

In Vitro Activities of Arylomycin Natural-Product Antibiotics against *Staphylococcus epidermidis* and Other Coagulase-Negative Staphylococci^{∇†}

Peter A. Smith,[‡] Michael E. Powers,[‡] Tucker C. Roberts, and Floyd E. Romesberg*

Department of Chemistry, The Scripps Research Institute, La Jolla, California 92037

Received 21 October 2010/Returned for modification 23 November 2010/Accepted 20 December 2010

The arylomycins are a class of natural-product antibiotics that act via the inhibition of type I signal peptidase (SPase), and we have found in diverse bacteria that their activity is limited by the presence of a resistance-conferring Pro residue in SPase that reduces inhibitor binding. We have also demonstrated that *Staphylococcus epidermidis*, which lacks this Pro residue, is extremely susceptible to the arylomycins. Here, to further explore the potential utility of the arylomycins, we report an analysis of the activity of a synthetic arylomycin derivative, arylomycin C₁₆, against clinical isolates of *S. epidermidis* and other coagulase-negative staphylococci (CoNS) from distinct geographical locations. Against many important species of CoNS, including *S. epidermidis*, *S. haemolyticus*, *S. lugdunensis*, and *S. hominis*, we find that arylomycin C₁₆ exhibits activity equal to or greater than that of vancomycin, the antibiotic most commonly used to treat CoNS infections. While the susceptibility was generally correlated with the absence of the previously identified Pro residue, several cases were identified where additional factors also appear to contribute.

The coagulase-negative staphylococci (CoNS) are a heterogeneous group of at least 15 different species of Gram-positive bacteria that have emerged in recent decades as important nosocomial pathogens (10, 31). A particularly problematic species is *Staphylococcus epidermidis*, which is responsible for a growing number of infections among hospital patients with compromised immune systems and is especially notorious for forming biofilms that adhere to surgical equipment and other hospital surfaces and indwelling devices (3, 18). Methicillin was traditionally the first-line antibiotic against CoNS, but its widespread use has resulted in resistance in 50% to 80% of CoNS infections and 75 to 90% of nosocomial *S. epidermidis* infections (18). As a result, vancomycin is now the first line agent for treating CoNS infections; however, isolates with reduced susceptibility to vancomycin have also been observed (11, 26), and the emergence of enterococci harboring mobile elements that confer vancomycin resistance has raised concerns that resistance might be transferred to *S. epidermidis* and/or other CoNS (17, 28). These concerns continue to motivate the search for new antibiotics that are active against CoNS, especially *S. epidermidis*.

The arylomycins (Fig. 1) are a novel class of natural-product antibiotics that act by inhibiting bacterial type I signal peptidase (SPase) (19, 25). SPase is a Ser-Lys dyad protease that removes N-terminal signal sequences from preproteins following their translocation across the cytoplasmic membrane (5, 20). SPase is an attractive target for antibiotic therapy because it is conserved, essential, and located in the relatively accessible outer leaflet of the cytoplasmic membrane. Furthermore,

because bacterial SPase acts via a catalytic mechanism that is distinct from that of its eukaryotic homologues, the arylomycins are unlikely to exhibit mechanistic toxicity in humans (5, 20).

Despite the apparent accessibility, essentiality, and conservation of SPase, initial reports suggested that the arylomycins were active against only a few Gram-positive bacteria, including *Streptococcus pneumoniae*, *Rhodococcus opacus*, and *Brevibacillus brevis* (15, 25), and not against other important Gram-positive pathogens or against any Gram-negative bacteria. However, after reporting the first synthesis of an arylomycin, arylomycin A₂, as well as the synthetic derivative arylomycin C₁₆ (Fig. 1), we found that each potently inhibits the growth of *S. epidermidis* (24) and that *S. epidermidis* evolves resistance to the arylomycins by mutating residue 29 of one of its two SPases, SpsIB, from Ser (Ser²⁹) to Pro (Pro²⁹) (29). Moreover, a Pro residue is naturally present at the analogous position in the homologous SPases of the pathogens *Staphylococcus aureus*, *Escherichia coli*, and *Pseudomonas aeruginosa*, and we showed that it imparts resistance by reducing the affinity with which the arylomycins bind. Furthermore, we found that a remarkably diverse range of both Gram-positive and Gram-negative bacteria whose SPases lack a Pro at the analogous position are susceptible to the arylomycins, including *Staphylococcus epidermidis*, *Streptococcus pyogenes*, *Helicobacter pylori*, *Chlamydia trachomatis*, and some strains of *Francisella tularensis* (29). In total, the results suggest that the identified SPase polymorphism is a major contributor to naturally occurring arylomycin resistance. However, we also showed that *Yersinia pestis* and some strains of *S. aureus* are susceptible to the arylomycins despite the presence of an analogous Pro, while others, such as many of the *Lactobacillales*, *Clostridia*, and *Bacteroidetes*, are resistant despite its absence, implying that in some cases, susceptibility must depend on additional factors, such as variable levels of toxicity associated with the inhibition of protein secretion.

