↑ Mitochondrial toxicity	[13, 32••, 91, 92]
			↑ Steatosis and fibrosis	[91, 92]
			↓ Response to anti-HCV therapies	[32••, 89]

HAART highly active antiretroviral therapy; IFN interferon; IRD immune restoration disease; NNRTI nonnucleoside reverse transcriptase inhibitor (efavirenz); NRTI nucleoside reverse transcriptase inhibitor (abacavir, didanosine, stavudine); PegIFN + RBV pegylated interferon + ribavirin; PI protease inhibitor (nelfinavir); RBV ribavirin; SVR sustained viral response to pegIFN + RBV; ZDV zidovudine



associated with increased risk of mitochondrial toxicity (MT) and worsening steatosis/fibrosis [13, 91, 92]. MT-associated laboratory abnormalities are frequent during pegIFN + RBV therapy in combination with HAART, especially when high RBV doses are used [91]. However, patients with signs of MT show faster decreases in HCV RNA levels and achieve higher SVR rates, probably because MT reflects increased intracellular RBV levels. Clinicians should be aware of potential interactions between NRTIs and RBV in coinfected patients, and increased lactate levels might be useful to adjust RBV dosage to optimal efficacy [91].

Co-infected patients may experience liver enzyme elevation (LEE) following HAART initiation [93]. CD4 Tcell increases early after HAART initiation were higher among co-infected patients who developed LEE than coinfected or monoinfected patients who did not develop LEE, suggesting that LEE early in HAART is a form of immune restoration disease (IRD) or immune reconstitution inflammatory syndrome (IRIS) involving an immune reconstitution-induced inflammatory response to HCVspecific antigens [94]. The risk of LEE after HAART initiation is lower in co-infected patients with SVR to anti-HCV therapy, arguing for HCV treatment before commencing HAART [76]. In contrast, an Italian multicenter study found that HAART is not a risk factor for LEE in coinfected patients and does not modify the association between coinfection and the risk of LEE [95].

Novel New Therapies

Because pegIFN + RBV leads to successful outcome in only 40% to 50% of monoinfected and 30% of co-infected patients, more effective drugs are being developed [14, 15•]. As in HIV, triple-combination therapies may be more effective in achieving virologic cure and less prone to resistance development in monoinfected and co-infected patients. NS3 protease and NS5B polymerase enzymes, essential for HCV replication, are primary targets. Results for inhibitors targeting these enzymes (combined with pegIFN + RBV) are positive in clinical trials [11, 96, 97] and preclinical studies [98–100]. The most promising of these are the protease inhibitors telaprevir and boceprevir, which are expected to be approved this year [101, 102]. Taribavirin, an oral RBV prodrug with significantly lower anemia rates, is in phase III trials [103].

Anti-HCV Treatment in Acute HCV Infection

Increasing acute HCV infections among HIV-infected populations were reported in Europe in the early 2000s, most due to sexual transmission [4]. Because HIV-infected persons have delayed HCV antibody production, screening with third-generation ELISA tests in the first 6 months is inadequate [104]. A recent study found the median time from HCV

infection to antibody production was 158 days but the median ALT at the first positive PCR was 65 IU/mL, suggesting that elevated ALTs in at-risk patients should indicate HCV testing by PCR to diagnose and treat infection early [104].

Treatment data for acute HCV infection in co-infected patients have emerged only recently, and questions remain regarding optimal time, indication, therapy duration, and whether parallel initiation of HAART is necessary [13]. The Australian Trial in Acute Hepatitis C (ATAHC) found high treatment success after 24 weeks of therapy for both acute and early chronic HCV infections in co-infected patients, with an SVR of 80% [105]. RVR had a 100% positive predictive value, suggesting that RVR is an important predictor of SVR in acute and chronic HCV infection. Results for three patients suggest that an SVR is possible even during primary HIV infection [106].

Conclusions

HIV-infected patients should be screened regularly for HCV co-infection, particularly if they are in high-risk groups. Sexual transmission of HCV needs to be included in patient counseling both for HIV-infected men and women. Increased progression rates to AIDS and liver disease in co-infected individuals may require earlier and more aggressive treatment of both infections. However, following treatment initiation, patients should be monitored for AEs, with prompt intervention to support continued treatment. For most co-infected patients, HAART should be initiated before anti-HCV therapy to slow liver progression and increase CD4 counts. Promising new therapies available in the near future will increase SVR in co-infected individuals.

Acknowledgment The work of the authors for this review was supported by NIH/NIAID grant 2 R01 AI 052065-05A2.

Disclosure No potential conflicts of interest relevant to this article were reported.

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