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Practical Synthesis of PC190723, An Inhibitor of the Bacterial Cell Division Protein FtsZ

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Abstract

$$CI$$
 N
 S
 F
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 NH_2
 NH_2

A high-yielding and practical synthesis of the bacterial cell division inhibitor PC190723 is described. The synthesis is completed in a longest linear sequence of five steps from commercially available starting materials and can be readily executed on a multi-gram scale.

The emergence of pathogenic bacteria that are resistant to current antibiotics requires the exploration of new methods for treating infections caused by these organisms. Recent efforts have included finding new ways to modify the structures of existing antibiotics as well as discovering and developing new leads and new molecular targets. One promising area of development involves inhibiting bacterial cell division. 1 The central protein required for bacterial cytokinesis is FtsZ, a tubulin-like GTPase.2 Although FtsZ and tubulin share little sequence homology, X-ray crystallography revealed a high level of structural similarity. 3 Tubulin's function is effectively inhibited by a wide range of natural and synthetic inhibitors, some of which have become effective treatments for cancer (e.g. taxol). Binding sites for three potent tubulin inhibitors (colchicine, taxol, and *Vinca* alkaloids) have been elucidated by crystallography. 4 In contrast, few selective inhibitors of FtsZ are known, and most exhibit modest potency as judged by inhibition of GTP hydrolysis. 5 Moreover, there has been no direct elucidation (e.g. X-ray crystallography, NMR) of the binding sites for any FtsZ inhibitors.

We have initiated a program to synthesize and study naturally-occurring6 and synthetic inhibitors of FtsZ with a long term goal of elucidating their mechanism of inhibition. Researchers at Prolysis, Inc. recently described the activity of PC190723, a heterocyclic inhibitor of FtsZ that exhibits a high level of bacterial lethality against Gram-positive organisms and promising antimicrobial activity against Staphylococcus aureus infection in a mouse model.7 This compound represents an impressive optimization of the known cell-division inhibitor 3-methoxybenzamide (3MBA).8 3MBA shows activity *in vivo* at 10 mM (10,000 μ M) and analysis of resistant mutants at the time of its discovery indicated that FtsZ

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was the target. Optimization of this modest lead revealed PC190723, which inhibits the GTPase activity of FtsZ with an IC $_{50}$ of 0.154 μ M. In the absence of direct evidence, analysis of the structure of FtsZ from resistant mutants suggests that this compound interacts with FtsZ at a site outside of the GTP binding pocket.7a In addition, a recent report documents stabilization of FtsZ polymers by 1.9 Herein we report a practical synthesis10 of this compound that will enable more detailed studies of this compound's function and a broader understanding of the structural biology of FtsZ.

Retrosynthetic analysis reveals that PC190723 could be accessed by alkylation of phenol **3–5** with chloromethyl-substituted heterocycle **2** (Scheme 1). Phenols **3–5** are readily available from 2,4-difluorophenol by lithiation and acylation. The required chloroacetamide-substituted pyridine was prepared in two steps from commercially available 2,5-dichloro-3-aminopyridine **8**.

The phenol fragment of 1 could be introduced as either aldehyde 3, amide 4, or ester 5 (Scheme 2). Aldehyde 3 was prepared from difluorophenol in two steps.11 Protection with ethoxymethyl chloromethyl ether (EOM-Cl) followed by regioselective lithiation, acylation with DMF and EOM group removal provided aldehyde 3 in moderate yield. Quenching arylithium reagent 12 10 with CO $_2$ yielded acid 11, which was converted to the requisite amide 4 in two steps. Treatment of 11 with methanolic H_2SO_4 effected both esterification and deprotection to provide 5 in high yield.13

We initially explored several different methods for displacing the 2-chloro group of **8** with NaSH and a variety of nucleophilic sulfur sources. Ultimately this route was abandoned in favor of *N*-acylation followed by sulfurization of the amide with concomitant ring closure (Scheme 3). Acylation of 2,5-dichlo-3-aminopyridine proceeded smoothly under standard conditions. Sulfurization with Lawesson's reagent14 in toluene provided the desired thiazole **2** as well as the dechlorinated product **15** as determined by NMR spectroscopy and mass spectrometry. Further proof of the identity of **15** was provided by synthesis of a reference sample. Reaction of **2** with Lawesson's reagent did not produce **15** and the starting material was recovered unchanged, demonstrating that **2** is stable toward dechlorination. Attempts to convert **15** to **2** by radical chlorination were unsuccessful. Finally, switching from toluene to *p*-xylene for the reaction with Lawesson's reagent increased the yield of **2** and only trace amounts of **15** were observed.

