



Published in final edited form as:

Eur J Clin Invest. 2010 May ; 40(5): 440–450. doi:10.1111/j.1365-2362.2010.02276.x.

Calorie Restriction: What Recent Results Suggest for the Future of Aging Research

Daniel L. Smith Jr.^{1,2}, Tim R. Nagy^{1,2}, and David B. Allison^{1,2,3}

¹Department of Nutrition Sciences, University of Alabama at Birmingham, Birmingham AL, 35294 USA

²Nutrition and Obesity Research Center, University of Alabama at Birmingham, Birmingham AL, 35294 USA

³Department of Biostatistics, University of Alabama at Birmingham, Birmingham AL, 35294 USA

Abstract

Background—Calorie Restriction (CR) research has expanded rapidly over the past few decades and CR remains the most highly reproducible, environmental intervention to improve health and extend lifespan in animal studies. Although many model organisms have consistently demonstrated positive responses to CR, it remains to be shown whether CR will extend lifespan in humans. Additionally, the current environment of excess caloric consumption and high incidence of overweight/obesity illustrate the improbable nature of the long-term adoption of a CR lifestyle by a significant proportion of the human population. Thus, the search for substances that can reproduce the beneficial physiologic responses of CR without a requisite calorie intake reduction, termed CR mimetics (CRMs), has gained momentum.

Material & Methods—Recent articles describing health and lifespan results of CR in nonhuman primates and short-term human studies are discussed. Additional consideration is given to the rapidly expanding search for CRMs.

Results—The first results from a long-term, randomized, controlled CR study in nonhuman primates showing statistically significant benefits on longevity have now been reported. Additionally, positive results from short-term, randomized, controlled CR studies in humans are suggestive of potential health and longevity gains, while test of proposed CRMs (including rapamycin, resveratrol, 2-deoxyglucose and metformin) have shown both positive and mixed results in rodents.

Conclusion—Whether current positive results will translate into longevity gains for humans remains an open question. However, the apparent health benefits that have been observed with CR suggest that regardless of longevity gains, the promotion of healthy aging and disease prevention may be attainable.

Keywords

Calorie Restriction; Dietary Restriction; Aging; Longevity; Lifespan; Mortality

Corresponding Author: Daniel L. Smith, Jr., PhD, Department of Nutrition Sciences, Volker Hall, Room G005, University of Alabama at Birmingham, 1530 University Avenue, Birmingham, Alabama, 35294. Phone: 205-934-4008, Fax: 205-996-2074, dsmithjr@uab.edu..

Disclosures DLS and TRN have no conflicts of interest to disclose.

Introduction

Research interest in aging and age-related disease progression has rapidly increased during the last half century. Particularly over the last 2 decades, model organisms including yeast, worms, flies and mice have produced a wealth of information demonstrating an interaction between genes and environment in determining longevity. One particularly active area of research has been the influence of diet on longevity and age-related disease. In this field, calorie (energy) restriction (CR), sometimes referred to as dietary restriction (DR), has been repeatedly shown to significantly increase lifespan and reduce age-related disease compared with *ad libitum* (AL) feeding conditions. Other works report much of the background and historical context for the benefits observed with CR [1;2]. The focus of this article is the expected results of CR in primate models, including human outcomes, as well as the potential of alternatives to CR, particularly the rapidly growing area of calorie restriction mimetic (CRM) research, to improve health and delay death.

Should CR be expected to produce health and longevity benefits in nonhuman primates?

Most CR research on longevity in mammals has been performed in rodents, with laboratory mice, *Mus musculus*, predominant during recent history [1–3]. However, it should be noted that a host of other organisms have shown similar benefits including yeast, nematodes, flies, rotifers, spiders, fish, rats, hamsters and dogs [1–4]. Considering the breadth of organisms that respond positively to CR, should it be expected that nonhuman primates would likewise show similar results? There are two active randomized, nonhuman primate studies testing the benefits of long-term CR on longevity and disease in rhesus monkeys, *Macaca mulatta* – one at the University of Wisconsin at Madison and another at the National Institute on Aging (NIA) [5–7]. A third, non-randomized study at the University of Maryland with a smaller number of restricted monkeys on a weight maintenance diet has interpreted results in the context of CR as well. [8–11]. The two randomized studies were begun approximately two decades ago, such that results currently being reported benefit from the forethought of multiple researchers [5;7].

The recent results reported by Colman et al. (2009) are the first results from the Wisconsin CR study showing a significant benefit in reducing age-related mortality and disease with CR in rhesus monkeys [12]. When measuring mortality in a longevity study, consideration should be given to the cause of death, when possible. This can be exemplified by a study subject who dies in an accident (e.g. an automobile collision), which results in a mortality event, but not necessarily a result of the experimental treatment or aging process. In a similar way, even in a well-controlled longevity study, animals can encounter “accidents” which result in mortality, potentially independent of their underlying biological aging process. Thus, after censoring monkeys for what were considered non-age-related mortality events, like gastric bloat, anesthesia complications, endometriosis and injury (7 control and 9 CR monkeys), a significant lifespan benefit with CR ($P=0.03$ [Cox Regression Analysis]) was observed (age-related mortality events/group: Control:n=14, CR:n=5) [12]. However, when assessing “all-cause” mortality in all monkeys in the study and considering the interim mortality results for each group (all cause mortality events/group: n=21/38 control and 14/38 CR), CR does not currently provide a statistically significant lifespan increase ($P=0.16$ [Cox Regression]), although there is a difference in the expected direction [12]. The significance of the lifespan benefit observed on “age-related” mortality with CR is noteworthy, considering the reduced power of this analysis due to the relatively small sample size [12].

Although the demonstrated health and lifespan benefits are significant findings, a number of previously published interim reports and other studies have suggested the plausibility of this

outcome. Similar to rodents, CR in rhesus monkeys results in reduced circulating glucose and improved insulin function, decreased core body temperature, decreased body weight and fat, improved blood lipids and maintenance of dehydroepiandrosterone levels [8;9;13–23]. Despite the delay in knowing the final outcome of the full longevity study, the positive outcomes of the available data merit consideration. For example, CR resulted in a significant reduction of age-related diseases, when considering neoplasias, glucoregulatory impairment and cardiovascular disease (respective incidence controls: 8, 16, 4 vs. CR: 4, 0, 2) [12]. This reduction of age-related disease and a potential increase in longevity are promising, although the results on total mortality are not yet definitive with CR [12]. Moreover, results demonstrating a significant benefit on longevity in the NIA's CR monkey study are not yet available [24;25]. However, available data point to a reduction in disease risk and incidence. Based on these prospects, if CR does indeed improve health and potentially increase lifespan in monkeys, will it do so in humans?

Even if CR works in monkeys, will CR work in humans?

Extensive knowledge exists about human responses to energy restriction or CR. However, with a few notable exceptions [26–28], the supporting data are largely derived from the implementation of a dietary reduction to cause weight loss among overweight or obese persons. As informative as this may be, the ultimate question of whether CR will extend longevity and slow age-related disease in humans cannot be answered in the context of pre-existent obesity. By CR, one could simply be returning an unhealthy, disease-promoting state back to the norm without altering the underlying aging process [29]. Others have proposed this may be the case in rodent CR studies [30–33] and could potentially influence the interpretation of the non-human primate studies as well. Although this may be a potential confounder in laboratory studies of CR, it also has implications for the majority of the human population in the developed world, particularly with the rise in overweight/obesity prevalence in modern times [34]. Nevertheless, the ultimate question of whether CR alters aging and disease in otherwise healthy individuals has much less data available. One argument proposes that CR is not a universal phenomenon, and in combination with the variability of the response, the life history theory of longevity suggests there is no reason to believe that the relatively small reproductive costs of humans will result in a favorable tradeoff of lifespan extension [29;35;36]. On the other hand, because of the breadth of organisms that respond favorably to CR, the potentially conserved molecular and cellular mechanisms, and the evidence from the nonhuman primate and short- to medium-term human studies, it is reasonable to expect that the observed health benefits would translate into longevity gains [37–39].

