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## Arylethynyl receptors for neutral molecules and anions: emerging applications in cellular imaging†

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### Abstract

This *critical review* will focus on the application of shape-persistent receptors for anions that derive their rigidity and optoelectronic properties from the inclusion of arylethynyl linkages. It will highlight a few of the design strategies involved in engineering selective and sensitive fluorescent probes and how arylacetylenes can offer a design pathway to some of the more desirable properties of a selective sensor. Additionally, knowledge gained in the study of these receptors in organic media often leads to improved receptor design and the production of chromogenic and fluorogenic probes capable of detecting specific substrates among the multitude of ions present in biological systems. In this ocean of potential targets exists a large number of geometrically distinct anions, which present their own problems to the design of receptors with complementary binding for each preferred coordination geometry. Our interest in targeting charged substrates, specifically how previous work on receptors for cations or neutral guests can be adapted to anions, will be addressed. Additionally, we will focus on the design and development of supramolecular arylethynyl systems, their shape-persistence and fluorogenic or chromogenic optoelectronic responses to complexation. We will also examine briefly how the “chemistry in the cuvet” translates into biological media.

### Introduction

This *critical review* highlights the role functionalized ethynylarenes play in supramolecular sensor design. We begin with a non-comprehensive survey of representative examples of ethynyl- and butadiynyl-based receptors for neutral and biologically relevant guests and the translation of these systems into sensors for anionic species. We conclude with an overview of related work in our lab and provide a brief survey of the burgeoning field of arylethynyl probe development for biologically relevant anions.

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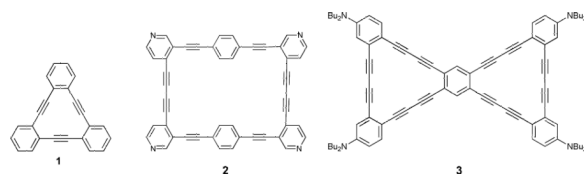
## 1.1 Arylethynyl receptors

Many synthetic receptors rely on some degree of preorganization in their approach to achieving high affinities and selectivities. Flexible receptors can achieve increased size-selectivity and preorganization through macrocyclization, and numerous, in-depth reviews have been written which cover the usefulness of the macrocyclic effect in increasing binding affinities.<sup>1-4</sup> However, macrocyclic receptors tend to exhibit slow binding kinetics<sup>1,2</sup> and indeed many sensor applications rely more upon kinetic selectivity rather than a large contribution from preorganization of the receptor. In addition it can be the case that the guest bound most strongly is not the guest that gives rise to the largest colorimetric or fluorometric response, which is difficult to predict. This gives rise to a delicate balance in designing probes capable of binding a target selectively, while also exhibiting a response for that specific analyte.

There exist many examples of conformationally rigid receptors,<sup>5-7</sup> most of which exploit the inherent rigidity in conjugated  $\pi$ -systems. Expanded porphyrins,<sup>8,9</sup> calixpyrroles or related calixarenes,<sup>10-12</sup> and innumerable other nitrogen<sup>13,14</sup> or oxygen containing heterocycles<sup>15</sup> have been synthesized and their affinities for both ionic and neutral guests studied. In much the same fashion phenylacetylenes have provided structure and optoelectronic handles to a multitude of coordination and host-guest complexes.<sup>16</sup> Macrocyclic host molecules as well as shape-persistent acyclic ligands for metal ions have benefited from their linear, rigid geometries and relatively simple derivatization.<sup>17-20</sup>

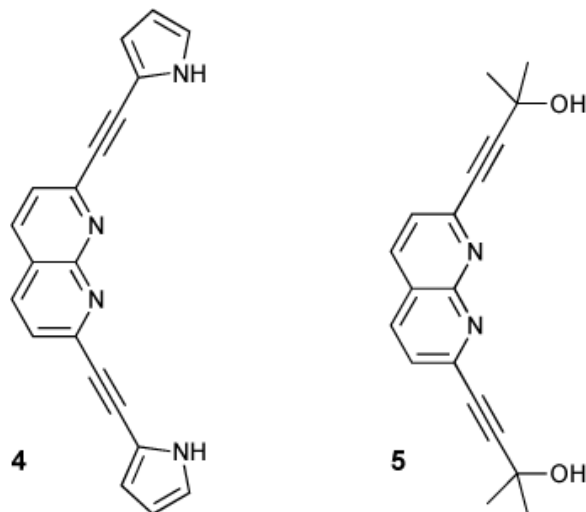
In some cases the acetylenes themselves have served as the binding site for transition metal guests.<sup>21</sup> The synthesis and characterization of a  $\text{Ni}^0$  complex of dehydroannulene **1** was reported in 1985.<sup>22</sup> More often, however, the rigid acetylenic linker serves to reinforce a desired binding conformation. The synthesis and characterization of phenylacetylenic macrocycles capable of differentiating transition metals as well as serving as rudimentary proton sensors has also been achieved.<sup>23-25</sup> Twistophane **2** was shown to signal  $\text{Pd}^{\text{II}}$  and  $\text{Hg}^{\text{II}}$  by a distinct fluorescence quenching response, and to signal  $\text{H}^+$  by bathochromic shifting and quenching of the fluorescence emission.<sup>23</sup>

Phenylacetylenes have well-studied fluorescence emission properties,<sup>26, 27</sup> and the optoelectronic response to perturbation of their ground-state conformations can be a useful spectroscopic handle. The conjugation of donor and acceptor groups via alkyne linkages<sup>28</sup> is much studied and this charge transfer process has been used as a fluorescent handle for the visualization of binding events.<sup>29</sup> Dibutylamine-functionalized dehydrobenzoannulene **3** was found to shift fluorescence emission based upon  $\text{H}^+$  concentration (as trifluoroacetic acid). Interestingly, it was found that emission shifting was dependent upon stepwise protonation of the dibutylaniline moieties, which indicated independent manipulation of the frontier molecular orbital energies and thus tunable charge transfer pathways.



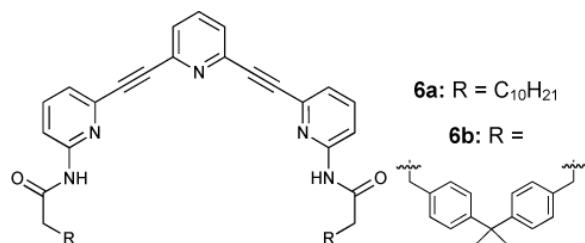
In 2002 ethynyl-linked pyrrole-naphthyridine compound **4** was found to selectively bind glucopyranoside.<sup>30</sup> The free receptor was found to adopt a slightly twisted conformation that exhibited a fluorescence emission maximum at 475 nm ( $\tau_f \approx 1.25$  ns), which decreased upon addition of octyl  $\beta$ -D-glucopyranoside (OGU). A new emission band at 535 nm ( $\tau_f \approx 0.95$  ns) grew in intensity as a 1:1 complex was formed. The association constants for this complex determined by both fluorescence and UV-Vis absorbance measurements were in

good agreement ( $K_a = 5.3 \times 10^3 \text{ M}^{-1}$  and  $K_a = 4.8 \times 10^3 \text{ M}^{-1}$  in  $\text{CH}_2\text{Cl}_2$ , respectively). Interestingly, the association constant for octyl  $\beta$ -D-galactopyranoside was found to be only  $1800 \text{ M}^{-1}$ , which is impressive considering these saccharides differ only in the orientation of the 4-hydroxyl group. The optoelectronic response in this system was attributed to a rigidification and planarization of the pyrrole-naphthyridine moieties, which served to enhance the charge transfer in this D- $\pi$ -A-A- $\pi$ -D system.



To test this an analogous receptor **5** was synthesized in which the binding sites were not conjugated through the core. Binding of OGU to this receptor was weaker due to the relative differences in acidity at the binding sites, but fluorescence emission intensity still increased, albeit not bathochromically shifted as could be expected by rigidification in this system. Additionally, modification of the pyrrolyl moieties of **4** to indolyl units gave the normally CD-silent receptor strong fluorescence-detected CD spectra upon binding, due to chirality transfer from the substrates.<sup>31</sup>

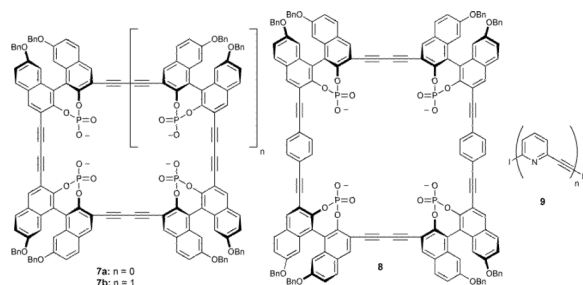
A simple illustration of how flexible macrocyclization of rigid arylolethynyl scaffolds can increase binding constants is found in two pyridine based systems **6a** and **6b**.<sup>32</sup> Acyclic **6a** bound ribofuranosides poorly in  $\text{CDCl}_3$  ( $K_a = 30 \text{ M}^{-1}$ ) but macrocyclization of this cleft-like receptor increased binding constants two orders of magnitude ( $K_a = 2400 \text{ M}^{-1}$ ). Related poly(ethyleneglycol) linked derivatives of this receptor motif allowed the expanded terpyridine core to adopt a slightly wider conformation while maintaining preorganization.<sup>32a</sup> Due to this longer, more flexible linker these receptors exhibited still higher affinities for larger monosaccharides.



A number of phosphorylated binol-based macrocycles have been investigated as receptors for neutral guests, such as saccharides.<sup>33-35</sup> The polyanionic cavities in receptors **7a** and **7b** were targeted at the hydroxyl moieties of the sugars, a strategy which proved effective even in slightly competitive media. It was found that macrocyclization can be a hindrance in these

very rigid systems; cyclotrimer **7a** was found to be incapable of binding monosaccharides in the cavity, but was still capable of binding OGU ( $K_a = 3500 \text{ M}^{-1}$  in  $\text{CD}_3\text{CN}$ ) ostensibly through a face-to-face interaction.<sup>34</sup> However, cyclotetramer **7b** was large enough to bind pyranose in its cavity, with a corresponding increase in association constant ( $K_a = 4500 \text{ M}^{-1}$  for OGU), while extended cyclotetramer **8** was selective for disaccharides over monosaccharides even in very competitive media ( $K_a$ 's of 10,700 – 12,500  $\text{M}^{-1}$  in 88:12  $\text{CD}_3\text{CN}$ - $\text{CD}_3\text{OD}$ ).<sup>35</sup> The rigidity of the macrocycles in these cases helped to reinforce their size selectivity between guests.