We attempted to solve the problem of dehalogenation during sulfurization by changing the order of events. In an attempt to alkylate 7 directly with aldehyde 3, an interesting cyclization occurred to produce lactam 17. Under the reaction conditions for alkylation, subsequent ring closure by S_N Ar displacement of the ortho fluorine could not be avoided.15 The structure of 17 was suspected based on the signals for diastereotopic methylene protons and confirmed by X-ray crystallographic analysis. Although alkylation of 7 with amide 4 did lead to the desired O-alkylation product (not shown), the subsequent sulfurization-cyclization resulted in a complex mixture of products.

The synthesis of the PC190723 from chloromethyl thiazole **2** was completed in high yield after exploring three possible routes. When alkylation of aldehyde **3** with chloromethyl heterocycle **2** was attempted (Cs_2CO_3 , THF or CH_3CN), multiple products were observed. Reaction of amide **4** with **2** produced **1** in modest yield under mild conditions. Finally, we demonstrated that ester **5** underwent alkylation in higher yield (62%, unoptimized) relative to amide **4**. Unfortunately, the resultant ester could not be converted to **1** by aminolysis under a variety of conditions.16 An alternative route is the conversion of chloride **2** to iodide **20** (78% yield),17 which alkylates **4** in good yield (70%). Although the reaction via **20** is one step longer, the overall yield is slightly higher (55%) when compared to the one-step

process. Characterization data for 1/PC190723 (¹HNMR, ¹³CNMR, ¹⁹FNMR, HRMS, IR) was consistent with published structure of PC190723.

In summary, we have demonstrated a practical synthesis of PC190723 in a longest linear sequence of five steps from 2,4-difluorophenol using a single protecting group. Although the synthesis requires metallation of an intermediate with *n*-BuLi, it is easily scaleable for the production of multi-gram quantities. A principle contrast between the previously reported route and ours is the use of low-cost starting materials, specifically avoiding 2,6-difluoro-3-methoxybenzamide and benzyloxyacetyl chloride. Access to this potent inhibitor of bacterial cell division will enable detailed studies of this essential process18 and pave the way for the development of new inhibitors that might serve as medicinal leads.

Experimental Section

1-Ethoxymethoxy-2,4-difluorobenzene (9)

2,4-difluorophenol (0.999 g, 7.68 mmol) was dissolved in 10 mL of dry CH₂Cl₂. (*i*-Pr)₂NEt (1.58 mL, 9.22 mmol) was added and the mixture was cooled to 0 °C. This was followed by the addition of chloromethyl ethyl ether (9.86 mL, 9.26 mmol). After 2 h of stirring at room temperature, 3M NaOH (20 mL) was added and extracted 2x with CH₂Cl₂. The organic layers were combined and 1M HCl was added (20 mL) and extracted 2x with CH₂Cl₂. The organic layers were combined and washed with brine and dried over K₂CO₃. The solvent was evaporated *in vacuo* yielding a colorless oil (1.32 g, 91%): ¹H NMR (300 MHz, CDCl₃) δ 7.16 (td, J = 9.1, 5,4 Hz, 1H), 6.90-6.71 (m, 2H), 5.19 (s, 2H), 3.76 (q, J = 7.1 Hz, 2H), 1.22 (t, J = 7.1 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 157.4 (dd, J = 242.7, 10.5 Hz), 153.2 (dd, J = 248.8, 12.1 Hz), 141.7 (dd, J = 10.9, 3.6 Hz), 119.1 (dd, J = 9.4, 2.6 Hz), 110.6 (dd, J = 22.5, 3.9 Hz), 104.8 (dd, J = 26.8, 22.6 Hz), 95.0, 64.8, 14.9; R_f 0.851 in 30% EtOAc/hexanes; HRMS (ESI) m /z calcd for C₉H₁₀F₂O₂ (M - H)⁻ 187.0574, found 187.0579.