To better address this question, the NIA has recently funded a multi-site human randomized clinical research study to assess the effects of two years of CR (~25% restriction) in non-obese, healthy individuals through the CALERIE study (Comprehensive Assessment of Long-Term Effects of Reducing Intake of Energy) (<http://calerie.dcri.duke.edu/index.html>). Preliminary results from these human studies are reproducing many of the metabolic and physiologic responses observed in rodents and monkeys. These include reduced body weight, along with reductions in subcutaneous fat, visceral fat and lean muscle mass, reduced insulin and improved lipid profiles, reduced energy expenditure and core body temperature [40–50]. Assuming this type of dietary restraint is sustainable beyond the short term (six-month to two-year) duration of the study one would expect these physiological changes to predict a reduction in age-related disease. Researchers have also studied volunteers who have adopted a self-restricted lifestyle and maintained it for longer durations than the current CALERIE study [51–56]. In agreement with the research results from animal studies, voluntary CR in humans results in significant improvements in cardiovascular profiles, glucose control, body composition and circulating hormones [51–

56]. The combined results from these randomized control studies and the self-restriction groups demonstrate that CR has beneficial short-term physiological effects in humans, particularly contributing to a reduction in cardiovascular and metabolic-related diseases risk factors, conditions which account for a significant proportion of the healthcare related costs and morbidity/mortality in the US [57;58]. Whether long-term CR would significantly increase longevity in humans will likely remain a matter of debate. The advancement of alternative restriction paradigms may ultimately aid in understanding this potential. Two types of DR, alternate day/intermittent fasting, sometimes called every other day feeding, and single nutrient restriction (e.g. protein or methionine restriction), are increasingly reported to produce positive health and longevity benefits similar to sustained, daily CR [59–68]. Although current data are intriguing and suggestive of potential implications for human health, these interventions also lack a clear, defined mechanism of action, much like CR. Future studies in a variety of organisms with varied dietary compositions will be necessary to further validate the significance of these findings.

Is there a short-cut to CR? Progress in mimetics research

As it is impractical and of questionable desirability to maintain long-term CR, starting in early life in humans and sufficient to produce beneficial effects on health and longevity commonly observed in the laboratory models, other alternatives have been pursued. It is proposed that identifying the genetic and physiological mediators of CR could aid in the discovery of compounds/treatments that would act on those pathways, thereby mimicking the positive aspects of CR without imposed food restriction [24;69–73]. An ideal calorie restriction mimetic (CRM) is proposed to: i) produce metabolic, hormonal and physiological effects similar to CR, ii) not induce a significant reduction of long-term food intake, iii) activate stress response pathways similar to CR iv) while providing beneficial effects on mortality and age-related disease [70]. This area of research has progressed rapidly over the past decade as large-scale genomic, proteomic and metabolomic studies are performed in model organisms, attempting to unravel the complex interactions of genetics and nutrition that regulate aging and disease [74–85]. The National Institute on Aging has established the Interventions Testing Program

(<http://www.nia.nih.gov/ResearchInformation/ScientificResources/InterventionsTestingProgram.htm>) as a multi-institutional program to collaborate with partner researchers and test substances predicted to “extend lifespan and delay disease and dysfunction” [86–91]. Thus far approximately a dozen agents have been investigated in the program, yielding both promising and mixed results (see Table II). The structure of the program utilizes three separate test sites (the University of Michigan, the Jackson Laboratories and the University of Texas Health Sciences Center at San Antonio), with each candidate compound being tested for longevity effects in a total of 108 female and 132 male mice (n=36/F, 44/M per site) and untreated control groups with twice as many mice (n=72/F, 96/M per site) [87;88]. This provides a sufficient sample size to detect a 10% change in mean lifespan with 80% power even if data from one site is for some unforeseen reason unusable [87;88;90]. Results from one of the most recent test compounds, rapamycin, are discussed below.

Resveratrol

Of the CRMs thus far investigated, few have received more attention than resveratrol. Resveratrol is a plant-derived polyphenol, most well known for its presence in the skins of red grapes. Studies in yeast, worms and flies over the past decade have suggested that CR works by activating members of the Sirtuin family of protein de-acetylases to mediate the lifespan benefits [92–96]. Resveratrol is reported to activate Sir2 [97], thus mimicking the benefits of CR in the absence of actual nutrient alteration [97]. Additionally, treatment with resveratrol is reported to mimic CR by increasing lifespan in yeast, worms, flies and fish, potentially through the activation of sirtuins [97–100]. However, the fundamental role of the

sirtuins in mediating the benefits of CR in yeast has been challenged by demonstrations that CR can extend lifespan in the absence of Sir2 or other sirtuins [101–104], while the *in vivo* activation of Sir2 by CR or resveratrol to extend lifespan has been challenged in multiple organisms [105–113]. Recent reports have expanded previous work which showed the *in vitro* activation of SIRT1 by resveratrol is substrate specific, challenging the basic mechanism of sirtuin activating compounds currently being tested [105;114–116].

Despite these disparate data, it appears resveratrol treatment produces a transcriptional response similar to CR [117], and in the presence of a high-fat diet, both health and longevity benefits have been reported [118]. However, when resveratrol was added to a normal diet, no significant lifespan benefits were observed in mice [119], suggesting it is not a true CRM. Based on the current data, resveratrol supplementation produces a variety of physiological benefits [120], but whether these are mediated by the sirtuins and are a *bona fide* mimic of CR is questionable and will require further data and clarification [105;114–116].

Rapamycin

Rapamycin (RAP), another proposed CRM, is an antibiotic and inhibitor of TOR (Target of Rapamycin) signaling in cells, with known immunosuppressive and anti-proliferative effects [121]. TOR has been identified as a mediator of nutrient signaling in cells, and is proposed to play a role in aging and the CR response [121–131]. A recent ITP study reported a significant mean lifespan extension in both male (9%, $P < 0.0001$ [log-rank test]) and female (13%, $P < 0.0001$ [log-rank test]) mice fed a standard diet and administered RAP beginning at approximately 20 months of age [90]. This is the first compound to provide such robust lifespan benefit in the ITP. Interestingly, CR is usually initiated prior to 6 months of age, and although CR can extend lifespan even when started at older ages [1;132] the effect at older ages is less pronounced and less reliable [3]. Notwithstanding the increase in lifespan, no significant differences were observed in the distribution of lesions found at necropsy with RAP treatment, suggesting the longevity benefits of RAP treatment may be mediated by pathways partially independent of the normal CR response [90]. However, no measures of glucose, insulin or body temperature were reported to permit a comparison of RAP treatment with the expected results of CR, although body weight was not reduced with RAP treatment [90]. The authors recognized and reported differences in the composition of the pre-study diets, which although all were based on the same standard (NIH-31), varied in the specific formulations [90]. Nevertheless, one of the test sites (The Jackson Laboratory) utilized the control diet for the duration of the study and observed a significant increase in lifespan in both male ($P = 0.02$ [log-rank test]) and female ($P < 0.0001$ [log-rank test]) mice [90]. An additional cohort of mice with RAP treatment initiated at 9 months of age will likely further validate the potential benefit of RAP on lifespan [90]. A fundamental question that arises from these results is whether CR, when combined with RAP treatment, would provide additional health and lifespan benefits, particularly if RAP is acting on the pathways mediating CR's longevity effect? Likewise, these results should be tempered with the reality that RAP is used as an immunosuppressant, of limited consequence in rodent longevity studies since mice are maintained in specific pathogen free facilities. However, its utility for administration to healthy humans, which rely on a robust immune system in daily life, is currently unclear.

Other Potential CRM

Metformin—A hallmark of the CR response is reduced circulating glucose and insulin, while the role of insulin/IGF-1 in aging has received much support from model organism studies [133–137]. Therefore, it was proposed that drugs that could reduce insulin and glucose would be potential CRM candidates [24;70;133]. The biguanide metformin is used

in the treatment of diabetes where it functions to suppress gluconeogenesis and increase insulin sensitivity [138], suggesting it could mimic CR. Metformin is also reported to partially mimic the CR transcriptional response in mice [80] and increase median lifespan in *C. elegans* [139]. In addition, a number of studies have shown that metformin and related biguanides, phenformin and buformin, delay the incidence and development of cancers and other disease conditions [134;140–149]. However, a test of metformin as a CRM with a normal diet and in a non-disease rodent model has not been reported. To address this deficiency, a longevity study of healthy male Fischer-344 rats fed a standard diet with metformin supplementation (300 mg/kg/day) has been performed. Metformin did not significantly increase lifespan compared to control rats (unpublished data), although only one dose of metformin was tested and the CR group did not extend maximum lifespan in the study. Therefore, we await data showing a significant lifespan benefit in the absence of a disease state with metformin supplementation before a final verdict regarding its status as a true CRM.

