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## Incorporation of Fluorinated Phenylalanine Generates Highly Specific Inhibitor of Proteasome's Chymotrypsin-like Sites

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### Abstract

Proteasomal processing is conducted by three individual catalytic subunits, namely  $\beta 1$ ,  $\beta 2$ , and  $\beta 5$ . Subunit-specific inhibitors are useful tools in dissecting the role of these individual subunits and are leads toward the development of antitumor agents. We here report that the presence of fluorinated phenylalanine derivatives in peptide based proteasome inhibitors has a profound effect on inhibitor potency and selectivity. Specifically, compound **4a** emerges as one of the most  $\beta 5$  specific inhibitors known to date.

### Introduction

The majority of all cytosolic and nuclear proteins in eukaryotic cells are degraded by the ubiquitin-proteasome pathway. In this system, proteins destined for degradation are modified with a polyubiquitin chain as a recognition tag for the 26S proteasome where proteolysis occurs. The 26S proteasome contains one or two 19S regulatory caps together with the proteolytically active, cylindrical 20S core. Within the mammalian constitutive 20S core, three pairs of proteolytically active sites are present displaying different substrate specificity. Of these, the  $\beta 1$  subunits (caspase-like) cleave after acidic residues, the  $\beta 2$  subunits (trypsin-like) cleave after basic residues, and the  $\beta 5$  subunits (chymotrypsin-like) cleave after bulky, hydrophobic residues.<sup>1,2</sup> The peptidyl boronic acid proteasome inhibitor PS-341 (**8**)<sup>3</sup> is used for the treatment of multiple myeloma and targets the  $\beta 5$  and  $\beta 1$  subunits. To study the role of the three individual active subunits, subunit-specific inhibitors are needed. Inhibitors with moderate to good selectivity for either one of the subunits have been developed,<sup>4</sup> however there is still room for improvements.

The search for subunit selective inhibitors is predominantly conducted by either screening of natural products,<sup>5</sup> rational design,<sup>6</sup> or compound library building.<sup>4b,7</sup> We observed that in these studies the effect of fluorine functionality in proteasome inhibitors is relatively uncharted.<sup>8</sup> In contrast, fluorine has found wide interest in bioorganic and structural chemistry over the past decade and has become an important feature in drug design.<sup>9</sup> This is predominantly due to the typical characteristics of fluorine (when bound to carbon) such as

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Supporting Information Available: Complete synthetic details and characterization of all compounds and additional biological experiments. This material is available free of charge via the Internet at <http://pubs.acs.org>.

its comparable size to hydrogen, its electron withdrawing ability, superhydrophobicity of fluorocarbons, and self-association between fluorinated moieties. In protein structure design, introduction of fluorine can mimic functional groups, alter structural properties, and thereby (de)stabilize protein structures or function as recognition motifs.<sup>10</sup> In addition, the beneficial <sup>19</sup>F nuclear magnetic characteristics have found their use in structure analysis by (solid state) <sup>19</sup>F NMR spectroscopy or <sup>19</sup>F MRI to study, for example, protein aggregation.<sup>11</sup>

The set of fluorinated proteasome inhibitors prepared in the context of the here presented studies are depicted in Figure 1. Compounds **2a** and **2b** containing pentafluoroPhe (PheF<sub>5</sub><sup>a</sup>) and 3,5-bis(trifluoromethyl)Phe (Phe(*m*-CF<sub>3</sub>)<sub>2</sub>), respectively, are based on compound **8** derivative **1** (having a comparable potency toward the  $\beta$ 1 and  $\beta$ 5 proteasome subunits with respect to **8**)<sup>12</sup> and differ in that the phenylalanine in **8** is replaced by the corresponding fluorinated analogue. In addition, incorporation of fluorinated phenylalanines at different positions in tripeptide epoxyketones<sup>2</sup> led to compounds **3–6** in which systematically either one or both of the P2 and P3 positions were altered. We opted for the use of Phe(*m*-CF<sub>3</sub>)<sub>2</sub> and PheF<sub>5</sub> for the dual reason that these are readily available and that hydrophobic amino acids (that is the nonfluorinated analogues) are in principle accepted by all proteasome active sites. The epoxyketone electrophilic trap was selected based on the natural product epoxomicin. The epoxyketone war-head featured by epoxomicin displays a specific and selective reactivity toward the N-terminal threonine residue that makes up the proteasome catalytic active sites.<sup>2,5</sup> For this reason, synthetic peptide epoxyketones are now much studied leads in medicinal chemistry studies in which the proteasome plays a role.<sup>13</sup> The tripeptide epoxyketones **3–6** feature an azide moiety at the N-terminal end for future modifications (for instance, coupling to a fluorophore or biotin in either one- or two-step labeling experiments).<sup>14</sup>

