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Abdominal MR Elastography

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Abstract

MR Elastography (MRE) is an MRI-based technique for quantitatively assessing the mechanical properties of tissues, based on the propagation of shear waves. Multiple studies have described many potential applications of MRE, from characterizing tumors to detecting diffuse disease processes. Studies have shown that MRE can be successfully implemented to assess abdominal organs. The first clinical application MRE to be well documented is the detection and characterization of hepatic fibrosis, which systematically increases the stiffness of liver tissue. In this diagnostic role, offers a safer, and less expensive, and potentially more accurate alternative to invasive liver biopsy. Emerging results suggest that measurements of liver and spleen stiffness may provide an indirect way to assess portal hypertension. Preliminary studies have demonstrated that is possible to use MRE to evaluate the mechanical properties of other abdominal structures, such as the pancreas and kidneys. Steady technical progress in developing practical protocols for applying MRE in the abdomen and pelvis provides opportunities to explore many other potential applications of this emerging technology.

Keywords

Abdominal MR Elastography; MRI

Elasticity Imaging

Many disease processes, either focal or diffuse, cause marked changes in tissue mechanical properties. For instance, hard masses detected by manual palpation in the thyroid, breast and prostate are known to be suspicious for malignancy. Generations of physicians have experienced that a fibrotic/cirrhotic liver typically is very hard. There is significant interest in precisely characterizing the mechanical properties of normal and abnormal tissues and using that information as potential indicators for diagnosing diseases and monitoring disease progression and treatment efficacy. With recent advances in biomedical imaging, the ability to quantify *in vivo* stiffness by detecting wave propagation velocity through human tissue (elastography) using noninvasive techniques like ultrasound and MRI has now became possible. At present, there are two well-developed noninvasive approaches to assess the mechanical properties of abdominal tissues *in vivo*. One is the Transient Ultrasound Elastography (TUE) technique^{1, 2}, which can provide localized quantitative shear stiffness based on 1-D measurements. The other is the MR Elastography (MRE), which is an MR-based method that uses a modified phase-contrast MRI acquisition to determine the tissue stiffness distribution throughout a 2-D or 3-D region of interest ³.

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Compared to ultrasound-based techniques for elasticity imaging, MR-based techniques offer many significant advantages and have been the focus of significant developments over the last few years. The measurement of motion with MR in medical applications has been motivated significantly by the desire for blood flow quantification and vascular system imaging ⁴. In the 1990's, measurement of tissue mechanical properties using MR methods was demonstrated ^{3, 5}. MR elastography uses a modified phase-contrast imaging sequence to detect propagating shear waves within the tissue of interest. The technique has been well documented and can be implemented on a conventional MRI system with minimal modifications to the hardware and software.

The hardware required for MRE is an acoustic driving system, which is used to generate shear waves in tissue, typically at frequencies between 40 and 200 Hz. Acoustic waves have been created in targeted tissues using active electromechanical voice coils ⁶, ⁷, passive rigid rod drivers ⁸, ⁹ and passive pneumatic drum drivers ^{10–13}. There have also been a number of other application-specific drivers, such as a needle-based acoustic driver system used to study the liver tissue of small animal models *in vivo* ¹⁴. The main driving system used in several studies involves a cylindrical, nonmetallic passive drum driver 18.5 cm in diameter and 3 cm in height. It is activated with varying acoustic pressure conducted via a 7.6-meter long plastic tube (PVC, polyvinyl chloride) from a distant active audio device located outside the scanner room. Among the aforementioned designs, the passive pneumatic driver has several advantages. It can be easily applied against body (such as against the thoracic and abdominal walls) in any orientation, even adjacent to the imaging coils, while avoiding problems with artifacts and heat buildup typical of electromechanical drivers.

