

# Note

## Polymorphic Regions Affecting Human Height Also Control Stature in Cattle

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### ABSTRACT

Orthologous positions of 55 genes associated with height in four human populations were located on the bovine genome. Single nucleotide polymorphisms close to eight of these genes were significantly associated with stature in cattle (*Bos taurus* and *Bos indicus*). This suggests that these genes may contribute to controlling stature across mammalian species.

**U**NDERSTANDING the genetic basis for variation in complex traits has been advanced by many genome-wide association studies (GWAS) conducted in humans (DONNELLY 2008) and, to a lesser extent, in other species (e.g., KARLSSON *et al.* 2007). However, there is still considerable debate on the interpretation of these studies. Most of the single nucleotide polymorphisms (SNPs) associated with complex traits account for a very small proportion of the genetic variance in the trait (VISSCHER 2008). This could be because they are in incomplete linkage disequilibrium (LD) with the causative polymorphism, because the causative polymorphism has a small effect on the trait, or because the minor allele at this polymorphism is rare. The first and third of these explanations tend to apply to the same cases because a rare causative allele cannot be in complete LD with a common allele at a SNP on a standard chip. Other explanations include genotype by environment interactions, epistasis, and allelic heterogeneity. It has even been suggested that most of these associations are artifacts caused by cryptic stratification of the sample of individuals used (McCLELLAN and KING 2010). One of the reasons for skepticism is that there is often no known mechanism linking the genes found by a GWAS to the complex trait with which they are associated.

There are many examples of traits affected by single genes where mutations in the same gene cause a similar

phenotype in different species. Classic examples include genes with a role in pigmentation, such as the *Mc1r* gene responsible for fair hair color in humans (VALVERDE *et al.* 1995) and coat or feather color in species as diverse as horse, pig, and chicken (ANDERSSON 2003). For polygenic traits, where there are many genes with small-to-moderate effect, there is little information on the extent to which the orthologous genes cause variation in different mammalian species. However, there is evidence to suggest a high degree of conservation of certain gene classes among mammalian species, e.g., milk protein genes and mammary genes (LEMAY *et al.* 2009). Evidence that the same genes are involved in controlling a complex trait would be important for several reasons. First, it would act as the ultimate validation study because stratification is unlikely to cause the same artifact in different species. Second, it would be strong evidence for a physiological connection between a gene in LD with an associated SNP and the trait. Third, it would provide some insight into the forces controlling genetic variation in complex traits.

Stature is an easy-to-measure phenotype used as a model complex trait in humans (VISSCHER 2008) and also measured in some domesticated cattle breeds (BARWICK *et al.* 2009). In four separate GWAS of human stature, a total of 58 loci were significantly associated with stature in Caucasians (GUDBJARTSSON *et al.* 2008; LETTRE *et al.* 2008; WEEDON *et al.* 2008) and a further 15 loci have been reported in a Korean population (KIM *et al.* 2010). To test if these loci were controlling stature in cattle, we first identified the positions of orthologous genes that could be matched to Bovine Genome Build 4.0 in the NCBI database (<http://www.ncbi.nlm.nih.gov/projects/genome/guide/cow/>). We were able to suc-

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cessfully map the positions of 55 genes (some genes were close neighbors and were considered to be at the same location). There were 879 SNPs that were 500 kbp either side of these genes on the BovineSNP50 BeadChip (Illumina, San Diego, CA; MATUKUMALLI *et al.* 2009), which comprises around 50,000 approximately equally spaced SNPs. The names of the orthologous genes and their bovine map positions are presented in supporting information, Table S1. We tested these SNPs for their effect on bovine stature in two cattle populations (dairy and beef cattle). The dairy data included 1832 Holstein dairy bulls with estimated breeding values for stature (hip height) based on at least 80 milking daughters per sire. The beef data set comprised hip height on 1224 Brahman and tropical composite beef heifers (*Bos taurus* × *Bos indicus*) measured at the end of the first post-weaning wet season when the heifers were aged ~18 months (BARWICK *et al.* 2009). We observed a skew in the quantile-quantile plot (see Figure 1), which indicates that more SNPs are associated with stature than would be expected by chance. Of 879 SNPs tested, 10 and 12 were associated with stature ( $P < 0.001$ ) in dairy and beef data sets, respectively (Table 1).