## Results and Discussion

The C-terminally modified oligopeptides were produced following synthesis protocols we reported previously.<sup>7</sup> The amino acids used were either commercially available or prepared following established procedures.<sup>15</sup> See for full experimental data on the synthesis of the compounds the Supporting Information (SI). The inhibition potential of compounds **2a** and **2b** in comparison with their nonfluorinated analogue boronic ester **1** (the pinanediol analogue of the clinical drug **8**) was assessed in a competition assay employing human embryonic kidney (HEK-293) cell lysates in combination with the fluorescent broad spectrum proteasome probe MV151 (**9**).<sup>12</sup> Cell lysates were incubated with each of the three compounds at 0.05, 0.1, and 1  $\mu$ M final concentrations prior to treatment with 0.5  $\mu$ M final concentration of **9**. The samples were denatured, resolved by SDS-PAGE, and the wet gel slabs were scanned on a fluorescence scanner. The gel image is shown in Figure 2. Lysates treated with the fluorescent probe display three bands that correspond to the three active subunits ( $\beta$ 1,  $\beta$ 2, and  $\beta$ 5) as depicted in Figure 2 lane 1. The ability of a compound to inhibit the proteasome active sites is reflected by disappearance of the bands. As apparent from this image, the fluorinated compounds **2a** and **2b** are at least as potent as their nonfluorinated counterpart **1** showing complete inhibition of the  $\beta$ 1 and  $\beta$ 5 subunits between 0.1 and 1  $\mu$ M. In addition, incorporation of fluorinated Phe leaves the selectivity of  $\beta$ 1 and  $\beta$ 5 over  $\beta$ 2 subunits for this type of inhibitor unchanged. This result is also apparent from the activity (IC<sub>50</sub>) measurements of the inhibitors toward the different subunits in purified rabbit 26S

<sup>a</sup>Abbreviations: Phe(F<sub>5</sub>), pentafluorophenylalanine; Phe(*m*-CF<sub>3</sub>)<sub>2</sub>, 3,5-bis(trifluoromethyl)phenylalanine, HEK-293, human embryonic kidney cell lysates; EL-4, mouse lymphoma cell line; DiPEA, *N,N*-diisopropylethylamine; DTT, dithiothreitol; Tris, tris(hydroxymethyl)-aminomethane; amc, 7-amido-4-methylcoumarin; TFA, trifluoroacetic acid; DMF, *N,N*-dimethylformamide; DCM, dichloromethane.

proteasome as shown in Table 1. Experiments in murine EL4 cell lysates (mouse lymphoma cell line) which contain both the immuno- and constitutive proteasome subunits gave similar results (see SI).

Next, the inhibition properties of the seven epoxyketone containing compounds were determined in a similar competition assay against **9** at 1 and 100  $\mu\text{M}$  final concentrations employing HEK-293 cell lysates (Figure 3) and by measuring inhibition of purified proteasomes for the most active compounds (Table 1). When comparing nonfluorinated compound **3** to the fluorinated ones (**4–6**), it becomes apparent that none of these compounds inhibit the  $\beta 1$  subunit at concentrations up to 100  $\mu\text{M}$  and that introducing fluorines (in either position) leads to a decrease in the inhibition of the  $\beta 2$  subunit and hence to an increase in  $\beta 5$  specificity. When comparing Table 1 with Figure 3, it appears that the results from the two independent assays deviate in places. For example the  $\beta 2$ -IC<sub>50</sub> value for **4a** is <15  $\mu\text{M}$  (Table 1), whereas the majority of the corresponding band in Figure 3 is gone at 1  $\mu\text{M}$ . Intrinsic differences in both assays, neither of which deliver  $k_i$  values are at the basis of these small but distinct differences. The complementary assays, however, both show similar trends, which is why we prefer to report both.<sup>16</sup> For instance, in Figure 3, there appears to be almost no difference in inhibition potential of  $\beta 5$  between the nonfluorinated compound **3** and either P2 or P3 fluorinated compounds **4a,b** and **5a,b**. Although there is a big difference in IC<sub>50</sub> values, all compounds are at least submicromolar inhibitors.