2-Deoxyglucose—2-Deoxyglucose (2DG) is a non-metabolizable glucose analogue, which is taken up by cells where it accumulates while inhibiting glycolysis [69;133]. Thus, 2DG is proposed to reduce the metabolic flux of glucose, resulting in a reduced energetic flow in cellular metabolism resembling CR [69;133]. However, one difficulty with 2DG is the dose dependent inhibition of basic cellular function, resulting in toxicity with increasing concentration [70;133]. Preliminary experiments with 2DG supplementation resulted in lowered plasma insulin and body temperature, similar to CR [133]. When administered long-term by diet supplementation (either 0.2% or 0.4% 2DG) in male Fischer 344 rats, rather than increasing lifespan, a dose-dependent reduction in lifespan was observed [150], suggesting there is a fine line between pharmaceutically mimicking the effects of CR with 2DG without causing toxicity and death [72].

Longevity and Quality of Life, do we desire one without the other?

Should we be concerned about the potential for increased human lifespan? Clearly life expectancy has risen over the last century and appears to be continuing to do so, although disagreement exists regarding the potential of future increases [151–156]. However, some reports warn of a potential plateau or impending reversal of these lifespan gains in developed countries as a result of multiple factors, perhaps most notably the current obesity epidemic [157]. What is of most concern is the potential that lifespan will increase, while the onset of age-related disease and co-morbidity will remain the same, resulting in the unpleasant outcome of reduced quality of life for a greater duration in old age. Although much of aging research has focused on lifespan, the number of days of life until death, an alternate measure of aging termed the healthspan, or the length of time prior to the onset of age-related disease, has been considered and may be of particular importance to humans as life expectancy continues to increase [158–161]. The results from the Wisconsin Rhesus monkey CR study [12], as well as other nonhuman primate studies, suggest even if longevity benefits are realized with CR, they may be secondary to the health gains achieved. Thus, the academic or esoteric question of whether lifespan can be truly extended by CR in humans may not be as important as the potential prolongation of healthspan. If these types of health-promoting and disease-reducing results can be achieved in humans, as short-term CR studies suggest, the answer may be that quality of life can be extended, potentially into advanced age. Likewise, the search for CRMs that extend lifespan, without altering the underlying disease pattern, would be of little utility. Therefore, careful examination of multiple outcomes beyond lifespan should be considered in any CRM intervention study to assess the effect on disease. Further study to identify the central pathways which mediate the beneficial responses of CR should be of high priority as these may serve as useful targets for interventions to improve health and possibly lifespan.

Concluding Remarks

The recent reports of Colman et al. (2009) and Harrison et al. (2009) illustrate the potential of translating fundamental discoveries across organisms in the effort to retard aging and disease. Due to the nature of a longevity study in primates, the final answer regarding the effect of CR on total and maximal lifespan is still probably a decade away. Whether these types of interventions will reduce disease incidence/severity and increase lifespan in humans is still unknown. Nevertheless, these results are pointing in a positive direction and suggest that finding a means to implement or mimic the CR response in humans could significantly affect the health and well-being of our species, regardless of the eventual lifespan result.

Acknowledgments

We thank Matt Giddings for critically reading the manuscript. Supported in part by NIH grants P01AG011915 and P30DK056336. DLS is supported by T32DK062710. The opinions expressed herein are those of the authors and not necessarily those of the NIH or any other organization with which the authors are affiliated.

DBA has received grants, consulting fees, and donations from multiple profit and non profit entities with interests in obesity and CR mimetics.

Reference List

- [1]. Weindruch, R.; Walford, RL. *The Retardation of Aging and Disease by Dietary Restriction*. C. C. Thomas Publisher; Springfield, IL: 1988.
- [2]. Masoro EJ. Subfield history: caloric restriction, slowing aging, and extending life. *Sci Aging Knowledge Environ*. Feb 26.2003 2003:RE2. [PubMed: 12844547]
- [3]. Masoro EJ. Overview of caloric restriction and ageing. *Mech Ageing Dev*. 2005; 126:913–922. [PubMed: 15885745]
- [4]. Barrows CH Jr. Kokkonen G. The effect of various dietary restricted regimes on biochemical variables in the mouse. *Growth*. 1978; 42:71–85. [PubMed: 669401]
- [5]. Ingram DK, Cutler RG, Weindruch R, Renquist DM, Knapka JJ, April M, Belcher CT, Clark MA, Hatcherson CD, Marriott BM. Dietary restriction and aging: the initiation of a primate study. *J Gerontol*. 1990; 45:B148–B163. [PubMed: 2394908]
- [6]. Lane MA, Ingram DK, Cutler RG, Knapka JJ, Barnard DE, Roth GS. Dietary restriction in nonhuman primates: progress report on the NIA study. *Ann N Y Acad Sci*. Dec 26.1992 673:36–45. [PubMed: 1485732]
- [7]. Kemnitz JW, Weindruch R, Roecker EB, Crawford K, Kaufman PL, Ershler WB. Dietary restriction of adult male rhesus monkeys: design, methodology, and preliminary findings from the first year of study. *J Gerontol*. 1993; 48:B17–B26. [PubMed: 8418134]
- [8]. Bodkin NL, Alexander TM, Ortmeyer HK, Johnson E, Hansen BC. Mortality and morbidity in laboratory-maintained Rhesus monkeys and effects of long-term dietary restriction. *J Gerontol A Biol Sci Med Sci*. 2003; 58:212–219. [PubMed: 12634286]
- [9]. Bodkin NL, Ortmeyer HK, Hansen BC. Long-term dietary restriction in older-aged rhesus monkeys: effects on insulin resistance. *J Gerontol A Biol Sci Med Sci*. 1995; 50:B142–B147. [PubMed: 7743393]
- [10]. Hansen BC, Bodkin NL, Ortmeyer HK. Calorie restriction in nonhuman primates: mechanisms of reduced morbidity and mortality. *Toxicol Sci*. 1999; 52:56–60. [PubMed: 10630591]
- [11]. Hansen BC, Bodkin NL. Primary prevention of diabetes mellitus by prevention of obesity in monkeys. *Diabetes*. 1993; 42:1809–1814. [PubMed: 8243827]
- [12]. Colman RJ, Anderson RM, Johnson SC, Kastman EK, Kosmatka KJ, Beasley TM, Allison DB, Cruzen C, Simmons HA, Kemnitz JW, Weindruch R. Caloric restriction delays disease onset and mortality in rhesus monkeys. *Science*. Jul 10.2009 325:201–204. [PubMed: 19590001]
- [13]. Lane MA, Baer DJ, Tilmont EM, Rumpler WV, Ingram DK, Roth GS, Cutler RG. Energy balance in rhesus monkeys (*Macaca mulatta*) subjected to long-term dietary restriction. *J Gerontol A Biol Sci Med Sci*. 1995; 50:B295–B302. [PubMed: 7671021]