3-Ethoxymethoxy-2,6-difluorobenzoic acid (11)

Compound **9** (3.93 g, 20.9 mmol) was dissolved in dry THF (80 mL) and *n*-BuLi (10 mL, 25.0 mmol, 2.5M in hexanes) was added dropwise at -78 °C. After stirring at -78 °C for 2 h, the mixture was poured onto freshly crushed CO₂ followed by the addition of another layer of freshly crushed CO₂. The mixture was allowed to warm to room temperature overnight. Water was added to the mixture (50 mL) and extracted with EtOAc. The aqueous layer was acidified to pH 1 and extracted immediately with CH₂Cl₂. The organic layers were combined and dried over Na₂SO₄. The solvent was evaporated *in vacuo*, yielding a white solid (4.51 g, 93%): mp 154-154 °C; ¹H NMR (400 MHz, CDCl₃) δ 7.37 (m, 1H), 6.91 (dd, J = 10.1, 8.3 Hz, 1H), 5.25 (s, 2H), 3.80 (q, J = 7.1 Hz, 2H), 1.25 (t, J = 7.1 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 166.4, 155.5 (dd, J = 253.5, 4.4 Hz), 151.9 (dd, J = 260.0, 5.9 Hz), 142.2 (dd, J = 11.0, 3.7 Hz), 122.2 (dd, J = 9.9, 3.2 Hz), 111.6 (dd, J = 23.1, 4.4 Hz), 110.5 (dd, J = 17.7, 14.5 Hz), 95.1, 65.0, 15.1; IR (neat) 3536, 3476, 1671 cm⁻¹; R_f 0.340 in 50% EtOAc/hexanes; HRMS (ESI) m/z calcd for C₁₀H₁₀F₂O₄ (M - H)⁻ 231.0469, found 231.0479.

3-Ethoxymethoxy-2,6-Difluoro-benzamide (12)

Compound **11** (1.20 g, 5.18 mmol) was dissolved in 50 mL of dry CH₂Cl₂ followed by the addition of (*i*-Pr)₂NEt (0.976 mL, 5.70 mmol). Ethyl chloroformate (0.600 mL, 6.30 mmol) was added at 0 °C. After 1.5 h at 0 °C, 6 mL of 12M NH₄OH was added and stirred for an additional 30 min. The reaction was monitored by TLC. After 3 h, the solvent was evaporated *in vacuo* and then 1M HCl (30 mL) was added to the mixture and extracted 2x with ethyl acetate. The organic layers were combined, washed with brine and concentrated. The crude mixture was purified by flash chromatography (40 to 70% EtOAc/hexanes). The

solvent was evaporated *in vacuo* yielding a white solid (1.13 g, 94%): mp 87.4–87.5 °C; 1 H NMR (400 MHz, CDCl₃) δ 7.27 (td, J = 9.4, 5.3, 1H), 6.98 – 6.80 (m, 1H), 6.33 (brs, 1H), 6.06 (brs, 1H), 5.23 (s, 2H), 3.78 (q, J = 7.1, 2H), 1.23 (t, J = 7.1, 3H); 13 C NMR (75 MHz, CDCl₃) δ 162.7 , 154.2 (dd, J = 247.3, 5.2), 150.5 (dd, J = 253.9, 7.2), 142.0 (dd, J = 11.1, 3.4), 120.3 (d, J = 9.3), 114.4-113.8 (m), 111.3 (dd, J = 23.5, 3.6), 94.9 , 64.9, 15.1; IR (neat) 3381, 3198, 1656 cm $^{-1}$; R_f 0.409 in 60% EtOAC/hexanes; HRMS (ESI) m / z calcd for $C_{10}H_{11}F_2NO_3$ (M + H) $^+$ 232.0785, found 232.0765.

2,6-Difluoro-3-hydroxy-benzamide (4)

The EOM-protected amide **12** (0.875 g, 3.78 mmol) was dissolved in a 50:50 mixture of 6M HCl and MeOH (30 mL total volume). The reaction was monitored by TLC. After ~2 h, the solvent was evaporated *in vacuo* yielding an amorphous white solid (0.622 g, 95%): 1 H NMR (400 MHz, DMSO-d) δ 9.96 (brs, 1H), 8.09 (brs, 1H), 7.79 (brs, 1H), 6.93 (m, 2H); 13 C NMR (75 MHz, DMSO-d) δ 162.5, 151.3 (dd, J = 238.6, 6.3), 147.6 (dd, J = 245.2, 8.6), 142.1 (d, J = 14.9), 118.1 (dd, J = 9.0, 3.6), 117.1 (dd, J = 24.6, 20.7), 111.6 (dd, J = 22.8, 3.4); IR (neat) 3433, 3371, 1687 cm $^{-1}$; R_f 0.185 in 50% EtOAc/hexanes; HRMS (ESI) m/z calcd for $C_7H_5F_2NO_2$ (M + H) $^+$ 174.0366, found 174.0354. 1 H NMR spectrum was in accordance with the literature10.