Interestingly, the presence of fluorine substituents at both P2 and P3 positions (**6a,b**, Figure 3) has a dramatic effect on the inhibition. This effect is most pronounced for the Phe-(*m*-CF<sub>3</sub>)<sub>2</sub> analogues (**b** series). In general, the PheF<sub>5</sub> compounds (**a** series) are more active against the  $\beta 5$  subunit than their hexafluoro-Phe analogues (**b** series). Experiments in the lysates of murine EL4 cells gave similar results, but the difference in inhibition potential between the compounds was even more pronounced (see SI). Thus, it appears that introduction of Phe(F<sub>5</sub>) in the P2 position generates a highly specific inhibitor of the  $\beta 5$  site. This most potent and  $\beta 5$  selective inhibitor (compound **4a**) was further investigated. The inhibitory potential at much lower concentrations (1 nM to 1  $\mu\text{M}$ ) is shown in Figure 4 (competition assay against **9**). Already, at 5 nM, inhibition of  $\beta 5$  is apparent, and between 100 and 250 nM, all  $\beta 5$  subunits are saturated while  $\beta 1$  and  $\beta 2$  are unaffected or even upregulated (a phenomenon which is not fully understood but has been observed by others as well).<sup>17</sup> Having an IC<sub>50</sub> value of 2 nM for the  $\beta 5$  subunit against >15  $\mu\text{M}$  for  $\beta 1$  and  $\beta 2$ , this compound is one of the most  $\beta 5$  selective inhibitors known to date. For instance, this compound compares well with NC005, a  $\beta 5$  selective inhibitor we recently found.<sup>6</sup> Comparison of nonfluorinated compound **3** with **4a** reveals that both compounds are equally active toward the  $\beta 5$  subunit. Enhanced selectivity for  $\beta 5$  arises by the dramatic drop in activity for the  $\beta 2$  subunit when fluorine is introduced, as in **4a**.

For the direct labeling of  $\beta 5$ , a new fluorescent probe was made by reacting compound **4a** with Bodipy-FL-alkyne<sup>18</sup> in a Cu(I) mediated Huisgen 1,3-dipolar cycloaddition, giving green fluorescent probe **7** (Figure 5). The potential of this probe to label the  $\beta 5$  subunit was explored in a competition assay against **9** as explained before. The wet gel slab was scanned on a fluorescence scanner at two different wavelengths, allowing visualization of one of the two fluorescent dyes at a time. Figure 5A shows the read-out at 520 nm visualizing the appearance of one band: labeling of the  $\beta 5$  subunit by compound **7**. The labeling is already visible at a concentration of 1 nM, and the subunit appears to be saturated (no more increase in the bands intensity) between 50 and 100 nM. At this point, only a faint  $\beta 2$  band is visible.<sup>19</sup> The fluorescence read-out at 560 nm in Figure 5B (displaying labeling with **9** of remaining activities) reveals that the  $\beta 5$  band disappears while leaving the remaining subunit-bands intact.

## Conclusions

In summary, the effect of incorporation of fluorinated Phe in proteasome inhibitors was studied. We found that substitution of nonfluorinated Phe in compound **8** analogue **1** with fluorinated versions does not affect its selectivity for the different active subunits, whereas the potency is slightly increased for the Phe(*m*-CF<sub>3</sub>)<sub>2</sub> version. In addition, we found that the effect of incorporation of fluorinated Phe in peptide epoxyketone proteasome inhibitors depends on the site of substitution. Fluorination of both the P2 and P3 sites decreases potency dramatically, however, fluorinated Phe at the P2 position hardly affects the potency but instead yields much more  $\beta$ 5 selective inhibitors. Comparison of the results obtained with the boronic esters with those from the epoxyketone studies invites the tentative conclusion that  $\beta$ 2 is most sensitive toward fluorine substituents. Compound **1** is ineffective toward  $\beta$ 2 and introduction of fluorines into this sequence has no apparent effect. When comparing epoxyketone **3** with **4–6**, however, the major difference is that the latter, fluorinated analogues leave  $\beta$ 2 largely intact. Further studies, for instance, making use of different fluorine amino acids, are needed to substantiate this finding. Finally, compound **4a** was identified as one of the most  $\beta$ 5 selective inhibitors known to date and was converted to a  $\beta$ 5 selective fluorescent probe (**7**), which can be used to label and visualize the  $\beta$ 5 subunit selectively.