The potent activity of the arylomycins against a strain of *S.*

* Corresponding author. Mailing address: Department of Chemistry, The Scripps Research Institute, 10550 North Torrey Pines Road, La Jolla, CA 92037. Phone: (858) 784-7290. Fax: (858) 784-7472. E-mail: floyd@scripps.edu.

[‡] These authors contributed equally to this work.

[†] Supplemental material for this article may be found at <http://aac.asm.org/>.

[∇] Published ahead of print on 28 December 2010.

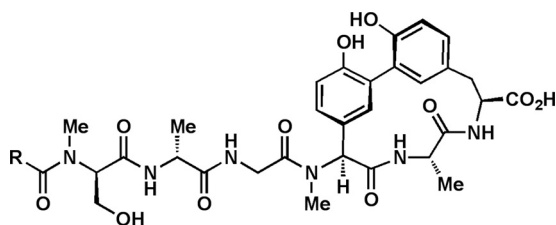


FIG. 1. Structures of arylomycin A_2 ($R = iso-C_{12}$) and arylomycin C_{16} ($R = iso-C_{16}$) [$iso-C_{12} = CH_2(CH_2)_7CH(CH_3)_2$ and $iso-C_{16} = CH_2(CH_2)_{11}CH(CH_3)_2$].

epidermidis (RP62A) suggests that they might be useful in the treatment of this and perhaps other CoNS. Here, to examine the spectrum of activity of the arylomycins against clinical isolates of *S. epidermidis* and other CoNS, we report the activity of arylomycin C_{16} against two panels of isolates from hospitals in geographically diverse locations and compare the activity to that of vancomycin. The results reveal that the arylomycins have potent antibacterial activity against a range of important CoNS species whose SpsIB orthologs lack the previously identified resistance-conferring Pro, while less activity is observed against species where Pro is present. While we generally observed similar susceptibilities for different isolates within a species, significant differences were observed in several cases, with one atypical instance of susceptibility resulting from the presence of a Ser in place of the resistance-conferring Pro. Significant differences in susceptibility between isolates of the same species are usually observed with clinically deployed antibiotics where selection for resistance has occurred during therapy (2, 9, 14, 21), and therefore, these results may be relevant to understanding the natural evolution of arylomycin resistance in nature.

MATERIALS AND METHODS

A total of 282 nonduplicate, clinical isolates of CoNS, identified to species level, were obtained from the London Health Services Centre, London, Ontario, Canada (13). Of these, 143 isolates were *S. epidermidis*, while the remaining 139 consisted of 12 groups comprised of various numbers of isolates of *Staphylococcus capitis*, *Staphylococcus caprae*, *Staphylococcus cohnii*, *Staphylococcus haemolyticus*, *Staphylococcus lugdunensis*, *Staphylococcus saprophyticus*, *Staphylococcus simulans*, *Staphylococcus warneri*, and *Staphylococcus hominis*. This panel of CoNS is a subset of a larger panel whose susceptibility to quinupristin-dalfopristin, linezolid, telithromycin, and vancomycin has been reported (13). A second set of nonduplicate clinical isolates of CoNS not identified to species level was obtained from the Department of Microbiology, University of California San Diego Medical Center Hillcrest, San Diego, CA. These strains were collected from patients with infections of various coagulase-negative bacteria between April and June of 2008.

