

Visual discrimination between two sexually deceptive *Ophrys* species by a bee pollinator

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Abstract Almost all species of the orchid genus *Ophrys* are pollinated by sexual deception. The orchids mimic the sex pheromone of receptive female insects, mainly hymenopterans, in order to attract males seeking to copulate. Most *Ophrys* species have achromatic flowers, but some exhibit a coloured perianth and a bright, conspicuous labellum pattern. We recently showed that the pink perianth of *Ophrys heldreichii* flowers increases detectability by its pollinator, males of the long-horned bee *Eucera berlandi*. Here we tested the hypothesis that the bright, complex labellum pattern mimics the female of the pollinator to increase attractiveness toward males. In a dual-choice test we offered *E. berlandi* males an *O. heldreichii* flower and a flower from *O. dictynnae*, which also exhibits a pinkish perianth but no conspicuous labellum pattern. Both flowers were housed in UV-transmitting acrylic glass boxes to exclude olfactory signals. Males significantly preferred *O. heldreichii* to *O. dictynnae* flowers. In a second experiment, we replaced the perianth of both flowers with identical artificial perianths made from pink card, so that only the labellum differed between the two flower stimuli. Males then chose between both stimuli at random, suggesting that the presence of a labellum pattern does not affect their choice. Spectral measurements revealed higher colour contrast with the background of the perianth of

O. heldreichii compared to *O. dictynnae*, but no difference in green receptor-specific contrast or brightness. Our results show that male choice is guided by the chromatic contrast of the perianth during the initial flower approach but is not affected by the presence of a labellum pattern. Instead, we hypothesise that the labellum pattern is involved in aversive learning during post-copulatory behaviour and used by the orchid as a strategy to increase outcrossing.

Keywords *Ophrys heldreichii* · Colour vision · Sexual deception · Signal evolution · Object detection · Pollination

Introduction

Long before the discovery of sexual deception as pollination syndrome, some authors had already recognised the apparent visual resemblance between flowers of the orchid genus *Ophrys* and insect bodies (Linnaeus 1745; Darwin 1862). One early hypothesis, for instance, suggested that the orchids mimic a feeding insect to advertise a potential food source to conspecifics of the species being imitated (Darwin 1862 and citations therein). From the 1920s on, when Pouyanne discovered that the orchids in fact achieve pollination by means of sexual deception of male insects, the insect-like appearance of the labellum was regarded as imitating various female signals, such as the body shape, wings, or abdominal markings (Correvon and Pouyanne 1916; Kullenberg 1961; Paulus and Gack 1990; Paulus 2006, 2007).

Nearly 100 years later, pollination by sexual deception has been described from all continents except Antarctica. The syndrome has so far only been discovered in the orchid family (Orchidaceae) and has evolved several times

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independently (van der Pijl and Dodson 1966; Nilsson 1992). The orchids mimic species-specific mating signals in order to attract male insects, especially bees and other hymenopterans, and elicit courtship behaviour. Whilst attempting to copulate with the flower's labellum, the males come into contact with the sticky viscidia and the pollinaria become attached to their bodies. Should the male subsequently be deceived by another orchid, this pollen is transferred to the stigma of the next flower and pollination occurs (Kullenberg 1961; Paulus and Gack 1990; Cioteck et al. 2006).

Attraction of the target-male can only be successful if the orchid mimics the key signals deployed in insect mating communication. In hymenopterans, the major communication channel is olfaction. Virgin females produce a sex pheromone that usually consists of a blend of semiochemicals that signals mating readiness (Ayasse et al. 2001; Paxton 2005). The olfactory communication is highly specific and thus constitutes a private communication channel, i.e. only males of the target species can decipher the message sent by the female (Endler 1993; Ayasse et al. 2001; Schaefer et al. 2004). The chemical profile of the flower closely resembles the behaviourally active substances of the model insect; as a consequence, pollinator attraction in sexually deceptive orchids is highly specific (Schiestl et al. 1999; Ayasse et al. 2000; Schiestl 2005; Mant et al. 2005).

