

## Study of Protective Effect of Date and *Nigella Sativa* on Aflatoxin B<sub>1</sub> Toxicity

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### Abstract

**Background:** Many medicinal plants and their purified constituents have been shown beneficial therapeutic potentials. Seeds of *Nigella sativa*, a dicotyledon of the Ranunculaceae family, have been utilized for thousands of years as a spice and food preservative.

**Methods:** the toxic effect of aflatoxin-B<sub>1</sub> (AFB<sub>1</sub>) and the possible cytoprotective effect of *Nigella sativa* (NS) oil and aqueous extract of date were studied on 40 male rats. The animals were divided into 4 groups (10 rats each) and treated daily for two weeks. Group 1 received normal saline as controls. Group 2 treated via intraperitoneal (IP) route with AFB<sub>1</sub> (50µg/kg BW). Group 3 treated with AFB<sub>1</sub> and NS oil via IP. Group 4 treated with AFB<sub>1</sub> and received orally aqueous extract of date (15mg/15ml). The liver and kidneys of each animal were histological examined and biochemical evaluation of the liver and kidney functions was performed.

**Results:** Group 2 showed severe degenerative and necrotic changes in the liver and kidney. The plasma levels of alanine transaminase (ALT), aspartate transaminase (AST), creatinine and urea in AFB<sub>1</sub> group were significantly higher than the control group. Livers and kidneys of rats, treated with AFB<sub>1</sub> and NS showed less histopathological changes in comparison with the AFB<sub>1</sub> treated group. Livers and kidneys of rats treated with AFB<sub>1</sub> and date group showed only mild histopathological changes in comparison with AFB<sub>1</sub> treated group. These histopathological changes seen in animals treated with AFB<sub>1</sub> and dates were associated with a significant reduction in levels of ALT, AST, creatinine and urea. Likewise, histopathological changes in the AFB<sub>1</sub> and NS group were associated with significant reduction in the levels of beforementioned indices. Moreover, AFB<sub>1</sub> and date group showed significant improvement in liver function comparing with AFB<sub>1</sub> and NS group.

**Conclusion:** our study revealed that treatment with AFB<sub>1</sub> induced histopathological changes in the tissues of liver and kidney associated with dysfunction of these organs. Both NS and date reduce the toxic effects of AFB<sub>1</sub> in liver and kidney. But date treatment was more cytoprotective for liver than NS treatment against aflatoxicosis in rats.

**Keywords:** *Nigella sativa*; Date, Aflatoxin B1 toxicity; Liver, Kidney, rats.

### Introduction

Mycotoxins, are structurally diverse toxic fungal metabolites. They represent the most important category of biologically produced natural toxins relative to human health and economic impact worldwide. <sup>(1, 2)</sup> Spurred by the discovery of aflatoxin in the 1960s

the first cases of mycotoxicosis were noted leading to the identification of more than 100 toxigenic fungi and in excess of 300 mycotoxins worldwide. <sup>(3,4)</sup> The diverse chemical structures of mycotoxins account for their differing biological properties and effects. Depending upon the toxins' precise biochemical nature, it may have any of a number of toxic properties including being carcinogenic, tetratogenic, mutagenic, oestrogenic, neurotoxic, or immunotoxic.

Aflatoxins represent a group of closely related difuranocoumarin compounds produced as secondary fungal metabolites of the common molds *Aspergillus flavus*, *Aspergillus parasiticus* and to a lesser extent *Aspergillus nominus*. There are three strains of *Aspergillus* from which four major aflatoxins B<sub>1</sub>, B<sub>2</sub>, G<sub>1</sub>, and G<sub>2</sub> (AFB<sub>1</sub>, AFB<sub>2</sub>, AFG<sub>1</sub> and AFG<sub>2</sub>) are produced. Among these aflatoxins, AFB<sub>1</sub> is the most prevalent and toxic with acute toxicity demonstrated in all species of animals. AFB<sub>1</sub> is also known as being one of the most potent genotoxic agents and hepatocarcinogens. <sup>(4,5)</sup>

The toxicity and carcinogenicity of AFB<sub>1</sub> is thought to be directly linked to its bioactivation, that results in formation of a highly reactive AFB<sub>1</sub> 8,9-epoxide. The bioactivation of AFB<sub>1</sub> occurs primarily by a microsomal cytochrome P450. It is dependent on epoxidation of the terminal furan ring of AFB<sub>1</sub> and is responsible for binding to cellular macromolecules such as DNA, RNA and other protein constituents. <sup>(6,7)</sup> Damage of hepatocytes and the kidney is believed to be the result of this process. <sup>(8)</sup>

Interest in medicinal plants has burgeoned by the increased efficiency of new plant-derived drugs and the growing interest in natural products. Because of the concerns about the side effects of conventional medicine, the use of natural products as an alternative to conventional treatment in healing and treatment of various diseases has been on the rise in the last few decades. The use of plants as medicines dates to the dawn of history. <sup>(9, 10)</sup> Medicinal plants serve as therapeutic alternatives, safer choices, or in some cases, as the only effective treatment. People in separate cultures and places are known to have used the same plants for similar medical problems. A larger number of these plants and their isolated constituents have shown beneficial therapeutic effects, including anti-oxidant, anti-inflammatory, anti-cancer, anti-microbial, and immunomodulatory effects. <sup>(10-13)</sup>

Among the promising medicinal plants, *N. sativa*, a dicotyledon of the Ranunculaceae family, is an amazing herb with a rich historical and religious background. <sup>(14)</sup> The seeds of *N. sativa* are the source of the active ingredients of this plant. It is the black seed referred to by the Prophet Mohammed (pbuhs) as having healing powers. <sup>(14)</sup> Black seed is also identified as the curative black cumin in the Holy Bible and is described as the Melanthion of Hippocrates and Discroides and as the Gith of Pliny. <sup>(15)</sup>

Historically, it has been recorded that *N. sativa* seeds were prescribed by ancient Egyptian and Greek physicians to treat headache, nasal congestion, toothache, and intestinal worms. It was used as a diuretic, and to promote menstruation and increase milk production. <sup>(14)</sup> The seeds of *N. sativa*, known as black seed, black cumin or "Habatul-Barakah," have long been used in folk medicine in the Middle and Far East as a traditional medicine for a wide range of illnesses, including bronchial asthma, headache, dysentery, infections, obesity, back pain, hypertension and gastrointestinal problems. <sup>(16)</sup> Its use in some skin conditions such as eczema has also been recognized worldwide. <sup>(14)</sup> The seeds can be ground to a powder, mixed with a little flour as a binder, and applied

directly to abscesses, nasal ulcers, and rheumatic joints. <sup>(17)</sup> One of the potential properties of the *N. sativa* seed is the ability of one or more of its constituents to reduce toxicity due to its anti-oxidant activities. <sup>(17)</sup>

The fruits of the date palm (*Phoenix dactylifera* L.-Areceae) are commonly consumed in many parts of the world and are a vital component of the diet in most of the Arabian countries. The dates, either fresh or dried, have high sugar content, low fats and protein contents, as well as iron, potassium and antioxidant flavonoids. Moreover, the utilization of date in medicine was reported. <sup>(18)</sup>

The aim of the present study was to study the pathological and biochemical effect of AFB<sub>1</sub> on the rat liver and kidney and to investigate the possible protective effect of the date fruits and *Nigella sativa* seeds on these organs.