Arylomycin C_{16} was synthesized as described previously (24). The MICs of arylomycin C_{16} were determined for isolates from both sets of CoNS using a modified CLSI agar dilution method. Bacterial inocula were prepared from log-phase cultures or from suspensions of colonies grown on solid medium, diluted appropriately, and spotted onto tryptic soy agar at a concentration of 1×10^4 CFU/spot. Spots were then incubated for 24 h at 37°C, and the MIC was defined as the concentration at which there was no visible growth. MICs of vancomycin were also determined, in an identical manner, to provide a reference activity. Several of the isolates demonstrated heterogeneous growth on solid medium, and in these cases, individual colonies of each morphotype were analyzed separately and identified to species level based on sequencing and phylogenetic analysis of a portion of the *dnaJ* gene, which has previously been demonstrated to serve as a sensitive measure to identify staphylococci to species level (see Fig. S1 in the supplemental material) (27). Species-level identification was also confirmed by *dnaJ* sequence analysis for isolates whose arylomycin MICs

deviated from the MIC₅₀ by more than 4-fold (see Fig. S1 in the supplemental material). Additionally, the *spsIB* genes from these isolates were sequenced to determine whether any polymorphisms were present that could account for the observed variation in susceptibility. Primers for the PCR amplification of *spsIB* genes were designed based on regions of highly conserved sequence within the upstream open reading frame SERP0551 (*S. epidermidis* RP62A numbering) and the downstream *rexB* gene (see Fig. S2 in the supplemental material). Degeneracies were included at positions that varied within the sequenced strains of the CoNS in order to maximize the likelihood of annealing (see Table S1 in the supplemental material).

To determine with high confidence the history of speciation within the CoNS examined in this study, a phylogenetic analysis was performed using regions of four essential genes that have previously been validated as speciation markers (16S rRNA genes, *rpoB*, *groEL*, and *dnaJ*) (see Table S2 in the supplemental material) (1, 6, 7, 27). For each protein-coding gene, published DNA sequences from strains identified to species level were translated into protein sequences and aligned using MUSCLE (8). The DNA sequences were then mapped onto the amino acid alignment using the program TranAlign (23). Aligned 16S rRNA gene sequences were obtained from the Ribosomal Database Project (4). For each organism, the four aligned sequences were concatenated, and phylogenetic analysis was conducted by the maximum-likelihood method with PhyML 3.0 (12), using the HKY85 nucleotide substitution model, 20 rate categories, and SPR (subtree pruning and regrafting) branch improvement. The phylogeny of SPase genes was determined similarly, using sequences deposited in the NCBI database as well as sequences obtained in the present study. Trees were displayed using MEGA4 (30).

RESULTS

A total of 143 isolates of *S. epidermidis* from the London Health Services Center were analyzed. The MIC₅₀ and MIC₉₀ of arylomycin C_{16} against these 143 *S. epidermidis* isolates were found to be 0.5 μg/ml and 1 μg/ml, respectively (Table 1). These susceptibilities are similar to that reported previously for the *S. epidermidis* laboratory strain RP62A (MIC = 0.5 μg/ml) (24) and compare favorably to the MIC₅₀ and MIC₉₀ of vancomycin, both of which we found to be 2 μg/ml, in agreement with values reported previously (13). Additionally, the range of MICs observed among the 143 isolates was narrow, with the most resistant isolate having an arylomycin C_{16} MIC of 4 μg/ml.

The remaining 9 species of CoNS examined displayed arylomycin C_{16} susceptibilities ranging from highly susceptible to resistant (Table 1). Specifically, we found that *S. haemolyticus*, *S. lugdunensis*, and *S. hominis* are highly susceptible to arylomycin C_{16} , with MIC₅₀ and MIC₉₀ values of 0.25 to 2 μg/ml. In contrast, we found that *S. capitis*, *S. caprae*, and *S. cohnii* are only moderately susceptible, with MIC₅₀ values between 8 and 16 μg/ml and MIC₉₀ values between 16 and 32 μg/ml, and that *S. warneri* and *S. saprophyticus* are resistant, with MIC₅₀ values equal to or greater than 64 μg/ml and MIC₉₀ values of more than 64 μg/ml. Finally, while many *S. simulans* isolates were sensitive to the arylomycins (MIC₅₀ of 2 μg/ml), a significant number of isolates were highly resistant, resulting in a MIC₉₀ of >64 μg/ml.

Under laboratory conditions, *S. epidermidis* RP62A evolves arylomycin resistance by mutation of Ser²⁹ to Pro²⁹ in SpsIB, one of two active SPases in this organism; in the related organism *S. aureus*, as well as in the more distantly related *E. coli* and *P. aeruginosa*, an analogous Pro²⁹ is naturally present and contributes significantly to arylomycin resistance (29) (here and throughout the remainder of the manuscript, unless otherwise indicated, Ser²⁹ and Pro²⁹ refer to the residue in SpsIB orthologs at the position corresponding to 29 in *S. epidermidis*).

