# Hexamoll DINCH plasticised PVC containers for the storage of platelets

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#### **Abstract:**

Introduction: Containers for the storage of platelets are made using polyvinyl chloride plasticised with di, (2-ethyl hexyl) phthalate, n-butyryl, tri (n-hexyl) citrate and tri (2-ethyl hexyl) mellitate or using special poly olefins without plasticiser. Of these, the first two have disadvantages such as plasticiser leaching and impairment of platelet function. Polyolefin bags cannot be HF welded or steam sterilized. Mellitate plasticised bags can store platelets well for five days but they are not completely phthalate free. Research and Development: We have developed a new generation of containers made of PVC plasticised with the non DEHP, non aromatic plasticiser,1,2- Cyclohexanedicarboxylic acid, diisononyl ester (Hexamoll DINCH) which can store platelets without loss of function for at least six days. Observation: The present studies show that DINCH plasticised PVC bags (TPL-167) are well suited for the storage of platelet concentrates for more than five days. Conclusion: The present studies show that the PVC plasticised with the non phthalate, non aromatic, non toxic plasticiser DINCH is a viable alternative to other existing containers for the storage of platelets for more than five days.

#### Key words:

Hexamoll DINCH plasticiser, Non DEHP plasticizer, platelets, platelet storage bags

#### Introduction

Platelet concentrates have a very important role in ensuring adequate hemostasis under various clinical situations and have made possible notable advances in the medical and surgical fields. In consequence, the demand for uncontaminated viable platelets, preferably with improved storage periods is rising rapidly.

Platelets get 85% of their energy requirements by aerobic metabolism in which glucose undergoes glycolysis followed by oxidative phosphorylation of the products. Substrates such as free fatty acids and amino acids are also involved in the process. The residual 15% of the energy requirements are met by anaerobic glycolysis in which glucose is converted to lactate. The conversion of glucose to ATP by the oxidative mechanism is 18 times more efficient than anaerobic glycolysis. The carbon dioxide produced during the oxidative pathway gets converted to bicarbonate which acts as the buffer system of plasma. The containers in which platelets are stored should be carefully crafted to maintain the oxidative metabolism of the platelets. This can be achieved only if the permeability to oxygen of the containers is high. At the same time, the permeability to carbon dioxide should be high enough to permit a good part of the carbon dioxide to diffuse out but it should not be very high which would cause too much of the carbon dioxide formed to leave the container, thereby compromising the production of the bicarbonate buffer.

The first generation containers for storing RBC and platelets were made of PVC plasticised with DEHP {di-(2- ethyl hexyl) phthalate}. DEHP continues to be the plasticiser of choice for blood bags for the storage of RBCs particularly because the DEHP leached into blood plasma has a distinct protective effect on the RBC membrane which enables storage of RBC concentrates for up to 42 days. Several studies have, however, shown that leached DEHP has deleterious effect particularly for newborns, very young children and patients who require frequent blood transfusions. Hence, there is a strong move to replace DEHP with other plasticisers. DEHPplasticised PVC containers have comparatively low permeability to oxygen and carbon dioxide and this restricts the storage period for platelets. [1-3] The leached DEHP also causes reduced aggregation responses of platelets.<sup>[4]</sup> The second generation bags overcame the permeability problem by using thinner sheets of PVC plasticised with DEHP [Teroflex XT-612 (Terumo)], and by using the plasticiser TOTM (CLX, Cutter), PL-1240 (Baxter). M/s Baxter also developed special polyolefin bags without plasticiser (PL-732). Platelet concentrates could be stored in such bags for up to five days with better preservation of function and viability.[5-11]

PVC plasticised with nbutyryl tri-(n-hexyl) citrate (BTHC) was introduced in 1989. Such bags have been shown to be acceptable for the storage of platelet concentrates and RBC. [12-15] M/s Terumo Corporation introduced a new PVC bag which was plasticised with di-(ndecyl) phthalate [16] which had very low leachability into blood plasma and was suitable for



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the five day storage of platelet concentrates.

The platelet storage bags in present day use have shortcomings such as phthalate contamination, compromised platelet aggregation, unpleasant odour and allergic reactions.