The Mediterranean genus *Ophrys* comprises about 260 species of which almost all are sexually deceptive and usually attract males of only one species (Paulus and Gack 1990; Ayasse et al. 2000). Due to the highly specific olfactory attraction, visual signals are assumed to play only a minor role (Kullenberg 1961; Kullenberg and Bergström 1976). Most *Ophrys* flowers are indeed dull (Delforge 2006), probably to avoid visual attraction of unspecific visitors and thus the risk of pollen loss. However, some *Ophrys* species exhibit brightly-coloured sepals and petals besides the labellum (henceforth referred to as perianth for simplicity). Recent investigations have shown that such potential visual signals can increase detectability and attractiveness to the pollinator on the one hand, but may also impose additional costs (Spaethe et al. 2007, 2010; Streinzer et al. 2009). Moreover, some *Ophrys* species show a bright and conspicuous pattern on the labellum which has been hypothesised to mimic traits of the pollinator's female, such as the wings or body-markings to make the flower more attractive for the males (Kullenberg 1961; Paulus 2006, 2007). However, the labellum pattern is relatively small and the compound eyes of hymenopterans allow only a coarse spatial resolution (Land 1997) and thus it is not clear whether males can indeed perceive these patterns and mistake them for female traits.

Here we tested the hypothesis that the conspicuous labellum pattern mimics behaviourally-relevant parts of the

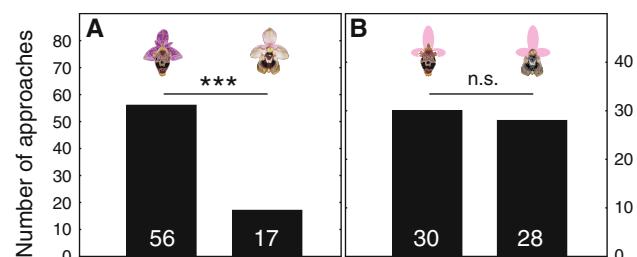


Fig. 1 Male choices for **a** intact flowers of *Ophrys heldreichii* (left column) and *O. dictynnae* (right column) (73 approaches in total), and for **b** flowers where the original perianth was replaced by an artificial one made from pink card (58 approaches). Number of approaches are given within the column. *** $P < 0.001$, n.s., $P > 0.05$; χ^2 -test

female's body and thus increase attractiveness to males. We conducted dual-choice experiments with two *Ophrys* species that both are pollinated by long-horned bees. *Ophrys heldreichii*, which is pollinated by *Eucera berlandi*, possesses a bright and complex labellum pattern, while *O. dictynnae* lacks such a conspicuous pattern (Figs. 1, 2). Both species have a pink perianth, although they differ slightly with respect to the colour for a human observer. We presented both flower types in UV-transmitting acrylic glass boxes to the pollinator of *O. heldreichii*, *Eucera berlandi*. If the proposed hypothesis is correct, we expect that males prefer flowers with the conspicuous labellum pattern over flowers without such a pattern.

Materials and methods

Study sites and species

Experiments were carried out on Crete, about 5 km northeast of Neapolis ($N 35^{\circ}15'13''$, $E 25^{\circ}38'03''$) with Heldreich's bee orchid, *Ophrys heldreichii* SCHLECHTER (Orchidaceae), which is member of the *O. oestrifera* group, and its specific pollinator, males of the long-horned bee *Eucera (Synhalonia) berlandi* DUSMET (Apoidea, Apidae, Eucerini). As second orchid species, *Ophrys dictynnae* DELFORGE, a recently described representative of the *O. tenthredinifera* group, was chosen for the discrimination experiments. This orchid differs from *O. heldreichii* in the absence of a conspicuous labellum pattern and in perianth colour, which appears slightly brighter to a human observer (Figs. 1, 2). *O. dictynnae* is also pollinated by males of a long-horned bee species, *Eucera nigrita* FRIESE (Streinzer and Paulus, unpublished). For the experiments, intact flowers were collected from various sites within an area of about 40 km around the experimental site. *O. heldreichii* and *O. dictynnae* do not occur at the experimental site and thus all tested *E. berlandi* males were naïve to both species.