We used a randomized permutation test to evaluate how likely it was to retrieve this number of SNPs with  $P < 0.001$  by chance. We repeated the tests between bovine SNPs and stature by selecting 55 genes at random (SNPs in the 1-Mbp region surrounding each gene) and counted the number of associations that were  $P < 0.001$ . Figure 2 shows the distribution of 10,000 random tests (for each data set). Only 0.19% and 0.38% of the randomized permutation tests in beef and dairy cow data, respectively, had fractions equal to or exceeding the results observed for stature. Therefore, it is unlikely that the results reported here for stature in dairy and beef cattle arose by chance. Note that the genes identified as stature orthologs were included in the data set used for the randomized permutation test. However, the same pattern of results was observed when the stature orthologs were excluded.

Of 55 orthologous genes tested, the SNPs that were associated with stature in cattle were close to 10 genes and 8 genomic regions (Table 1). Six of these had significant SNPs in either beef or dairy cattle, while significant SNPs in both dairy and beef cattle were observed in two regions containing the gene *NCAPG* and a cluster on chromosome 14 (*PLAG1*, *CHCHD7*, and *RDHE2*). *NCAPG* has previously been associated with fetal growth (EBERLEIN *et al.* 2009) and carcass size (SETOGUCHI *et al.* 2009) in cattle and is thought to have a role in cell division (MURPHY and SARGE 2008). The strongest signal in our study was observed for a SNP close to *HMG2A2* in the beef data set ( $2.7 \times 10^{-10}$ ). This gene has consistently been found to be associated with stature in human studies (VISSCHER 2008), and it is interesting that it seems likely that it has a conserved

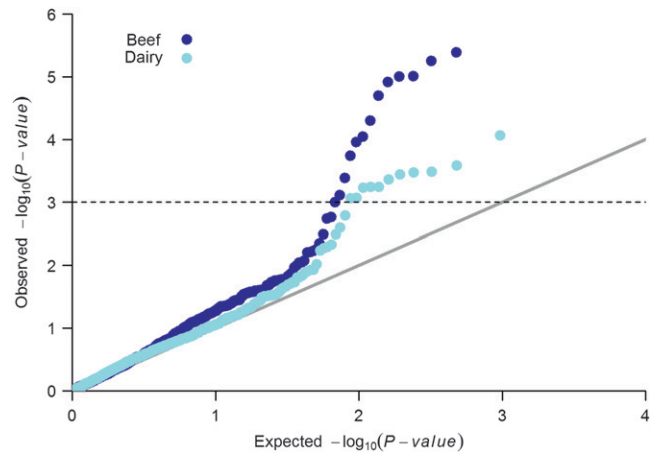


FIGURE 1.—Quantile-quantile plot of  $P$ -values of 879 SNPs that were 500 kbp either side of 55 orthologous genes found to be associated with height in human populations (LETTRE *et al.* 2008; GUDBJARTSSON *et al.* 2008; WEEDON *et al.* 2008; KIM *et al.* 2010). Using the dairy and beef data sets, the phenotype (stature) was regressed on each SNP by using a mixed model that included pedigree (ASReml, GILMOUR *et al.* 2006) and the same approach as PRYCE *et al.* (2010). The model used was  $y = I_n \mu + Wg + Zu + e$ , where  $y$  is the vector of phenotypes,  $I_n$  is a vector of 1s of length  $n$  animals,  $W$  is a vector allocating SNP effects where each SNP was fitted individually,  $g$  is the fixed effect of the SNP,  $u$  is the vector of polygenic residual breeding values not accounted for by the effect of the SNP sampled from the distribution  $N(0, A\sigma_a^2)$  where  $A$  is the additive relationship matrix constructed from the pedigree,  $\sigma_a^2$  is the additive genetic variance, and  $e$  is the vector of random deviates. The purpose of including pedigree is to control for relationships and population structure. Observed and expected  $P$ -values would fall on the gray solid line if there were no association. The dashed horizontal line is the threshold selected for significance ( $P < 0.001$ ). Note that a 1-Mbp window was used from which to select SNPs because, in contrast to humans, where LD is expected to persist over only 10s of kilobase pairs (TENESA *et al.* 2007), non-zero levels of LD have been observed up to 1 Mbp in cattle (BOVINE HAP-MAP CONSORTIUM 2009).