## Methods

### *Chemicals*

AFB<sub>1</sub> was obtained from Sigma-Aldrich (St. Louis, MO, USA). *Nigella sativa* oil with high degree of quality assurance was bought from the local market. Dates (Sokkare) were purchased from Qassim area. All other chemicals used were of the analytical grade.

### *Animal treatment*

Forty healthy male albino rats (*Rattus norvegicus*) with average body weight 150–170 gm were used for this study. They were obtained from the Animal House of the Faculty of Medicine, King Saud University. All animals were conditioned at room temperature at natural photoperiod for 1 week before the start of the experiment. A commercial balanced diet and tap water ad libitum were provided. Animals were divided into four groups (10 animals each) and received the tested compounds daily for 2 weeks. The control group received normal saline via intra peritoneal (I.P.) injection. The second group received AFB<sub>1</sub> at a dose 50 µg/kg via I.P injection (19). The third group received AFB<sub>1</sub> and *Nigella sativa* (NS) oil (6mg/Kg) via I.P. injection. The fourth group received AFB<sub>1</sub> and aqueous date extract (15 gm/15 ml) by the oral route (gastric tube) (18). At the end of the experiment, the rats were scarified and livers and kidneys were excised immediately for histopathological studies.

The blood samples from all groups were collected from the orbital plexus of veins in heparinized tubes and were centrifuged at 5000 rpm for 10 min for plasma separation.

### *Histopathological Studies*

The liver and kidney tissue samples were collected and fixed in 10% neutral buffered formalin and processed for histopathology using haematoxylin and eosin staining method according to Carleton et al. (20).

### *Biochemical parameters*

The levels of alanine aminotransferase (ALT) and aspartate aminotransferase (AST) in plasma were estimated by the method of King, (21). Protein content was determined by the method of Lowry et al. (22) using bovine albumin as a standard. The plasma levels of creatinine and urea were estimated according to Fabiny and Ertingshausen (23) and Chaney and Marbach (24) respectively.

### *Statistical analysis*

The results are expressed as mean±standard error (SE). Differences between groups were assessed by one-way analysis of variance (Bonferroni test) using the Prism version 4 software package for Windows. The level of significance was *accepted* with  $P \leq 0.05$ .

## **Results**

### *Biochemical results:*

As shown in Table 1, the plasma liver function indices, ALT & AST in AFB<sub>1</sub> group were significantly higher than control group. The levels of ALT and AST activities were reduced significantly in both AFB<sub>1</sub> and NS group and AFB<sub>1</sub> and date group in comparison with the AFB<sub>1</sub> group. The AFB<sub>1</sub> and NS group & AFB<sub>1</sub> and date group showed significant differences in the levels of transaminases activities.

On the other hand, the levels of creatinine and urea were significantly elevated in the AFB<sub>1</sub> group in comparison with controls. The levels of creatinine and urea were lower significantly in both AFB<sub>1</sub> and date group and AFB<sub>1</sub> and NS group in comparison with the AFB<sub>1</sub> group. The levels of total protein did not change significantly among different groups.

### *Pathological Results:*

*Gross picture:* The liver of male albino rats, intoxicated with AFB<sub>1</sub> alone showed mild hepatomegaly in all animals. Severe haemorrhage and pale, depressed necrotic friable focal lesions were observed in seven cases and yellowish fatty areas with firm nodular structures were observed in three cases. Cut surface showed areas of congestion and haemorrhage. The kidney of the rats intoxicated with AFB<sub>1</sub> alone showed congestion, haemorrhage, swelling and yellowish-white and depressed areas. The liver as well as kidney of rats treated with AFB<sub>1</sub> and NS or AFB<sub>1</sub> and date showed nearly the same gross features. No morphological gross changes are detected either in liver or in kidney of the control group.

*Microscopic picture:* No remarkable histological changes were detected in the control cases either in liver or kidney. The liver of rats, treated with AFB<sub>1</sub> alone, showed severe pan lobular degenerative changes represented by cloudy swelling, vacuolar, hydropic and fatty degenerations in addition to diffuse necrosis of hepatocytes that were represented by nuclear pyknosis, karyorrhexis, cytoplasmic fine granularity and marked eosinophilia. Congestion, haemorrhage, kupffer cell hyperplasia, interstitial edema with perivascular mononuclear cell infiltration were also detected (Fig.: 1-3). In three cases, isolated eosinophilic necrotic masses or patches were present with connective tissue proliferation and mononuclear cell infiltration "Nodular like grossly". Portal tracts showed biliary hyperplasia, perivascular and interstitial edema, congestion and hemorrhage (Fig. 4-6). The kidneys of rats treated with AFB<sub>1</sub> alone, showed cloudy swelling, vacuolar, and hydropic degeneration in the tubular cells. Hyaline casts were seen inside the renal tubules that showed cystic dilatation in some sections, and necrotic changes in others. Renal tubules showed sloughed epithelial cells in their lumina and were surrounded by

polymorph nuclear leukocytes. Blood vessels showed dilatation, congestion and haemorrhage. Glomeruli showed cystic dilatation of glomerular Bowman's spaces, hypercellularity of some glomeruli and sclerosis of others. Interstitial edema, fibrosis and haemorrhages were also observed (Fig.: 7-11). The liver of rats treated with AFB<sub>1</sub> plus NS oil showed less morphological changes, in comparison to those treated by AFB<sub>1</sub> alone. They showed few necrotic cells, biliary hyperplasia, congestion, perivascular as well as intercellular edema with kupffer cell hyperplasia. In two cases, the hepatocytes were nearly normal (Fig.12,13). Kidneys of male albino rats that treated with AFB<sub>1</sub> and NS, showed sub and capsular edema, congestion with perivascular edema and few mononuclear leukocytic cells infiltration. The renal tubules showed mild degenerative changes and few scattered necrotic debris (Fig.: 14). The liver of rats treated with AFB<sub>1</sub> and date were more or less normal. They showed mild congestion and kupffer cell hyperplasia (Fig.:15), whereas the kidneys of the same group were also more or less normal. They occasionally showed hydropic degeneration of the cells of proximal convoluted tubules, hyaline casts, congestion with few mononuclear cells infiltration (Fig. 16).