2-Chloro-N-(2,5-dichloro-pyridin-3-yl)-acetamide (7)

3-amino-2,5-dichloropyridine **8** (1.02 g, 6.25 mmol) was dissolved in dry CH₂Cl₂ (50 mL). The mixture was cooled to 0 °C and chloroacetyl chloride (0.75 mL, 9.43 mmol) was added dropwise. This was followed by the addition of (*i*-Pr)₂NEt (1.6 mL, 9.35 mmol). The reaction was monitored by TLC. After 3 h, the solvent was evaporated *in vacuo* and the crude mixture was purified by flash chromatography (30 to 50% EtOAc/hexanes) yielding an off-white solid (1.44 g, 96%): mp 121–122 °C; ¹H NMR (300 MHz, CDCl₃) δ 8.94 (s, 1H), 8.82 (d, J = 2.4, 1H), 8.14 (d, J = 2.4, 1H), 4.26 (s, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 164.7, 143.3, 137.9, 131.6, 131.4, 128.1, 42.7; IR (neat) 3366, 2946, 1682 cm⁻¹; R_f 0.833 in 50% EtOAc/hexanes; HRMS (ESI) m / z calcd for C₇H₅Cl₃N₂O (M + H)⁺ 238.9545, found 238.9512.

6-Chloro-2-chloromethyl-thiazolo[5,4-b]pyridine (2)

Compound **7** (0.561 g, 2.34 mmol) was dissolved in dry toluene (25 mL) and Lawesson's reagent (0.946 g, 4.68 mmol) was added. The reaction mixture was refluxed for 12 h at 110 °C. The reaction was monitored by mass spectroscopy for disappearance of starting material. After ~12 h, the color of the reaction mixture turned slightly red. At this point, the reaction was stopped and cooled to room temperature. The crude mixture was passed through a plug of silica gel and washed with CH₂Cl₂. The solvent was evaporated *in vacuo* and the crude mixture purified by flash chromatography (30 to 50% EtOAc/hexanes) yielding a solid (0.307 g, 60%): mp 89–90 °C; ¹H NMR (600 MHz, CDCl₃) δ 8.58 (d, J = 2.1, 1H), 8.23 (d, J = 2.1, 1H), 4.92 (s, 2H); ¹³C NMR (150 MHz, CDCl₃) δ 170.4, 156.6, 146.8, 146.5, 130.3, 130.1, 42.4; IR (neat) 3038, 1736, 1682 cm⁻¹; R_f 0.833 in 50% EtOAc/hexanes; HRMS (ESI) m/z calcd for C₇H₄Cl₂N₂S (M + H)⁺ 218.9850, found 218.9839.

3-((6-Chloropyrido[3,2-d][1,3]thiazol-2-yl}methoxy)-2,6-difluoro- benzamide (1)

Compound **4** (0.250 g, 1.44 mmol) was dissolved in 20 mL of dry CH₃CN and Cs₂CO₃ (0.94 g, 2.88 mmol) was added. Compound **2** (0.211 g, 0.963 mmol) was added to the mixture and stirred at 65 °C for ~3 h. The solution was filtered, concentrated *in vacuo* and purified by flash chromatography (30 to 70% EtOAc/hexanes) yielding an off-white solid (0.135 g, 40%): m.p. 212.5–214.5 (lit¹⁰. 218°C); R_f 0.333 in 50% EtOAc/hexanes; ¹H NMR (600 MHz, DMSO-d) δ 8.73 (d, J = 2.2, 1H), 8.68 (d, J = 2.2, 1H), 8.18 (brs, 1H), 7.90 (brs,

2H), 7.41 (m, 2H), 7.12 (m, 2H), 5.73 (s, 2H); 13 C NMR (100 MHz, DMSO-d) δ 171.5, 161.8, 156.2, 154.3–154.4 (dd, J = 235.49, 7.0), 152.1-152.0 (dd, J = 239.7, 9.0), 149.9 (d, J = 8.3), 147.4 (d, J = 8.3Hz), 142.5 (d, J = 10.9), 130.6, 129.9, 117.4 (dd, J = 41.7, 17.0), 117.0 (d, J = 8.8), 111.9 (d, J = 23.1), 69.6; 19 F NMR (282 MHz, DMSO-d) δ -123.4, -134.1; IR 3393, 3198, 1671 cm⁻¹; HRMS (ESI) m/z calcd for $C_{14}H_{8}ClF_{2}N_{3}O_{2}S$ (M + H)⁺ 356.0072, found 356.0051. 1 H NMR spectrum was in accordance with the literature10.