## Experimental Section

All compounds tested are >95% pure on the basis of LC-MS. LC-MS analysis was performed on a Jasco HPLC system with a Phenomenex Gemini 3  $\mu$ m C18 50 mm  $\times$  4.60 mm column (detection simultaneously at 214 and 254 nm), coupled to a PE Sciex API 165 mass spectrometer with ESI.

Compounds **2a** and **2b** were synthesized via the same method described in literature for compound **1**.<sup>12</sup>

### General Procedure for the Synthesis of Compounds 3–6

Hydrazine hydrate (20 equiv) was added to N<sub>3</sub>Phe-Phe-PheOMe (1 equiv) in MeOH (20 mL/mmol) and refluxed until TLC analysis revealed complete consumption of the starting material (usually after 3 h). Toluene was added, and the mixture was concentrated under reduced pressure followed by coevaporation with toluene (2 $\times$ ). The acylhydrazide (1 equiv) was dissolved in a 10/1 (v/v) mixture of dichloromethane/*N,N*-dimethylformamide (DCM/DMF) (10 mL/mmol) and cooled to  $-35$  °C under argon. To this were added *tert*-butylnitrite (1.1 equiv) and HCl (2.8 equiv as a 4M solution in 1,4-dioxane) and the mixture was stirred for 3 h at  $-35$  °C. Next, a mixture of the deprotected amine-epoxyketone war-head (1.1 equiv, as a trifluoroacetic acid (TFA) salt) and *N,N*-diisopropylethylamine (DiPEA) (5 equiv) in DMF (1 mL) were added. The reaction was slowly warmed to room temperature and stirred for another 12 h before being diluted with DCM and extracted with 1 M HCl (2 $\times$ ), saturated NaHCO<sub>3</sub> (2 $\times$ ) and brine. After drying (MgSO<sub>4</sub>) and concentrating, the obtained crude product was purified by column chromatography, applying a 1% $\rightarrow$ 15% MeOH/DCM eluent system.

### Competition Experiments

Whole cell lysates of HEK-293T were made by sonication in 3 volumes of lysis buffer containing 50 mM Tris pH 7.5, 1 mM dithiothreitol (DTT), 5 mM MgCl<sub>2</sub>, 250 mM sucrose, and 2 mM ATP. Protein concentration was determined by the Bradford assay. Cell lysates (13.5  $\mu$ g total protein) were exposed to the inhibitors for 1 h prior to incubation with **9** (0.5  $\mu$ M) for 1 h at 37 °C. Reaction mixtures were boiled with Laemmli's buffer containing  $\beta$ -mercaptoethanol for 3 min before being resolved on 12.5% SDS-PAGE. In-gel detection of

residual proteasome activity was performed in the wet gel slabs directly on a Typhoon variable mode imager (Amersham Biosciences) using the Cy3/Tamra settings ( $\lambda_{\text{ex}}$  532,  $\lambda_{\text{em}}$  560 nm).

### IC<sub>50</sub> Determinations

Purified 26S proteasome (~10 ng/mL) was incubated with various concentrations of inhibitors at 37 °C for 30 min in the assay buffer (50 mM Tris-HCl, pH 7.5, 40 mM KCl, 2 mM EDTA, 1 mM DTT, 100  $\mu$ M ATP, 50  $\mu$ g/mL BSA). In the meantime, 100  $\mu$ M solution of the fluorogenic peptide substrates (Suc-LLVY-7-amido-4-methyl-coumarin (amc) for the  $\beta$ 5 site, Ac-nLPnLD-amc for the  $\beta$ 1 site, and Ac-RLR-amc or Ac-RQR-amc for the  $\beta$ 2 site) in the assay buffer were pre-incubated at 37 °C. Immediately after the end of this incubation, an aliquot of the inhibitor-treated proteasome was mixed with the substrate, and fluorescence of released amc was measured continuously for 30 min at 37 °C. The rate of reaction was determined from the slope of the reaction progress curves. Mock-treated proteasomes served as control. Residual activity in inhibitor treated samples were plotted against concentration of inhibitors, and IC<sub>50</sub> values were determined from these plots.

### Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

### Acknowledgments

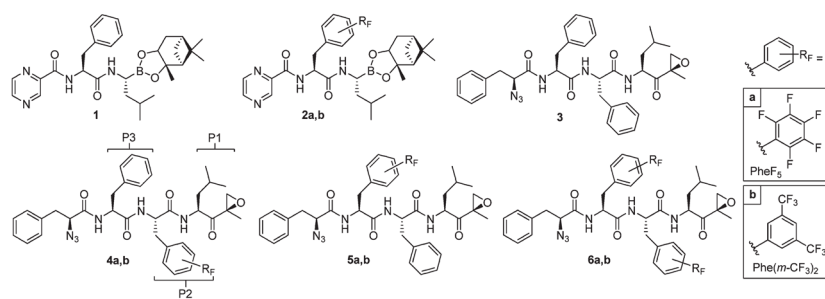
This work was supported by The Netherlands Organization for Scientific Research (NWO), The Netherlands Genomics Centre Initiative (NGI) and The NCI (grant RO1CA124634). We thank Hans van den Elst and Nico Meeuwenoord for HPLC and LC-MS assistance.

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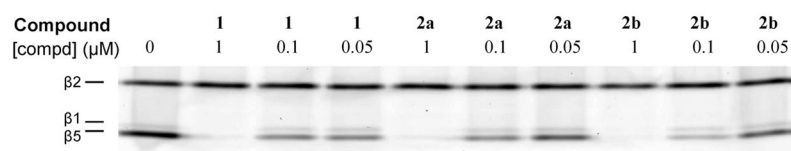
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19. The  $\text{IC}_{50}$  values of compound 7 for each active subunit were determined in the same manner as for the other compounds:  $\beta 1 > 10 \mu\text{M}$ ,  $\beta 2 > 10 \mu\text{M}$ ,  $\beta 5 \ 0.30 \mu\text{M}$ .

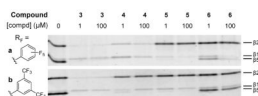


**Figure 1.** Synthesized fluorinated proteasome inhibitors. Indicated are the enzyme pockets (P1, P2, P3).

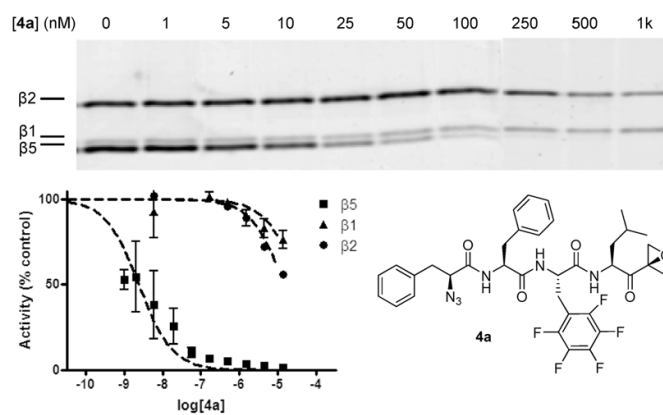


**Figure 2.** Characterization of the specificity of the fluorinated dipeptide boronates. Competition assay in HEK-293 lysate. Lysates were incubated for one hour with compounds **1**, **2a**, and **2b** at the indicated final concentrations. Residual proteasome activity was labeled with  $0.5 \mu\text{M}$  **9** for 1 h.