## Discussion

The liver is the target organ for AFB<sub>1</sub>. Ingestion of this mycotoxin, is known to be capable of inducing acute poisoning, aflatoxicosis, and is believed to be participated in the development of primary liver cancer. <sup>(25)</sup> AFB<sub>1</sub> was shown to be converted into its epoxide. This derivative produces DNA adduction causing DNA strand breaks and point mutations. <sup>(26)</sup> Moreover, under this pathological condition, the active process of cellular self-destruction, apoptosis, can occur. <sup>(27)</sup>

In the present study, the liver enzymes (ALT and AST) in AFB<sub>1</sub> -treated group were significantly higher than in the control group. This finding was confirmed by histopathological examination of liver tissues of AFB<sub>1</sub> -treated rats. The morphological changes in rat hepatocytes, in the current study, were previously described by many investigators. <sup>(28-31)</sup> The morphological changes included cloudy swelling, vacuolar, hydropic and fatty degeneration, diffuse necrosis as well as apoptosis of hepatocytes. Congestion, hemorrhage, kupffer cell hyperplasia, and biliary hyperplasia were also included.

On the other hand, AFB<sub>1</sub> -treated group showed higher levels of creatinine and urea than controls. This finding is keeping in line with findings of Rati et al <sup>(32)</sup> Histological examination of kidney tissues of the same group showed previously mentioned microscopic changes that were in agreement with the kidney dysfunction shown by biochemical tests.

Souza et al<sup>(33)</sup> reported that the oxidative stress is the principle manifestations of AFB<sub>1</sub>-induced toxicity which could be mitigated by antioxidants. Previously, many researchers investigated the effect of different antioxidants such as melatonin on AFB<sub>1</sub>. <sup>(27)</sup> Recently, Abdel-Wahhab and Aly <sup>(34)</sup> found that treatment with *Nigella sativa* oil of rats fed with aflatoxin-contaminated diet resulted in significant protection against aflatoxicosis. In the current study, the liver enzymes (ALT and AST) in AFB<sub>1</sub> and NS group were significantly higher than controls but these levels were reduced significantly in comparison with AFB<sub>1</sub> treated group. Histopathologically, the liver and kidney treated

with AFB<sub>1</sub> and NS showed less morphological changes in comparison to the changes seen in AFB<sub>1</sub> alone, which is an indication for partial protection.

The antioxidant effects of *N. Sativa* have been examined using different hepatic and kidney toxicity in vivo murine models induced by many toxic compounds such as carbon tetrachloride, tetra-butyl hydroperoxide, potassium bromate and schistosoma mansoni infection. <sup>(17)</sup> The *N. sativa* induced protection against hepatotoxicity via decreasing the elevated lipid peroxidation and levels of the liver enzymes and increasing the antioxidant enzymes levels. <sup>(35)</sup> Also Khan et al. <sup>(36)</sup> found *N. sativa* oil significantly decreases oxidative stress that coincide with marked recovery of renal glutathione content and antioxidant enzymes.

To the best of our knowledge, there are no previous studies exploring the effect of date on the aflatoxicosis. Date palms (*Phoenix dactylifera* L., Palmae) have been cultivated in the Middle East over at least 6000 years ago. <sup>(37)</sup> For the natives in this region, dates are considered a staple carbohydrate food. <sup>(38)</sup> Date fruits are also used in the production of local beverages and spirits. In local medicinal practices, dates are considered a “tonic” and “aphrodisiac”, and in some communities they are thought to be useful against ulcer. <sup>(38)</sup> In fact, Muslims believe that “He who eats seven dates every morning will not be affected by poison or magic on the day he eats them”. <sup>(39)</sup> Moreover, Vayalil <sup>(18)</sup> showed that date fruit has antioxidant and antimutagenic activity and this implicates the presence of compounds with potent free-radical-scavenging activity.

In the AFB<sub>1</sub> and date group, the plasma levels of liver function enzymes (ALT and AST), creatinine and urea were significantly lower than the AFB<sub>1</sub> group. Histopathologically, the livers and kidneys of rats treated with AFB<sub>1</sub> and Date, showed nearly normal and the mild changes were just vacuolation of hepatocytes, hyaline casts of renal tubules, congestion and few mononuclear cells infiltration.

The histopathological and biochemical actions of date may be due to its antioxidant effects. Vayalil <sup>(18)</sup> showed that the induced liver protection against aflatoxicosis occurred via decreased the level of liver enzyme activity as well as decreased the free radical propagation, also besides its lowering the pathological lesions resulted from AFB<sub>1</sub>. The previous mentioned histological changes due to AFB<sub>1</sub> and NS as well as AFB<sub>1</sub> and Date were in agreement with those reported by many researchers. <sup>(18, 40-41)</sup>

Our study concludes that AFB<sub>1</sub> induced histopathological changes in the tissues of liver and kidney associated with dysfunction of these organs. Both NS and Date reduce the toxic effect of AFB<sub>1</sub> in liver and kidney which may be related to their cytoprotective and antioxidant properties, and their effect on some mediators of inflammation. The date treatment was most likely cytoprotective for liver than NS treatment against aflatoxicosis in rats.

## References

1. Cole R.J. and Cox, R.H. Handbook of Toxic Fungal Metabolites, Academic Press, New York. 1981
2. Ciegler A, Lee LS, Dunn JJ. Production of Naphthoquinone Mycotoxins and Taxonomy of *Penicillium viridicatum*. Appl Environ Microbiol. 1981;42(3):446-449 .