role in cattle, too. There were only two SNPs that had associations with stature in both beef and dairy cattle data sets. This is not surprising as the SNP density of the Illumina BovineSNP50 BeadChip is insufficient to expect the phase of LD to persist across breeds as diverse as beef and dairy cattle (DE ROOS *et al.* 2008).

These results provide a unique confirmation of the significant associations found in human GWAS for height. The power to detect associations in the cattle experiments described is not high (there were <2000 animals in each study) so associations with all the SNPs tested is not expected. Furthermore, the physiological control of height is probably not identical in the two species. However, by using a series of 10,000 randomized permutation tests, we were able to show that achieving the number of associations listed in Table 1 by chance would be very unlikely (Figure 2). Together, these results add considerable weight to the conclusion that several genes identified in human GWAS for

TABLE 1

Dairy and beef cattle SNPs associated with stature within 500 kbp of genes associated with human height

Gene associated with height in humans	Bovine chromosome	Orthologous start and stop position of gene (bp) in cattle	Position of significant SNPs (bp)	Dairy		Beef	
				P-value	Effect size (%) <sup>a</sup>	P-value	Effect size (%) <sup>a</sup>
<i>HMGA2</i> <sup>b,c,d</sup>	5	51,740,850–51,890,260	51,770,120	0.69	0.01	$2.7 \times 10^{-10}$	<b>4.03</b>
			52,214,922	0.72	0.00	$2.0 \times 10^{-5}$	<b>1.38</b>
<i>LCORL/NCAPG</i> <sup>c,d,e</sup>	6	38,153,047–38,199,154	38,256,889	0.05	0.34	$1.1 \times 10^{-4}$	<b>0.97</b>
			38,326,147	0.45	0.07	$5 \times 10^{-5}$	<b>1.07</b>
			38,479,643	<b><math>5.68 \times 10^{-4}</math></b>	<b>0.40</b>	0.76	0.01
			38,500,209	<b><math>5.68 \times 10^{-4}</math></b>	<b>1.53</b>	0.76	0.01
			38,558,526	<b><math>2.57 \times 10^{-4}</math></b>	<b>0.50</b>	0.27	0.09
<i>FBP2</i> <sup>d</sup>	8	85,344,905–85,387,652	84,943,316	<b><math>8.61 \times 10^{-4}</math></b>	<b>0.06</b>	0.01	0.41
<i>PTCHI</i> <sup>e</sup>	8	86,551,192–86,621,018	86,576,819	0.60	0.03	$9.9 \times 10^{-4}$	<b>0.70</b>
<i>PAPPA</i> <sup>b</sup>	8	110,722,545–110,982,014	111,423,182	<b><math>8.53 \times 10^{-4}</math></b>	<b>1.86</b>	0.66	0.01
<i>GPR126</i> <sup>b</sup>	9	82,555,167–82,704,883	82,609,868	0.59	0.02	$7.7 \times 10^{-4}$	<b>0.87</b>
<i>PLAG1, CHCHD7, RDHE2</i> <sup>b,d,e</sup>	14	23,219,718–23,221,723 <sup>PLAG1</sup>	22,720,374	0.19	0.33	$1.2 \times 10^{-5}$	<b>1.47</b>
			22,768,981	0.02	1.14	$9.7 \times 10^{-6}$	<b>1.50</b>
			22,803,367	<b><math>3.27 \times 10^{-4}</math></b>	<b>2.53</b>	$9.9 \times 10^{-6}$	<b>1.50</b>
			22,838,802	0.11	0.55	$4.1 \times 10^{-6}$	<b>1.62</b>
			23,365,427–23,398,159 <sup>RDHE2</sup>	0.19	0.29	$9.0 \times 10^{-5}$	<b>1.26</b>
<i>CABLES1</i> <sup>d,e</sup>	24	34,619,497–34,683,911	23,519,449	<b><math>8.63 \times 10^{-5}</math></b>	<b>2.83</b>	$5.5 \times 10^{-6}$	<b>1.66</b>
			34,436,971	<b><math>4.35 \times 10^{-4}</math></b>	<b>0.55</b>	0.08	0.24
			34,457,809	<b><math>5.86 \times 10^{-4}</math></b>	<b>1.47</b>	1.00	0.00
			34,637,479	<b><math>3.59 \times 10^{-4}</math></b>	<b>0.57</b>	0.22	0.12