TABLE 1. MICs of arylomycin C₁₆ and vancomycin for coagulase-negative staphylococcus strains, identified to species level, obtained from the London Health Services Centre

Species	No. of isolates	MIC ($\mu\text{g/ml}$) of:					
		Arylomycin C ₁₆			Vancomycin		
		MIC ₅₀	MIC ₉₀	Range	MIC ₅₀	MIC ₉₀	Range
<i>S. epidermidis</i>	143	0.5	1	0.12–4	2	2	1–2
<i>S. lugdunensis</i>	10	0.25	0.25	0.03–32	2	2	2
<i>S. hominis</i>	17	0.25	0.5	0.06–0.5	2	4	1–4
<i>S. haemolyticus</i>	10	2	2	1–2	2	2	2–4
<i>S. simulans</i>	12	2	>64	2–>64	1	2	1–2
<i>S. caprae</i>	10	8	16	4–16	2	2	1–2
<i>S. cohnii</i>	27	8	16	4–16	2	2	1–2
<i>S. capitis</i>	24	16	32	4–64	2	2	1–2
<i>S. warneri</i>	19	64	>64	2–>64	4	4	1–4
<i>S. saprophyticus</i>	10	>64	>64	>64	1	2	1–2
<i>S. xylosum</i> ^a				4			
<i>S. aureus</i> ^a				>64 ^b			

^a Reference strains *S. xylosum* ATCC 29971 and *S. aureus* NCTC 8325 are included for comparison.

^b From reference 24.

In contrast, we did not observe mutations in the second *S. epidermidis* SPase gene, *spsI*, in any of the resistant isolates. To determine whether similar mutations might be responsible for the differences in arylomycin susceptibility observed among the different CoNS, we examined the sequences of their *spsI* and *spsIB* genes (Fig. 2; also see Table S3 in the supplemental material). To identify SPase sequences, we used the NCBI BLAST interface to search the sequenced genomes of *S. capitis*, *S. haemolyticus*, *S. hominis*, *S. lugdunensis*, *S. saprophyticus*, and *S. warneri*, and found that, except for *S. saprophyticus*, each

possesses both *SpsI* and *SpsIB* homologs. We also included the recently reported sequence of the *spsIB* gene from *Staphylococcus xylosum* (22), which does not have an *SpsI* SPase (*S. Leroy*, personal communication). Because no correlation between susceptibility and the sequence of *spsI* was observed and because *S. epidermidis* evolves resistance by mutation of *spsIB*, we PCR amplified and sequenced the *spsIB* orthologs from *S. caprae*, *S. cohnii*, and *S. simulans*, whose genomes have not been fully sequenced (see the supplemental material). Overall, the data support the model that Pro²⁹ contributes significantly to arylomycin resistance within CoNS. *S. lugdunensis*, *S. haemolyticus*, *S. hominis*, and a significant percentage of *S. simulans* isolates are extremely susceptible, and these species lack Pro²⁹. In contrast, the CoNS species most resistant to the arylomycins, *S. warneri* and *S. saprophyticus*, each harbor an SPase with Pro²⁹. The correlation between Pro²⁹ and arylomycin susceptibility is less clear for species that display intermediate susceptibilities; for example, while *S. caprae* and *S. capitis* have Pro²⁹ and *S. cohnii* has Ser²⁹, all three display MIC₅₀ values between 4 and 16 $\mu\text{g/ml}$.

Most of the species examined have the narrow range of susceptibilities to arylomycin C₁₆ that is expected for an antibiotic that has not been used clinically (2, 9, 14, 21) (Table 1). However, for several species, we found a greater range in susceptibilities or we found outliers with susceptibilities that differed significantly from those of the other isolates (Table 1; also see Fig. S3 in the supplemental material). For example, one isolate of *S. lugdunensis* is significantly more susceptible than the species average (its MIC is 8-fold below the MIC₅₀), while a second isolate is unusually resistant (its MIC is 128-fold above the MIC₅₀). Sequencing of the *spsIB* genes revealed identical sequences at the amino acid level (including at residue 29) (see Fig. S4 in the supplemental material). In addition, a bimodal distribution of MICs was observed for isolates of *S. warneri* and *S. simulans*, with peaks at 16 $\mu\text{g/ml}$ and >64 $\mu\text{g/ml}$ and at 2 $\mu\text{g/ml}$ and >64 $\mu\text{g/ml}$, respectively (see Fig. S3 in the supplemental material). Sequencing the *spsIB* genes from representative isolates of each group revealed nearly identical SPase sequences and no variation at residue 29 (see Fig. S4 in