3. Sharma R.P. and Salunkhe D.K. *Mycotoxins and Phytotoxins*, CRC Press, Boca Raton, FL. 1991
4. Miller J.D. and Trenholm H.L. *Mycotoxins in Grain: Compounds other than Aflatoxins*, Eagan Press, St Paul, MN. 1994
5. Wang J.-S., Kensler T.W. and Groopman J.D. Toxicants in food: fungal contaminants. In: C. Ioannides, Editor, *Current Toxicology Series: Nutrition and Chemical Toxicity*. Wiley, New York.1998:29–57.
6. Massey T.E.,Stewart R.K.,Daniels J.M. and Liu L. Biochemical and molecular aspects of mammalian susceptibility to aflatoxin B1 carcinogenicity, *Proc. Soc. Exp. Biol. Med.* 1995;208:pp. 213–227.
7. Wang, J.-S. and Groopman J.D. DNA damage by mycotoxins, *Mutation Res.* 1999;424 :167–181.
8. Eaton DL, Gallagher EP. Mechanisms of aflatoxin carcinogenesis. *Annul Rev Pharmacol Toxicol.* 1994;34:135–172.
9. Fong H.H. Integration of herbal medicine into modern medical practices: issues and prospects, *Integr Cancer Ther.*2002;1:287–293.
10. Dattner A.M. From medical herbalism to phytotherapy in dermatology: back to the future, *Dermatol Ther.* 2003;16 :106–113.
11. Huffman M.A. Animal self-medication and ethno-medicine: exploration and exploitation of the medicinal properties of plants, *Proc Nutr Soc* 2003;62:371–381.
12. Parab S., Kulkarni R. and Thatte U. Heavy metals in ‘herbal’ medicines, *Indian J Gastroenterol* 2003;22 :111–112.
13. Salem M.L. and Hossain, M.S. Protective effect of black seed oil from *Nigella sativa* against murine cytomegalovirus infection, *Int J Immunopharmacol* 2000;22(9): 729–740.
14. Goreja, W.G. *Black Seed: Nature's Miracle Remedy*, Amazing Herbs Press, New York, NY. 2003
15. Junemann M. *Three great healing herbs*, Lotus Light Publications, Twin Laked,WI, 1998; p. 45.
16. Schleicher P. and Saleh M. *Black seed cumin: the magical Egyptian herb for allergies, asthma, and immune disorders*, Healing Arts Press, Rochester, Vermont, 1998;p. 90.

17. Salem ML Immunomodulatory and therapeutic properties of the *Nigella Sativa* L. Seed. *Int Immunopharmacol.* 2005;5(13-14):1749-70.
18. Vayalil PK. Antioxidant and antimutagenic properties of aqueous extract of date fruit (*Phoenix dactylifera* L. *Areaceae*). *J Agric Food Chem.* 2002;30;50(3):610-7.
19. Meki AR, Abdel-Ghaffar, S. and El-Gabaly I., Aflatoxin B1 induces apoptosis in rat liver: protective effect of melatonin. *Neuroendocrinol. Lett.* 2001;22, pp. 417–426.
20. Carleton M.A.; Drury, G.A.; Willington, E.A. and Cammeron, H. Carleton,s histological technique 4<sup>th</sup> Ed.Oxford Univ.Press.New York,Toronto,London. 1967
21. King J. The transferases- alanine and aspartate transaminases.In:King, J.(Ed) *Practical Clinical Enzymology* Van Nostrand Company Ltd,London.1965;pp.121-128.
22. Lowry, O.H., Rosebrough, N.J., Farr, A.L., Randall, R.J. Protein measurement with the Folin phenol reagent. *J. Biol. Chem.* 1951;193, 265–275.
23. Fabiny DL, Ertingshausen G. Automated reaction-rate method for determination of serum creatinine with the CentrifChem. *Clin Chem.* 1971;17(8):696-700.
24. Chaney, A.L. Marbach E.P. Modified reagents of determination of urea and ammonia.*Clin.Chem.* 1962;8:130-132.
25. Neal GE. Genetic implications in the metabolism and toxicity of mycotoxins. *Toxicol Lett;* 1995;82:861–867.
26. Eaton D.L. and Groopman J.D. The toxicology and aflatoxin.Human health,Veterinary and Agricultural significance, Academic Press, SanDiego,CA. 1994
27. Meki AR, Esmail Eel-D, Hussein AA, Hassanein HM. Caspase-3 and heat shock protein-70 in rat liver treated with aflatoxin B1: effect of melatonin. *Toxicon.* 2004;43(1):93-100.
28. Bondy GS, Armstrong CL, Curran IH, Barker MG, Mehta R. Retrospective evaluation of serum ornithine carbamyltransferase activity as an index of hepatotoxicity in toxicological studies with rats. *Toxicol Lett.* 2000;3;114(1-3):163-71.
29. Dennis,M.H.; Myers,M.J.; Raybourner,A.; Joseph,S.A. and Ming,W.C. Immunotoxicity of aflatoxin-B1 in rats: Effect on lymphocytes and the inflammatory response in a chronic intermittent dosing study.*Toxicological Science.* 2003;73(2):362-377.
30. Wangikar P.B., Dwivedi F, Shrma A.K. and Neeraj S. Effect in rats simultaneous prenatal exposure to ochratoxin A and aflatoxin B1 .II.Histopathological features of



teratological anomalies induced in fetus. Birth defect research part B: Developmental and Reproductive toxicology. 2004;71(6):352-358.

31. Tessari E.N, Oliveira C.A, Cardoso AL and Rottinghaus GE (2006): Effects of aflatoxin B1 and fumonisin B1 on body weight of antibody titres and histology of broiler chicks. Br. Poult. Sci. 2006;47(3):357-364.

32. Rati ER, Shantha T, Ramesh HP. Effect of long term feeding and withdrawal of aflatoxin B1 and ochratoxin A on kidney cell transformation in albino rats. Indian J Exp Biol. 1991;29(9):813-7.

33. Souza MF, Tome AR, Rao VS. (1999) Inhibition by the bioflavonoid termination of aflatoxin B1-induced lipid peroxidation in rat liver. J Pharm Pharmacol; 51:125-129.

34. Abdel-Wahhab MA and Aly SE. Antioxidant property of *Nigella sativa* (black cumin) and *Syzygium aromaticum* (clove) in rats during aflatoxicosis. J Appl Toxicol. 2005;25(3):218-23.

35. Kanter, M. Meral I, Dede S. and Ozbek H. Effect of *Nigella sativa* L. and *Urtrica dioica* L. on lipid peroxidation, antioxidant enzyme system and some liver enzymes in CCL4- treated rats. J. Vet. Med. A. Physiol. Pathol. Clin. Med. 2003;50:264-268.