This size regulation can also be extended to polymers of phenylacetylenic subunits. Polymers of **9** were found to exhibit saccharide-dependent induction of chirality.<sup>36</sup>



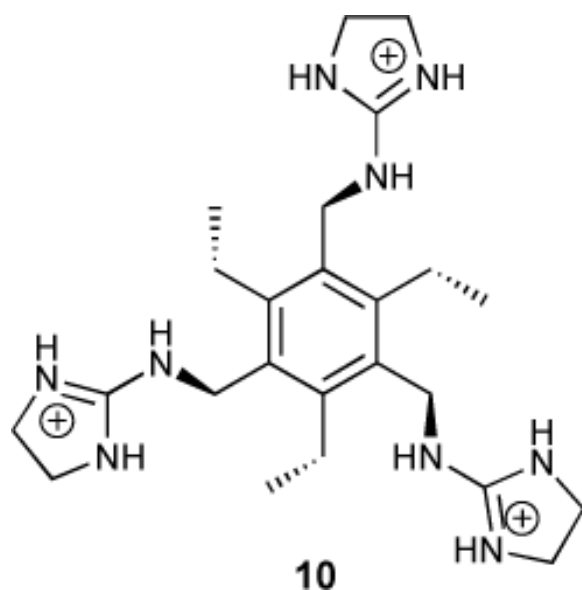
Hydrogen bonding with saccharide guests was reinforced by the rigidity and relative angle of the alkyne linkers between pyridine units, which necessarily directed all of the hydrogen-bond accepting lone pairs to the interior of the cavity. This reinforcement also biased the polymer for the 2,3,4,6-OH groups of  $\beta$ -glucoside from other monosaccharides or their derivatives. The above examples, while by no means exhaustive, illustrate a few of the design strategies involved in engineering a selective and sensitive fluorescent probe, and how phenylacetylenes offer one such viable design pathway.

## 1.2 Approaches to receptor design

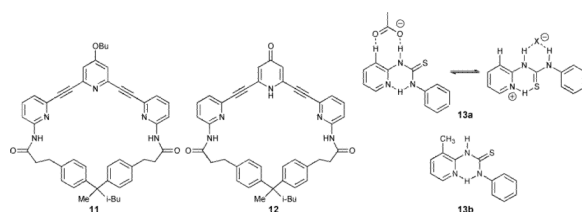
A viable non-covalent receptor must strike a balance between selectivity, solubility, robustness and, in the case of sensors, signalling or response.<sup>1-3</sup> There are a few distinct strategies for designing effective non-covalent, fluorionophores. These can be classified simply as either “FSR” (fluorophore-spacer-receptor) or displacement.<sup>37-39</sup>

In the FSR system, an independently tunable binding site is linked covalently to a signalling unit. The role of the signalling unit is simply to transduce the chemical information upon coordination from the binding site into a specific fluorescent response. This historically has been the most common approach to engineering fluorogenic hosts to complex ionic targets, and many reviews exist.<sup>37,39</sup> There has been considerable success with what could be called a noncovalent version of FSR.<sup>38</sup> This dye-displacement method relies upon careful modulation of a bound fluorophore or chromophore that is designed to have a weaker affinity for the host than the target analyte.

In 1998 there was reported the synthesis of the hexa-substituted benzene-based tripodal receptor **10** that used three guanidinium groups as binding sites.<sup>40</sup> Carboxyfluorescein was found to bind with a  $K_a = 4.7 \times 10^3 \text{ M}^{-1}$  in aqueous buffer, while citrate bound with  $K_a = 2.9 \times 10^5 \text{ M}^{-1}$ . Modulation of the  $\text{p}K_a$  of the phenolic proton of carboxyfluorescein was implicated in the decreased fluorescence of free dye versus the host-dye complex.



Alternatively, replacement of a non-conjugated spacer with a conjugated linker has proven an interesting approach to receptor design. It allows for discrete modulation of the optoelectronic response of a fluorogenic receptor via conformation rather than just electronic effects from guest inclusion, as well as rigidifying the receptor. This provides an element of size-based recognition and can be exploited to modulate the fluorescent response, either through twisting of the  $\pi$ -system<sup>41-43</sup> or extension of excitation/emission into the near-IR for use in biological systems.<sup>44</sup> Regardless of the desired properties, this approach is closely related to traditional FSR receptors, although in this instance the spacer plays a more active role in enforcing receptor conformation. Integration of these components can access smaller molecular sensing systems as well as provide additional information about the binding geometry via modulation of optoelectronic response.



### 1.3 Modularity

Convergent syntheses are a common route in molecule assembly. The well-studied techniques of arene-acetylene cross-coupling can allow the supramolecular chemist to synthesize and derivatize a wide array of functional subunits with spacers that transduce signal themselves, rather than separating these into independent moieties in the receptor.<sup>45</sup> These can then be easily linked together in a convergent fashion through a multitude of cross-coupling reactions, most of which are characterized by their relatively benign reaction conditions.<sup>46</sup> This modular approach to synthesis allows for quick and effective screening of candidates by allowing quick and subtle changes to each building block. Most of the phenylacetylene work mentioned thus far has made use of sequential cross-couplings of independently synthesized subunits, and the budding field of alkyne metathesis may open up another efficient and modular strategy in the synthesis of ethynyl or butadiynyl-containing small molecules.<sup>47,48</sup> In tuning binding parameters such as cavity size, bite angle or the number or relative position of binding sites, secondary properties (i.e., optoelectronic

response, photostability or solubility) of the receptor scaffold can be adjusted as well, and simplicity when modulating all of these parameters is key to quick and efficient discovery of “Goldilocks” candidates for viable receptors.

#### 1.4 Switchability in Sensitivity

Controlling the affinity of a host for a specific analyte via allosteric modulation (e.g., pH or ionic environment) is an interesting tool for modifying the selectivity of a binding site.<sup>49,50</sup> This allosteric behaviour is relatively rare in synthetic receptors, but has particular implications for designing biomimetic receptors that will operate within the diminished pH windows in biological systems, where either “turn-on” or “turn-off” binding may be desired in response to pH changes in cellular compartments. Notably, switchable conformational control has been shown in rotaxanes<sup>51</sup> and in “FSR” type hydrogen-bonding receptors.<sup>52,53</sup> As an example, the switchable complexation of uracil in two amine-triazine-crown ether receptors has been recently reported.<sup>52</sup> Protonation of the amine initiated an intramolecular hydrogen-bonding interaction between the ammonium moiety and the crown ether that sterically blocked the triazine binding site.

Switchability in a sensor can also come from conformational change in induced-fit sensors. In these cases clever design can limit the conformational degrees of freedom such that only a single analyte gives the most intense response. For example, this can be accomplished by appending fluorophores such that conformational change upon binding brings them into proximity for excimer formation (e.g., FRET)<sup>54</sup> or redox active groups can have their environments changed in such a way as to give rise to a known modulation in potential for non-optical sensing.<sup>55,56</sup> Phenylacetylenic systems can be exploited when engineering optically responsive receptors in much the same way as discussed below.

Extended bis-ethynylpyridine compounds **11** and **12** were reported to bind deoxyribosides in  $\text{CDCl}_3$  ( $K_a = 690 \text{ M}^{-1}$  for **11** and  $K_a = 19000 \text{ M}^{-1}$  for **12**).<sup>57</sup> The modification of the hydrogen bond accepting pyridine lone pair to a hydrogen bond donating pyridinone was responsible for this significant increase in affinity. This switchability, although engineered into the receptor during the synthesis, has interesting implications for the design of receptors capable of binding anionic guests.

As an example, pyridylthiourea **13a** exhibits switchable affinities for either acetate or halide anions.<sup>58</sup> Protonation as a switch in this case provides an additional hydrogen bonding contact which further stabilizes the spherical halides. Treating the protonated receptor with excess acetate led to deprotonation and binding of the residual acetate anion. Replacing the arene C-H with a methyl group yielded **13b**, which had no affinity for acetate or hydrogen halides, thus highlighting the role arene C-H hydrogen bonding can play in designing receptors.

#### 2.1 Tuning Receptors for Anionic Targets

As anions have become the focus of more and more research efforts, the modification and application of known cation receptor design criteria for anionic targets has grown as well. For many years the importance of anions in the natural world was overlooked. Relatively benign chloride, carbonate and sulfate, as well as toxic arsenate anions, are found naturally in water the world over,<sup>61</sup> or in runoff and acid rain in the case of industrial pollutants.<sup>62</sup> Anthropogenic anions can be expanded to include phosphate and nitrates from agriculture,<sup>63</sup> or pertechnetate and perchlorate from industrial waste streams.<sup>64,65</sup> Their ubiquity in the natural world has led to an increase of research in this field with modest advances made in the characterization and sequestration of electron-rich wastewater contaminants. This has led



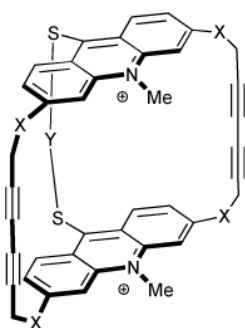
to elegant designer nanomaterials that offer selectivity for some of these myriad problematic anionic pollutants present in both developing and developed countries.<sup>66,67b</sup>

In spite of their prevalence, targeting anionic substrates has its own inherent challenges. Their low charge to radius ratio, high solvation energies, polarizability and the large range of preferred geometries (or lack thereof) make them difficult targets in competitive media. <sup>1-3</sup> Protic solvents tend to form extremely stable hydrogen bonds with most anions, which mandates an extremely strong host-guest interaction if any binding is to be accomplished. On the other hand, relatively nonpolar solvents give rise to ion-pairing, which can have a significant negative influence on binding ability. In addition, many biological anions exist only within narrow pH windows. Nature has many strategies to overcome these difficulties, a fact illustrated beautifully by the complexity of many of the natural receptors whose function depends upon their selectivity for specific anions. A particularly elegant example was the elucidation of the StCIC and EcCIC chloride ion channels via X-ray crystallography in 2002.<sup>68</sup> These CIC ion channels are selective for Cl<sup>-</sup> and Br<sup>-</sup> through partial positive charges within the channels, rather than a fully electrostatic interaction with the nearby lysine and arginine residues which would bind too strongly and inhibit the function of the channel. Interestingly, the gating mechanism necessary for the channel to function is hypothesized to depend on two chloride binding events, and that even small changes in the anionic substrate, i.e., from chloride to bromide anion, alter the operational efficiency of the channel.<sup>69</sup> In spite of insights such as these into the complexity of Nature's use of anion coordination chemistry in cell regulation, our understanding of how the properties of specific anions affect changes in these systems is severely limited.