- [14]. Lane MA, Ball SS, Ingram DK, Cutler RG, Engel J, Read V, Roth GS. Diet restriction in rhesus monkeys lowers fasting and glucose-stimulated glucoend points. *Am J Physiol.* 1995; 268:E941–E948. [PubMed: 7762649]
- [15]. Lane MA, Baer DJ, Rumpler WV, Weindruch R, Ingram DK, Tilmont EM, Cutler RG, Roth GS. Calorie restriction lowers body temperature in rhesus monkeys, consistent with a postulated anti-aging mechanism in rodents. *Proc Natl Acad Sci U S A.* Apr 30.1996 93:4159–4164. [PubMed: 8633033]
- [16]. Lane MA, Ingram DK, Ball SS, Roth GS. Dehydroepiandrosterone sulfate: a biomarker of primate aging slowed by calorie restriction. *J Clin Endocrinol Metab.* 1997; 82:2093–2096. [PubMed: 9215277]
- [17]. Mattison JA, Lane MA, Roth GS, Ingram DK. Calorie restriction in rhesus monkeys. *Exp Gerontol.* 2003; 38:35–46. [PubMed: 12543259]
- [18]. Kemnitz JW, Roecker EB, Weindruch R, Elson DF, Baum ST, Bergman RN. Dietary restriction increases insulin sensitivity and lowers blood glucose in rhesus monkeys. *Am J Physiol.* 1994; 266:E540–E547. [PubMed: 8178974]
- [19]. Ramsey JJ, Colman RJ, Binkley NC, Christensen JD, Gresl TA, Kemnitz JW, Weindruch R. Dietary restriction and aging in rhesus monkeys: the University of Wisconsin study. *Exp Gerontol.* 2000; 35:1131–1149. [PubMed: 11113597]
- [20]. Colman RJ, Roecker EB, Ramsey JJ, Kemnitz JW. The effect of dietary restriction on body composition in adult male and female rhesus macaques. *Aging (Milano).* 1998; 10:83–92. [PubMed: 9666188]
- [21]. Colman RJ, Beasley TM, Allison DB, Weindruch R. Attenuation of sarcopenia by dietary restriction in rhesus monkeys. *J Gerontol A Biol Sci Med Sci.* 2008; 63:556–559. [PubMed: 18559628]
- [22]. Gresl TA, Colman RJ, Roecker EB, Havighurst TC, Huang Z, Allison DB, Bergman RN, Kemnitz JW. Dietary restriction and glucose regulation in aging rhesus monkeys: a follow-up report at 8.5 yr. *Am J Physiol Endocrinol Metab.* 2001; 281:E757–E765. [PubMed: 11551852]
- [23]. Lane MA, Ingram DK, Roth GS. Calorie restriction in nonhuman primates: effects on diabetes and cardiovascular disease risk. *Toxicol Sci.* 1999; 52:41–48. [PubMed: 10630589]
- [24]. Ingram DK, Anson RM, de CR, Mamczarz J, Zhu M, Mattison J, Lane MA, Roth GS. Development of calorie restriction mimetics as a prolongevity strategy. *Ann N Y Acad Sci.* 2004; 1019:412–423. [PubMed: 15247056]
- [25]. Mattison JA, Roth GS, Lane MA, Ingram DK. Dietary restriction in aging nonhuman primates. *Interdiscip Top Gerontol.* 2007; 35:137–158. [PubMed: 17063037]
- [26]. Keys, A.; Brozek, J.; Henschel, A.; Mickelsen, O.; Taylor, HL. *The Biology of Human Starvation.* University of Minnesota Press; 1950.
- [27]. Walford RL, Mock D, Verdery R, MacCallum T. Calorie restriction in biosphere 2: alterations in physiologic, hematologic, hormonal, and biochemical parameters in humans restricted for a 2-year period. *J Gerontol A Biol Sci Med Sci.* 2002; 57:B211–B224. [PubMed: 12023257]
- [28]. Walford RL, Mock D, MacCallum T, Laseter JL. Physiologic changes in humans subjected to severe, selective calorie restriction for two years in biosphere 2: health, aging, and toxicological perspectives. *Toxicol Sci.* 1999; 52:61–65. [PubMed: 10630592]
- [29]. Shanley DP, Kirkwood TB. Caloric restriction does not enhance longevity in all species and is unlikely to do so in humans. *Biogerontology.* 2006; 7:165–168. [PubMed: 16858629]
- [30]. Austad SN. Does caloric restriction in the laboratory simply prevent overfeeding and return house mice to their natural level of food intake? *Sci Aging Knowledge Environ.* Nov 7.2001 2001:e3.
- [31]. Austad SN, Kristan DM. Are mice calorically restricted in nature? *Aging Cell.* 2003; 2:201–207. [PubMed: 12934713]
- [32]. Harper JM, Leathers CW, Austad SN. Does caloric restriction extend life in wild mice? *Aging Cell.* 2006; 5:441–449. [PubMed: 17054664]
- [33]. Speakman JR, Hambly C. Starving for life: what animal studies can and cannot tell us about the use of caloric restriction to prolong human lifespan. *J Nutr.* 2007; 137:1078–1086. [PubMed: 17374682]

- [34]. Ogden CL, Carroll MD, Curtin LR, McDowell MA, Tabak CJ, Flegal KM. Prevalence of overweight and obesity in the United States, 1999–2004. *JAMA*. Apr 5.2006 295:1549–1555. [PubMed: 16595758]
- [35]. Phelan JP, Rose MR. Caloric restriction increases longevity substantially only when the reaction norm is steep. *Biogerontology*. 2006; 7:161–164. [PubMed: 16858630]
- [36]. Phelan JP, Rose MR. Why dietary restriction substantially increases longevity in animal models but won't in humans. *Ageing Res Rev*. 2005; 4:339–350. [PubMed: 16046282]
- [37]. Weindruch R. Will dietary restriction work in primates? *Biogerontology*. 2006; 7:169–171. [PubMed: 16680522]
- [38]. Yu BP. Why calorie restriction would work for human longevity. *Biogerontology*. 2006; 7:179–182. [PubMed: 16676136]
- [39]. Roth GS, Ingram DK, Lane MA. Calorie restriction in primates: will it work and how will we know? *J Am Geriatr Soc*. 1999; 47:896–903. [PubMed: 10404938]
- [40]. Civitarese AE, Carling S, Heilbronn LK, Hulver MH, Ukropcova B, Deutsch WA, Smith SR, Ravussin E. Calorie restriction increases muscle mitochondrial biogenesis in healthy humans. *PLoS Med*. 2007; 4:e76. [PubMed: 17341128]
- [41]. Heilbronn LK, de JL, Frisard MI, Delany JP, Larson-Meyer DE, Rood J, Nguyen T, Martin CK, Volaufova J, Most MM, Greenway FL, Smith SR, Deutsch WA, Williamson DA, Ravussin E. Effect of 6-month calorie restriction on biomarkers of longevity, metabolic adaptation, and oxidative stress in overweight individuals: a randomized controlled trial. *JAMA*. Apr 5.2006 295:1539–1548. [PubMed: 16595757]
- [42]. Heilbronn LK, Smith SR, Martin CK, Anton SD, Ravussin E. Alternate-day fasting in nonobese subjects: effects on body weight, body composition, and energy metabolism. *Am J Clin Nutr*. 2005; 81:69–73. [PubMed: 15640462]
- [43]. Larson-Meyer DE, Newcomer BR, Heilbronn LK, Volaufova J, Smith SR, Alfonso AJ, Lefevre M, Rood JC, Williamson DA, Ravussin E. Effect of 6-month calorie restriction and exercise on serum and liver lipids and markers of liver function. *Obesity (Silver Spring)*. 2008; 16:1355–1362. [PubMed: 18421281]
- [44]. Larson-Meyer DE, Heilbronn LK, Redman LM, Newcomer BR, Frisard MI, Anton S, Smith SR, Alfonso A, Ravussin E. Effect of calorie restriction with or without exercise on insulin sensitivity, beta-cell function, fat cell size, and ectopic lipid in overweight subjects. *Diabetes Care*. 2006; 29:1337–1344. [PubMed: 16732018]
- [45]. Lefevre M, Redman LM, Heilbronn LK, Smith JV, Martin CK, Rood JC, Greenway FL, Williamson DA, Smith SR, Ravussin E. Caloric restriction alone and with exercise improves CVD risk in healthy non-obese individuals. *Atherosclerosis*. 2009; 203:206–213. [PubMed: 18602635]
- [46]. Redman LM, Heilbronn LK, Martin CK, Alfonso A, Smith SR, Ravussin E. Effect of calorie restriction with or without exercise on body composition and fat distribution. *J Clin Endocrinol Metab*. 2007; 92:865–872. [PubMed: 17200169]
- [47]. Fontana L, Villareal DT, Weiss EP, Racette SB, Steger-May K, Klein S, Holloszy JO. Calorie restriction or exercise: effects on coronary heart disease risk factors. A randomized, controlled trial. *Am J Physiol Endocrinol Metab*. 2007; 293:E197–E202. [PubMed: 17389710]
- [48]. Holloszy JO, Fontana L. Caloric restriction in humans. *Exp Gerontol*. 2007; 42:709–712. [PubMed: 17482403]
- [49]. Racette SB, Weiss EP, Villareal DT, Arif H, Steger-May K, Schechtman KB, Fontana L, Klein S, Holloszy JO. One year of caloric restriction in humans: feasibility and effects on body composition and abdominal adipose tissue. *J Gerontol A Biol Sci Med Sci*. 2006; 61:943–950. [PubMed: 16960025]
- [50]. Weiss EP, Racette SB, Villareal DT, Fontana L, Steger-May K, Schechtman KB, Klein S, Ehsani AA, Holloszy JO. Lower extremity muscle size and strength and aerobic capacity decrease with caloric restriction but not with exercise-induced weight loss. *J Appl Physiol*. 2007; 102:634–640. [PubMed: 17095635]