36. Khan N., Sharma S. and Sultana S. *Nigella sativa* (black cumin) ameliorates potassium bromate-induced early events of carcinogenesis: diminution of oxidative stress, Hum Exp Toxicol. 2003;22:193-203.

37. Copley, M.S., Rose, P.J., Clampham, A., Edwards, D.N., Horton, M.C., Evershed, R.P. Detection of palm fruit lipids in archaeological pottery from Qasr Ibrim, Egyptian Nubia. Proceedings of the Royal Society, London. 2001;268:593-597.

38. Al-Shahib, W., Marshall, R.J. The fruit of the date palm: its possible use as the best food for the future. International Journal of Food Science and Nutrition. 2003;54:247-259.

39. Miller, C.J., Dunn, E.V., Hashim, I.B. The glycaemic index of dates and date/yoghurt mixed meals. Are dates "the candy that grows on trees"? European Journal of Clinical Nutrition. 2003;57:427-430.

40. AL-Sagair, O.A. and EL-Daly, E.S. Influence of combined antioxidants, *Nigella sativa* and Tamoxifen pretreatment on mycotoxins induced neurotoxicity in rats. J Egypt. Ger. Soc. Zool. Comparative physiol. 2006:207-236.

41. Maduka, A.O.; Uwaifo, J.O. and Nwankwo, J.O. The modulating effects of red palm oil on aflatoxin induced toxicity in weaning rats. Asian Network for Scientific Informa. 2001;3(1):91-96.

Table (1) Plasma bioindices of liver and kidney functions in different groups of male rats.

<b>Variables</b>	<b>Controls</b>	<b>AFB1 group</b>	<b>AFB1 + Date group</b>	<b>AFB1 + NS group</b>
Total protein (g/dL)	7.34±0.41	6.74±0.21 <sup>a</sup> P>0.05(NS)	7.19±0.26 <sup>a</sup> P>0.05(NS) <sup>b</sup> P>0.05(NS)	7.21±0.11 <sup>a</sup> P>0.05(NS) <sup>b</sup> P>0.05(NS) <sup>c</sup> P>0.05(NS)
ALT (U/L)	29.65±0.73	51.67±3.59 <sup>a</sup> P<0.01	37.38±0.83 <sup>a</sup> P<0.001 <sup>b</sup> P<0.01	40.87±1.01 <sup>a</sup> P<0.001 <sup>b</sup> P<0.01 <sup>c</sup> P<0.001
AST (U/L)	25.88 ± 1.84	51.49 ± 2.08 <sup>a</sup> P<0.001	32.95 ± 2.12 <sup>a</sup> P<0.05 <sup>b</sup> P<0.001	44.36 ± 1.04 <sup>a</sup> P<0.001 <sup>b</sup> P<0.01 <sup>c</sup> P<0.001
Creatinine (mg/dL)	0.814 ± 0.08	1.829 ± 0.19 <sup>a</sup> P<0.001	1.000±0.044 <sup>a</sup> P>0.05 (NS) <sup>b</sup> P<0.001	0.850±0.277 <sup>a</sup> P>0.05(NS) <sup>b</sup> P<0.01 <sup>c</sup> P>0.05(NS)
Urea (mg/dL)	24.10±0.861	59.07±1.919 <sup>a</sup> P<0.001	50.63±3.383 <sup>a</sup> P<0.001 <sup>b</sup> P<0.05	33.36±1.086 <sup>a</sup> P<0.001 <sup>b</sup> P<0.001 <sup>c</sup> P<0.001

Values are means±SE for 10 rats (N= 10 for each group). P values are shown as (a) for comparison AFB1-treated group, AFB1+Date group and AFB1+ NS group versus controls. (b) for comparison AFB1+ Date group and AFB1+ NS group versus AFB1-treated group. (c) for comparison AFB1+ Date group versus AFB1+ NS group. ALT, alanine transaminase; AST, aspartate transaminase; NS, Nigella sativa; AFB1, aflatoxin B1. Other details are given in materials and methods section.

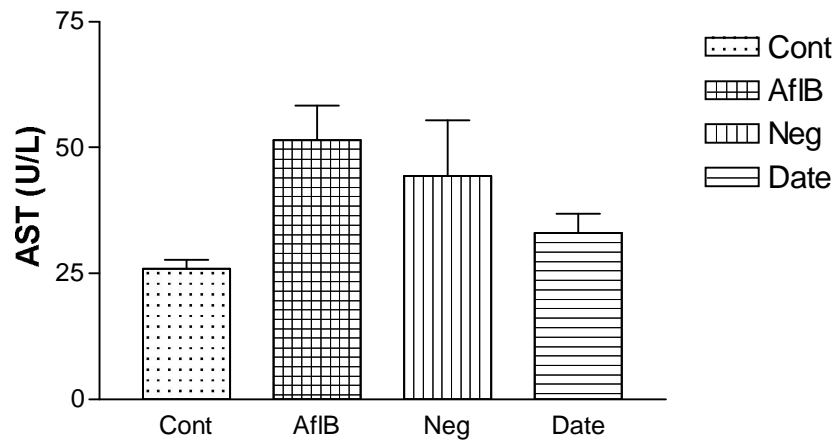


Fig. ( 1 ) Plasma levels of aspartate transaminase in controls and different treated rat groups.

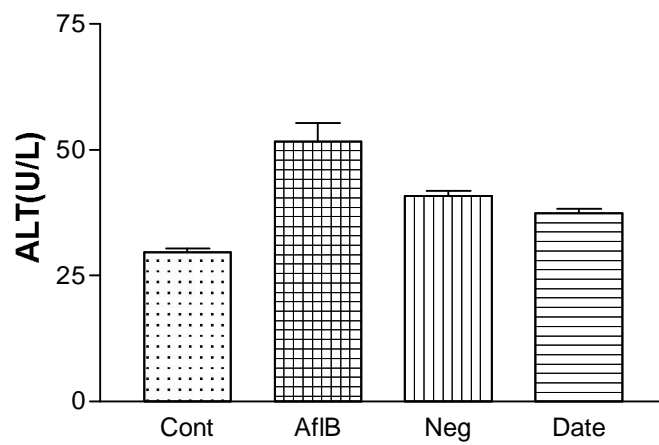


Fig. ( 2 ) Plasma levels of alanine transaminase in controls and different treated rat groups.