## 2.2 Common functionalities for anion receptors and probes

This review focuses on receptors with linked sp-carbon atoms to which are appended a variety of functional groups: arenes such as pyrrole, indole, carbazole, or carbonyl-containing amide, urea, thiourea or sulfonamide functionalities.<sup>78</sup> This combination can translate into well-defined binding sites, whether they be based strictly on polycyclic arenes or involve extension via acetylenic linkers, as well as impart distinct optoelectronic properties to these receptors. While the inclusion of metal centers is an important design motif,<sup>59</sup> contributions from organometallic receptors for anions have been recently reviewed<sup>60</sup> and are beyond the scope of this review.

An early example of an arene-alkyne based receptor attempted to reinforce face-to-face interactions between the host and guest via multiple butadiyne linkages that were not in conjugation. Investigation of the binding affinities for heterotricyclic receptors **14a-d** found that these bound large, flat substrates well with the highest binding constants observed for the largest substrates (e.g., terephthalate<sup>2-</sup> > 2,6-naphthalenedicarboxylate<sup>2-</sup> > 2,6-antraquinonedisulfonate<sup>2-</sup>) as these fit the large, reinforced binding pocket better.<sup>70</sup>

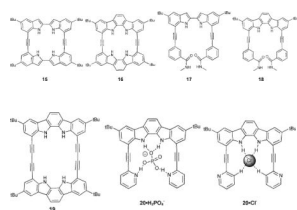


- 14a:** X = O  
Y = -(CH<sub>2</sub>)<sub>6</sub>-
- 14b:** X = O  
Y = -(CH<sub>2</sub>)<sub>2</sub>O(CH<sub>2</sub>)<sub>2</sub>-
- 14c:** X = O  
Y = -(CH<sub>2</sub>CH<sub>2</sub>O)<sub>2</sub>CH<sub>2</sub>-
- 14d:** X = NH  
Y = -(CH<sub>2</sub>)<sub>2</sub>O(CH<sub>2</sub>)<sub>2</sub>-

The binding constants with neutral adenosine were measured in an aqueous buffer ( $\log K_a = 4.00, 3.87, 3.86$  and  $4.00$ , **14a-d** respectively) and followed the same trend with doubly charged AMP ( $\log K_a = 4.08, 3.79, 3.92$  and  $4.08$ ).

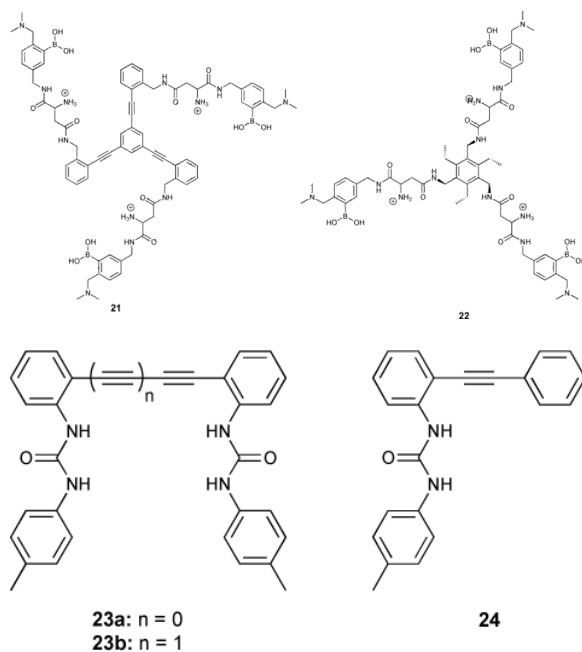
Alkyne-linked macrocyclic indole and indolocarbazole receptors **15** and **16** were synthesized and their anion affinities probed in acetonitrile.<sup>71</sup> It was found that further rigidification of receptor **15** to **16** very modestly increased binding constants (e.g.  $\text{Cl}^- K_a = 1.5 \times 10^6 \text{ M}^{-1}$  to  $2.1 \times 10^6 \text{ M}^{-1}$ ,  $\text{N}_3^- K_a = 8.8 \times 10^5 \text{ M}^{-1}$  to  $9.1 \times 10^5 \text{ M}^{-1}$ ,  $\text{H}_2\text{PO}_4^- K_a = 2.1 \times 10^6 \text{ M}^{-1}$  to  $3.2 \times 10^6 \text{ M}^{-1}$ ). A crystal structure of the **16**• $\text{Cl}^-$  complex revealed that the binding pocket was slightly too small to incorporate the  $\text{Cl}^-$  anion completely. Both of these receptors exhibited increased affinities over previously studied acyclic receptors **17** and **18**.<sup>72</sup> Rigidification in this system had the same effect as in the macrocyclic versions (**17**:  $K_a = 5100 \text{ M}^{-1}$  to **18**:  $110,000 \text{ M}^{-1}$  for  $\text{Cl}^-$ ). In more recent work, **19** was used to study the preferred binding geometry of azide, halide and oxoanionic guests.<sup>73</sup>

Changing the ethynyl linker in **16** to butadiynyl in **19** decreased the affinity for  $\text{Cl}^-$  anion over four fold, although the affinity for  $\text{Br}^-$  and  $\text{I}^-$  anions was greater for the larger cavity of **19**. There was no affinity for acetate, but other polyoxo-anions ( $\text{H}_2\text{PO}_4^-$ ,  $\text{NO}_3^-$  and  $\text{HSO}_4^-$ ) exhibited increased affinities of one to two orders of magnitude. Azide bound in an orthogonal fashion (normal to the macrocycle plane) in ethynyl linked **16** with one N atom in the cavity, but butadiynyl **19** had a large enough binding pocket to fully accommodate azide in a linear fashion, with a concomitant increase in binding constant ( $K_a = 2300 \text{ M}^{-1}$  to  $81,000 \text{ M}^{-1}$ ). The binding events in these receptors were followed by UV-Vis or  $^1\text{H-NMR}$  spectroscopy; the change in their fluorescence emission was not reported. The cleft-like derivative **20** was studied as well and found to strongly and selectively bind  $\text{H}_2\text{PO}_4^-$  ( $K_a = 1.1 \times 10^5 \text{ M}^{-1}$ ) due to the inclusion of two additional hydrogen bond acceptors in the ethynyl “arms”.<sup>74</sup> Although the binding constant for this system is smaller than for their previously reported macrocyclic receptor **16**, the selectivity increased, highlighting the well-designed binding pocket for the geometry of the desired guest. As expected, when binding guests with hydrogen bond donating capability the pyridine nitrogens were pointed into the cavity (e.g., **20**• $\text{H}_2\text{PO}_4^-$ ). The pyridine nitrogens rotated out when binding guests lacking hydrogen bond donating ability (e.g., **20**• $\text{Cl}^-$ ), and this was accompanied by a downfield shift of the C-H protons at the 3 position in the arene rings indicating C-H...anion hydrogen bonding. This underscores the need for some flexibility in the conformation of the binding site for selective receptors, as well as the contribution of an increased degree of flexibility in the scaffold.

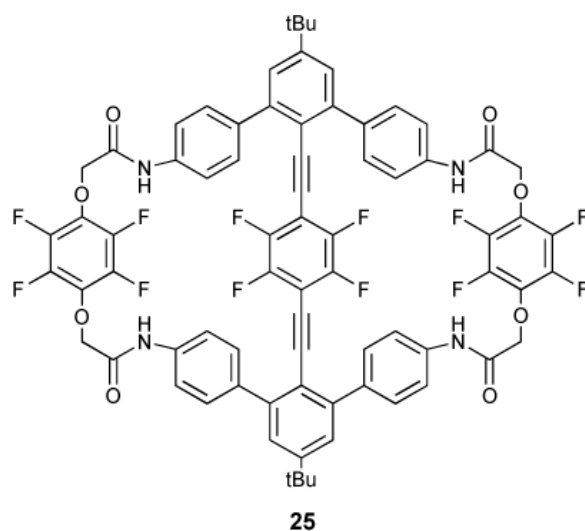


Invariably receptor systems must contain some signalling unit to function as a sensor, as showcased in the phenylacetylene-based tripodal receptor for heparin in serum **21**.<sup>75</sup> In **21** the three ammonium-bearing arms were extended from the fluorophore body by ethynyl linkages, which proved to be the structural criterion necessary for effective binding of anionic heparin. In contrast, **22**, with its closely packed tripodal arms, proved insufficiently sensitive to be viable in biological media such as serum.<sup>76</sup> This was attributed to non-specific binding of proteins in serum which displaced the dye bound in the receptor. Additionally, this provided elegant precedent for the ability of small-molecule synthetic receptors to be viable probes for complex biological substrates in competitive media.