Behavioural experiments

To test whether the males discriminate between the two orchid species we conducted a dual-choice experiment. Two flowers, one of each species, were presented simultaneously to individual males of *E. berlandii* and choices were recorded. To eliminate odour-induced bias the flowers were presented in UV-transmitting acrylic glass boxes at a height of approx. 30 cm and 10 cm apart from each other (Spaethe et al. 2007). Since visual stimuli alone are not sufficient to attract pollinators, we placed 20–25 flowers of *O. heldreichii* in a 2-l acrylic jar. Air was channelled through the jar by means of an aquarium pump (SCHEGO optimal, Offenbach am Main, Germany) at a rate of 250 l/h. The odour-saturated outflow was then distributed equally to both acrylic glass boxes via a Y-tube (Spaethe et al. 2007). The flowers were presented in front of a piece of green leaf of a sea onion (*Urginea maritima*).

In the first experiment we presented original flowers of both species to test whether males discriminate between them. In a second experiment, to separate the role of the perianth and the labellum, we removed the original sepals and petals in both flowers and replaced them with an identical artificial perianth cut from coloured card that approximately resembled the original perianth of *O. heldreichii* in colour, size and shape (Fig. 3, Streinzer et al. 2009). In this test, the only difference between both test flowers was the labellum.

All approaches of individual males that resulted in contact with one of the acrylic glass boxes were counted as a decision. Males were then immediately caught and marked with a coloured marker on the thorax or abdomen. Repeat visits by individual males were excluded from the analysis. The position of the flowers was interchanged after every fifth visit to exclude any bias due to side. Moreover, the orchid flowers were replaced every couple of hours from a random sample of flowers collected from different populations (see below).

Flower morphology

Flowers from both species differ slightly in shape and size. To account for size effects on choice behaviour we measured sepal and labellum surface of *O. heldreichii* and *O. dictynnae* flowers. Specimens were collected from the same populations where we took the flowers for the behavioural experiments (one population of *O. heldreichii*, two of *O. dictynnae*). We randomly chose one flower from each plant ($N_{O. heldreichii} = 15$, $N_{O. dictynnae} = 27$) for morphological and spectral measurements.

The sepals were separated from the labellum and all parts were transferred onto a piece of adhesive tape together with a size reference, and photographed with a digital

camera (Nikon D70s, AF 105 mm/2,8 D; Nikon, Japan). Pictures were subsequently transferred to a personal computer and analyzed with ImageJ (<http://rsbweb.nih.gov/ij/>). We measured the total area of the three sepals and of the labellum (without the stylum).

Spectral measurements and bee colour space calculations

Spectral reflectance measurements of the perianth were carried out on the same flowers as for morphological measurements. Spectra were obtained by means of a USB 2000 spectrometer with a balanced deuterium/halogen light source (Ocean Optics B.V., Duiven, The Netherlands) between 300 and 700 nm on an approx. 0.25 cm² area of the median sepal. The spectrometer was calibrated with a white PTFE standard (Diffuse Reflectance Standard, WS-1, Ocean Optics). Spectral data were processed in Microsoft Excel 2002.

To estimate the bee specific receptor contrasts and perceptual colour distance between the perianth and the stimulus background we applied the bee colour hexagon (Chittka 1992). Since the receptor sensitivities of *E. berlandii* are unknown, we used the spectral sensitivity functions of *Apis mellifera* (Peitsch et al. 1992). This approach seems appropriate since most bees have been found to share similar receptor properties with respect to number and shape of the sensitivity curves (Peitsch et al. 1992). Colour loci and receptor-specific excitations were calculated using standard procedures (Spaethe et al. 2001; Chittka and Kevan 2005). It is assumed that photoreceptors adapt, depending on the ambient illumination, to the background (Chittka 1992). We therefore chose an averaged reflection spectrum of several leaves from different flowers which are abundant in the experimental area (including *Urginea maritima*) as adaptation background (Fig. 3a). Colour distance between colour loci of the flower perianths and the stimulus background were calculated as the Euclidean distance in the colour hexagon. Green receptor-specific contrast was calculated as the absolute value of the difference between the calculated excitation of the long-wavelength receptor by the background and the flower. Brightness was calculated as the sum of all three receptor excitations (Spaethe et al. 2001).

Statistics

All statistical analyses were performed using SPSS 11.5 (SPSS Inc., Chicago, IL). To test if behavioural responses of the males differ significantly from chance a χ^2 -test was applied. For morphological measurements and colour contrast we first validated whether the two populations of *O. dictynnae* differ from each other. All measured

parameters were found to be not significantly different and thus data from both populations were pooled (data not shown). To test whether spectral and morphological parameters differ between the two orchid species we applied a Mann–Whitney *U*-test. All *P*-values above 0.05 were considered as statistically non-significant.