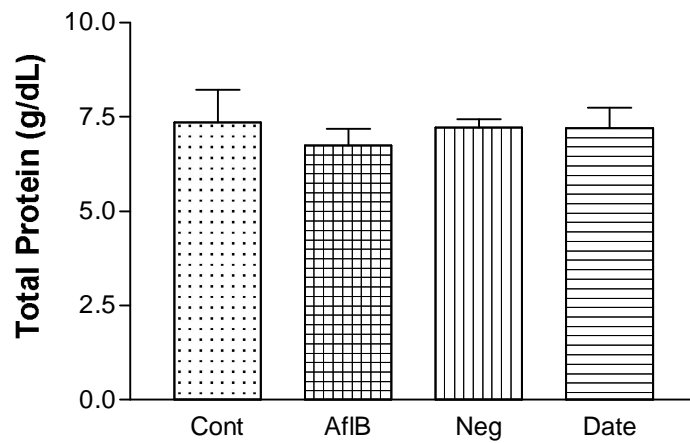


Fig. ( 3 ) Plasma levels of total protein in controls and different treated rat groups.

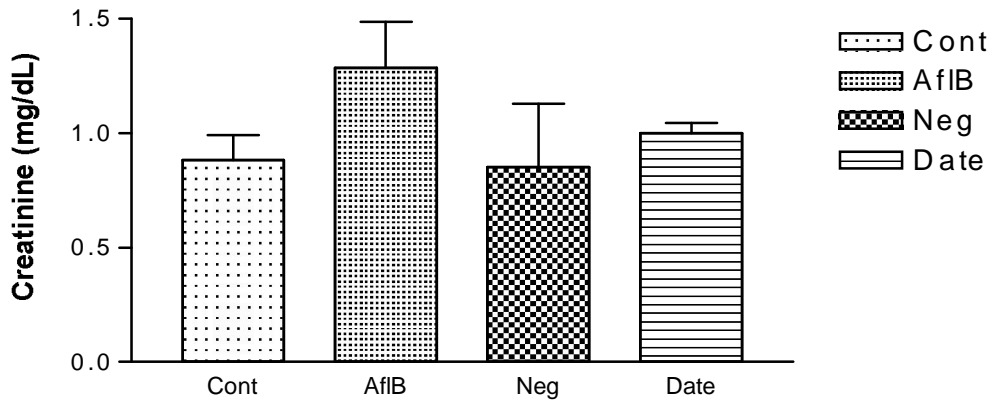


Fig. ( 4 ) Plasma levels of creatinine in controls and different treated rat groups.

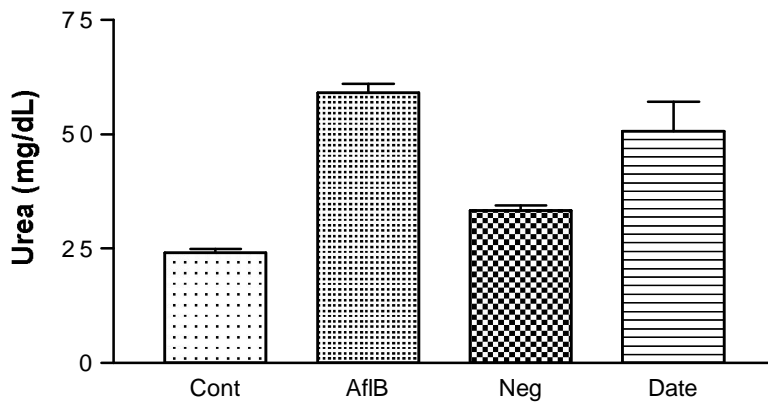


Fig. ( 5 ) Plasma levels of urea in controls and different treated rat groups.

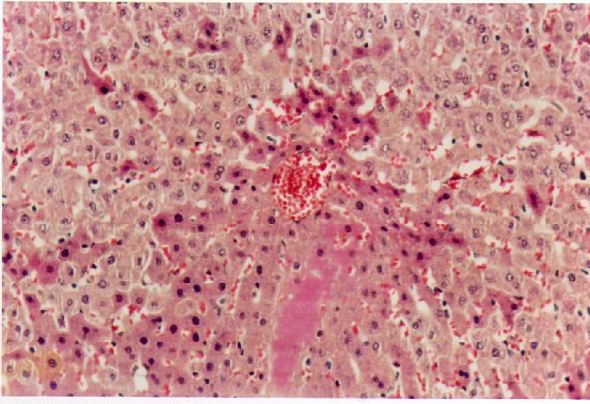


Fig 6: Liver of rats, intoxicated with AFB1, shows multifocal necrosis of hepatocytes, congestion and intercellular edema (H&E. X200).

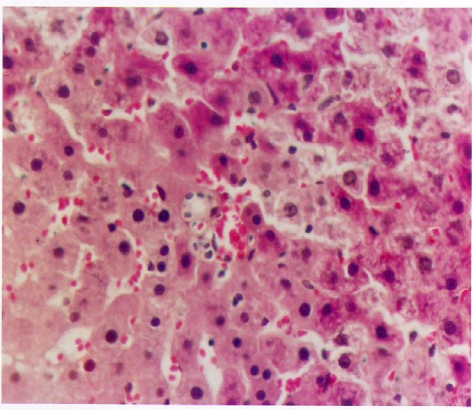


Fig. 7: Liver of rats, intoxicated with AFB1 , showing: diffuse hepatocellular necrosis with marked cytoplasmic eosinophilia.(H&E.X400).

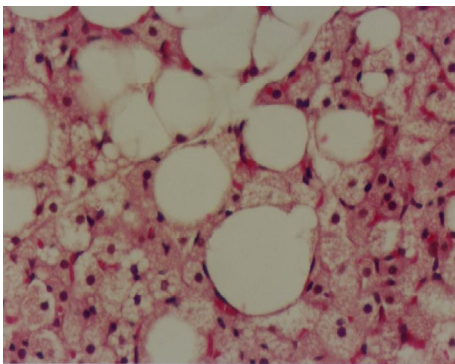


Fig.8: Liver of rats, intoxicated with AFB1 , showing: fatty and vacuolar degenerations of hepatocytes (H&E.X400)