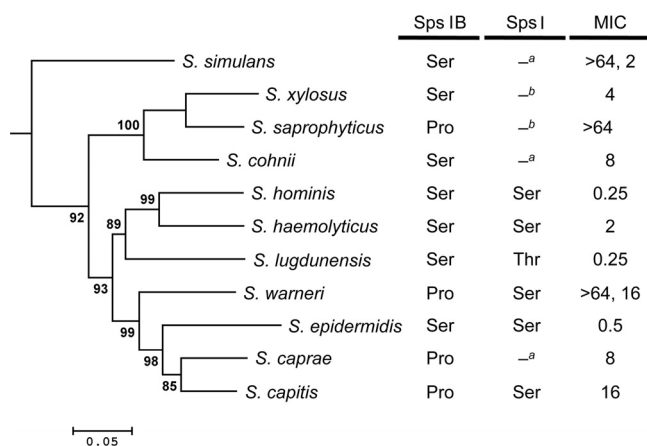


FIG. 2. Phylogenetic relationship, SPase sequence, and arylomycin susceptibility of CoNS species. The tree was constructed from the concatenated DNA sequence of several conserved genes using the method of maximum likelihood (see Materials and Methods). *Macrococcus caseolyticus* was used as an outgroup to root the tree, and alternative likelihood ratio test values are indicated at the branches to reflect the confidence of the branching pattern. The scale bar length corresponds to 0.05 substitutions per nucleotide position. For each species, the amino acids in *SpsIB* and in *SpsI* at the positions corresponding to residue 29 in *S. epidermidis* *SpsIB* are indicated. MIC values reflect the MIC₅₀ except for *S. simulans* and *S. warneri*, where the values listed correspond to peaks in a bimodal distribution of MICs. *a* and *b* indicate that the presence and/or sequence of an *SpsI* homologue is unknown (*a*) or that the species does not have an *SpsI* homologue (*b*).

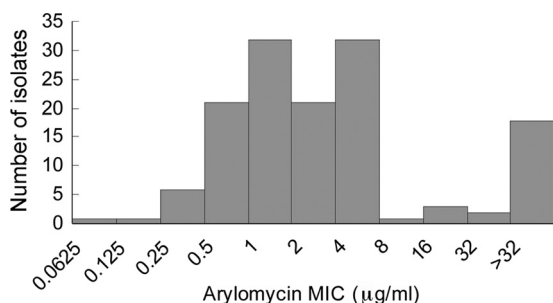


FIG. 3. Distribution of arylomycin susceptibilities of CoNS isolates not identified to species level, obtained from the University of California San Diego Medical Center.

the supplemental material). Finally, we identified a particularly susceptible isolate of *S. warneri*, with a MIC that is 32-fold below the species MIC₅₀. Interestingly, the particularly susceptible strain of *S. warneri* had Ser²⁹ as opposed to Pro²⁹, which is typically present in *S. warneri* isolates, further supporting the genotype/phenotype correlation.

The observed differences in MICs that cannot be accounted for by differences in SpsIB sequence, both within and between species, suggest that additional factors contribute to arylomycin susceptibility. To determine whether these additional factors are shared among related organisms, we examined MICs as a function of phylogeny (Fig. 2). The phylogenetic analysis, based on four highly conserved genes (see Materials and Methods), suggests that *S. simulans* was the earliest species to diverge from the common ancestor of the examined CoNS, with the other species forming two groups, one containing *S. xylosus*, *S. saprophyticus*, and *S. cohnii* and the second containing the remaining species. Notably, SpsI SPases have only been shown to be present in the second group, although their presence cannot be ruled out in the unsequenced species.