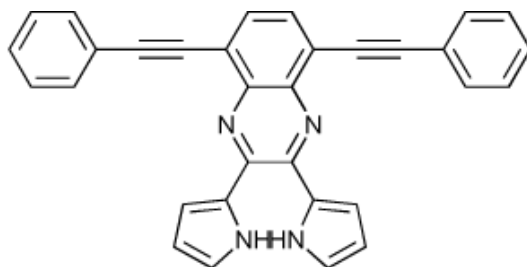
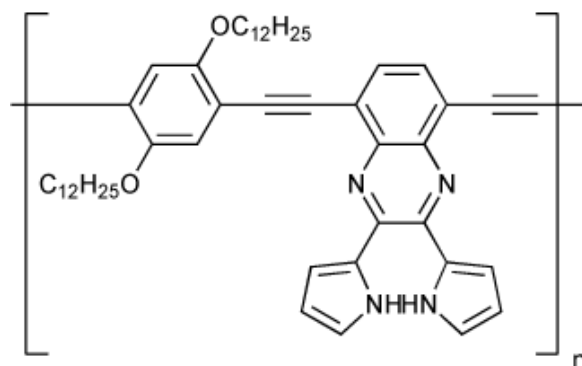




Recent work has made use of fluorescent “turn-off” anion sensing modes<sup>77</sup> and phenylacetylene subunits for both “turn-on” signalling and binding site size enforcement.<sup>78</sup> The chloride binding properties of diphenylacetylene-based anion receptors **23a,b** exhibited “turn-off” fluorescence in the presence of a suitable guest. Bisurea **23a** exhibited quenched fluorescence emission in its unbound “open” state. The DFT minimized conformation had the typical low fluorescence observed in diphenylacetylenes due to the dark  $\pi_y^* \leftarrow \pi_x$  ( $^1A_{1u}$ ) transition being close in energy to the emissive  $\pi_x^* \leftarrow \pi_x$  ( $^1B_{1u}$ ) transition. Fitting a suitably sized guest in the receptor cleft (in this case  $\text{Cl}^-$  anion) induced planarity by rotation around the alkynyl bond. This increased the energy of the  $^1A_{1u}$  transition, and the bound receptor thus became emissive. Binding constants for model systems **23b** and **24** were also measured against halide and oxoanions. Receptor **23a** had the highest affinity for  $\text{Cl}^-$  ( $\log \beta = 2.557$  versus 1.831 for **23b** and  $<1$  for **24**). The length of the phenylacetylenic linker influenced size selectivity, as  $\text{Br}^-$  was bound by **23a** and not by **23b**. None of the receptors exhibited any affinity for  $\text{NO}_3^-$ .



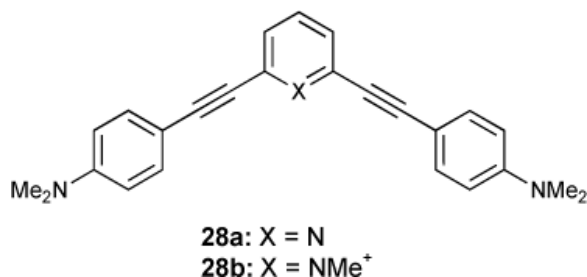
The allosteric binding of **25**, a macrocyclic receptor with a central tetrafluorophenyl “turnstile” suspended in the cavity by ethynyl bridges, was studied by NMR spectroscopy.<sup>79</sup> The turnstile inhibited both amide binding sites in the free receptor. Upon binding one equivalent of a guest the tetrafluorophenyl gate was opened and allowed access to the second binding site. Acetate,  $\text{Br}^-$ , and phosphate bound cooperatively (Hill coefficients of 1.4, 1.5 and 1.4, respectively). Interestingly,  $\text{Br}^-$  anion was large enough ( $1.82\text{\AA}$  in an octahedral environment) to exhibit cooperative binding, but  $\text{Cl}^-$  ( $1.67\text{\AA}$ ) ostensibly did not, although chloride had the higher  $K_{a1}$  with approximately equivalent second binding constants. This was reflected in the Hill coefficient for  $\text{Cl}^-$  anion (1.1).

**26****27a:**  $n = 7$ **27b:**  $n = 110$ 

Based on earlier work,<sup>80</sup> dipyrrolylquinoxaline derivative **26** was reported to bind anions with both a chromogenic and fluorogenic response.<sup>81</sup> This monomer exhibited affinity for anions ( $\text{F}^- > \text{HP}_2\text{O}_7^{3-} > \text{CN}^- > \text{OAc}^- > \text{H}_2\text{PO}_4^- \approx \text{Cl}^- \approx \text{Br}^- \approx \text{I}^- \approx \text{NO}_3^-$ ) correlating to the relative basicity of the guest, although fluoride and pyrophosphate were found to deprotonate the receptor. Polymerization to **27a** and **27b** was accompanied by a 34-fold increase in optoelectronic response, which translates into increased sensitivity with increased repeat units.

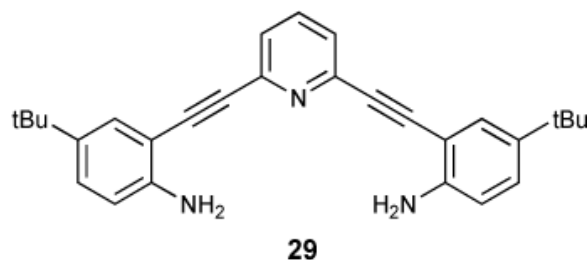
Although beyond the scope of this review, it is worth noting the two-photon absorption (TPA) properties that some phenylacetylenes can lend to a probe. Cross-conjugated D- $\pi$ A- $\pi$ -D bis(anilinoethynyl)-functionalized pyridine **28** was synthesized and its properties as a two-photon induced polymerization (TPIP) initiator were studied.<sup>82</sup> While free-base **28a** was too poor an acceptor to provide a good TPA cross section, cationic **28b** was assumed to have a much higher value, though this ultimately could not be verified due to the poor solubility of

**28b.** While applications for a target specific polymerization initiator may exist, the application of these to two-photon imaging techniques also warrants interest. It has also been reported that TPA fluorophores of the A- $\pi$ -A type can be modulated via their planarity, which has also been a demonstrated handle for supramolecular sensors.<sup>83</sup> A combination of selective targeting and low energy excitation and emission is a promising application for phenylacetylene-based sensors.



### 3.1 2,6-bis(arylethynyl)pyridine-based receptors

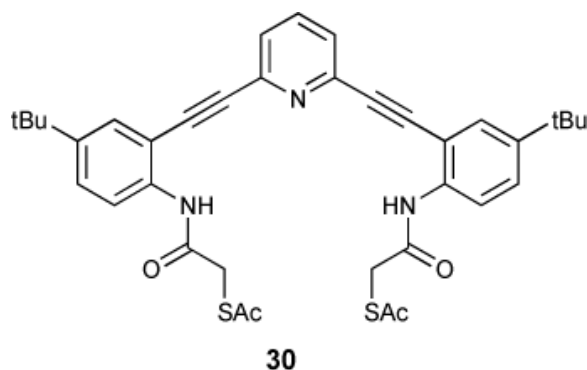
2,6-Bis(arylethynyl)pyridines have found utility in numerous areas of chemistry. The inherent properties of these ethynylpyridines (conjugation, absorption/emission, pH dependence, metal binding capability, rigidity etc.) have been exploited for applications in: liquid crystals, light-emitting materials, rotaxane-type structures, molecular magnets, antiangiogenic activity, polymer composites and coordination complexes.<sup>84-87</sup> In contrast, the supramolecular chemistry of 2,6-bis(arylethynyl)pyridines has received little attention, perhaps in part due to the scarcity of uniting the fields of molecule/ion recognition with the synthesis of highly-conjugated, carbon-rich materials. Most of the host/guest studies reported have focused on exploiting the pyridine lone pair to bind metal ion,<sup>88</sup> or organoiodides.<sup>85</sup> Alternatively, we hypothesized that the unique absorption/emission properties of arylethynylpyridines in tandem with their structural rigidity aptly positions them to function as small molecule or ion receptors. Moreover, we envisioned that the 2,6-bis(arylethynyl)pyridine scaffold and its derivatives would serve as a versatile building block for the development of receptor molecules that target a variety of guests depending on the protonation state of the pyridine and the type of functional group appended to the arylethynyl unit.



Our work thus far has focused on derivatives of the 2,6-bis(anilinoethynyl)pyridine **29**.<sup>89-90</sup> Recent investigations in the Haley lab of the structural and electronic properties of pyridine-based cyclines<sup>27</sup> and metallacycles<sup>91</sup> prompted our consideration of their use in molecular recognition applications. A highly conjugated, structurally rigid phenylacetylene scaffold incorporating the inward-directed and tunable (protonated or free base) pyridine functionality affords a pre-organized receptor capable of an optoelectronic response (e.g., change in emission) upon guest complexation. By opening up the macrocycle to provide a binding pocket for guest molecules, our acyclic design is now capable of adapting to a range of guest sizes. This is achieved via a three-point binding motif where the amide hydrogens

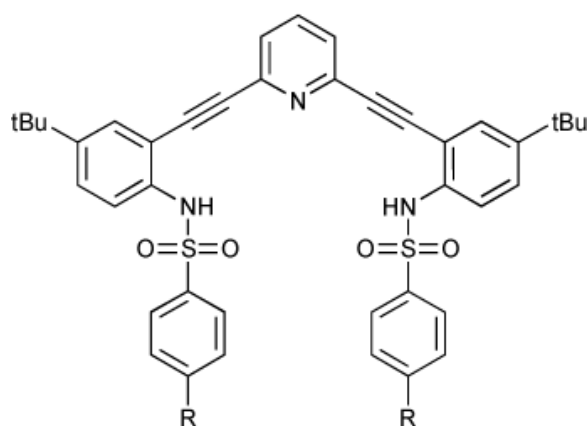
offer tunable hydrogen-bond donating sites along with protonation of the pyridine lone-pair as a switchable hydrogen-bond accepting or donating site. Additionally, exploiting the easily derivatized aniline functionality as a synthetic handle makes the modular approach to receptor design feasible.

As an example, acetyl-protected mercaptoamide **30** crystallizes in a polymeric chain as the neutral compound.<sup>92</sup> Protonation of the receptor with HCl gas bubbled through solution provides a racemic mixture of crystals with induced helicity from the bound anion. Low temperature <sup>1</sup>H-NMR spectroscopy indicated rapid interconversion between the two possible helicities by significant broadening of the methylene proton signals.