The software required for MRE includes a specialized phase-contrast MRI pulse sequence and an inversion algorithm which are used to image and process the wave images, respectively. Depending on the specific application, the MRE sequence uses a conventional MR imaging sequence (e.g., gradient-recalled echo (GRE), spin echo (SE), balanced steadystate free precession (bSSFP), or echo planar imaging (EPI)) with the inclusion of additional motion-encoding gradients (MEGs), which allow shear waves with amplitudes in the micron range to be readily imaged ^{3, 5, 15–20}. The MEGs are imposed along a specific direction and switched in polarity at an adjustable frequency. Trigger pulses synchronize the imaging sequence with the driving system to induce shear waves in the tissue, usually at the same frequency as the MEGs. In the received MR signal, a measurable phase shift caused by the cyclic motion of the spins in the presence of these MEGs can be used to calculate the displacement at each voxel, which provides a snapshot of the mechanical waves propagating within the tissue. By adjusting the phase offset between the mechanical excitation and the oscillating MEGs, wave images can be obtained for various time points of the vibration. Four to eight phase offsets evenly spaced over 1 cycle of the motion are obtained in most experiments. This allows for the extraction of the harmonic component of the phase data at the driving frequency, giving the amplitude and the phase of the harmonic displacement at each point in space, which is the input to most MRE inversion algorithms.

MRE uses propagating mechanical shear waves rather than a static stress as a means to study tissue elasticity. This provides an important advantage for calculating elasticity in that dynamic MRE does not require the estimation of the regional static stress distribution either inside or on the boundaries of the tissue. The acquired wave images are processed using an inversion algorithm to generate quantitative images that depict tissue stiffness – called elastograms. Many different inversion algorithms have been developed to process MRE wave data based on different assumptions about the nature of the wave propagation. Preprocessing algorithms for the wave data include phase unwrapping, removal of concomitant gradient field effects, and directional filtering ²¹ to enhance the accuracy of the elastograms. The inversion algorithms themselves can be performed using measures of the

phase gradient or spatial frequency content, direct inversions of the differential equations of motion, or iterative methods involving finite element models ^{22–25}.

Over the years, MRE has been developed for many applications to quantitatively assess the viscoelastic properties of various soft tissues, including brain ^{26–28}, breast ^{29–32}, heart ^{33, 34}, lung ³⁵, muscle ³⁶, liver ³⁷, spleen ¹³, kidneys ³⁸ and pancreas ³⁹. A preliminary study has shown that abdominal MRE is feasible for the assessment of most abdominal organs ⁴⁰. The purpose of this review work is to summarize the current status of MRE applications for the abdomen, including the current reported diagnostic capabilities of MRE and the demonstrated feasibility of studying select organs, disease models and patient populations.

MRE Assessment of Hepatic Fibrosis

Essential for life, the liver assists in the maintenance of metabolic homeostasis by processing dietary carbohydrates, amino acids, lipids, and vitamins; metabolizing cholesterol and toxins; producing clotting factors; and storing glycogen. Chronic liver disease is a worldwide problem, which has a variety of causes including viral, autoimmune, drug-induced, cholestatic, metabolic and hereditary ⁴¹. In response to inflammation or direct toxic insult to the liver, zonal cellular injury forms and eventually leads to progressive hepatic fibrosis, which is scar tissue that develops progressively by an excessive production of extracellular matrix proteins ⁴². The end stage of hepatic fibrosis is cirrhosis, which is characterized by severe complications such as variceal bleeding and ascites as a result of portal hypertension. It is also associated with a 50% 5-year mortality due to hepatic failure and the development of hepatocellular carcinoma ⁴³. In the United States during 2004, disease-related complications were associated with nearly 40,000 deaths and greater than 1.4 billion dollars spent on medical services ^{44, 45}. Furthermore, these trends are expected to increase based on an aging population, the growing epidemic of obesity, and the continued emergence of clinical manifestations among individuals with long-standing chronic hepatitis C infection ^{46, 47}. In terms of individual patients as well as populations, there remains a great need to develop and identify methods of risk stratification and prognosis for individuals with chronic liver disease.

Fibrosis previously was thought to be an irreversible scarring process formed in response to inflammation or direct toxic insult to the liver. However, increasing evidence has demonstrated that, unlike cirrhosis, the early stages of fibrosis are treatable and reversible if appropriate anti-fibrotic treatment is given ^{48, 49}. As anti-fibrotic therapies evolve, a reliable, noninvasive and cost-effective technique for diagnosis and follow-up assessment of hepatic fibrosis is needed to manage patients with chronic liver disease. Being able to noninvasively monitor the progression of liver fibrosis would help in understanding the natural history of liver fibrosis in patients with chronic liver disease, determining which patients require antiviral therapy, predicting the approximate time to the development of cirrhosis, and discovering new directions of scientific inquiry. Consequently, the degree and rate of fibrosis progression are important prognostic indicators for many chronic liver diseases and hepatic fibrosis should be regarded as a distinct clinical problem, amenable to specific diagnostic tests and therapies that are independent of the etiology. Therefore, there is a need for a reliable, noninvasive method to assess liver fibrosis, not only to detect and stage the disease itself, but also to monitor treatment efficacy and optimize dosing.