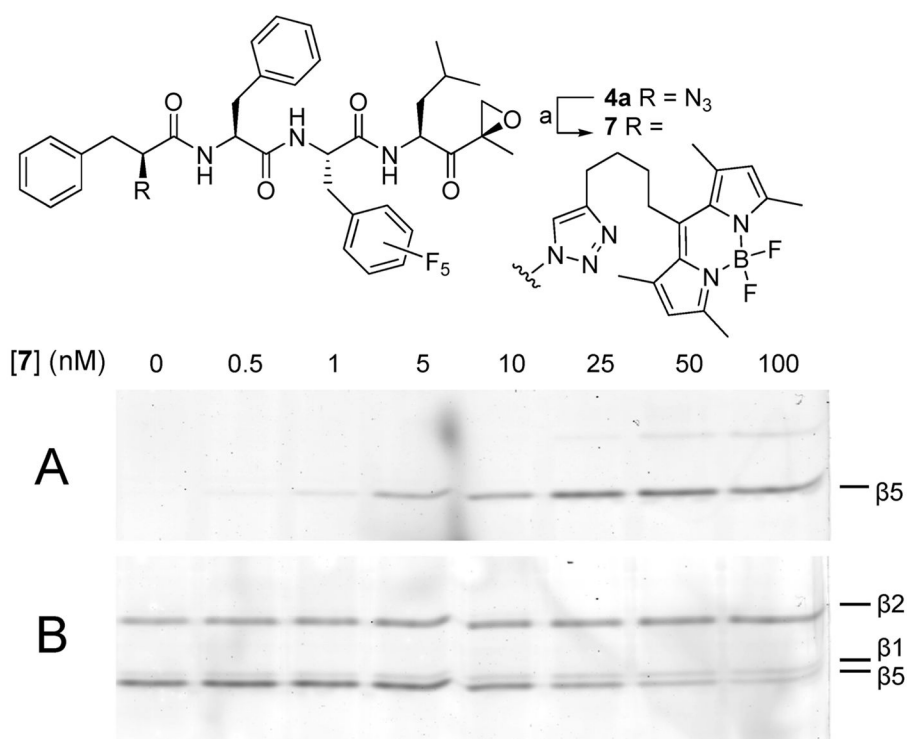




**Figure 3.** Characterization of specificity of fluorinated peptide epoxyketones. Competition assay in HEK-293 lysate. Lysates were incubated for 1 h with compounds **3**, **4**, **5**, or **6** at the indicated final concentrations. Residual proteasome activity was labeled with 0.5  $\mu\text{M}$  **9** for 1 h. N.B. compound **3a** = **3b** = **3**.



**Figure 4.** Characterization of specificity of compound **4a**. Upper panel: Competition assay in HEK-293 lysate. Lysates were incubated for 1 h with compound **4a** at the indicated final concentrations. Residual proteasome activity was labeled with  $0.5 \mu\text{M}$  **9** for 1 h (This image was created from two separate gel images which, together, show a larger concentration series. The original images are shown in Figure S-4 in the SI). Lower panel: Remaining subunit activity after inhibition with compound **4a** determined with fluorogenic peptides.



**Figure 5.** Novel probe for the  $\beta 5$  site. Competition assay in HEK-293 lysate. Lysates were incubated for 1 h with compound **7** at the indicated final concentrations. Residual proteasome activity was labeled with 0.5  $\mu\text{M}$  **9** for 1 h. Fluorescence readout at (A)  $\lambda_{\text{ex}}$  488 nm,  $\lambda_{\text{em}}$  520 nm (compound **7**) and (B)  $\lambda_{\text{ex}}$  532 nm,  $\lambda_{\text{em}}$  560 nm (compound **9**). Reagents and conditions: (a) Bodipy-FL-alkyne,<sup>14</sup> 10 mol % CuSO<sub>4</sub>, 15 mol % sodium ascorbate, toluene/H<sub>2</sub>O/*t*-BuOH 1/1/1, 80 °C, yield 87%.

**Table 1**Activity (IC<sub>50</sub> in  $\mu\text{M}$ ) of **2–5** against the Three Active Constitutive 26S Proteasome Subunits<sup>a</sup>

compd	$\beta$ 1 nLPnLD	$\beta$ 2 RLR	$\beta$ 5 LLVY
<b>2a</b>	0.44	>15	0.031
<b>2b</b>	0.16	11	0.0030
<b>3</b>	>15	1.8	0.0010
<b>4a</b>	>15	>15	0.0020
<b>4b</b>	>15	>15	0.10
<b>5a</b>	>15	>15	0.20
<b>5b</b>	>15	>15	0.13

<sup>a</sup>Determined with the indicated subunit specific fluorogenic peptide substrates. All values are averages of two experiments.