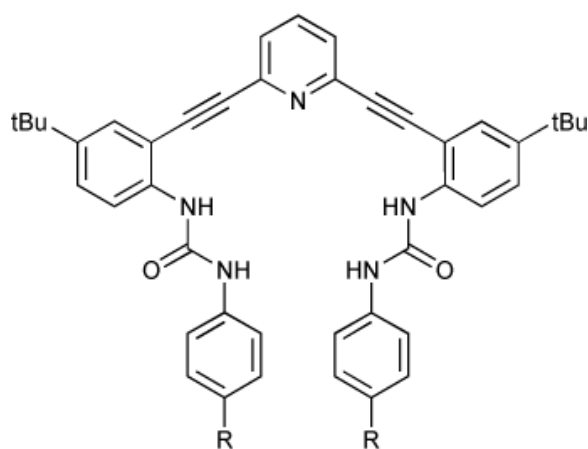


### 3.2 Sulfonamide and urea as hydrogen bond-donating binding sites

Both urea and sulfonamide derivatives **31** and **32** were investigated as anion receptors by UV-Vis, <sup>1</sup>H-NMR and fluorescence spectroscopic studies.<sup>89</sup>



**31a** R = Me  
**31b** R = OMe  
**31c** R = NO<sub>2</sub>



**32a** R = H  
**32b** R = OMe  
**32c** R = NO<sub>2</sub>

Sulfonamide receptors yielded [2 + 2] complexes from numerous solvents. Crystallization by either diffusion or evaporation yielded complexes with two water guests or two halide guests (if the receptors were protonated). Surprisingly, a heterodimer (**H31a**<sup>+</sup>•Cl<sup>-</sup>)•(**31a**•H<sub>2</sub>O) was obtained from treatment of the receptor with concentrated HCl, where one chloride and one water guest filled the binding pocket between the two receptor units (Figure 1). This dimeric species contained one protonated receptor whose pyridinium hydrogen shared a strong hydrogen bond with the chloride guest, which itself shared a hydrogen bond to a water guest. A final hydrogen bond from the water to the lone pair of the freebase receptor stitched the heterodimeric complex together. An additional four hydrogen bonds from the sulfonamide units of each receptor formed a network of helical hydrogen bonds running between the two receptors. Both the water and the halide guest freely exchanged in the system, and hence the heterodimeric structure exhibited properties intermediate to either of the homogenous dimers. In addition to the stabilization of the seven total hydrogen bonds,  $\pi$ -stacking interactions between the two receptors also contributed to the stabilization of the complex. Both hydrogen halide and H-OH shared the same structural

role in this self-assembled system, a synergistic effect that has recently become more widely appreciated as an important structural role in protein folding.

These studies have prompted us to exploit the modular nature of our synthetic strategy to vary the linking arms, functional groups, cores and binding units in the multitopic receptors.

In contrast to the dimeric structures obtained in the sulfonamide analogues, urea receptors **32a,b** crystallized in [4 + 4] tetrameric fashion with solvent molecules in the binding pockets (Figure 2).<sup>90</sup> Crystals obtained from DMSO/MeOH/toluene mixture or pure ethanol were found in either an “S” or “W” conformation, and these stacked in an “SWWS” repeat unit. Protonation of receptor **32a** with HCl yielded single crystals of a [1 + 1] complex (Figure 3).

Binding constants in water saturated chloroform with  $\text{NBu}_4^+$  salts followed the Hofmeister series in the neutral receptor (treated as [1 + 1] for fitting):  $2100 \text{ M}^{-1}$  for  $\text{Cl}^-$ ,  $400 \text{ M}^{-1}$  for  $\text{Br}^-$ , iodide had no measurable binding. In contrast trifluoroacetic acid protonated receptor bound halides in water saturated chloroform with the opposite affinity:  $41,700 \text{ M}^{-1}$  for  $\text{Cl}^-$ ,  $61,700 \text{ M}^{-1}$  for  $\text{Br}^-$  and  $83,200 \text{ M}^{-1}$  for iodide.

Geminate “turn on” and “turn off” fluorescence emission was observed for these receptors. In both sulfonamide and urea derivatives, protonation of the receptors in solution led to either quenching or enhancement of fluorescence based upon the pendant functionality. Electron-rich sulfonamide **31b** was fluorescent in the neutral state, but protonation by oxoacids (TFA or acetic acid), or hydrogen halides (HCl or HBr) quenched fluorescence emission and bathochromically shifted the residual emission band. In electron-poor **31c**, the opposite behaviour was observed: non-fluorescent neutral receptors fluoresced with an emission maximum at 515 nm (Figure 4).<sup>93</sup>

This trend also held true for the urea receptors **32b,c** which were investigated for their signalling ability *in vitro*. Methoxy-substituted phenylurea **32b** fluoresced in NIH3t3 murine embryo fibroblasts treated with a high  $\text{Cl}^-$  buffer, while  $\text{NO}_2$ -substituted phenylurea **32c** did not behave in the “OFF-ON” fashion expected, most likely due to solubility issues. Control experiments with nitrate buffers (no  $\text{Cl}^-$ ) exhibited minimal fluorescence (Figure 5).

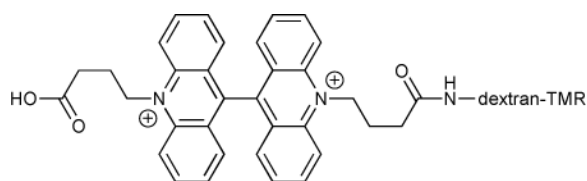
These results underscore the need for a full understanding of how a promising receptor in the cuvet can be successfully translated to biological media.

#### 4.1 Biological applications of phenylacetylene-based sensors

There are several biochemical factors to consider in the design of new intracellular ion indicators. These include the loading of the indicator(s) into target cells,<sup>94-98</sup> maximizing optimal detection parameters<sup>99</sup> with minimal toxicity<sup>100</sup> and the ability to selectively quantitate the ion concentration in question within the cells or tissues being examined.<sup>101</sup> Within these areas are several additional factors that affect utility including compartmentalization or partitioning within the cell into specific organelles,<sup>102</sup> matching optical excitation/emission wavelengths to match common instrumentation in use<sup>103</sup> and the use of ratiometric determination methods to help quantitate binding and ion levels.<sup>104,105</sup> Finally, factors such as specificity versus other intracellular ions,<sup>106</sup> water solubility,<sup>107</sup> relative binding constants for the intracellular ion or ions<sup>108</sup> and the effect of pH on binding<sup>99,109</sup> and fluorescence emission<sup>101</sup> are also important factors to be optimized for biochemical probe optimization. Of course, it is impossible to optimize all parameters, but of ultimate importance are ion selectivity and the ability to quantitate intracellular concentration.

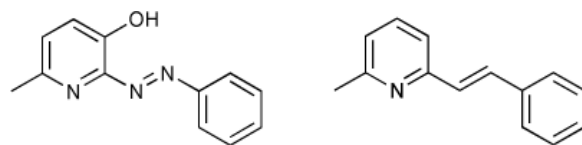
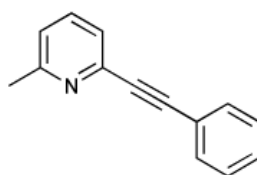
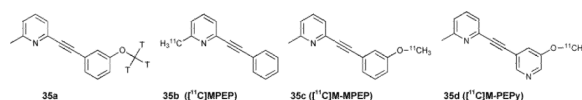


Anions in cells are present at roughly 70% of all enzymatic sites and are responsible for everything from the activation of signal transduction pathways to maintaining osmotic pressure and cell volume.<sup>1,110</sup> They are involved in many disease pathways, from cystic fibrosis to osteoporosis.<sup>111-113</sup> In mitochondria, at least 14 different anion transport pathways have been identified<sup>114</sup> and are responsible for the regulation and trafficking of ADP, ATP, citrate, maleate and halide anions, among others.<sup>115,116</sup> The ubiquity of these molecules in living organisms, and their importance in regulating life systems can best be summed up in the realization of the polyanionic character of both DNA and RNA. The acidity of intracellular compartments has been implicated in some disease pathways, notably in the cellular breach of anthrax lethal toxin and edema toxin.<sup>117</sup>

**33**

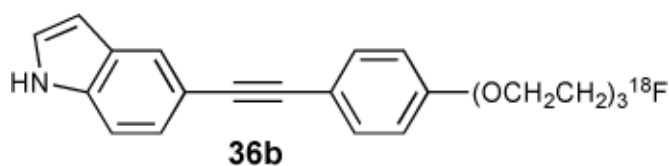
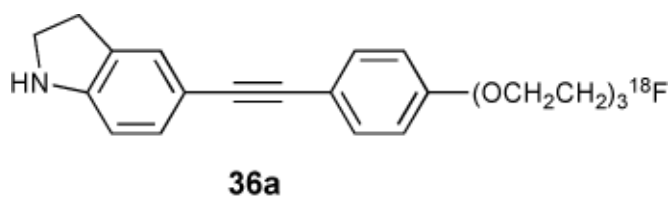
This breach has been correlated with decreased endosomal pH, which can be achieved either by concomitant movement of  $K^+$ , or an influx of  $Cl^-$ .

One recently reported example of a ratioable fluorescent  $Cl^-$  probe was dextran-tethered acridinium system **33**, used for charting endosomal  $Cl^-$  concentration in tandem with endosomal pH.<sup>109</sup> The biacridinium subunit of the receptor was found to be quenched by  $Cl^-$  anion with a Stern-Volmer constant of  $36 M^{-1}$ , and to be insensitive to non-halide anions (nitrate, phosphate, bicarbonate and sulfate). By tethering this system to tetramethylrhodamine, which is insensitive to  $Cl^-$ , they developed a ratioable sensor for *in vivo* application that allowed them to observe the change in  $Cl^-$  concentration in endosomes with decreasing pH. Using this same technique, it was also shown that the intracellular CIC-3  $Cl^-$  channel regulated the vacuolar  $H^+$  pump during endosomal acidification.<sup>118</sup> The ease with which a well designed small molecule sensor can visualize these complex biological responses demonstrates the import role these receptors will play in furthering our understanding of cellular processes.