Needle liver biopsy, analyzed with connective tissue stains, has long been considered the "gold standard" to detect and quantify hepatic fibrosis. However, it is an invasive procedure with some inherent risk (mortality rate 0.1~0.01%) with many potential complications (leading to poor acceptance by patients) and the likelihood of sampling errors (due to only 1/50,000th of the liver mass being sampled). The biopsy histological examination largely

depends on sample quality (the length and size of the tissue specimen) and the subjective evaluation of morphological changes including grading of necro-inflammatory activity and staging extent of fibrotic organ transition. Existing histologic scoring systems such as the Ishak ⁵⁰ and Metavir ⁵¹ classifications provide a semiquantitative assessment of fibrosis extent which, in theory, has the potential to allow for serial assessments to document progression. However, there has been little documentation of strong reproducibility for existing methods despite their widespread use in clinical practice and controlled trials. A number of studies have demonstrated excessive rates of sampling error (25%–40%) resulting in poor reproducibility of the needle liver biopsy regardless of underlying liver disease etiology ⁵². In addition, the extent of variation in observer interpretation by expert histopathologists may be as high as 20% ⁵³. Therefore, the diagnostic value of the biopsy as "gold standard" in the detection of fibrosis/fibrogenesis must be questioned. Therefore, there remains an opportunity to improve the assessment of individuals with progressive fibrosis who might benefit from early intervention.

Animal studies

It is difficult to precisely describe the development of fibrosis and its impact on tissue mechanical properties in humans due to unreliable liver biopsy data and impractical or impossible longitudinal studies. Therefore, animal models were used to provide progressively increased degrees of hepatic fibrosis which could be accurately assessed with both MRE and histological analysis. Two different animal models (a CCl₄-induced fibrosis mouse model and a knockout mouse model of autosomal recessive polycystic kidney disease (ARPKD)) have been investigated to evaluate the relationship between liver stiffness and the extent of hepatic fibrosis ⁷, ¹⁴. It was validated that the shear stiffness of the liver tissue increases systematically with the extent of hepatic fibrosis in both animal models. Since there may be an increase in the lipid content of fibrotic livers, which is very common in patients with chronic liver diseases, an additional goal of the animal study was to determine whether fat infiltration could potentially interfere with the MRE assessment of liver fibrosis. In order to evaluate this factor, Yin et al. quantitatively assessed fat infiltration using a threepoint Dixon method in the ARPKD mouse model, which can develop obesity due to diet, stress, or malfunction of lipid metabolism because of liver disease. It demonstrated that there is no significant correlation between the liver stiffness measurements and the liver fat-towater ratio at different stages of hepatic fibrosis (Figure 1)¹⁴.

Nonalcoholic fatty liver disease (NAFLD) refers to a wide spectrum of liver disease ranging from simple steatosis, to nonalcoholic steatohepatitis (NASH), to irreversible cirrhosis. All of the stages of NAFLD result in the accumulation of fat in the liver cells. In NASH, the fat accumulation is associated with varying degrees of inflammation and hepatic fibrosis. Recently, Salameh et al. demonstrated that MRE-assessed liver elasticity increased before fibrosis development (linked to myofibroblast activation) in a well-established NAFLD rat model ⁵⁴. If these results are confirmed in NAFLD/NASH patients, MRE may be a potential diagnostic method to discriminate between simple steatosis and steatohepatitis. Therefore, MRE might have a new role in early detection and treatment assessment in NASH patients.