3-((6-Chloropyrido[3,2-d][1,3]thiazol-2-yl}methoxy)-2,6-difluoro- benzamide (1)

Compound **4** (0.100 g, 0.579 mmol) was dissolved in 20 mL of dry CH₃CN and K_2 CO₃ (0.241 g, 1.73 mmol) was added. Iodide **20** (0.120 g, 0.386 mmol) was added and stirred at 65 °C for ~3 h. The solution was filtered, concentrated *in vacuo* and purified by flash chromatography (30 to 70% EtOAc/hexanes) yielding an off-white solid (0.097 g, 70%). ¹H NMR spectrum was in accordance with the literature 10.

4-(2,5-Dichloro-pyridin-3-yl)-6-fluoro-3-oxo-3,4-dihydro-2H-benzo[1,4]oxazine-5-carbaldehyde (17)

Compound **7** (0.314 g, 1.31 mmol) was dissolved in dry DMF (15 mL) and NaI (0.197 g, 1.31 mmol) was added and stirred at 45 °C for 40 min. Then compound **3** (0.249 g, 1.57 mmol) and K_2CO_3 (1.0 g, 7.23 mmol) were added and the mixture was stirred for 2–3 h. The solution was filtered, concentrated *in vacuo* and purified by flash chromatography (30 to 50% EtOAc/hexanes) yielding a white solid (0.295 g, 66%); mp 190–191 °C; R_f 0.833 in 50% EtOAc/hexanes; ¹H NMR (300 MHz, CDCl₃) δ 9.90 (s, 1H), 8.34 (d, J = 2.4, 1H), 7.90 (d, J = 2.4, 1H), 7.37 (dd, J = 9.0, 5.1, 1H), 7.00 (m, 1H), 4.82 (d, J = 14.3, 1H), 4.63 (d, J = 14.3, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 185.2 (d, J = 10.6), 164.3, 162.7, 159.3, 148.0, 145.5 (d, J = 14.7), 141.3, 132.9, 130.5, 129.2, 123.4 (d, J = 10.5), 115.9 (d, J = 11.3), 113.2 (d, J = 23.8), 68.8; ¹⁹F NMR (282 MHz, CDCl₃) δ -125.0; IR (neat) 1724, 1694 cm⁻¹; HRMS (ESI) m/z calcd for $C_{14}H_7Cl_2FN_2O_3$ (M + H)+ 340.9896, found 340.9897.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

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References

- 1. a) Kapoor S, Panda D. Expert Opin. Ther. Targets. 2009; 13:1037–1051. [PubMed: 19659446] b) Vollmer W. Chem. Biol. 2008; 15:93–94. [PubMed: 18291311]
- a) Romberg L, Levin PA. Ann. Rev. Microbiol. 2003; 57:125–154. [PubMed: 14527275] b)
 Margolin W. Nat. Rev. Mol. Cell Biol. 2005; 6:862–871. [PubMed: 16227976] c) Dajkovic A,
 Lutkenhaus J. J. Mol. Microbiol. Biotechnol. 2006; 11:140–151. [PubMed: 16983191] d) Callaway
 E. Nature. 2008; 451:124–126. [PubMed: 18185562]
- a) Lowe J. J. Struct. Biol. 1998; 124:235–243. [PubMed: 10049809] b) Leung AK, Lucile White E, Ross LJ, Reynolds RC, DeVito JA, Borhani DW. J. Mol. Biol. 2004; 342:953–970. [PubMed: 15342249] c) Oliva MA, Trambaiolo D, Lowe J. J. Mol. Biol. 2007; 373:1229–1242. [PubMed: 17900614]
- a) Lowe J, Li H, Downing KH, Nogales E. J. Mol. Biol. 2001; 313:1045–1057. [PubMed: 11700061] b) Gigant B, Wang C, Ravelli RB, Roussi F, Steinmetz MO, Curmi PA, Sobel A, Knossow M. Nature. 2005; 435:519–522. [PubMed: 15917812] c) Gigant B, Wang C, Ravelli RB,