P-values that are < 0.001 are shown in bold-face; P-values that are > 0.001 are shown in non-bold-face

<sup>a</sup> Effect size (percentage of variation explained)  $(2pq\beta^2)/V_A$ , where  $p$  and  $q$  are allele frequencies,  $\beta$  is the SNP solution estimate, and  $V_A$  is the genetic variance of stature.

<sup>b</sup> LETTRE *et al.* (2008).

<sup>c</sup> WEEDON *et al.* (2008).

<sup>d</sup> KIM *et al.* (2010).

<sup>e</sup> GUDBJARTSSON *et al.* (2008).

stature also have a conserved role in the physiology of growth in cattle, including a role in regulating cell division and cell cycle.

If many complex traits have a similar architecture in different species, then humans could in fact be used as a model for identifying genes for complex traits in non-

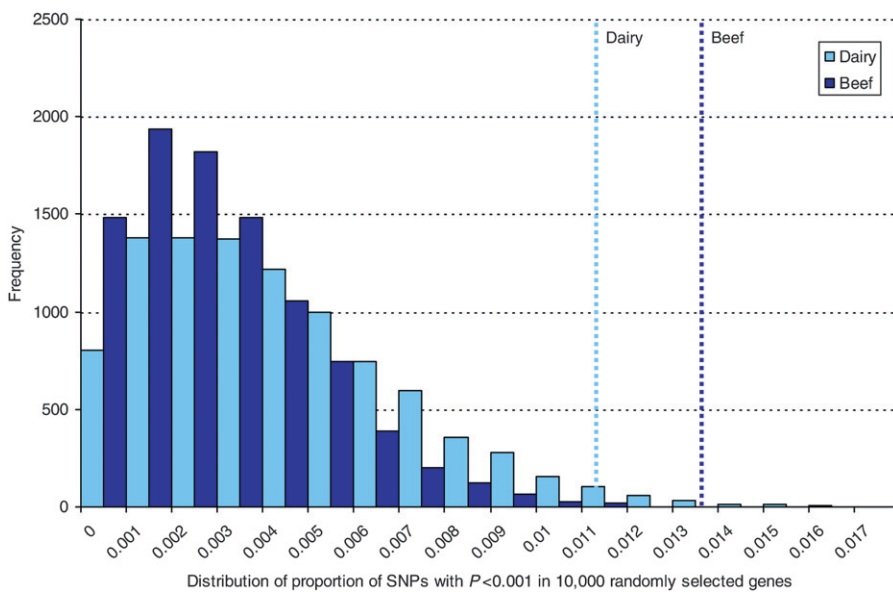


FIGURE 2.—A frequency distribution of the proportion of SNPs with a  $P < 0.001$  effect on stature in dairy and beef cattle data sets in 10,000 control tests. To make the tests comparable to the results reported in Figure 1, 55 genes were randomly selected from 16,850 bovine genes that were orthologs of human genes downloaded from BioMart (<http://www.ensembl.org/biomart/index.html>). The SNPs for the random control tests were in a 1-Mbp region centered on each of the 55 genes. The vertical lines represent the number of bovine SNPs with  $P < 0.001$  association with stature in each data set. The percentage of control tests with an equivalent or greater proportion of significant SNPs associated with stature in the beef and dairy cow data was 0.19 and 0.38%, respectively. For each test, the number of SNPs significantly ( $P < 0.001$ ) associated with stature was counted. The number of significant SNPs divided by the number of SNPs tested is represented along the x-axis.

model species such as primates, marsupials, and cetaceans. Comparing GWAS across species, as done here, will aid understanding of gene pathways that contribute to complex traits.

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# GENETICS

## Supporting Information

<http://www.genetics.org/cgi/content/full/genetics.110.123943/DC1>

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TABLE S1

**Orthologous bovine genome positions of genes associated with stature in humans (Lettre *et al.* 2008; Gudbjartsson *et al.* 2008; Weedon *et al.* 2008; Kim *et al.* 2010). Also shown are the number (N) of bovine SNP in the 1 Mbp region surrounding each gene and the number of SNP that were significant ( $P < 0.001$ ) (the gene is highlighted in bold) in dairy and beef cattle.**

**Genes that are italicized have SNP that overlap with neighboring genes**

Gene	Chromosome	Gene start position (bp)	Gene end position (bp)	SNP (N)	P<0.001 dairy (N)	P<0.001 beef (N)