Human studies

The practical *in vivo* hepatic MRE technique for clinical investigations in humans is the culmination of technology development and optimization involving driver design, pulse sequence design, imaging parameters and inversion algorithms. Because of the limited accessibility of the liver, many researchers have investigated the primary mechanism of shear wave production in the abdominal organs using vibrations from external sources. Instead of transversely orientated drivers, which have poor depth penetration and orientation flexibility, longitudinal surface drivers were deemed to be more optimal by generating shear

waves via mode conversion. Several application-specific longitudinal drivers that can deliver acoustic waves to liver tissue have been investigated ^{6, 9, 10}. Currently, there are three types of liver MRE drivers. The electromechanical drivers ⁶ deliver longitudinal vibrations through the posterior body wall. This driver design has the limitation that the driver is fixed within the imaging coil and may have disadvantages related to heat deposition, artifacts and inflexible orientation. The subcostally located, extended-piston driver design ⁹ applies longitudinal motion to the anterior body surface. The nonmetallic materials employed in this design avoid problems with artifacts and heat buildup when it is driven continuously. However, this type of design lacks flexibility in its orientation and how it can contact the body habitus. Without the protection of the rib cage, the subcostal liver tissue may suffer from an unexpected preload from the driver that may lead to overestimation of the stiffness in that region.

The development of a pressure-activated driver system ¹⁰ has provided a mechanism for performing liver MRE without the aforementioned limitations. Placed against the lower right rib cage, the pressure-activated driver can effectively and efficiently generate shear waves throughout the entire liver tissue as well as within the spleen even though it is located on the opposite side of the abdominal cavity from where the driver is placed. This driver system even works well in obese patients (30 < BMI < 42) and those with other complications such as ascites ¹¹. A study comparing MRE and TUE in a series of 141 patients with chronic liver disease showed that while TUE has become a useful clinical tool, the technical success rate for MRE was higher (94%) than that of TUE (84%). This was due to technical limitations of the TUE technique which make it difficult to perform on patients with ascites (since fluid perturbs the wave propagation being measured) and obese patients (BMI > -28) since TUE measures the stiffness at a fixed location approximately 6 cm below the skin surface, which may not reach the liver in obese patients. MRE also proved to have better diagnostic accuracy in that study ⁵⁵. The reproducibility of MRE has also been assessed in 5 healthy volunteers ⁶ and 56 patients with chronic liver diseases ⁵⁵, which resulted in 9% within-subject coefficient of elasticity variance and 0.97 (95% CI: 0.92-0.99) interclass correlation coefficient. This high reproducibility for MRE is essential for the design of future longitudinal studies.

The MRE visualization of the full 3-D vector displacement field within the liver produced by longitudinal drivers has also proven to offer insights and improvements in both experimental design and 2-D imaging plane selection for *in vivo* MRE applications ^{56, 57}. These studies showed that in a large portion of the liver, the propagation direction of shear waves is approximately parallel with the transverse imaging plane when using a pressure-activated drum driver on the anterior chest wall. This allows accurate elastograms to be generated from 2-D wave image data (which can be acquired in a breath-hold), rather than requiring 3-D wave imaging, which requires longer acquisition times and more complex measures to address respiratory motion. With the acoustic driver system, the 2-D wave approximation is generally valid in the axial imaging planes distanced 2–10 cm away from the superior margin of the liver, assuming the liver has average vertical height of 15 cm. Compared with 3-D hepatic MRE analysis, 2-D wave analysis based on 2-D MRE data provides accurate results in a significant part of the liver volume with a substantially reduced calculation time (Figure 2) ⁵⁸.

Motivated by the results of the pilot volunteer and patient studies ^{6, 9, 10}, multiple investigators have studied the diagnostic ability of liver MRE for detecting hepatic fibrosis in patients with chronic liver disease from various etiologies ^{11, 55, 59–61}. The consistent findings have established that liver MRE has excellent diagnostic accuracy for assessing hepatic fibrosis. MRE has been shown to be effective for distinguishing normal from fibrotic livers with a very high negative predictive value of 97%. ROC analysis has shown that, with

a shear stiffness cut-off value of 2.93 kPa, the predicted sensitivity and specificity for detecting all grades of liver fibrosis is 98% and 99%, respectively ¹¹. ROC analysis has also provided evidence that MR elastography can discriminate between patients with moderate and severe fibrosis (grades 2–4) and those with mild fibrosis, as shown in Table 1 and Figure 3 ^{11, 55, 59}. Hepatic stiffness assessed by MRE has been shown to not be systematically influenced by the presence of steatosis ¹¹, which agrees with studies involving TUE in which the correlation between liver stiffness and fibrosis stage were not affected by steatosis as well ^{62, 63}.