A new generation of PVC-based platelet storage bag [TPL 167] was developed in 2006 by M/s Terumo Penpol Ltd<sup>[17,18]</sup> in which the non DEHP, non aromatic plasticiser 1,2-cyclohexane-di carboxylic acid, diisononylester subsequently referred to as DINCH, was used. These bags were comparable to TPL's standard TEHTM plasticised platelet storage bag (TPL-157) and other well-known platelet storage bags plasticised with TEHTM.

A second comparative assessment of TPL's DINCH and TEHTM plasticised platelet storage bags was done at the Apollo Hospital, Chennai.

The results of these studies are presented in this paper:

## Materials and Methods

The platelet storage bags used for the study were as follows.

#### Bags evaluated

Details	Plasticiser used
TPL – 157	TEHTM
TPL - 167	DINCH

These bags were part of top and bottom quadruple bag systems consisting of:

- Main bag 450 ml
- SAGM bag 400 ml containing 100 ml SAGM solution attached to the bottom of the main bag.
- Platelet storage bag 400 ml (xperimental bag)
- Transfer bag 400 ml.

## Characteristics of test bags

The characteristics of the material of the test bags are given in Table 1. The bags were also tested for physical, chemical and biological requirements for conformance with the ISO 3826 standard for blood bags.

## Blood collection and component separation

Blood of the same blood group was collected from four volunteer

Table 1: Characteristics of sheets used in this study

Table II ellaracteriotics of circuit		
Property	TPL 157	TPL 167
	TOTM	DINCH
Average thickness (mm)	0.4	0.4
Hardness – (shore A)	78.5	78
ASTM D 2240		
Tensile strength (Kg/cm²)	170	155
ASTM D 882		
Elongation at break (%)	500	480
ASTM D 882		
Permeability		
Oxygen (g/m²/24h/37°C)	1020	1070
Carbon dioxide (g/m²/24hr/37°C)	4850	4600
ASTM D 1434		
Brittle point (°C)	-36	-38
ASTM D7028		

donors in SB 450 bags containing CPD and was kept without disturbance at room temperature for 1 h. The blood was pooled in a 2L capacity Terumo pooling bag using Terumo's sterile tube sealing device (TSCD). The blood was mixed by gentle tilting. 450 ml lots of the mixed blood were then transferred into the main bags of the two QB blood bag systems (as described earlier) using TSCD. Separation of the components was done by the Buffy coat method. After a holding time of 1 h, the bags were centrifuged at 3300 rpm for 13 min at 20°C in a Hitachi Rotosilenter 630 RS centrifuge.

Three layers separated as follows:

- · Top layer of platelet poor plasma
- · Middle layer of Buffy coat
- RBC concentrate

#### PC preparation

The RBC concentrate was transferred to the bottom bag containing 100 ml of SAGM solution and the plasma was transferred to the corresponding plasma bag. These bags were separated after sealing off the connecting tube. The Buffy coat rich in platelets remained in the main bag. It was kept suspended for 24 h at 20 - 24°C. This bag was given a soft spin at 600 rpm for 5 min at 20°C and the separated platelet concentrate was transferred to the platelet bag under test. The platelet volume was adjusted to 70 ml by the addition of plasma. The platelet bags were stored in a platelet agitator with horizontal shaking at 22°C  $\pm$  2°C. PC samples were drawn on zero, one, three, five and seven days after separation to components and the holding time of 24 h.

#### Parameters studied

- RBC count, leucocyte count and platelet count
   These were measured using the automatic blood cell counter
   Beckman LH 750- 5 part differential cell counter.
- pH, pO<sub>2</sub>, pCO<sub>2</sub>, HCO<sub>3</sub>
   These were measured using blood gas analyzer model Bayer, Rapid Lab 248.
  - Lactate
  - Measured using Hitachi 911 UV spectrophotometer.
- . Glucose, plasma Na $^{\scriptscriptstyle +}$  and K $^{\scriptscriptstyle +}$  Measured using L X 20 procedure in an automatic analyzer.
- Aggregation

PC in which the number of platelets was adjusted to 2.5 lakhs/µl with fresh frozen autologous plasma was measured at the maximum aggregation induced by collagen at a concentration of  $16\,\mu\text{g/ml}$  (8 µl per test) and ADP at a concentration of  $80\,\mu\text{mol/ml}$  (40µl per test). Chrono -Log platelet aggregometer was used for the measurements.