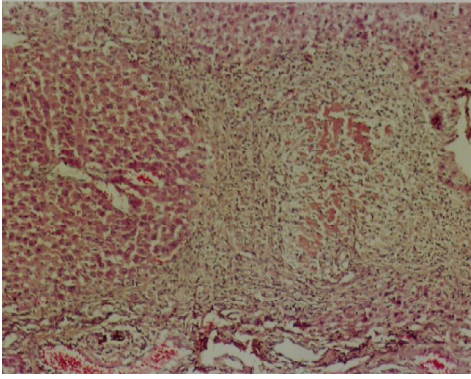


Fig 9: Liver of rats, intoxicated with AFB1 , showing normal liver cells to the left and hepatocellular necrosis and fibrous proliferation to the right (H&E.X 100)

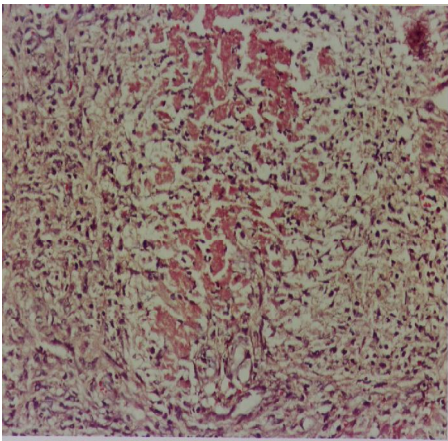


Fig. 10: Sequestration and fibrosis and mononuclear cells infiltrations (H&E.X200).

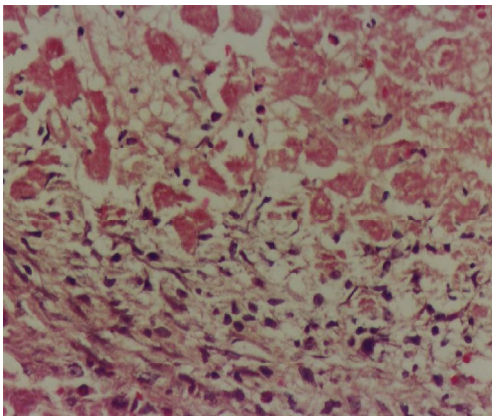


Fig. 11: Sequestration and fibrosis and mononuclear cells infiltrations (H&E.X400).

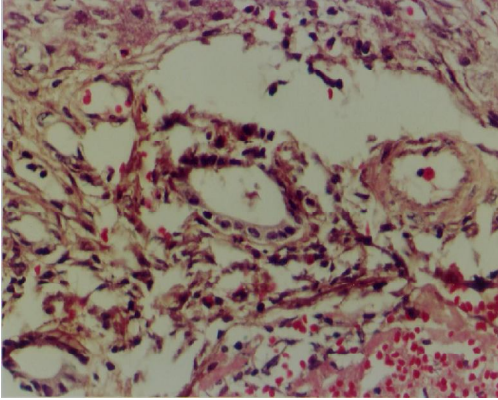


Fig.12: Liver of rat, intoxicated with AFB1, showing: biliary hyperplasia, congestion, hemorrhage, perivascular edema and mononuclear leukocytic cell infiltration (H&E.X400).

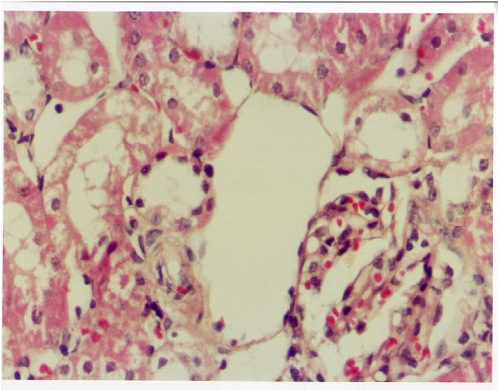


Fig. 13: Kidney of male albino rat, treated with AFB1 alone, showing cloudy swelling, vacuolar degeneration and necrosis of the renal tubular cells. Some tubules are cystically dilated (H&E.X400).

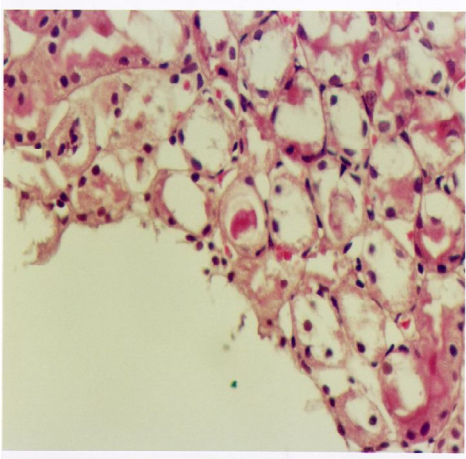


Fig. 14: Kidney of male albino rat, treated with AFB1 alone, showing hyaline casts and tubular necrosis (H&E.X400).

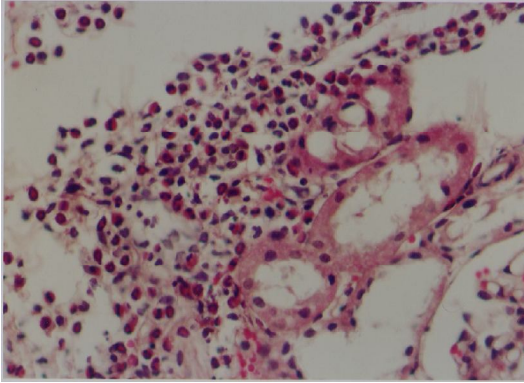


Fig. 15: Kidney of male albino rat, treated with AFB1 alone, showing: infiltration by eosinophils and tubular necrosis (H&E.X400).

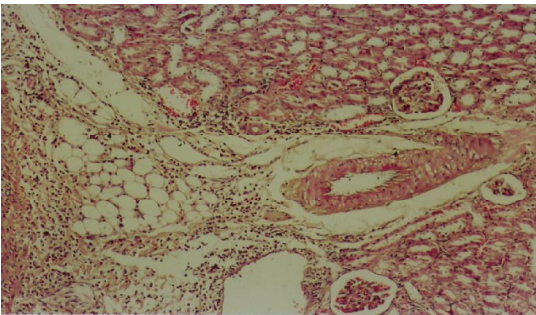


Fig. 16: Kidney of male albino rat, treated with AFB1 alone, showing fatty change, and inflammatory cell infiltration on top of tubular necrosis (H&E.X100).