Motivated by the successful implementation of MRE for the study of diffuse changes in hepatic stiffness due to fibrosis, a pilot study has been conducted to evaluate the potential role of MRE in characterizing hepatic tumors. In this preliminary work ⁶⁴, patients with 44 hepatic masses were evaluated with MRE and the results were correlated with pathological diagnosis or other accepted diagnostic criteria. The stiffness of benign masses (9 hemangiomas, 3 focal nodular hyperplasia, and 1 hepatic adenoma) averaged 2.7 kPa, slightly higher than the mean stiffness of normal liver parenchyma (2.3 kPa). The mean stiffness of the malignant tumors was 10.1 kPa. In this series, a cut-off value of 5 kPa completely separated all benign liver masses from malignant lesions. Figure 4 shows the results from an expanded cohort of patients from that trial including 91 hepatic tumors. The results indicate that MRE shows substantial promise for aiding the characterization of liver tumors which provides motivation for exploring the potential for evaluating other mass lesions in the abdomen as well.

MR Elastography of Other Abdominal Organs – Preliminary Results

Spleen

Measurements of the stiffness of the spleen, incidentally imaged in exploratory hepatic MRE studies in patients with chronic liver disease, led to an intriguing and potentially important observation about the correlation between spleen and liver mechanics. The preliminary study of 12 normal volunteers and 38 liver patients with biopsy-proven chronic liver diseases demonstrated higher spleen stiffness in patients with chronic liver disease and a very strong correlation between hepatic and splenic stiffness in these patients ($R^2 = 75$, pvalue < 0.001)¹³. This may suggest that the bulk stiffness of the spleen is strongly affected by the portal venous pressure through a poroelasticity effect. To study this effect, a preliminary study is underway involving a well-developed canine model of portal hypertension. MRE has been performed on two adult mongrel dogs immediately after and 4 weeks after initiating cholestatic liver disease by common bile duct ligation. Prior work has established that this model reliably results in portal hypertension within 4 weeks. Subcutaneous vascular access ports were placed with catheter tips in the portal vein and the right hepatic vein allowing measurement of hepatic venous pressure gradient (HVPG). As shown in Figure 5, the HVPG increased from a mean value of 4.5 mmHg in these dogs to a mean value of 13.5 mmHg. The threshold for development of varices in humans is thought to an HVPG of 10-12 mmHg. The MRE-assessed splenic stiffness in these dogs rose from a mean value of 1.8 kPa to an average of 3.4 kPa. Although this study is still ongoing, these results provide preliminary evidence to support the hypothesis that in the absence of confounding effects, the MRE-assessed stiffness of the spleen reflects the magnitude of the HVPG. This also provides motivation for the development of improved MRE techniques for the assessment of the spleen as well as the development and validation of poroelastic models that may allow the HVPG to be estimated noninvasively from MRE-based measurements of splenic stiffness. This would be a very significant development because knowledge of the HVPG is considered to be very important in the management of chronic liver disease and is very difficult to determine noninvasively.

Kidneys and Pancreas

Exploratory MRE of the upper abdomen in healthy volunteers has demonstrated that shear waves can be readily generated and imaged in other organs, such as the kidneys, pancreas, gastric wall, and bowel. Unlike the liver and spleen, the patterns of wave propagation in these structures are extremely complex and include significant components propagating at oblique angles relative to an axial plane of section. Given these characteristics, it is necessary to image the pattern of wave propagation in 3 dimensions and to obtain data from all 3 polarizations of motion, which requires longer imaging times. This kind of 3-D/3-axis MRE acquisition is problematic if the imaging is to be conducted during suspended respiration. One approach to this is to use respiratory gating ⁶⁵, though total acquisition times can still be quite long. As an alternative, EPI-based techniques can be used to significantly reduce acquisition times. One prototype EPI MRE sequence was investigated that was capable of acquiring 12 wave images (4 phase offsets at each of 3 motion-encoding directions) with an acquisition time of 8 seconds per slice. In limited volunteer studies to date, this technique successfully generated 3-D wave data sets with typical voxel dimensions of 4 mm ⁶⁶. With a 3-D extension of the inversion software, including 3-D spatial filtering ⁶⁷, the preliminary results indicate that it is quite feasible to image tissue stiffness throughout a larger 3-D region of interest in the abdomen and has yielded provocative preliminary results in the pancreas and kidneys that motivate further development in these areas (Figure 6).