Results

Behavioural experiments

When males of the long-horned bee *E. berlandi* were allowed to choose between intact flowers of *O. heldreichii* and *O. dictynnae*, they significantly preferred *O. heldreichii* over *O. dictynnae* ($\chi^2 = 20.84$, $P < 0.001$, $N = 73$; Fig. 1a). Since the olfactory signal was the same for both stimuli, the result clearly shows that males use visual cues or signals to discriminate between the two species of orchids. To test whether the preference of *O. heldreichii* flowers is due to differences of the perianth and/or the labellum, we replaced the original perianth with an artificial one, which was identical for both flower stimuli. We found that when only the labellum differs, males do not show a preference for either of the two flower types ($\chi^2 = 0.07$, $P = 0.79$, $N = 58$; Fig. 1b).

Flower morphology measurements

Perianth surface did not differ between both orchid species ($Z = 0.25$, $P = 0.80$; $N_{O. heldreichii} = 15$, $N_{O. dictynnae} = 27$; Fig. 2a). When we compared the labellae, *O. dictynnae* showed a significantly larger labellum than *O. heldreichii* ($Z = -4.19$, $P < 0.001$; $N_{O. heldreichii} = 15$, $N_{O. dictynnae} = 27$; Fig. 2b). However, the behavioural data indicate that

neither labellum size nor the presence/absence of a labellum pattern affect male choice.

Spectral measurements

The spectral reflectance curves of the *O. heldreichii* and *O. dictynnae* sepals show a different shape (Fig. 3a–c). To examine whether the two species also differ in their appearance to the bee's visual system we plotted the perianth colours in a bee specific colour space (Chittka 1992; Fig. 3d; Table 1). The colour loci of the two populations of *O. dictynnae* were found to overlap and were therefore pooled for further analysis (data not shown). *O. heldreichii* flowers exhibit a higher colour contrast to the background compared to *O. dictynnae* ($Z = 4.55$, $P < 0.001$). Beside the difference in colour contrast, the two species also differ in hue, i.e. the angle they subtend from the centre ($\alpha_{heldreichii} = 100.6 \pm 10.9^\circ$; $\alpha_{dictynnae} = 124.8 \pm 22.2^\circ$; Mean \pm SD; Fig. 3d; $Z = -3.54$, $P < 0.001$; where 0° corresponds to the vector between centre and the UV corner of the colour hexagon). Green receptor contrast, a sensory channel used by bees to detect objects at small angular sizes, and total brightness did not differ between the two species ($Z = -0.46$, $P = 0.65$ and $Z = 1.38$, $P = 0.17$, respectively; Table 1).

Discussion

In this study we tested whether *E. berlandi* males use visual cues to discriminate between flowers of two species of the sexually deceptive orchid genus *Ophrys*. We chose two species that differ both in the appearance of the labellum and also the colour of the perianth. In dual-choice experiments we found that males are clearly able to discriminate between both species visually. However, when we replaced the coloured perianth with an artificial one, males chose both flower types at random. From these results we can conclude that (1) the labellum pattern of *O. heldreichii* is not involved in male attraction and that (2) when males can choose between flowers they prefer flowers with a perianth with the higher chromatic contrast.

Various types of labellum patterns are present in the genus *Ophrys* and were assumed to mimic female traits to increase attractiveness to the males (Paulus 1988, 2006, 2007). However, whether the males indeed mistake these patterns as females or at least key parts of a female's body, has not yet been shown. *O. heldreichii* exhibits a whitish, highly contrasting pattern on a dark labellum, which is also found of similar type in all species of the *Ophrys oestrifera* group (Delforge 2006). At a first glance the labellum pattern shows no similarity to the female of the pollinator species, which is reddish–brownish and without any conspicuous