S. saprophyticus, which has Pro²⁹ and is extremely resistant to the arylomycins (MIC₅₀ > 64 µg/ml), is most closely related to *S. cohnii*, which lacks Pro²⁹, and yet is only moderately susceptible to the arylomycins (MIC₅₀ = 8 µg/ml). To test whether other species that are closely related to *S. saprophyticus* and *S. cohnii* share this intrinsically lower susceptibility, we determined the MIC of a typed strain of *S. xylosus*, which is closely related to *S. saprophyticus* but, like *S. cohnii*, lacks Pro²⁹. As expected, the strain of *S. xylosus* is moderately susceptible to the arylomycin (MIC = 4 µg/ml). Thus, this group of related bacteria appears to have a lower basal level of arylomycin susceptibility. In contrast, *S. epidermidis*, *S. hominis*, *S. lugdunensis*, and *S. haemolyticus*, which lack Pro²⁹ and display extreme arylomycin susceptibility (MIC₅₀ values of 0.25 to 2 µg/ml), as well as *S. capitis* and *S. caprae*, which have Pro²⁹ but remain moderately susceptible (MIC₅₀ values of 8 to 16 µg/ml), are more related to one another than to the other CoNS species examined. Despite the fact that some *S. warneri* isolates are extremely resistant, the remaining isolates have MICs of ~16 µg/ml, which is consistent with the level of resistance observed for the other CoNS with Pro²⁹ within the more susceptible phylogenetic group.

Finally, we evaluated the activity of arylomycin C₁₆ against strains from a CoNS panel that has not been identified to the

species level (obtained from the University of California San Diego Medical Center) and observed MIC₅₀ and MIC₉₀ values of 2 µg/ml and >32 µg/ml, respectively (Fig. 3). In addition to revealing a portion of resistant isolates, the distribution of MICs shows significant variations in the susceptibility of the remaining strains. Although interpretation of these results is complicated by the lack of species identification, the MIC distribution is consistent with a model in which both the presence of Pro²⁹ and a second, yet-to-be-determined factor combine to yield tiers of arylomycin susceptibility.

DISCUSSION

Members of the arylomycin class of natural-product antibiotics act via a novel mechanism of action, the inhibition of SPase and, thus, the inhibition of the essential process of protein secretion. We found that, in addition to excellent activity against *S. epidermidis* (24), arylomycin C₁₆ has potent activity against *S. haemolyticus*, *S. lugdunensis*, and *S. hominis*. Each of these members of the CoNS has an SPase with Ser²⁹, consistent with the central role of this residue in determining arylomycin susceptibility that we observed previously with other bacteria (29).

The remaining species examined range from only moderately susceptible to extremely resistant, and Pro²⁹ clearly makes a significant contribution to resistance in many of these cases. Interestingly, phylogenetic analysis suggests that Ser²⁹ was prevalent during speciation but that Pro²⁹ evolved in at least two independent instances, once in *S. saprophyticus* and once in the common ancestor of *S. caprae* and *S. capitis* (Fig. 2). Pro²⁹ in *S. warneri* may represent a third instance, although it might instead result from common ancestry with *S. caprae* and *S. capitis*; this distinction is beyond the resolution of the current analysis. Interestingly, many synonymous and nonsynonymous mutations differentiate the *spsIB* genes of the different staphylococcal species, suggesting that ample mutational diversity has been sampled, but only Ser and Pro appear to be tolerated at position 29. Importantly, the multiple independent instances where Pro²⁹ was introduced during staphylococci speciation suggest that there may be a natural selective pressure for Pro²⁹, and the variation within isolates of an extant species suggests that this pressure may have occurred recently and may even still be present. While the nature of the selection pressure is currently unknown, it is tempting to consider that it may be related to the presence of arylomycins in nature (29).

The observed differences in susceptibilities between some of the staphylococcal species reveal that, in addition to SpsIB residue 29, there must be other factors that also contribute to overall susceptibility, and these other factors generally appear to be shared among closely related species, resulting in groups of species having higher or lower basal susceptibilities. One possibility is that the presence of a second SPase gene, orthologs of *S. epidermidis* *spsI*, in some species of CoNS could contribute to their different arylomycin susceptibilities. In principle, expressing two arylomycin-sensitive SPase proteins could result in hypersusceptibility if each SPase recognized a different subset of essential preprotein substrates. Alternatively, expressing one sensitive and one resistant SPase could result in either a susceptible or resistant phenotype, depending on the exact mechanism of arylomycin-induced cell death, which is yet

to be determined. Interestingly, all of the species with higher basal susceptibilities have two SPases, while *S. xyloso* (S. Leroy, personal communication) and *S. saprophyticus* have only one SPase and display lower basal susceptibilities. Consistent with a possible role for a second SPase contributing to arylomycin susceptibility, *S. aureus* strain 8325 is highly resistant to arylomycin (MIC > 128 µg/ml) (24) despite being closely related to the more susceptible group of CoNS, and this species has only a single SPase, apparently due to a recent deletion of its *sp*S**1 homologue. Further experiments will be required to elucidate the potential role of multiple SPases in CoNS susceptibilities and/or to identify other factors contributing to susceptibility, such as differences in the composition of secreted proteins or the presence of modifying enzymes. Comparison of resistant and susceptible isolates of the same species may provide a valuable approach to identifying additional factors that contribute to arylomycin susceptibility.