Influence of Perfusion on Abdominal Tissue Stiffness

Preliminary results from the study relating splenic stiffness to HVPG suggests that the stiffness of abdominal organs may have two components: a static component reflecting intrinsic structural properties and a dynamic component reflecting extrinsic perfusion changes. To investigate the influence of perfusion on the shear stiffness of abdominal organs, studies of two specific conditions of the liver and kidneys are being pursued.

Postprandial Liver Stiffness Augmentation

Food intake is known to cause an increase in mesenteric blood flow, which may lead to a postprandial increase in hepatic stiffness that is different in patients with hepatic fibrosis than in normal volunteers ⁶⁸. It has been observed that MRE-assessed liver stiffness increases significantly (average increment of 18% with ranges from 5% up to 48%) following a test meal in patients with advanced hepatic fibrosis, whereas fasting and postprandial liver stiffness are similar in the normal state ⁶⁸ Figure 7. This finding suggests that there is a dynamic component to the liver stiffness that is dependent on the portal pressure. Therefore, it is very important to have hepatic MRE examinations performed consistently in a fasting state. The postprandial augmentation in hepatic stiffness after a test meal known to increase mesenteric blood flow is likely due to the transiently increased portal pressure in patients with hepatic fibrosis. It is thought that mechanical distortion of the intrahepatic vasculature caused by fibrosis impairs the autoregulatory mechanism for the portal venous pressure, which may cause acceleration of the development of portal systemic varies and stretching of hepatic parenchyma and stellate cells that are instrumental in the progression of hepatic fibrosis. This promising observation provides motivation for further studies to determine the potential value of assessing postprandial hepatic stiffness augmentation for predicting progression of fibrotic disease and the development of portal varices. It may also provide new insights into the natural history and pathophysiology of chronic liver disease.

Stenotic Kidney MRE

The kidney is a richly perfused organ receiving 25% of the cardiac output. Renal occlusions, such as renal arterial stenosis (RAS), threaten the viability of the kidney by diminishing blood flow leading to irreversible tissue fibrosis and ultimately kidney failure. Preliminary research has been performed to study the impact of RAS on kidney stiffness in a porcine model of acute renal aterial stenosis ⁶⁹. MRE measurements showed that the stiffness of the kidney progressively decreased as the renal artery stenosis was increased. Surprisingly, the stiffness of the contralateral kidney was observed to increase progressively. This indicates that hemodynamics can significantly affect the mechanical properties of renal parenchyma. Fibrosis is associated with elevated tissue stiffness. However, previous studies have indicated the rich perfusion of the kidney may affect its distension and, in turn, stiffness ⁶⁹, 70

In order to determine the interdependent relationship between perfusion, fibrosis and stiffness, an acute RAS model was established in adult pigs by isolating the renal artery to insert a vascular occluder and an embedded Doppler flow probe. The renal blood flow (RBF) was gradually reduced from the baseline level to total occlusion of 100% in with MRE acquisitions performed at each step. The cortex of the acutely stenotic kidney decreased in stiffness as the degree of stenosis was increased to 40% and above (p<0.05), as shown in the Figure 8. The systemic blood pressure also rose during the each decrement in the RBF (from 75 ± 3 to 96 ± 3 mmHg). These renal MRE preliminary results encourage further evaluation of renal hemodynamics on tissue stiffness, which may be due in part to perfusion pressure applied to the organ. These factors may play a complicating role in detecting the presence of fibrosis due to RAS, and may lead to employing new techniques to assess tissue stiffness. The use of MRE to assess changes in tissue mechanics associated with the dynamic perfusion of tissue may also provide new insights into the natural history and pathophysiology of renal diseases.

Conclusion

MRE has emerged as an increasingly powerful method for noninvasively assessing the mechanical properties of soft tissues. This information can provide valuable information to clinicians and researches interested in distinguishing normal and diseased tissue, improving the diagnosis of various diseases, and monitoring the progression of disease. Numerous investigations have shown that it is readily possible to perform MRE in abdominal organs for detecting specific diseases, such as hepatic fibrosis and portal hypertension, which increase the stiffness of the liver and spleen. Other preliminary studies have demonstrated that is possible to evaluate the mechanical properties of other abdominal structures, such as the pancreas and kidneys. These results will further motivate future studies incorporating MRE to study the normal and pathological mechanics and physiology of abdominal organs.