- [51]. Fontana L, Meyer TE, Klein S, Holloszy JO. Long-term calorie restriction is highly effective in reducing the risk for atherosclerosis in humans. *Proc Natl Acad Sci U S A*. Apr 27.2004 101:6659–6663. [PubMed: 15096581]
- [52]. Fontana L, Klein S, Holloszy JO. Long-term low-protein, low-calorie diet and endurance exercise modulate metabolic factors associated with cancer risk. *Am J Clin Nutr*. 2006; 84:1456–1462. [PubMed: 17158430]
- [53]. Fontana L, Meyer TE, Klein S, Holloszy JO. Long-term low-calorie low-protein vegan diet and endurance exercise are associated with low cardiometabolic risk. *Rejuvenation Res*. 2007; 10:225–234. [PubMed: 17518696]
- [54]. Fontana L, Weiss EP, Villareal DT, Klein S, Holloszy JO. Long-term effects of calorie or protein restriction on serum IGF-1 and IGFBP-3 concentration in humans. *Aging Cell*. 2008; 7:681–687. [PubMed: 18843793]
- [55]. Fontana L. Modulating human aging and age-associated diseases. *Biochim Biophys Acta*. 2009; 1790:1133–1138. [PubMed: 19364477]
- [56]. Fontana L. The scientific basis of caloric restriction leading to longer life. *Curr Opin Gastroenterol*. 2009; 25:144–150. [PubMed: 19262201]
- [57]. Bray GA. Medical consequences of obesity. *J Clin Endocrinol Metab*. 2004; 89:2583–2589. [PubMed: 15181027]
- [58]. Friedman N, Fanning EL. Overweight and obesity: an overview of prevalence, clinical impact, and economic impact. *Dis Manag*. 2004; 7(Suppl 1):S1–S6. [PubMed: 15669572]
- [59]. Malloy VL, Krajcik RA, Bailey SJ, Hristopoulos G, Plummer JD, Orentreich N. Methionine restriction decreases visceral fat mass and preserves insulin action in aging male Fischer 344 rats independent of energy restriction. *Aging Cell*. 2006; 5:305–314. [PubMed: 16800846]
- [60]. Miller RA, Buehner G, Chang Y, Harper JM, Sigler R, Smith-Wheelock M. Methionine-deficient diet extends mouse lifespan, slows immune and lens aging, alters glucose, T4, IGF-I and insulin levels, and increases hepatocyte MIF levels and stress resistance. *Aging Cell*. 2005; 4:119–125. [PubMed: 15924568]
- [61]. Orentreich N, Matias JR, DeFelice A, Zimmerman JA. Low methionine ingestion by rats extends life span. *J Nutr*. 1993; 123:269–274. [PubMed: 8429371]
- [62]. Richie JP Jr, Leutzinger Y, Parthasarathy S, Malloy V, Orentreich N, Zimmerman JA. Methionine restriction increases blood glutathione and longevity in F344 rats. *FASEB J*. 1994; 8:1302–1307. [PubMed: 8001743]
- [63]. Sun L, Sadighi Akha AA, Miller RA, Harper JM. Life-span extension in mice by preweaning food restriction and by methionine restriction in middle age. *J Gerontol A Biol Sci Med Sci*. 2009; 64:711–722. [PubMed: 19414512]
- [64]. Zimmerman JA, Malloy V, Krajcik R, Orentreich N. Nutritional control of aging. *Exp Gerontol*. 2003; 38:47–52. [PubMed: 12543260]
- [65]. Anson RM, Guo Z, de CR, Iyun T, Rios M, Hagepanos A, Ingram DK, Lane MA, Mattson MP. Intermittent fasting dissociates beneficial effects of dietary restriction on glucose metabolism and neuronal resistance to injury from calorie intake. *Proc Natl Acad Sci U S A*. May 13.2003 100:6216–6220. [PubMed: 12724520]
- [66]. Martin B, Mattson MP, Maudsley S. Caloric restriction and intermittent fasting: two potential diets for successful brain aging. *Ageing Res Rev*. 2006; 5:332–353. [PubMed: 16899414]
- [67]. Mattson MP. Dietary factors, hormesis and health. *Ageing Res Rev*. 2008; 7:43–48. [PubMed: 17913594]
- [68]. Mattson MP, Wan R. Beneficial effects of intermittent fasting and caloric restriction on the cardiovascular and cerebrovascular systems. *J Nutr Biochem*. 2005; 16:129–137. [PubMed: 15741046]
- [69]. Lane MA, Ingram DK, Roth GS. 2-Deoxy-D-glucose feeding in rats mimics physiological effects of calorie restriction. *J Anti Aging Med*. 1998; 1:327–337.
- [70]. Ingram DK, Zhu M, Mamczarz J, Zou S, Lane MA, Roth GS, deCabo R. Calorie restriction mimetics: an emerging research field. *Aging Cell*. 2006; 5:97–108. [PubMed: 16626389]

- [71]. Lane MA, de CR, Mattison J, Anson RM, Roth GS, Ingram DK. The Roy Walford legacy: diet restriction from molecules to mice to monkeys to man and onto mimetics. *Exp Gerontol.* 2004; 39:897–902. [PubMed: 15217686]
- [72]. Lane MA, Roth GS, Ingram DK. Caloric restriction mimetics: a novel approach for biogerontology. *Methods Mol Biol.* 2007; 371:143–149. [PubMed: 17634579]
- [73]. Roth GS, Lane MA, Ingram DK. Caloric restriction mimetics: the next phase. *Ann N Y Acad Sci.* 2005; 1057:365–371. [PubMed: 16399906]
- [74]. Kennedy BK. The genetics of ageing: insight from genome-wide approaches in invertebrate model organisms. *J Intern Med.* 2008; 263:142–152. [PubMed: 18226092]
- [75]. Weindruch R, Kayo T, Lee CK, Prolla TA. Microarray profiling of gene expression in aging and its alteration by caloric restriction in mice. *J Nutr.* 2001; 131:918S–923S. [PubMed: 11238786]
- [76]. Weindruch R, Kayo T, Lee CK, Prolla TA. Gene expression profiling of aging using DNA microarrays. *Mech Ageing Dev.* 2002; 123:177–193. [PubMed: 11718811]
- [77]. Weindruch R, Kayo T, Lee CK, Prolla TA. Effects of caloric restriction on gene expression. *Nestle Nutr Workshop Ser Clin Perform Programme.* 2002; 6:17–28.
- [78]. Spindler SR. Use of microarray biomarkers to identify longevity therapeutics. *Aging Cell.* 2006; 5:39–50. [PubMed: 16441842]
- [79]. Spindler SR, Mote PL. Screening candidate longevity therapeutics using gene-expression arrays. *Gerontology.* 2007; 53:306–321. [PubMed: 17570924]
- [80]. Dhabhi JM, Mote PL, Fahy GM, Spindler SR. Identification of potential caloric restriction mimetics by microarray profiling. *Physiol Genomics.* Nov 17.2005 23:343–350. [PubMed: 16189280]
- [81]. Dhabhi JM, Tsuchiya T, Kim HJ, Mote PL, Spindler SR. Gene expression and physiologic responses of the heart to the initiation and withdrawal of caloric restriction. *J Gerontol A Biol Sci Med Sci.* 2006; 61:218–231. [PubMed: 16567370]
- [82]. Dhabhi JM, Kim HJ, Mote PL, Beaver RJ, Spindler SR. Temporal linkage between the phenotypic and genomic responses to caloric restriction. *Proc Natl Acad Sci U S A.* Apr 13.2004 101:5524–5529. [PubMed: 15044709]
- [83]. Kayo T, Allison DB, Weindruch R, Prolla TA. Influences of aging and caloric restriction on the transcriptional profile of skeletal muscle from rhesus monkeys. *Proc Natl Acad Sci U S A.* Apr 24.2001 98:5093–5098. [PubMed: 11309484]
- [84]. Lee CK, Klopp RG, Weindruch R, Prolla TA. Gene expression profile of aging and its retardation by caloric restriction. *Science.* Aug 27.1999 285:1390–1393. [PubMed: 10464095]
- [85]. Higami Y, Pugh TD, Page GP, Allison DB, Prolla TA, Weindruch R. Adipose tissue energy metabolism: altered gene expression profile of mice subjected to long-term caloric restriction. *FASEB J.* 2004; 18:415–417. [PubMed: 14688200]
- [86]. Warner HR, Ingram D, Miller RA, Nadon NL, Richardson AG. Program for testing biological interventions to promote healthy aging. *Mech Ageing Dev.* Jun 20.2000 115:199–207. [PubMed: 10906513]
- [87]. Nadon NL, Strong R, Miller RA, Nelson J, Javors M, Sharp ZD, Peralba JM, Harrison DE. Design of aging intervention studies: the NIA interventions testing program. *Age (Dordr).* 2008; 30:187–199. [PubMed: 19424842]
- [88]. Miller RA, Harrison DE, Astle CM, Floyd RA, Flurkey K, Hensley KL, Javors MA, Leeuwenburgh C, Nelson JF, Ongini E, Nadon NL, Warner HR, Strong R. An Aging Interventions Testing Program: study design and interim report. *Aging Cell.* 2007; 6:565–575. [PubMed: 17578509]
- [89]. Strong R, Miller RA, Astle CM, Floyd RA, Flurkey K, Hensley KL, Javors MA, Leeuwenburgh C, Nelson JF, Ongini E, Nadon NL, Warner HR, Harrison DE. Nordihydroguaiaretic acid and aspirin increase lifespan of genetically heterogeneous male mice. *Aging Cell.* 2008; 7:641–650. [PubMed: 18631321]
- [90]. Harrison DE, Strong R, Sharp ZD, Nelson JF, Astle CM, Flurkey K, Nadon NL, Wilkinson JE, Frenkel K, Carter CS, Pahor M, Javors MA, Fernandez E, Miller RA. Rapamycin fed late in life extends lifespan in genetically heterogeneous mice. *Nature.* Jul 16.2009 460:392–395. [PubMed: 19587680]