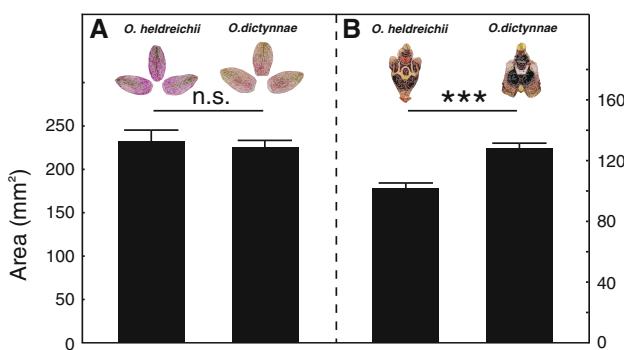


Fig. 2 Flower morphology of the perianth (a) and the labellum (b) of *O. heldreichii* (left column) and *O. dictynnae* (right column). Perianth size does not differ between the two species. However, *O. dictynnae* has a significantly larger labellum compared to *O. heldreichii*. $N_{O. heldreichii} = 15$, $N_{O. dictynnae} = 27$; means \pm SEM; *** $P < 0.001$, n.s., $P > 0.05$; Mann–Whitney *U*-test

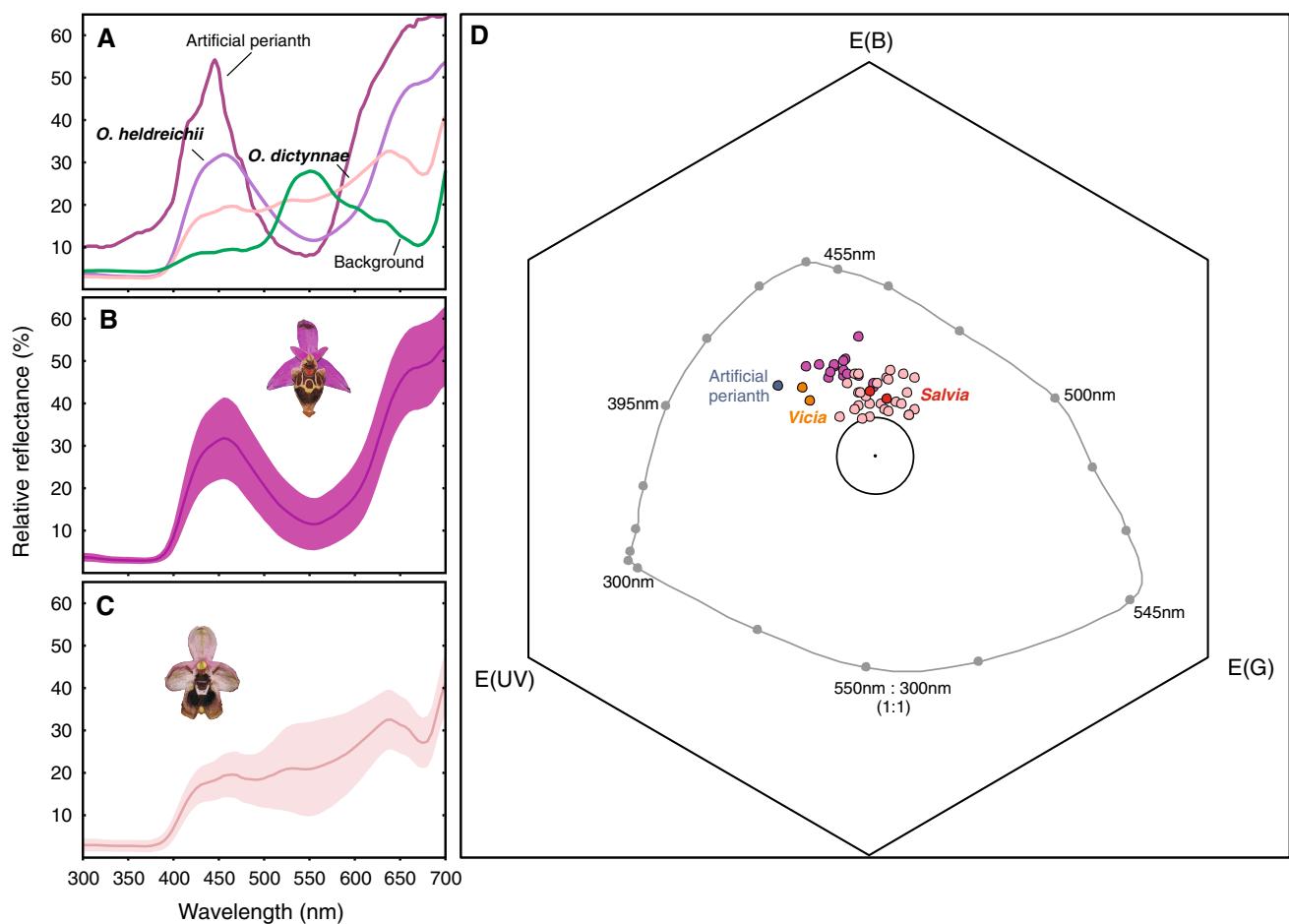


Fig. 3 **a–c** Spectral reflectance curves of the perianth of *Ophrys heldreichii* (dark violet), *Ophrys dictynnae* (pale pink), the artificial perianth (dark pink) and the background (green) as a function of wavelength. In **a** only mean reflectance of the flower perianth are given ($N_{O. heldreichii} = 15$, $N_{O. dictynnae} = 27$, $N_{\text{Background}} = 5$, see text). **b** and **c** show the mean reflectance of *O. heldreichii* and *O. dictynnae*, respectively, presented with the standard deviation of the mean to visualize the variation within the species. **d** The colour loci of *Ophrys heldreichii* (dark violet), *O. dictynnae* (light pink), the

artificial perianth and two major food plants (*Vicia cracca* and *Salvia fructicosa*) represented in the colour hexagon model (Chittka 1992). The large circle denotes 0.1 hexagon units around the centre. All loci within this circle are assumed to appear achromatic to the bee (Chittka and Kevan 2005). The grey line indicates the loci of pure spectral lights at background intensity. Colour distances in the hexagon space are calculated as the Euclidean distance between two loci (Chittka 1992)

Table 1 Chromatic contrast, green contrast and brightness in relation to the background

| | <i>Ophrys heldreichii</i> ($n = 15$) | <i>Ophrys dictynnae</i> ($n = 27$) | Mann–Whitney <i>U</i> -test |
|------------------------------------|--|--------------------------------------|-----------------------------|
| Chromatic contrast (Hexagon units) | 0.23 ± 0.03 | 0.16 ± 0.04 | $P < 0.001$ |
| Green contrast (dimensionless) | 0.09 ± 0.05 | 0.09 ± 0.05 | NS |
| Brightness (dimensionless) | 1.74 ± 0.21 | 1.62 ± 0.25 | NS |