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Figure 1.

In an ARPKD mouse model resulting in liver fibrosis, a significant linear correlation was observed between the liver stiffness and the histological analysis of fibrosis extent (p-value < 0.0001). No significant correlation was found between liver stiffness and the fat-to-water ratio in any age group for either the normal or diseased mice. Adapted from reference ¹⁴.



Figure 2.

Left: Schematic illustrations of the hepatic MRE setup (with passive driver position indicated) and shear wave generation mechanism; Right: A significant correlation was found between 2-D liver MRE measurements and 3-D liver MRE measurements with full wave information (p < 0.001). Adapted from reference ⁵⁸.



Figure 3.

Example wave images and shear stiffness maps (elastograms) in liver patients with four different biopsy-proven hepatic fibrosis stages.



Figure 4.

Left: The top row shows the results of an MRE exam of a 39-year-old man with a hepatic adenoma. The shear stiffness was measured to be 3.1 kPa in the benign tumor and 2.4 kPa in the surrounding normal liver tissue. The bottom row shows an exam of a 44-year-old man with a cholangiocarcinoma. The stiffness was measured to be 19 kPa in the malignant tumor and 5.4 kPa in the surrounding fibrotic liver tissue. Right: The graph shows a box plot of the shear stiffness of normal liver tissue and various types of hepatic tumors. A cutoff value of 5 kPa separates the malignant tumors from benign tumors and normal liver tissue. Adapted from reference 64 .



Figure 5.

Cholestatic liver disease and portal hypertension was established in 20-kg adult mongrel dogs by common bile duct ligation (CBDL). At surgery, two subcutaneous vascular access ports (VAPs) were placed with their catheter tips in the portal vein and right hepatic vein. MRE and measurement of hepatic venous pressure gradient (HVPG) were obtained before and 4 weeks after surgery. In the maximum intensity projection (MIP) elastograms shown on the right, the dotted lines illustrate locations of the canine spleen tissues. The mean spleen stiffness increased substantially (from 1.8 kPa to 3.4 kPa) with the severely elevated mean HVPG (from 4.5 mmHg to 13.5 mmHg).



Figure 6.

One slice from a 3-D/3-axis abdominal MRE exam of a healthy volunteer showing excellent depictions of the liver, spleen, pancreas, and kidneys.



Figure 7.

Left: Examples of hepatic MRE exams performed pre- and postprandially in a normal volunteer and in a patient with cirrhotic liver. The liver stiffness in the patient with the cirrhotic liver was increased markedly in the postprandial exam. Right: A summary of the percent change in liver stiffness for healthy volunteers and patients with various stages of fibrosis. Results in 18 normal subjects demonstrated no significant change in postprandial hepatic stiffness compared with the fasting state. However, results obtained in 19 patients with hepatic fibrosis demonstrated a statistically significant increase in postprandial liver stiffness (p<0.01).



Figure 8.

An acute RAS model was established in a 45-kg adult pig by isolating the renal artery to insert a vascular occluder and an embedded Doppler flow probe. The RBF was reduced from the baseline level to 40% and then to total occlusion of 100%. The MRE-assessed renal tissue stiffness measurements demonstrated that the renal tissue stiffness decreases during gradual reductions in RBF in pigs.

Table 1

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Reference	Z	Cut-off	Sensitivity	Specificity	AUR (Area under the ROC curve)	Liver Stiffness, kPa
		F0:F1-4	85	91	0.96	2.4
. 55	; ;	F0-1:F2-4	100	91	0.99	2.5
Huwart et al ^{JJ}	551	F0-2:F3-4	91	76	0.99	3.1
		F0-3 : F4	100	96	1.00	4.1
		F0:F1-4	83	100	0.95	2.4
	00	FO-1:F2-4	98	100	0.99	2.5
Huwart et al	00	F0-2:F3-4	95	100	0.99	3.1
		F0-3 : F4	100	100	1.00	4.3
		N : F0-4	98	66	0.99	2.9
Ц. ;	05	FO-1:F2-4	86	85	0.92	4.9
Yin et al 11	çõ	F0-2:F3-4	78	96	0.92	6.7
		F0-3:F4	89	90	0.92	6.7