**34a****34b****34c****35a****35b** ( $^{13}C$ ]MPEP)**35c** ( $^{13}C$ ]M-MPEP)**35d** ( $^{13}C$ ]M-PEPy)

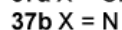
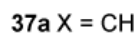
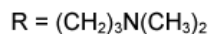
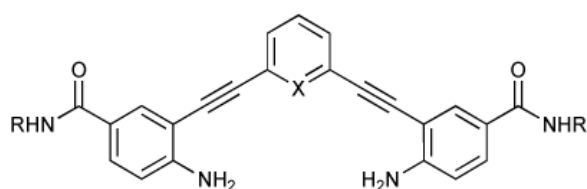
A series of linked pyridylarenes **34** were used as selective antagonists for the metabotropic glutamate receptor subtype 5 (mGlu5).<sup>119</sup> It was found that phenylethynyl derivative **34c** (MPEP) was the most active antagonist, and bound the receptor much more effectively than the non-ethynylated receptors. This work was continued with radiolabelled derivatives **35a**<sup>120</sup> and **35b-d**.<sup>121</sup> These radiolabelled compounds were thought to be appropriate for binding and following the distribution of mGluR5 receptors. Radiolabelled **35a** possessed a high affinity ( $K_d = 2$  nM) and was over 90% specific for mGlu5 receptor over the 1-10 nM range.

Expanding upon **35a**, ligands **35b-d** were also utilized as PET imaging agents in Sprague-Dawley rats.<sup>121</sup> Bioaccumulation of the radiolabelled agents were tracked by *in vivo* microPET imaging (Figure 6), and found to have the highest binding in the olfactory bulb, followed by striatum, hippocampus and cortex localization. It was hypothesized that the glutamate receptor mGluR5 might have major physiological function in the olfactory area as olfactory damage in neurodegenerative diseases is consistent and severe.<sup>122</sup> This would also suggest that early diagnosis of Parkinson's or Alzheimer's could be achieved via the olfactory bulb.



Related work with  $\beta$ -amyloid plaques ( $A\beta$  plaques) as pathological features of Alzheimer's onset made use of indolyl- and indolylphenylacetylenes as PET imaging agents.<sup>123</sup> Both **36a** and **36b** were found to bind  $A\beta$  plaques, with **36b** being slightly more selective in early trials (via autoradiography). No binding constants were reported, although relative binding was inferred from washout rates.

Phenylethynylamides **37a,b** were found to be potent G-quadruplex binders.<sup>124</sup> Binding was studied in this system by FRET melting assays, surface plasmon resonance and CD spectroscopy. Moderate stabilization with G-quadruplexes were observed for both ligands. No discernible duplex DNA stabilization was reported. As there is evidence that small molecules can bind G-quadruplexes and modulate transcription, selectivity like this is promising for future application of these acyclic compounds in this area. Additionally, the <sup>1</sup>H-NMR experiments reported were carried out in aqueous buffer, notable due to the low solubility for related analogues (no amide functionality) and water-solubility for many arylethynyl probes is a barrier to their use as bioimaging agents.



## Conclusions

Phenylacetylenes have emerged as important components in the design of an increasingly diverse array of exciting supramolecular host-guest complexes. Their unique combination of optoelectronic properties and shape persistence has played an important role in the development of many novel coordination compounds, fluorogenic and chromogenic probes and biological imaging agents. The relative insolubility of many of these compounds in aqueous media still remains a challenge in the development of this class of probe architectures. While metal-containing complexes have been extensively reviewed, host-guest complexes that make use of only simple organic phenylacetylenes have only recently received increasing attention. Their contribution to the rational and effective design of receptors able to target both neutral and ionic guests with concomitant optoelectronic response upon binding is at the core of this advancement. Additionally, commonly used synthetic techniques allow for facile and efficient preparation of a variety of novel candidates for study, which further drives innovation in this field, suggesting potential applications in molecular probe development, nanotechnology, and even recognition of biologically-relevant molecules and ions.<sup>125</sup>

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## Biographies



Calden N. Carroll is a native of Flagstaff, Arizona and received his extended B.S. in Chemistry in 2004 from Northern Arizona University. He is currently a Ph. D. candidate in the laboratories of Profs. Michael M. Haley and Darren W. Johnson at the University of Oregon where he studies the binding affinities of small molecule receptors for anionic guests when he is not on the river. He spends a lot of time rafting.



John J. Naleway received his Ph. D. in 1981 from Marquette University under the direction of Prof. Norman Hoffman. After completing postdoctoral research at The University of Alberta under Prof. Raymond Lemieux and at the University of Wisconsin under Prof. Laurens Anderson, he joined the staff at Monsanto Healthcare - Searle Pharmaceuticals, and was later the Laboratory Director and Director of Substrates Research at Molecular Probes, Inc. He is currently an Adjunct Faculty member in the Department of Chemistry at the University of Oregon and President of Marker Gene Technologies, Inc. in Eugene, Oregon where his research is centered around live cell assays for drug discovery and fluorescent chemosensors.



Michael M. Haley studied cyclopropene and cyclopropane chemistry with Prof. Ed Billups at Rice University where he received both his B.A. (1987) and Ph.D. degrees (1991). In 1991 he received a National Science Foundation Postdoctoral Fellowship to work with Prof. Peter Vollhardt on [N]phenylene chemistry at the University of California, Berkeley. In 1993 he joined the faculty at the University of Oregon where he is currently a Professor and Head of the Chemistry Department, as well as a member of the Materials Science Institute. Among the awards he has received are a National Science Foundation CAREER Award (1995), a Camille Dreyfus Teacher-Scholar Award (1998), an Alexander von Humboldt Research Fellowship (2000), Thomas F. Herman Distinguished Teaching Award (2002), and University of Oregon Fund for Faculty Members Excellence Award (2007). His current research focuses on the chemistry of dehydrobenzoannulenes, indenofluorenes, molecules based on phenyl-acetylene scaffolding and other novel carbon-rich systems.





Darren W. Johnson received his B.S. in Chemistry at the University of Texas at Austin in 1996, where he performed undergraduate research under the direction of Jonathan L. Sessler. He earned his Ph. D. in Chemistry in 2000 from the University of California at Berkeley working with Kenneth N. Raymond, and he then spent two years at the Scripps Research Institute as a National Institutes of Health post-doctoral fellow with Julius Rebek, Jr. He joined the chemistry faculty at the University of Oregon in 2003, where he is currently an Associate Professor. He is a Cottrell Scholar of Research Corporation for Science Advancement and a National Science Foundation CAREER awardee. Research in his group uses supramolecular chemistry as a tool to explore a variety of problems in coordination chemistry, molecule/ion recognition and inorganic cluster synthesis.

## Notes and references

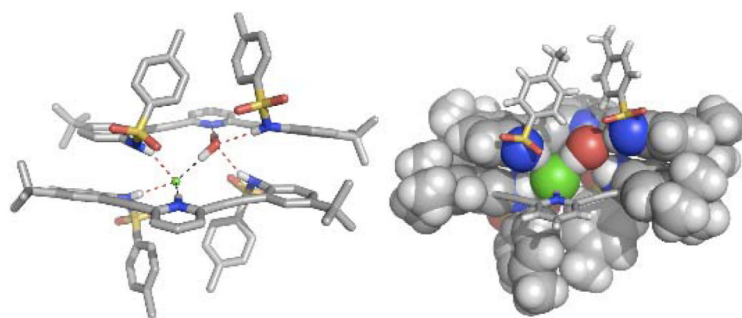
1. Sessler, J.L.; Gale, P.A.; Cho, W-S. *Anion Receptor Chemistry*. Royal Society of Chemistry; Cambridge: 2006.
2. Cho, W-S.; Sessler, J.L. *Functional Synthetic Receptors*. Schrader, T.; Hamilton, A.D., editors. Wiley-VCH; Weinheim: 2005. p. 165-256.
3. Bianchi, A.; Bowman-James, K.; Garcia-España, E., editors. *Supramolecular Chemistry of Anions*. Wiley-VCH; New York: 1997.
4. Houssain A. *Curr. Org. Chem.* 2008; 12:1231–1256.
5. Campbell, K.; Tykwinski, R.R. *Carbon-rich Compounds: From Molecules to Materials*. Haley, M.M.; Tykwinski, R.R., editors. Wiley-VCH; New York: 2006. p. 229-294.
6. Welti R, Diederich F. *Helv. Chim. Acta.* 2003; 86:494.
7. Huang C-Y, Cabell LA, Anslyn EV. *J. Am. Chem. Soc.* 1994; 116:2778.
8. Sessler J.L, Vivian A.E, Seidel D, Burrell A.K, Hoehner M, Mody T.D, Gebauer A, Weghorn S.J, Lynch V. *Coord. Chem. Rev.* 2000; 216-217:411–434.
9. Jasat A, Dolphin D. *Chem. Rev.* 1997; 97:2267–2340. [PubMed: 11848901]
10. Gale P.A, Anzenbacher P Jr, Sessler J.L. *Coord. Chem. Rev.* 2001; 222:57–102.
11. Anzenbacher P Jr, Nishiyabu R, Palacios M.A. *Coord. Chem. Rev.* 2006; 250:2929–2938.