- [91]. Nadon NL. Exploiting the rodent model for studies on the pharmacology of lifespan extension. *Aging Cell*. 2006; 5:9–15. [PubMed: 16441838]
- [92]. Lin SJ, Defossez PA, Guarente L. Requirement of NAD and SIR2 for life-span extension by calorie restriction in *Saccharomyces cerevisiae*. *Science*. Sep 22.2000 289:2126–2128. [PubMed: 11000115]
- [93]. Lin SJ, Kaerberlein M, Andalis AA, Sturtz LA, Defossez PA, Culotta VC, Fink GR, Guarente L. Calorie restriction extends *Saccharomyces cerevisiae* lifespan by increasing respiration. *Nature*. Jul 18.2002 418:344–348. [PubMed: 12124627]
- [94]. Anderson RM, Latorre-Esteves M, Neves AR, Lavu S, Medvedik O, Taylor C, Howitz KT, Santos H, Sinclair DA. Yeast life-span extension by calorie restriction is independent of NAD fluctuation. *Science*. Dec 19.2003 302:2124–2126. [PubMed: 14605207]
- [95]. Wang Y, Tissenbaum HA. Overlapping and distinct functions for a *Caenorhabditis elegans* SIR2 and DAF-16/FOXO. *Mech Ageing Dev*. 2006; 127:48–56. [PubMed: 16280150]
- [96]. Rogina B, Helfand SL. Sir2 mediates longevity in the fly through a pathway related to calorie restriction. *Proc Natl Acad Sci U S A*. Nov 9.2004 101:15998–16003. [PubMed: 15520384]
- [97]. Howitz KT, Bitterman KJ, Cohen HY, Lamming DW, Lavu S, Wood JG, Zipkin RE, Chung P, Kisielewski A, Zhang LL, Scherer B, Sinclair DA. Small molecule activators of sirtuins extend *Saccharomyces cerevisiae* lifespan. *Nature*. Sep 11.2003 425:191–196. [PubMed: 12939617]
- [98]. Wood JG, Rogina B, Lavu S, Howitz K, Helfand SL, Tatar M, Sinclair D. Sirtuin activators mimic caloric restriction and delay ageing in metazoans. *Nature*. Aug 5.2004 430:686–689. [PubMed: 15254550]
- [99]. Yang H, Baur JA, Chen A, Miller C, Adams JK, Kisielewski A, Howitz KT, Zipkin RE, Sinclair DA. Design and synthesis of compounds that extend yeast replicative lifespan. *Aging Cell*. 2007; 6:35–43. [PubMed: 17156081]
- [100]. Valenzano DR, Terzibasi E, Genade T, Cattaneo A, Domenici L, Cellierino A. Resveratrol prolongs lifespan and retards the onset of age-related markers in a short-lived vertebrate. *Curr Biol*. Feb 7.2006 16:296–300. [PubMed: 16461283]
- [101]. Jiang JC, Wawryn J, Shantha Kumara HM, Jazwinski SM. Distinct roles of processes modulated by histone deacetylases Rpd3p, Hda1p, and Sir2p in life extension by caloric restriction in yeast. *Exp Gerontol*. 2002; 37:1023–1030. [PubMed: 12213553]
- [102]. Kaerberlein M, Kirkland KT, Fields S, Kennedy BK. Sir2-independent life span extension by calorie restriction in yeast. *PLoS Biol*. 2004; 2:E296. [PubMed: 15328540]
- [103]. Smith DL Jr, McClure JM, Matecic M, Smith JS. Calorie restriction extends the chronological lifespan of *Saccharomyces cerevisiae* independently of the Sirtuins. *Aging Cell*. 2007; 6:649–662. [PubMed: 17711561]
- [104]. Kaerberlein M, Steffen KK, Hu D, Dang N, Kerr EO, Tsuchiya M, Fields S, Kennedy BK. HST2 mediates SIR2-independent life-span extension by calorie restriction. *Science*. Jun 2.2006 312:1312. [PubMed: 16741098]
- [105]. Kaerberlein M, McDonagh T, Heltweg B, Hixon J, Westman EA, Caldwell SD, Napper A, Curtis R, DiStefano PS, Fields S, Bedalov A, Kennedy BK. Substrate-specific activation of sirtuins by resveratrol. *J Biol Chem*. Apr 29.2005 280:17038–17045. [PubMed: 15684413]
- [106]. Kaerberlein M, Powers RW III. Sir2 and calorie restriction in yeast: a skeptical perspective. *Ageing Res Rev*. 2007; 6:128–140. [PubMed: 17512264]
- [107]. Kaerberlein M, Kennedy BK. Does resveratrol activate yeast Sir2 in vivo? *Aging Cell*. 2007; 6:415–416. [PubMed: 17635418]
- [108]. Smith DL Jr, Li C, Matecic M, Maqani N, Bryk M, Smith JS. Calorie restriction effects on silencing and recombination at the yeast rDNA. *Aging Cell*. Sep 2.2009
- [109]. Bass TM, Weinkove D, Houthoofd K, Gems D, Partridge L. Effects of resveratrol on lifespan in *Drosophila melanogaster* and *Caenorhabditis elegans*. *Mech Ageing Dev*. 2007; 128:546–552. [PubMed: 17875315]
- [110]. Greer EL, Brunet A. Different dietary restriction regimens extend lifespan by both independent and overlapping genetic pathways in *C. elegans*. *Aging Cell*. 2009; 8:113–127. [PubMed: 19239417]