Regardless of the origins of arylomycin susceptibility and resistance, it is clear that a broad and clinically important (16) range of CoNS strains are susceptible to the arylomycins. In fact, in these cases, the level of susceptibility compares favorably with the level of susceptibility to vancomycin, the antibiotic currently recommended for treatment of CoNS infections but for which resistance is a major concern. The availability of a novel antibiotic, with a novel mechanism of action, would significantly improve CoNS therapy. Along with previously reported data (29), the data also suggest that if the arylomycin class of natural products could be optimized to overcome the loss in affinity mediated by the presence of the resistance-conferring Pro²⁹, then they would have broad activity against CoNS and, possibly, other bacteria as well.

ACKNOWLEDGMENTS

We thank S. Leroy and Genoscope (Genomics Institute of CEA, Evry, France) for providing analysis of the unpublished *S. xyloso* genome sequence.

This work was supported by the Office of Naval Research (awards N000140310126 and N000140810478) and the National Institutes of Health (grant AI081126).

REFERENCES

- Barros, E. M., N. L. Iorio, C. Bastos Mdo, K. R. dos Santos, and M. Giambiagi-deMarval. 2007. Species-level identification of clinical staphylococcal isolates based on polymerase chain reaction-restriction fragment length polymorphism analysis of a partial *groEL* gene sequence. *Diagn. Microbiol. Infect. Dis.* **59**:251–257.
- Bowker, K. E., P. Caspers, B. Gaucher, and A. P. MacGowan. 2009. In vitro activities of three new dihydrofolate reductase inhibitors against clinical isolates of gram-positive bacteria. *Antimicrob. Agents Chemother.* **53**:4949–4952.
- Cheung, G. Y., and M. Otto. 2010. Understanding the significance of *Staphylococcus epidermidis* bacteremia in babies and children. *Curr. Opin. Infect. Dis.* **23**:208–216.
- Cole, J. R., et al. 2009. The Ribosomal Database Project: improved alignments and new tools for rRNA analysis. *Nucleic Acids Res.* **37**:D141–D145.
- Dalbey, R. E., M. O. Lively, S. Bron, and J. M. van Dijk. 1997. The chemistry and enzymology of the type I signal peptidases. *Protein Sci.* **6**:1129–1138.
- De Buyser, M. L., A. Morvan, S. Aubert, F. Dilasser, and N. el Solh. 1992. Evaluation of a ribosomal RNA gene probe for the identification of species and subspecies within the genus *Staphylococcus*. *J. Gen. Microbiol.* **138**:889–899.
- Drancourt, M., and D. Raoult. 2002. *rpoB* gene sequence-based identification of *Staphylococcus* species. *J. Clin. Microbiol.* **40**:1333–1338.
- Edgar, R. C. 2004. MUSCLE: multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Res.* **32**:1792–1797.
- Eliopoulos, G. M., et al. 1986. In vitro and in vivo activity of LY 146032, a new cyclic lipopeptide antibiotic. *Antimicrob. Agents Chemother.* **30**:532–535.
- Emori, T. G., and R. P. Gaynes. 1993. An overview of nosocomial infections, including the role of the microbiology laboratory. *Clin. Microbiol. Rev.* **6**:428–442.
- Garrett, D. O., et al. 1999. The emergence of decreased susceptibility to vancomycin in *Staphylococcus epidermidis*. *Infect. Control Hosp. Epidemiol.* **20**:167–170.
- Guindon, S., and O. Gascuel. 2003. A simple, fast, and accurate algorithm to estimate large phylogenies by maximum likelihood. *Syst. Biol.* **52**:696–704.
- John, M. A., C. Pletch, and Z. Hussain. 2002. In vitro activity of quinupristin/dalfopristin, linezolid, telithromycin and comparator antimicrobial agents against 13 species of coagulase-negative staphylococci. *J. Antimicrob. Chemother.* **50**:933–938.