The values represent means \pm standard deviations. Values were calculated using the colour hexagon model (Chittka 1992). For the calculation we used the spectral sensitivity curves of *Apis mellifera* (Peitsch et al. 1992). Note that for the green-receptor contrast the absolute values are given (Spaethe et al. 2001)

bright patterns on the body or wings. Our results show that the presence of such a pattern does not affect a male's initial flower choice, and thus it is most likely that the pattern does not mimic any female traits that increase attractivity to the

males. What then is the functional significance of the labellum pattern? We speculate that it may be involved in aversive learning during post-copulating behaviour, that is, the male associates the unsuccessful copulation attempt

with the labellum pattern and avoids further visits to flowers with an identical pattern (see Paulus 1988). Two observations support this hypothesis. First, males of *E. berlandi* do not fly away immediately after pseudocopulation but hover a few centimetres in front of the flower for periods of several seconds up to a minute (personal observation). Although the compound eyes limit the spatial resolution of diminutive and fine patterns, at this short distance the males might be able to resolve the labellum pattern (Lunau et al. 2009). Recent investigations have shown that bees are capable of learning and discriminating even very complex and fine-scale natural scenes (Dyer et al. 2008b). Second, pattern size and shape are found to be highly variable among flowers of a population, but very similar among flowers within a plant (Paulus 2007, personal observation). The high variation of this floral trait among *O. heldreichii* individuals might facilitate increased outcrossing, since, if the males are indeed able to learn the pattern, they would avoid other flowers with similar patterns to ones they had recently visited, but would be more willing to visit flowers from another plant with a different pattern. However, this hypothesis requires confirmation from behavioural studies showing that males are able to learn and discriminate patterns from different plants, and that males avoid flowers with patterns similar to those that have been encountered during a previous pseudocopulation.