12. Atwood JL, Holman KT, Steed JW. *J. Chem. Soc., Chem. Commun.* 1996:1401–1407.
13. Sessler JL, Rubin BL, Camiolo S, Cho W-S, Pantos GD, Lynch VM. *Supramolecular Chem.* 2006; 18:103–109.
14. Gale PA. *Chem. Commun.* 2005:3761–3772.
15. Evan-Salem T, Baruch I, Avram L, Cohen Y, Palmer LC, Rebek J Jr. *Proc. Nat. Acad. Sci.* 2006; 103:12296–12300. [PubMed: 16894154]
16. Leininger S, Olenyuk B, Stang PJ. *Chem. Rev.* 2000; 100:853–908. [PubMed: 11749254]
17. Anderson HL, Sanders JKM. *Chem. Commun.* 1996:946–947.
18. Ferrara JD, Tessier-Youngs C, Youngs WJ. *Organometallics.* 1987; 6:676–678.
19. Iyoda M, Sirinintasak S, Nishiyama Y, Vorasingha A, Sultana F, Nakao K, Kuwatani Y, Matsuyama H, Yoshida M, Miyake Y. *Synthesis.* 2004:1527–1531.
20. Ferrara JD, Tanaka AA, Fierro C, Tessier-Youngs C, Youngs WJ. *Organometallics.* 1989; 8:2089–2098.
21. Ferrara JD, Djebli A, Tessier-Youngs C, Youngs WJ. *J. Am. Chem. Soc.* 1988; 110:647–649.
22. Ferrara JD, Tessier-Youngs C, Youngs WJ. *J. Am. Chem. Soc.* 1985; 107:6719.
23. Baxter PNW. *Chem. Eur. J.* 2003; 9:2531–2541.
24. Baxter PNW. *Chem. Eur. J.* 2002; 8:5250–5264.
25. Baxter PNW, Dali-Youcef R. *J. Org. Chem.* 2005; 70:4935–4953. [PubMed: 15960491]
26. Samori S, Tojo S, Fujitsuka M, Spitler EL, Haley MM, Majima T. *J. Org. Chem.* 2007; 72:2785–2793. [PubMed: 17367190]
27. Spitler EL, McClintock SP, Haley MM. *J. Org. Chem.* 2007; 72:6692–6699. [PubMed: 17685654]
28. Zhang H, Wan X, Xue X, Li Y, Yu A, Chen Y. *Eur. J. Org. Chem.* 2010:1681–1687.
29. Spitler EL, Haley MM. *Tetrahedron.* 2008; 64:11469–11474.
30. Liao J-H, Chen C-T, Chou H-C, Cheng C-C, Chou P-T, Fang J-M, Slanina Z, Chow TJ. *Org. Lett.* 2002; 4:3107–3110. [PubMed: 12201728]
31. Fang JM, Selvi S, Liao JH, Slanina Z, Chen CT, Chou PT. *J. Am. Chem. Soc.* 2004; 126:3559. [PubMed: 15025485]
32. (a) Inouye M, Miyake T, Furusyo M, Nakazumi H. *J. Am. Chem. Soc.* 1995; 117:12416. (b) Inouye M, Chiba J, Nakazumi H. *J. Org. Chem.* 1999; 64:8170–8176. [PubMed: 11674733]
33. Anderson S, Neidlein U, Gramlich V, Diederich F. *Angew. Chem. Int. Ed. Engl.* 1995; 34:1596.
34. Neidlein U, Diederich F. *Chem. Commun.* 1996:1493.
35. Droz AS, Neidlein U, Anderson S, Seiler P, Diederich F. *Helv. Chim. Acta.* 2001; 84:2243.
36. Inouye M, Waki M, Abe H. *J. Am. Chem. Soc.* 2004; 126:2022. [PubMed: 14971935]
37. Martínez-Máñez R, Sancenón F. *Chem. Rev.* 2003; 103:4419–4476. [PubMed: 14611267]
38. Anslyn E. *J. Org. Chem.* 2006; 72:687–699. [PubMed: 17253783]
39. Gunnlaugsson T, Glynn M, Tocci GM, Kruger PE, Pfeffer FM. *Coord. Chem. Rev.* 2006; 250:3094–3117.
40. Metzger A, Anslyn EV. *Angew. Chem. Int. Ed.* 1998; 37:649–651.
41. Malashikhin SA, Baldrige KK, Finney NS. *Org. Lett.* 2010; 12:940–943. [PubMed: 20131818]
42. Kondo S, Sato M. *Tetrahedron.* 2006; 62:4844.
43. Anthony JE, Khan SI, Rubin Y. *Tetrahedron.* 1997; 38:3499–3502.
44. Taniguchi M, Cramer DL, Bhise AD, Kee HL, Bocian DF, Holten D, Lindsey JS. *New. J. Chem.* 2008; 32:947–958.
45. Droz AS, Diederich F. *J. Chem. Soc., Perkin Trans. 1.* 2000:4224–4226.
46. Young, JK.; Moore, JS. *Modern Acetylene Chemistry.* Stang, PJ.; Diederich, F., editors. Wiley-VCH; New York: 1995. p. 415-442.
47. Beer S, Hrib CG, Jones PG, Brandhorst K, Grunenberg J, Tamm M. *Angew. Chem. Int. Ed.* 2007; 46:8890–8894.
48. Zhang W, Moore JS. *Adv. Syn. Catal.* 2007; 349:93–120.
49. Scrimgeour, KG. *Chemistry and Control of Enzymatic Reactions.* Academic Press, Inc.; New York: 1977.

50. Schetz JA, Sibley DR. *J. Pharmacol Exp. Ther.* 2001; 296:359. [PubMed: 11160618]
51. (a) Martinez-Diaz M-V, Spencer N, Stoddart F. *Angew. Chem. Int. Ed. Engl.* 1997; 36:1904. (b) Huang Y-L, Hung W-C, Lai C-C, Liu Y-H, Peng SM, Chiu S-H. *Angew. Chem., Int. Ed.* 2007; 46:6629–6633.
52. Al-Sayah MH, Branda NR. *Org. Lett.* 2002; 4:881. [PubMed: 11893176]
53. Al-Sayah MH, Branda NR. *Angew. Chem., Int. Ed. Engl.* 2000; 39:945. [PubMed: 10760902]
54. Filby MH, Dickson SJ, Zaccheroni N, Prodi L, Bonacchi S, Montalti M, Paterson MJ, Humphries TD, Chiorboli C, Steed JW. *J. Am. Chem. Soc.* 2008; 130:4105–4113. [PubMed: 18314990]
55. Bucher C, Devillers CH, Moutet J-C, Royal G, Saint-Aman E. *New J. Chem.* 2004; 28:1584–1589.
56. Miyaji H, Gasser G, Green SJ, Molard Y, Strawbridge SM, Tucker JHR. *Chem. Commun.* 2005:5355–5357.
57. Inouye M, Takahashi K, Nakazumi H. *J. Am. Chem. Soc.* 1999; 121:341.
58. Rashadan S, Light ME, Kilburn JD. *Chem. Commun.* 2006:4578–4580.
59. Steed JW. *Chem. Soc. Rev.* 2008; 38:506–519. [PubMed: 19169464]
60. Amendola V, Bonizzoni M, Esteban-Goméz D, Fabrizzi L, Licchelli M, Sancenón F, Taglietti A. *Coord. Chem. Rev.* 2006; 250:1451–1470.
61. Cercla Priority List of Hazardous Substances. 2007. <http://www.atsdr.cdc.gov/cercla>
62. US EPA Case Study – Arsenic Treatment Technologies, Tucson, Arizona. 2003. <http://www.epa.gov/safewater/arsenic/publications.html>
63. Liu C-Q, Li S-L, Lang Y-C, Xiao H-Y. *Env. Sci. Technol.* 2006; 40:6928. [PubMed: 17153996]
64. Katayev EA, Kolesnikov GV, Sessler JL. *Chem. Soc. Rev.* 2009; 38:1572–1586. [PubMed: 19587953]
65. Cherimisinoff NP. *Pollution Engineering.* 2001; 33:38.
66. Maji SK, Pal A, Pal T. *J. Haz. Mat.* 2008; 151:811–820.
67. (a) Gu B, Brown GM, Bonnesen PV, Liang L, Moyer BA, Ober R, Alexandratos SD. *Env. Sci. Technol.* 2000; 34:1075. (b) Chuoyok W, Wiacek RJ, Pattamakomsan K, Sangvanich T, Grudzien RM, Fryxell GE, Yantasee W. *Env. Sci. Technol.* 2010; 44:3073. [PubMed: 20345133]
68. Dutzler R, Campbell EB, Cadene M, Chait BT, Mackinnon R. *Nature.* 2002; 415:287. [PubMed: 11796999]
69. Rychkov GY, Pusch M, Roberts ML, Jentsch TJ, Bretag AH. *J. Gen. Physiol.* 2001; 530:379–393.
70. Cudic P, Zinic M, Tomisic V, Simeon V, Vigneron J-P, Lehn J-M. *J. Chem. Soc., Chem. Commun.* 1995:1073.
71. Chang K-J, Moon D, Lah MS, Jeong K-S. *Angew. Chem., Int. Ed.* 2005; 44:7926.
72. Kim N-K, Chang K-J, Moon D, Lah MS, Jeong K-S. *Chem. Commun.* 2007:3401–3403.
73. Chang K-J, Chae MK, Lee C, Lee J-Y, Jeong K-S. *Tetrahedron Lett.* 2006; 47:6385.
74. Kwon TH, Jeong K-S. *Tetrahedron Lett.* 2006; 47:8539.
75. Wright AT, Zhong Z, Anslyn EV. *Angew. Chem. Int. Ed.* 2005; 44:5679–5682.
76. Zhong Z, Anslyn EV. *J. Am. Chem. Soc.* 2002; 124:9014–9015. [PubMed: 12148981]
77. Swinburne AN, Paterson MJ, Beeby A, Steed JW. *Org. Biomol. Chem.* 2010; 8:1010–1016. [PubMed: 20165790]
78. Swinburne AN, Paterson MJ, Beeby A, Steed JW. *Chem. Eur. J.* 2010; 16:2714–2718.
79. Hirata O, Takeuchi M, Shinkai S. *Chem. Commun.* 2005:3805–3807.
80. Black CB, Andrioletti B, Try AC, Ruiperez C, Sessler JL. *J. Am. Chem. Soc.* 1999; 121:10438–10439.
81. Wu C-Y, Chen M-S, Lin C-A, Lin S-C, Sun S-S. *Chem. Eur. J.* 2006; 12:2263–2269.
82. Pucher N, Rosspeinter A, Satzinger V, Schmidt V, Gescheidt G, Stampfl J, Liska R. *Macromolecules.* 2009; 42:6519–6528.
83. Porres L, Charlot M, Entwistle CD, Beeby A, Marder TB, Blanchard-Desce M. *Proc. SPIE.* 2005; 5943:559340F.
84. Ahn CM, Shin W-S, Woo HB, Lee S, Lee H-W. *Bio. Org. Med. Chem. Lett.* 2004; 14:3893.
85. Holmes BT, Deb P, Pennington WT, Hanks TW. *J. Polym. Res.* 2006:133.