- Jones, R. N., D. M. Johnson, and M. E. Erwin. 1996. In vitro antimicrobial activities and spectra of U-100592 and U-100766, two novel fluorinated oxazolidinones. *Antimicrob. Agents Chemother.* **40**:720–726.
- Kulanthaivel, P., et al. 2004. Novel lipoglycopeptides as inhibitors of bacterial signal peptidase I. *J. Biol. Chem.* **279**:36250–36258.
- Marshall, S. A., W. W. Wilke, M. A. Pfaller, and R. N. Jones. 1998. *Staphylococcus aureus* and coagulase-negative staphylococci from blood stream infections: frequency of occurrence, antimicrobial susceptibility, and molecular (*mecA*) characterization of oxacillin resistance in the SCOPE program. *Diagn. Microbiol. Infect. Dis.* **30**:205–214.
- Noble, W. C., Z. Virani, and R. G. Cree. 1992. Co-transfer of vancomycin and other resistance genes from *Enterococcus faecalis* NCTC 12201 to *Staphylococcus aureus*. *FEMS Microbiol. Lett.* **72**:195–198.
- Otto, M. 2009. *Staphylococcus epidermidis*—the “accidental” pathogen. *Nat. Rev. Microbiol.* **7**:555–567.
- Paetzel, M., J. J. Goodall, M. Kania, R. E. Dalbey, and M. G. Page. 2004. Crystallographic and biophysical analysis of a bacterial signal peptidase in complex with a lipopeptide-based inhibitor. *J. Biol. Chem.* **279**:30781–30790.
- Paetzel, M., A. Karla, N. C. Strynadka, and R. E. Dalbey. 2002. Signal peptidases. *Chem. Rev.* **102**:4549–4580.
- Petersen, P. J., N. V. Jacobus, W. J. Weiss, P. E. Sum, and R. T. Testa. 1999. In vitro and in vivo antibacterial activities of a novel glycylicline, the 9-t-butylglycylamido derivative of minocycline (GAR-936). *Antimicrob. Agents Chemother.* **43**:738–744.
- Planchon, S., et al. 2007. Proteomic analysis of cell envelope from *Staphylococcus xyloso* C2a, a coagulase-negative staphylococcus. *J. Proteome Res.* **6**:3566–3580.
- Rice, P., I. Longden, and A. Bleasby. 2000. EMBOSS: the European Molecular Biology Open Software Suite. *Trends Genet.* **16**:276–277.
- Roberts, T. C., P. A. Smith, R. T. Cirz, and F. E. Romesberg. 2007. Structural and initial biological analysis of synthetic arylomycin A₂. *J. Am. Chem. Soc.* **129**:15830–15838.
- Schimana, J., et al. 2002. Arylomycins A and B, new biaryl-bridged lipopeptide antibiotics produced by *Streptomyces* sp. Tü 6075. I. Taxonomy, fermentation, isolation and biological activities. *J. Antibiot. (Tokyo)* **55**:565–570.
- Schwalbe, R. S., J. T. Stapleton, and P. H. Gilligan. 1987. Emergence of vancomycin resistance in coagulase-negative staphylococci. *N. Engl. J. Med.* **316**:927–931.
- Shah, M. M., et al. 2007. *dnaJ* gene sequence-based assay for species identification and phylogenetic grouping in the genus *Staphylococcus*. *Int. J. Syst. Evol. Microbiol.* **57**:25–30.
- Sieradzki, K., R. B. Roberts, D. Serur, J. Hargrave, and A. Tomasz. 1999. Heterogeneously vancomycin-resistant *Staphylococcus epidermidis* strain causing recurrent peritonitis in a dialysis patient during vancomycin therapy. *J. Clin. Microbiol.* **37**:39–44.
- Smith, P. A., T. C. Roberts, and F. E. Romesberg. 2010. Broad spectrum antibiotic activity of the arylomycin natural products is masked by natural target mutations. *Chem. Biol.* **17**:1223–1231.
- Tamura, K., J. Dudley, M. Nei, and S. Kumar. 2007. MEGA4: Molecular Evolutionary Genetics Analysis (MEGA) software version 4.0. *Mol. Biol. Evol.* **24**:1596–1599.
- Wisplinghoff, H., et al. 2004. Nosocomial bloodstream infections in US hospitals: analysis of 24,179 cases from a prospective nationwide surveillance study. *Clin. Infect. Dis.* **39**:309–317.