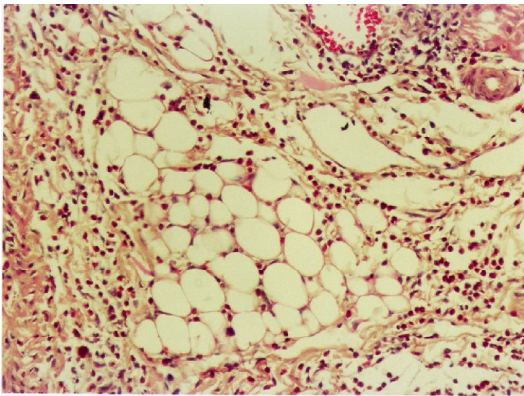


Fig. 17: Kidney of male albino rat, treated with AFB1 alone, showing fatty change, and inflammatory cell infiltration on top of tubular necrosis (a high power resolution).



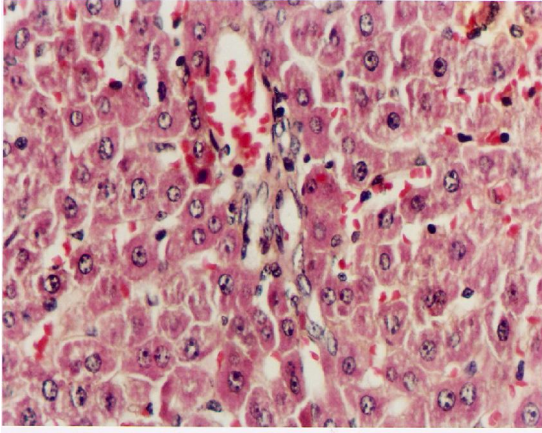


Fig. 18: Liver of male albino rat, treated with AFB1+N.sativa, showing:apoptotic hepatocytes and congestion (H&E.X400).

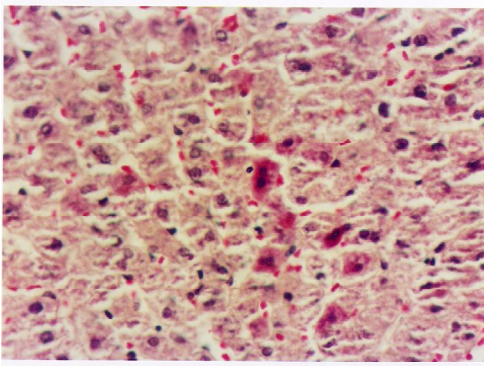


Fig. 19: Liver of male albino rat, treated with AFB1+N.sativa, showing focal liver cell necrosis (H&E.X400).

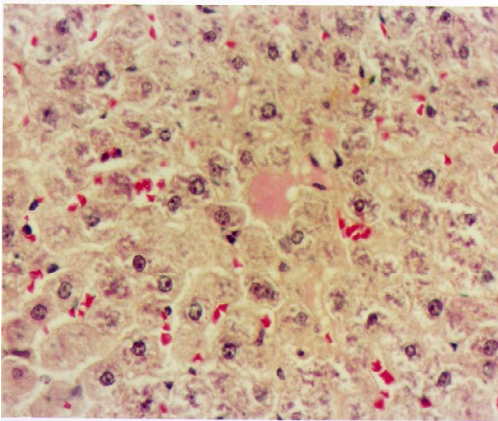


Fig. 20: Liver of male albino rats, treated with AFB1+N.sativa, showing mild degree of degeneration. Some hepatocytes contain multiple nucleoli (sign of regeneration and hyperplasia) (H&E.X400).

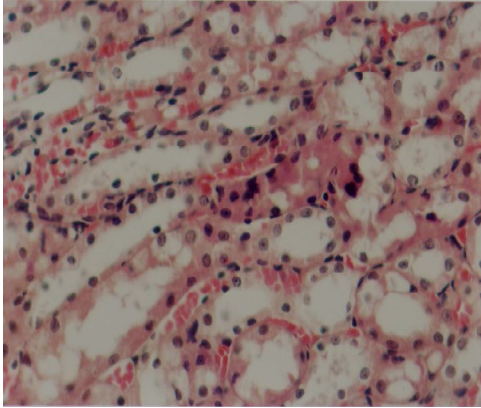


Fig. 21: Kidney of male albino rat, treated with AFB1+N.sativa, showing hydropic degeneration and congestion (H&E.X400).

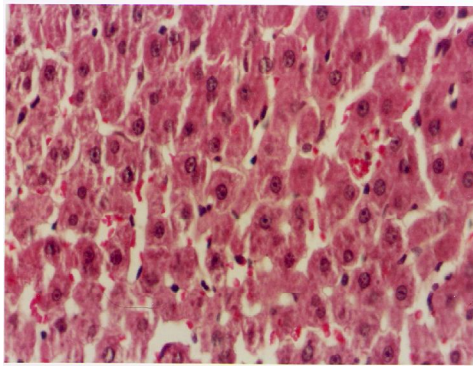


Fig. 22: Liver of male albino rat, treated with AFB1+Date, showing more or less normal liver except for mild congestion (H&E.X400).

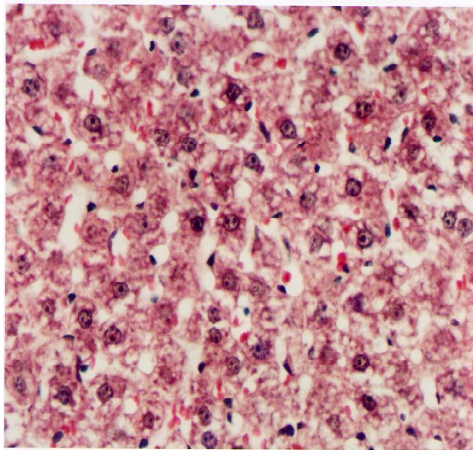


Fig. 23: Liver of male albino rat, treated with AFB1+Date, showing more or less normal hepatocytes “just mild degeneration”. (H&E.X400).

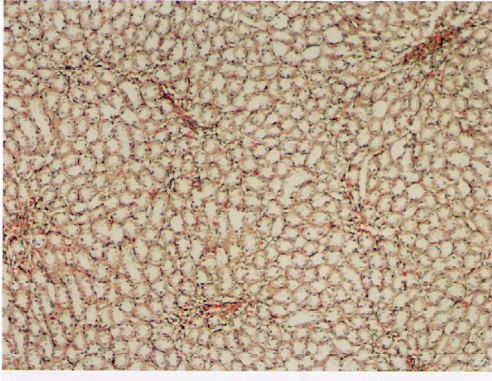


Fig. 24: Kidney of male albino rat, treated with AFB1+Date, showing nearly normal renal tubules (H&E.X200).