- [111]. Kaerberlein TL, Smith ED, Tsuchiya M, Welton KL, Thomas JH, Fields S, Kennedy BK, Kaerberlein M. Lifespan extension in *Caenorhabditis elegans* by complete removal of food. *Aging Cell*. 2006; 5:487–494. [PubMed: 17081160]
- [112]. Zou S, Carey JR, Liedo P, Ingram DK, Muller HG, Wang JL, Yao F, Yu B, Zhou A. The longevity effect of resveratrol depends on dietary composition and calorie intake in a tephritid fruit fly. *Exp Gerontol*. 2009; 44:472–476. [PubMed: 19264118]
- [113]. Riesen M, Morgan A. Calorie restriction reduces rDNA recombination independently of rDNA silencing. *Aging Cell*. 2009; 8:624–632. [PubMed: 19732046]
- [114]. Borra MT, Smith BC, Denu JM. Mechanism of human SIRT1 activation by resveratrol. *J Biol Chem*. Apr 29.2005 280:17187–17195. [PubMed: 15749705]
- [115]. Beher D, Wu J, Cumine S, Kim KW, Lu SC, Atangan L, Wang M. Resveratrol is not a direct activator of SIRT1 enzyme activity. *Chem Biol Drug Des*. 2009; 74:619–624. [PubMed: 19843076]
- [116]. Pacholec M, Chrunyk BA, Cunningham D, Flynn D, Griffith DA, Griffor M, Loulakis P, Pabst B, Qiu X, Stockman B, Thanabal V, Varghese A, Ward J, Withka J, Ahn K. SRT1720, SRT2183, SRT1460, and resveratrol are not direct activators of SIRT1. *J Biol Chem*. Jan 8.2010
- [117]. Barger JL, Kayo T, Vann JM, Arias EB, Wang J, Hacker TA, Wang Y, Raederstorff D, Morrow JD, Leeuwenburgh C, Allison DB, Saupe KW, Cartee GD, Weindruch R, Prolla TA. A low dose of dietary resveratrol partially mimics caloric restriction and retards aging parameters in mice. *PLoS ONE*. 2008; 3:e2264. [PubMed: 18523577]
- [118]. Baur JA, Pearson KJ, Price NL, Jamieson HA, Lerin C, Kalra A, Prabhu VV, Allard JS, Lopez-Lluch G, Lewis K, Pistell PJ, Poosala S, Becker KG, Boss O, Gwinn D, Wang M, Ramaswamy S, Fishbein KW, Spencer RG, Lakatta EG, Le CD, Shaw RJ, Navas P, Puigserver P, Ingram DK, de CR, Sinclair DA. Resveratrol improves health and survival of mice on a high-calorie diet. *Nature*. Nov 16.2006 444:337–342. [PubMed: 17086191]
- [119]. Pearson KJ, Baur JA, Lewis KN, Peshkin L, Price NL, Labinskyy N, Swindell WR, Kamara D, Minor RK, Perez E, Jamieson HA, Zhang Y, Dunn SR, Sharma K, Pleshko N, Woollett LA, Csiszar A, Ikeno Y, Le CD, Elliott PJ, Becker KG, Navas P, Ingram DK, Wolf NS, Ungvari Z, Sinclair DA, de CR. Resveratrol delays age-related deterioration and mimics transcriptional aspects of dietary restriction without extending life span. *Cell Metab*. 2008; 8:157–168. [PubMed: 18599363]
- [120]. Baur JA, Sinclair DA. Therapeutic potential of resveratrol: the in vivo evidence. *Nat Rev Drug Discov*. 2006; 5:493–506. [PubMed: 16732220]
- [121]. Crespo JL, Hall MN. Elucidating TOR signaling and rapamycin action: lessons from *Saccharomyces cerevisiae*. *Microbiol Mol Biol Rev*. 2002; 66:579–91. table. [PubMed: 12456783]
- [122]. Kaerberlein M, Powers RW III, Steffen KK, Westman EA, Hu D, Dang N, Kerr EO, Kirkland KT, Fields S, Kennedy BK. Regulation of yeast replicative life span by TOR and Sch9 in response to nutrients. *Science*. Nov 18.2005 310:1193–1196. [PubMed: 16293764]
- [123]. Martin DE, Hall MN. The expanding TOR signaling network. *Curr Opin Cell Biol*. 2005; 17:158–166. [PubMed: 15780592]
- [124]. Stanfel MN, Shamieh LS, Kaerberlein M, Kennedy BK. The TOR pathway comes of age. *Biochim Biophys Acta*. 2009; 1790:1067–1074. [PubMed: 19539012]
- [125]. Powers RW III, Kaerberlein M, Caldwell SD, Kennedy BK, Fields S. Extension of chronological life span in yeast by decreased TOR pathway signaling. *Genes Dev*. Jan 15.2006 20:174–184. [PubMed: 16418483]
- [126]. Jia K, Chen D, Riddle DL. The TOR pathway interacts with the insulin signaling pathway to regulate *C. elegans* larval development, metabolism and life span. *Development*. 2004; 131:3897–3906. [PubMed: 15253933]
- [127]. Chen D, Thomas EL, Kapahi P. HIF-1 modulates dietary restriction-mediated lifespan extension via IRE-1 in *Caenorhabditis elegans*. *PLoS Genet*. 2009; 5:e1000486. [PubMed: 19461873]
- [128]. Kapahi P, Zid B. TOR pathway: linking nutrient sensing to life span. *Sci Aging Knowledge Environ*. Sep 8.2004 2004:E34.

- [129]. Kapahi P, Zid BM, Harper T, Koslover D, Sapin V, Benzer S. Regulation of lifespan in *Drosophila* by modulation of genes in the TOR signaling pathway. *Curr Biol*. May 25.2004 14:885–890. [PubMed: 15186745]
- [130]. Toth ML, Sigmond T, Borsos E, Barna J, Erdelyi P, Takacs-Vellai K, Orosz L, Kovacs AL, Csikos G, Sass M, Vellai T. Longevity pathways converge on autophagy genes to regulate life span in *Caenorhabditis elegans*. *Autophagy*. Apr 1.2008 4:330–338. [PubMed: 18219227]
- [131]. Vellai T, Takacs-Vellai K, Zhang Y, Kovacs AL, Orosz L, Muller F. Genetics: influence of TOR kinase on lifespan in *C. elegans*. *Nature*. Dec 11.2003 426:620. [PubMed: 14668850]
- [132]. Weindruch R, Walford RL. Dietary restriction in mice beginning at 1 year of age: effect on lifespan and spontaneous cancer incidence. *Science*. Mar 12.1982 215:1415–1418. [PubMed: 7063854]
- [133]. Roth GS, Ingram DK, Lane MA. Caloric restriction in primates and relevance to humans. *Ann N Y Acad Sci*. 2001; 928:305–315. [PubMed: 11795522]
- [134]. Anisimov VN, Semenchenko AV, Yashin AI. Insulin and longevity: antidiabetic biguanides as geroprotectors. *Biogerontology*. 2003; 4:297–307. [PubMed: 14618027]
- [135]. Longo VD, Fabrizio P. Regulation of longevity and stress resistance: a molecular strategy conserved from yeast to humans? *Cell Mol Life Sci*. 2002; 59:903–908. [PubMed: 12169020]
- [136]. Bartke A. Insulin and aging. *Cell Cycle*. Nov 1.2008 7:3338–3343. [PubMed: 18948730]
- [137]. Bartke A. Minireview: role of the growth hormone/insulin-like growth factor system in mammalian aging. *Endocrinology*. 2005; 146:3718–3723. [PubMed: 15919742]
- [138]. Radziuk J, Bailey CJ, Wiernsperger NF, Yudkin JS. Metformin and its liver targets in the treatment of type 2 diabetes. *Curr Drug Targets Immune Endocr Metabol Disord*. 2003; 3:151–169. [PubMed: 12769787]
- [139]. Onken B, Driscoll M. Metformin induces a dietary restriction-like state and the oxidative stress response to extend *C. elegans* Healthspan via AMPK, LKB1, and SKN-1. *PLoS ONE*. 2010; 5:e8758. [PubMed: 20090912]
- [140]. Anisimov VN. Effect of buformin and diphenylhydantoin on the life span, estrous function and spontaneous tumor incidence in rats. *Vopr Onkol*. 1980; 26:42–48. [PubMed: 7189923]
- [141]. Anisimov VN. Life span extension and cancer risk: myths and reality. *Exp Gerontol*. 2001; 36:1101–1136. [PubMed: 11404054]
- [142]. Anisimov VN, Berstein LM, Popovich IG, Zabezhinski MA, Egormin PA, Tyndyk ML, Anikin IV, Semenchenko AV, Yashin AI. Central and peripheral effects of insulin/IGF-1 signaling in aging and cancer: antidiabetic drugs as geroprotectors and anticarcinogens. *Ann N Y Acad Sci*. 2005; 1057:220–234. [PubMed: 16399897]
- [143]. Anisimov VN, Egormin PA, Bershtein LM, Zabezhinskii MA, Piskunova TS, Popovich IG, Semenchenko AV. Metformin decelerates aging and development of mammary tumors in HER-2/neu transgenic mice. *Bull Exp Biol Med*. 2005; 139:721–723. [PubMed: 16224592]
- [144]. Anisimov VN, Berstein LM, Egormin PA, Piskunova TS, Popovich IG, Zabezhinski MA, Kovalenko IG, Poroshina TE, Semenchenko AV, Provinciali M, Re F, Franceschi C. Effect of metformin on life span and on the development of spontaneous mammary tumors in HER-2/neu transgenic mice. *Exp Gerontol*. 2005; 40:685–693. [PubMed: 16125352]
- [145]. Anisimov VN, Berstein LM, Egormin PA, Piskunova TS, Popovich IG, Zabezhinski MA, Tyndyk ML, Yurova MV, Kovalenko IG, Poroshina TE, Semenchenko AV. Metformin slows down aging and extends life span of female SHR mice. *Cell Cycle*. Sep 1.2008 7:2769–2773. [PubMed: 18728386]
- [146]. Dil'man VM, Anisimov VN. Increase in longevity and a decrease in the frequency of tumors in C3H/Sn mice under the influence of fenformin and diphenin. *Dokl Akad Nauk SSSR*. 1979; 245:753–757. [PubMed: 428319]
- [147]. Dilman VM, Anisimov VN. Effect of treatment with phenformin, diphenylhydantoin or L-dopa on life span and tumour incidence in C3H/Sn mice. *Gerontology*. 1980; 26:241–246. [PubMed: 7390164]
- [148]. Dilman VM, Anisimov VN, Ostroumova MN, Khavinson VK, Morozov VG. Increase in lifespan of rats following polypeptide pineal extract treatment. *Exp Pathol (Jena)*. 1979; 17:539–545. [PubMed: 575333]