Although we could clearly show that *E. berlandi* males are not attracted by the labellum pattern of *O. heldreichii* flowers in their initial flower choice, our results cannot simply be generalised for all *Ophrys* species. The labellum of *O. speculum*, for instance, possesses a conspicuous shiny blue mirror that also reflects in the ultraviolet (UV). In dual-choice tests, males of the sphecid wasp *Dasyscolia ciliata*, the pollinator of *O. speculum*, preferred flowers with an unmanipulated labellum to flowers whose mirror had been manipulated to reduce the UV reflectance (Paulus 2007, personal observations). However, for most *Ophrys* species with labellum patterns the functional significance of such markings in flower-pollinator communication is still unknown.

The only visual trait we could find that affects male choice behaviour was the colour of the perianth. The perianths of *O. heldreichii* flowers have a higher colour contrast (but identical green-receptor and brightness contrasts) and are more attractive compared to *O. dictynnae* (but see Vereecken and Schiestl 2009). In a recent study we found that the detection of *O. heldreichii* flowers by *E. berlandi* from a distance of approximately 30 cm was improved when the green contrast between the perianth and the background increased, but colour contrast was found to have no effect (Streinzer et al. 2009). However, in the present study, males could choose between the two flower stimuli within a range of a few centimetres which most

likely enabled them to use the chromatic channel that only works at large visual angles (Giurfa et al. 1996; Dyer et al. 2008a). Flower choice experiments with naïve bumblebees revealed a preference for colours providing a large colour contrast to the background and might reflect a more general preference in the flower choice behaviour of bees (Lunau 1990; Lunau et al. 1996). Besides colour contrast, the spectral purity of a target (i.e. saturation) may also affect colour choice (Lunau et al. 1996). Our experimental setup, however, does not allow to disentangle whether the choice behaviour of *E. berlandi* males is primarily based on the colour contrast between the perianth and the background or the saturation of the perianth or both. Alternatively, males might prefer the hue of the perianth of *O. heldreichii* flowers compared to that of *O. dictynnae* because it more closely resembles the flower colour of the females' most frequent host plants (e.g. *Vicia cracca* and *Salvia fruticosa*, see Spaethe et al. 2007 for spectral reflectance curves). However, the colour loci of the food plants overlap at least partially with the loci of both orchids, *O. heldreichii* and *O. dictynnae*, respectively (Fig. 3d). Additional behavioural experiments are necessary to disentangle the role of colour contrast, saturation and hue in male attraction.

Pollinator visitation rate is a critical factor determining reproductive success in *Ophrys* (Vandewoestijne et al. 2009). Thus, it is surprising that selection did not lead to a perianth colour that provides higher colour contrast (and is thus more attractive) in *O. dictynnae*, which is also pollinated by a long-horned bee. We suggest two non-mutually exclusive hypotheses which may account for this discrepancy. First, males of the two pollinator species, *E. berlandi* and *E. nigrita*, may exhibit preferences for different colour hues which resulted in a separation of the perianth colours of the two *Ophrys* species by natural selection. We observed males and females of *E. berlandi* foraging on various food plants at the experimental site (see above). However, we neither know the colour of the major host plants of *E. nigrita*, nor if the flowers visited by the two pollinator-species cover different areas in the bees' colour space. Second, differences in colour contrast (and also hue) between the two *Ophrys* species are caused by differences in the expression of floral pigments in the perianth. Anthocyanins are probably the major pigment in the pink perianths of *Ophrys* flowers (Arditti and Fisch 1974; Uphoff 1979), and an increased anthocyanin concentration may lead to a higher colour contrast. However, the production of flower pigments most probably imposes costs on the plant and *O. dictynnae* might be more limited in resources that can be allocated to the anthocyanin production compared to *O. heldreichii*. Again, further investigations are necessary to fully understand the ultimate and proximate mechanisms accounting for differences in perianth colour among *Ophrys*.

In conclusion, the conspicuous labellum pattern of *O. heldreichii* flowers does not increase pollinator attraction during the initial flower approach flight. Instead, we suggest that the pattern is involved in aversive learning by the males during post-copulatory behaviour and constitute a strategy of the orchid to increase outcrossing. Labellum patterns are commonly found within the *Ophrys* genus but differ largely in size, colour and complexity, and thus most likely in their function for flower-pollinator interaction. Further behavioural investigations could provide insights into whether a particular pattern-type is associated with a specific pollinator group, or mimic a particular female trait as previously suggested.

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