ZBTB38	1	129,739,602	129,746,475	22	0	0
ANAPC13	1	137,198,394	137,227,980	19	0	0
IHH	2	111,402,259	111,408,032	23	0	0
MTMR11	3	22,211,138	22,219,111	8	0	0
SPAG17	3	27,351,775	27,516,376	15	0	0
<b>CDK6 inhibitor</b>	<b>3</b>	<b>102,130,348</b>	<b>102,135,037</b>	<b>18</b>	<b>1</b>	<b>1</b>
SCMH1	3	112,037,874	112,166,384	21	0	0
<i>PEX1</i>	<i>4</i>	<i>10,032,603</i>	<i>10,114,126</i>	<i>23</i>	<i>0</i>	<i>0</i>
<i>CDK6</i>	<i>4</i>	<i>10,182,420</i>	<i>10,430,210</i>	<i>26</i>	<i>0</i>	<i>0</i>
WDR60	4	123,821,776	123,867,079	18	0	0
SOCS2	5	26,159,649	26,164,528	14	0	0
<b>HMGA2</b>	<b>5</b>	<b>51,740,850</b>	<b>51,890,260</b>	<b>8</b>	<b>0</b>	<b>2</b>
<i>NUP37</i>	<i>5</i>	<i>70,834,703</i>	<i>70,886,595</i>	<i>11</i>	<i>0</i>	<i>0</i>
<i>IGF1</i>	<i>5</i>	<i>71,126,213</i>	<i>71,198,012</i>	<i>11</i>	<i>0</i>	<i>0</i>
<b>NCAPG</b>	<b>6</b>	<b>38,153,047</b>	<b>38,199,154</b>	<b>21</b>	<b>3</b>	<b>2</b>
BMP2	6	96,943,071	97,033,845	24	0	0
DOT1L	7	20,061,134	20,113,023	20	0	0
FREM1	8	31,056,507	31,056,507	15	0	0
<b>FBP2</b>	<b>8</b>	<b>85,344,905</b>	<b>85,387,652</b>	<b>16</b>	<b>1</b>	<b>0</b>
<b>PTCH1</b>	<b>8</b>	<b>86,551,192</b>	<b>86,621,018</b>	<b>14</b>	<b>0</b>	<b>1</b>
ZNF462	8	101,706,529	101,860,713	23	0	0
PALM2-AKAP2	8	104,377,441	104,760,542	28	0	0
<b>PAPPA</b>	<b>8</b>	<b>110,722,545</b>	<b>110,982,014</b>	<b>25</b>	<b>1</b>	<b>0</b>
<b>BAT3/HLA class 3</b>	<b>9</b>	<b>35,902,575</b>	<b>35,902,575</b>	<b>23</b>	<b>0</b>	<b>1</b>
LIN28B	9	47,121,302	47,243,468	10	0	1
<b>GPR126</b>	<b>9</b>	<b>82,555,167</b>	<b>82,704,883</b>	<b>20</b>	<b>0</b>	<b>1</b>
EFEMP1	11	39,991,035	40,060,587	14	0	0
FUBP3	11	104,601,087	104,650,684	22	0	0
DLEU7	12	19,520,388	19,633,990	12	0	0
BMP2	13	49,391,620	49,402,920	9	0	0
<i>UQCC</i>	<i>13</i>	<i>65,156,129</i>	<i>65,239,080</i>	<i>22</i>	<i>0</i>	<i>0</i>
<i>GDF5</i>	<i>13</i>	<i>65,264,274</i>	<i>65,268,030</i>	<i>21</i>	<i>0</i>	<i>0</i>
LTBP1	14	16,297,282	16,520,432	20	0	0
<b>PLAG1</b>	<b>14</b>	<b>23,219,718</b>	<b>23,221,723</b>	<b>15</b>	<b>2</b>	<b>6</b>
<b>CHCHD7</b>	<b>14</b>	<b>23,265,198</b>	<b>23,271,074</b>	<b>15</b>	<b>2</b>	<b>6</b>
<b>RDHE2</b>	<b>14</b>	<b>23,365,427</b>	<b>23,398,159</b>	<b>15</b>	<b>1</b>	<b>2</b>
PXMP3	14	37,070,697	37,085,976	14	0	0