- Roussi F, Steinmetz MO, Curmi PA, Sobel A, Knossow M. Nature. 2005; 435:519–522. [PubMed: 15917812]
- a) Vollmer W. Appl. Microbiol. Biotechnol. 2006; 73:37–47. [PubMed: 17024474] b) Jaiswal R, Beuria TK, Mohan R, Mahajan SK, Panda D. Biochemistry. 2007; 46:4211–4220. [PubMed: 17348691] c) Domadia PN, Bhunia A, Sivaraman J, Swarup S, Dasgupta D. Biochemistry. 2008; 47:3225–3234. [PubMed: 18275156] d) Beuria TK, Singh P, Surolia A, Panda D. Biochem. J. 2009; 423:61–69. [PubMed: 19583568] e) Plaza A, Keffer JL, Bifulco G, Lloyd JR, Bewley CA. J. Am. Chem. Soc. 132:9069–9077. [PubMed: 20536175]
- a) Urgaonkar S, La Pierre HS, Meir I, Lund H, RayChaudhuri D, Shaw JT. Org. Lett. 2005; 7:5609–5612. [PubMed: 16321003] b) Kim MB, Shaw JT. Org. Lett. 2010:3324–3327. [PubMed: 20597470]
- 7. a) Haydon DJ, et al. Science (Washington, DC, U. S.). 2008; 321:1673–1675.b) Czaplewski LG, et al. Bioorg. Med. Chem. Lett. 2009; 19:524–527. [PubMed: 19064318]
- 8. Ohashi Y, Chijiiwa Y, Suzuki K, Takahashi K, Nanamiya H, Sato T, Hosoya Y, Ochi K, Kawamura F. J. Bacteriol. 1999; 181:1348–1351. [PubMed: 9973366]
- 9. Andreu JM, Schaffner-Barbero C, Huecas S, Alonso D, Lopez-Rodriguez ML, Ruiz-Avila LB, Nunez-Ramirez R, Llorca O, Martin-Galiano AJ. J. Biol. Chem. 2010; 285:14239–14246. [PubMed: 20212044]
- During the preparation of this manuscript, a report from Prolysis included a synthesis of 1 and analogs prepared by a closely related route: Haydon DJ, et al. J. Med. Chem. 2010; 53:3927–3936. [PubMed: 20426423]
- Chen GT, King M, Gusovsky F, Creveling CR, Daly JW, Chen BH, Nie JY, Kirk KL. J. Med. Chem. 1993:3947–3955. [PubMed: 8254623]
- 12. Marzi E, Gorecka J, Schlosser M. Synthesis. 2004:1609-1618.
- 13. Yamaguchi Y, Menear K, Cohen N-C, Mah R, Cumin F, Schnell C, Wood JM, Maibaum. J. Bioorg. Med. Chem. Lett. 2009:4863–4867.
- 14. 2,4-Bis(4-methoxyphenyl)-1,3,2,4-dithiadiphosphetane 2,4-disulfide.
- 15. For a related two-step transformation of an α-bromoamide, see: Coutts IGC, Southcott MR. J. Chem. Soc. Perkin Trans. 1990; 1:767–771.
- 16. Attempted aminolysis reagents included NH₄OH with various co-solvents, NH₃/methanol, and LiNH₂ in THF generated from NH₄Cl and *n*-BuLi.
- 17. Williams DR, Berliner MA, Stroup BW, Nag PP, Clark MP. Org. Lett. 2005; 7:4099–4102. [PubMed: 16146361]
- 18. Research quantities of this compound are available on request. Please contact JTS by (shaw@chem.ucdavis.edu).

Scheme 1. Retrosynthetic analysis of PC190723 (1).

Scheme 2. Synthesis of 2,6-difluoro-3-salical dehyde 3, amide 4 and methyl ester 5.

8

$$C_{1}$$
 C_{2}
 C_{3}
 C_{4}
 C_{5}
 C_{6}
 C_{6}
 C_{1}
 C_{1}
 C_{1}
 C_{2}
 C_{3}
 C_{4}
 C_{4}
 C_{5}
 C_{5}
 C_{6}
 C_{7}
 C

Scheme 3. Synthesis of chloromethyl thiazoline **2**.

Scheme 4. Attempted alkylation of 7 with phenol 3.

A 3,
$$Cs_2CO_3$$
 CI 4, Cs_2CO_3 CI N S F O NH₂
 CI 18 F O NH₄OH MeOH

 I 19 F O OCH₃
 I 10 OCH₃

Scheme 5.

A) Final alkylation to produce ${\bf 1}$. B) Improved alkylation yield from conversion of chloride ${\bf 2}$ to iodide ${\bf 20}$.