- [149]. Ma TC, Buescher JL, Oatis B, Funk JA, Nash AJ, Carrier RL, Hoyt KR. Metformin therapy in a transgenic mouse model of Huntington's disease. *Neurosci Lett*. Jan 10.2007 411:98–103. [PubMed: 17110029]
- [150]. Minor RK, Smith DL Jr, Sossong AM, Kaushik S, Poosala S, Spangler EL, Roth GS, Lane M, Allison DB, de CR, Ingram DK, Mattison JA. Chronic ingestion of 2-deoxy-d-glucose induces cardiac vacuolization and increases mortality in rats. *Toxicol Appl Pharmacol*. Dec 22.2009
- [151]. Oeppen J, Vaupel JW. Demography. Broken limits to life expectancy. *Science*. May 10.2002 296:1029–1031. [PubMed: 12004104]
- [152]. Vaupel JW, Carey JR, Christensen K, Johnson TE, Yashin AI, Holm NV, Iachine IA, Kannisto V, Khazaeli AA, Liedo P, Longo VD, Zeng Y, Manton KG, Curtsinger JW. Biodemographic trajectories of longevity. *Science*. May 8.1998 280:855–860. [PubMed: 9599158]
- [153]. Carnes BA, Olshansky SJ, Grahn D. Biological evidence for limits to the duration of life. *BioGerontology*. 2003; 4:31–45. [PubMed: 12652187]
- [154]. Olshansky SJ, Carnes BA, Desesquelles A. Demography. Prospects for human longevity. *Science*. Feb 23.2001 291:1491–1492. [PubMed: 11234076]
- [155]. Christensen K, McGue M, Petersen I, Jeune B, Vaupel JW. Exceptional longevity does not result in excessive levels of disability. *Proc Natl Acad Sci U S A*. Sep 9.2008 105:13274–13279. [PubMed: 18711139]
- [156]. Christensen K, Doblhammer G, Rau R, Vaupel JW. Ageing populations: the challenges ahead. *Lancet*. Oct 3.2009 374:1196–1208. [PubMed: 19801098]
- [157]. Olshansky SJ, Passaro DJ, Hershow RC, Layden J, Carnes BA, Brody J, Hayflick L, Butler RN, Allison DB, Ludwig DS. A potential decline in life expectancy in the United States in the 21st century. *N Engl J Med*. Mar 17.2005 352:1138–1145. [PubMed: 15784668]
- [158]. Kirkland JL, Peterson C. Healthspan, translation, and new outcomes for animal studies of aging. *J Gerontol A Biol Sci Med Sci*. 2009; 64:209–212. [PubMed: 19196900]
- [159]. Tatar M. Can we develop genetically tractable models to assess healthspan (rather than life span) in animal models? *J Gerontol A Biol Sci Med Sci*. 2009; 64:161–163. [PubMed: 19225031]
- [160]. Sprott RL. Biomarkers of aging and disease: Introduction and definitions. *Exp Gerontol*. Aug 3.2009
- [161]. Butler RN, Warner HR, Williams TF, Austad SN, Brody JA, Campisi J, Cerami A, Cohen G, Cristofalo VJ, Drachman DA, Finch CE, Fridovich I, Harley CB, Havlik RJ, Martin GM, Miller RA, Olshansky SJ, Pereira-Smith OM, Smith JR, Sprott RL, West MD, Wilmoth JR, Wright WE. The aging factor in health and disease: the promise of basic research on aging. *Aging Clin Exp Res*. 2004; 16:104–111. [PubMed: 15195984]

Table I

Abbreviations

AL	<i>Ad libitum</i>
CALERIE	Comprehensive Assessment of Long-Term Effects of Reducing Intake of Energy
CR	Calorie Restriction
CRM	Calorie Restriction Mimetic
2DG	2-Deoxyglucose
ITP	Interventions Testing Program
NIA	National Institute on Aging
RAP	Rapamycin
TOR	Target of Rapamycin

Table II

Interventions Testing Program Compounds

Compound	Concentration	Age of Treatment Initiation	Lifespan Effect Sex – Mean Effect* (p-value)	Description	Reference
Aspirin	20 ppm	4 months	M – 8% (p=0.001) F – ns	Non-steroidal anti-inflammatory, anti-thrombotic, anti-oxidant	[1;2]†
Nitroflurbiprofen (NFP)	200 ppm	4 months	M – ns F – ns	Nitric oxide releasing non-steroidal anti-inflammatory drug	[1;2]†
Nordihydroguaiaretic acid (NDGA)	2,500 ppm	9 months	M – 12% (p=0.006) F – ns	Anti-oxidant, anti-inflammatory, polyphenol	[1;2]†
4-OH- α -phenyl-tert-butylnitron (4-OH-PBN)	315 ppm	4 months	M – ns F – ns	Nitron-based free radical trap	[1;2]†
Caffeic acid phenethyl ester (CAPE)	30 ppm	4 months	M – ns F – ns	Anti-oxidant, anti-inflammatory, anti-tumorigenic	[3]†
Caffeic acid phenethyl ester (CAPE)	300 ppm	4 months	M – ns F – ns	Anti-oxidant, anti-inflammatory, anti-tumorigenic	[3]†
Enalapril Maleate	120 ppm	4 months	M – ns F – ns	Anti-hypertensive agent; Angiotensin converting enzyme inhibitor	[3]†
Rapamycin	14 ppm	20 months	M – 9% (p<0.0001) F – 13% (p<0.0001)	Anti-fungal, anti-cancer, immunosuppressant	[3]†
Rapamycin	14 ppm	9 months	In testing	Anti-fungal, anti-cancer, immunosuppressant	[3]†
Simvastatin	12 ppm	10 months	In testing	Hypolipidemic drug	†
Simvastatin	120 ppm	10 months	In testing	Hypolipidemic drug	†
Resveratrol	300 ppm	12 months	In testing	Phytoalexin, anti-fungal, anti-microbial, anti-cancer, anti-inflammatory, polyphenol	†
Resveratrol	1,200 ppm	12 months	In testing	Phytoalexin, anti-fungal, anti-microbial, anti-cancer, anti-inflammatory, polyphenol	†
Resveratrol	300 ppm	4 months	In testing	Phytoalexin, anti-fungal, anti-microbial, anti-cancer, anti-inflammatory, polyphenol	†
Oxaloacetic acid	2,200 ppm	4 months	In testing	Citric acid cycle metabolite	†
Green Tea Extract	2,000 ppm	4 months	In testing	Anti-oxidant, anti-inflammatory, polyphenol(s),	†
Curcumin	2,000 ppm	4 months	In testing	Anti-tumor, anti-oxidant, anti-inflammatory, polyphenol	†
Medium Chain Triglyceride Oil	60,000 ppm	4 months	In testing	Fat supplement	†
17 α -Estradiol	4.8 ppm	10 months	In testing	Neuroprotective, mitochondrial protective, muscle relaxant	†
Methylene Blue	28 ppm	4 months	In testing	Chemical dye, diverse biological activities	†
Acarbose	1,000 ppm	4 months	In testing	Glycoside hydrolase inhibitor, anti-diabetic	†

Compound	Concentration	Age of Treatment Initiation	Lifespan Effect Sex – Mean Effect* (p-value)	Description	Reference
Rapamycin LoPhase II	4.7 ppm	9 months	<i>In testing</i>	Anti-fungal, anti-cancer, immunosuppressant	[†]
Rapamycin MidPhase II	14 ppm	9 months	<i>In testing</i>	Anti-fungal, anti-cancer, immunosuppressant	[†]
Rapamycin HiPhase II	42 ppm	9 months	<i>In testing</i>	Anti-fungal, anti-cancer, immunosuppressant	[†]

M=Male, F=Female; *ns* – not significant

* Log-rank test for lifespan effects; significance set at $p < 0.05$ [1–3]

[†] <http://www.nia.nih.gov/ResearchInformation/ScientificResources/CompoundsInTesting.htm>