EXT1	14	43,445,832	43,759,712	27	0	0
GLT25D2	16	62,449,914	62,629,806	21	0	0
HHIP	17	14,401,494	14,504,964	16	0	0
ANKFN1	19	6,254,001	6,410,288	25	0	0
NOG	19	6,520,420	6,520,961	18	0	0
TBX4	19	10,865,638	10,892,047	15	0	0
BCAS3	19	10,957,337	11,543,629	21	0	0
CRLF3	19	17,923,506	17,985,486	25	0	0
KRT23	19	42,381,026	42,398,584	12	0	0
ADAMTS17	21	5,065,926	5,481,345	19	0	0
ACAN	21	20,147,595	20,216,259	23	0	0
ADAMTSL3	21	23,953,056	24,232,435	21	0	0
SH3GL3	21	24,247,955	24,303,732	19	0	0
<i>TRIP11</i>	21	57,166,325	57,172,931	9	0	0
<i>ATXN3</i>	21	57,269,545	57,289,440	12	0	0
<i>HMGA1</i>	23	8,601,043	8,609,914	12	0	0
<i>C6orf106</i>	23	8,916,789	9,013,752	15	0	0
ANKS1	23	9,152,009	9,152,818	16	0	0
SUPT3H	23	18,888,191	19,284,659	20	0	0
HLA class III	23	27,188,360	27,191,071	16	0	0
HIST1H1D	23	31,746,873	31,750,532	14	0	0
BMP6	23	48,405,475	48,435,730	24	0	0
<b>CABLES1/RBBP8</b>	<b>24</b>	<b>34,619,497</b>	<b>34,683,911</b>	<b>21</b>	<b>3</b>	<b>0</b>
DYM	24	50,390,159	50,791,554	17	0	0
GNA12	25	42,426,379	42,501,860	19	0	0

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**FILE S1**  
**SUPPORTING DATA**

File S1 is available for download as a compressed (.zip) file at <http://www.genetics.org/cgi/content/full/genetics.110.123943/DC1>.