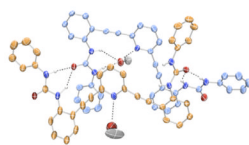
86. Rajadurai C, Ivanova A, Enkelmann V, Baumgarten M. *J. Org. Chem.* 2003; 68:9907. [PubMed: 14682682]
87. Yamaguchi Y, Kobayashi S, Wakamiya T, Matsubara Y, Yoshida Z-I. *Angew. Chem. Int. Ed.* 2005; 44:7040.
88. Phelps D, Crihfield A, Hartwell J, Hanks TW, Pennington WT, Bailey RD. *Mol. Cryst. Liq. Cryst.* 2000; 354:1111.
89. Berryman OB, Johnson CA, Zakharov LN, Haley MM, Johnson DW. *Angew. Chem. Int. Ed.* 2008; 47:117.
90. Carroll CN, Berryman OB, Johnson CA, Zakharov LN, Haley MM, Johnson DW. *Chem. Commun.* 2009:2520.
91. Johnson CA, Baker BA, Berryman OB, Zakharov LN, O'Connor MJ, Haley MM. *J. Organomet. Chem.* 2006; 691:413.
92. Johnson CA, Berryman OB, Sather AC, Zakharov LN, Haley MM, Johnson DW. *Cryst. Growth Des.* 2009; 9:4247.
93. Carroll CN, Coombs BA, Johnson CA, McClintock SP, Naleway JJ, Haley MM, Johnson DW. unpublished results.
94. Tsien RY. *Nature.* 1981; 290:527. [PubMed: 7219539]
95. Bush DS, Jones RL. *Plant Physiol.* 1990; 93:841. [PubMed: 16667590]
96. Steinberg TH, Newman AS, Swanson JA, Silverstein SC. *J. Biol. Chem.* 1987; 262:8884. [PubMed: 3597398]
97. Pilas B, Durack G. *Cytometry.* 1997; 28:316. [PubMed: 9266752]
98. Doyle AD, Lee J. *Biotechniques.* 2002; 33:358. [PubMed: 12188188]
99. Martínez-Zaguilán R, Parnami G, Lynch RM. *Cell Calcium.* 1996; 19:337. [PubMed: 8983854]
100. Katerinopoulos HE. *Curr. Pharm. Design.* 2004; 10:3835.
101. Lattanzio FA. *Biochem. Biophys. Res. Commun.* 1991; 177:184. [PubMed: 2043105]
102. Di Virgilio F, Steinberg TH, Silverstein SC. *Cell Calcium.* 1990; 11:57. [PubMed: 2191781]
103. Clark HA, Kopelman R, Tjalkens R, Philbert MA. *Anal. Chem.* 1999; 71:4837. [PubMed: 10565275]
104. Silver RB. *Methods Cell Biol.* 1998; 56:237. [PubMed: 9500141]
105. Bright GR, Fisher GW, Rogowska J, Taylor DL. *Methods Cell Biol.* 1989; 30:157. [PubMed: 2648109]
106. Carpenter RD, Verkman AS. *Org. Lett.* 2010; 12:1160. [PubMed: 20148571]
107. Biwersi J, Farah N, Wang YX, Ketchum R, Verkman AS. *Am. J. Physiol.* 1992; 262-1:C242. [PubMed: 1370743]
108. Gryniewicz G, Poenie M, Tsien RY. *J. Biol. Chem.* 1985; 260:3440. [PubMed: 3838314]
109. Sonawane ND, Thiagarajah JR, Verkman AS. *J. Biol. Chem.* 2002; 277:5506. [PubMed: 11741919]
110. Schroeder JI. *Plant Mol. Biol.* 1995; 28:353. [PubMed: 7543302]
111. Renkawek K, Bosman GJ. *NeuroReport.* 1995; 6:929. [PubMed: 7612885]
112. Anderson MP, Gregory RJ, Thompson S, Souza DW, Paul S, Mulligan RC, Smith AE, Welsh MJ. *Science.* 1991; 253:202. [PubMed: 1712984]
113. Kornak U, Kasper D, Bosl MR, Kaiser E, Schweizer M, Schulz A, Friederich W, Delling G, Jentsch TJ. *Cell.* 2001; 104:205. [PubMed: 11207362]
114. Kaplan RS. *J. Membr. Biol.* 2001; 179:165. [PubMed: 11246418]
115. Thompson RJ, Akana HCSR, Finnigan C, Howell KE, Caldwell JH. *Am. J. Physiol. Cell Physiol.* 2006; 290:C499. [PubMed: 16403948]
116. Okada SF, O'Neal WK, Huang P, Nicholas RA, Ostrowski LE, Craigen WJ, Lazarowski ER, Boucher RC. *J. Gen. Phys.* 2004; 124:513.
117. Zhang S, Finkelstein A, Collier RJ. *Proc. Natl. Acad. Sci.* 2004; 101:16756. [PubMed: 15548616]
118. Hara-Chikuma M, Yang B, Sonawane ND, Sasaki S, Uchida S, Verkman AS. *J. Biol. Chem.* 2005; 280:1241. [PubMed: 15504734]

119. Gasparini F, Lingenhöhl K, Stoehr N, Flor PJ, Heinrich M, Vranesic I, Biollaz M, Allgeier H, Heckendorn R, Urwyler S, Varney MA, Johnson EC, Hess SD, Sacaan AI, Santori EM, Veliçelebi G, Kuhn R. *Neuropharmacology*. 1999; 38:1493. [PubMed: 10530811]
120. Gasparini F, Andres H, Flor PJ, Heinrich M, Inderbitzin W, Lingenhöhl K, Müller H, Munk VC, Omilusik K, Stierlin C, Stoehr N, Vranesic I, Kuhn R. *Bioorg. Med. Chem. Lett.* 2002; 12:407. [PubMed: 11814808]
121. Yu M, Tueckmantel W, Wang X, Zhu A, Kozikowski AP, Brownell A-L. *Nucl. Med. Biol.* 2005; 32:631. [PubMed: 16026710]
122. Nores JM, Biacabe B, Bonfils P. *Ann. Med. Interne (Paris)*. 2000; 151:97. [PubMed: 10855362]
123. Qu W, Choi S-R, Hou C, Zhuang Z, Oya S, Zhang W, Kung M-P, Manchandra R, Skovronsky DM, Kung HF. *Bioorg. Med. Chem. Lett.* 2008; 18:4823. [PubMed: 18707879]
124. Dash J, Shirude PS, Hsu S-TD, Balasubramanian S. *J. Am. Chem. Soc.* 2008; 130:15950. [PubMed: 18980309]
125. Gale PA. *Chem. Commun.* 2010 ASAP: DOI:10.1039/C0CC00656D.

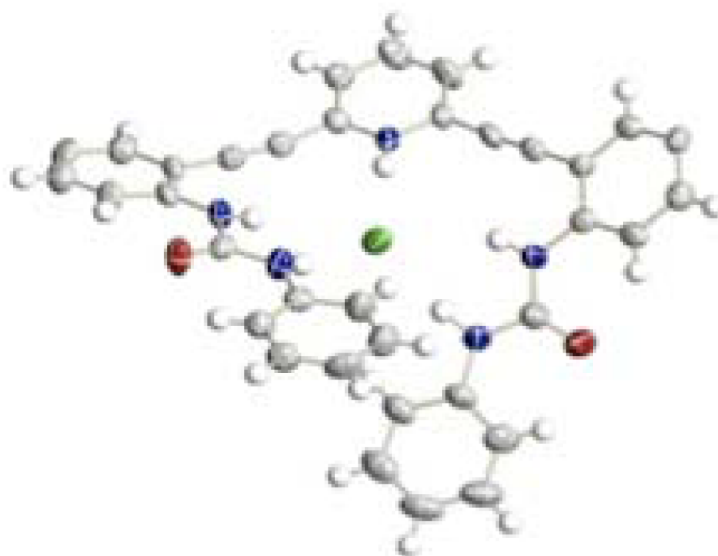


**Fig. 1.** Crystal structure representations (both stick and CPK) of  $(\mathbf{H31a}^+\cdot\mathbf{Cl}^-)\cdot(\mathbf{31a}\cdot\mathbf{H}_2\mathbf{O})$

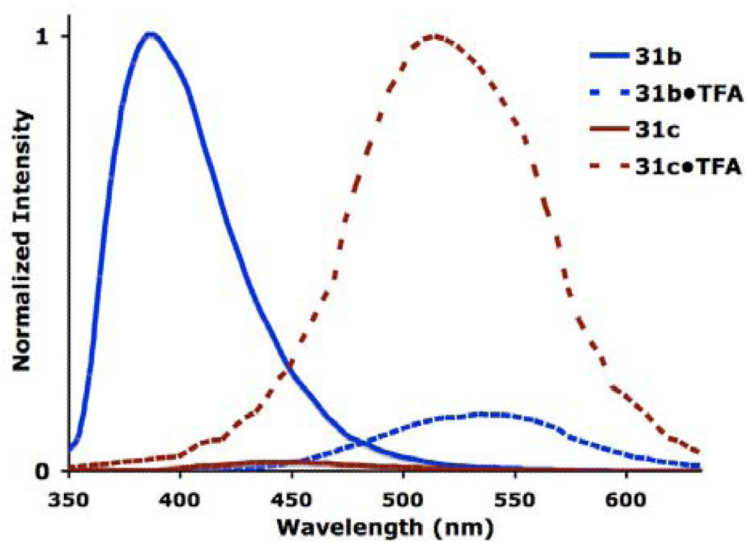




**Fig. 2.** Crystal structure representation of one half of **32a** (“S” in blue and “W” in orange) with bound MeOH solvent<sup>90</sup>



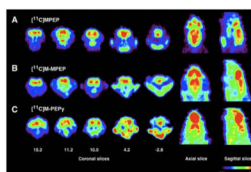
**Fig. 3.** ORTEP representation (50% probability level) of **H32a<sup>+</sup>•Cl<sup>-</sup>** (t-butyl groups are omitted for clarity)



**Fig. 4.** Normalized fluorescence emission of neutral and TFA-protonated bis-sulfonamides **31b,c**



**Fig. 5.** Epifluorescence microscope images of NIH3T3 murine embryo fibroblasts incubated in 10% aqueous DMSO with **32b** (a)  $\text{Cl}^-$  containing buffer, (b) Normarski phase image of same and (c)  $\text{NO}_3^-$  buffer (low  $\text{Cl}^-$ )<sup>93</sup>



**Fig. 6.** Coronal, axial and sagittal PET images of derivatives of **35** in anesthetized rat brain at 8-10 min after administration of radiolabelled ligand. The coronal level 15.2 corresponds to the olfactory area; 11.2 the cingulate level; 10.0 the striatal level; 4.2 the hippocampal level; and -2.8 the cerebellar level. The axial and sagittal views illustrate the distribution of the radioactivity at the mid-striatal level. (Reprinted from ref. <sup>121</sup> with permission from Elsevier)