## Results

Results of platelet storage evaluation studies conducted at Apollo Hospital, Chennai are given below.

# Hematological studies

The results of hematological studies are shown in Table 2. The platelet counts per experimental bag were in the range  $5.1 \text{ to } 9.2 \times 10^{10}$ .

#### **Biochemical studies**

The results of biochemical studies are shown in Table 3.

#### pH, pO<sub>2</sub>, pCO<sub>3</sub>

pH was above 7.0 in all cases. The partial pressure of oxygen

Table 2: Hematological studies

Study parameter	Sample	Test days				
		1	2	3	5	7
Platelet count	TOTM					
(x10³/µl)	TPL 157-a	1063	1167	1128	1073	991
	TPL 157-b	785	881	847	786	889
	TPL 157-c	728	805	690	849	889
	TPL 157-d	863	891	853	909	775
	DINCH					
	TPL 167-a	1154	1107	1248	1052	1062
	TPL 167-b	1034	876	870	894	910
	TPL 167-c	1072	1152	1078	1150	1112
	TPL 167-d	1162	1150	1184	1140	1098
NBC count	TOTM					
x10³/µl)	TPL 157-a	0.3	0.2	0.3	0.3	0.5
• ,	TPL 157-b	0.2	0.3	0.4	0.2	0.5
	TPL 157-c	0.3	0.3	0.4	0.3	0.3
	TPL 157-d	0.3	0.2	0.3	0.3	0.3
	DINCH					
	TPL 167-a	0.5	0.5	0.5	0.4	0.4
	TPL 167-b	0.2	0.1	0.2	0.2	0.2
	TPL 167-c	0.2	0.2	0.2	0.1	0.1
	TPL 167-d	0.8	0.8	0.8	0.8	0.8
RBC count	TOTM					
x10 <sup>6</sup> /µl)	TPL 157-a	0.01	0.01	0.01	0.01	0.02
• ,	TPL 157-b	0.01	0.01	0.01	0.01	0.02
	TPL 157-c	0.01	0.01	0.01	0.01	0.02
	TPL 157-d	0.01	0.01	0.01	0.01	0.02
	DINCH					
	TPL 167-a	0.02	0.02	0.02	0.02	0.02
	TPL 167-b	0.01	0.00	0.00	0.01	0.01
	TPL 167-c	0.02	0.02	0.02	0.02	0.01
	TPL 167-d	0.04	0.04	0.04	0.04	0.04

increased throughout preservation, while the partial pressure of carbon dioxide showed a gradual reduction. No significant differences were observed for both groups with regard to these parameters. It is clear that an oxidative atmosphere was maintained during the period of storage in both cases.

#### Glucose and lactic acid

The utilization of glucose during storage was similar in samples TPL - 157 and TPL -167.

The lactate level was comparable up to the fifth day above which increase was more marked for TPL - 157.

#### Bi carbonate

Decrease in bicarbonate is similar for the two samples studied.

#### Plasma K+, Na+

Plasma  $K^{\scriptscriptstyle +}$  and Plasma Na $^{\scriptscriptstyle +}$  remained fairly stable throughout the preservation.

#### LDH

LDH showed a slightly increasing pattern for both the samples.

The results are similar up to five days. Beyond five days, the DINCH plasticised bags appeared to be marginally better.

## Aggregation studies

The aggregation obtained for the various samples with ADP and collagen are given in Table 4. The results show that the aggregation was maintained well for more than five days.

# **Discussions**

The pH of plasma within both types of bags remained above 7.0 indicating good storage conditions. The pattern of  $\rm pO_2$  change indicates adequate oxygenation of the containers. The  $\rm HCO_3$  level is indicative of the presence of adequate buffer and stability consequent on the maintenance of optimum level of  $\rm pO_2$  within the containers. The lactate production was within limits and comparable for the DINCH and TEHTM plasticised bags up to the fifth day, beyond which it increased significantly for the TEHTM bags. Platelet aggregation, which is an indication of platelet function, was reasonably well maintained for more than five days. It may be noted that the Buffy coat was stored for 24 h before the platelets were separated indicating one more day of storage for platelets.

The overall pattern indicates that platelet concentrates are well preserved in DINCH plasticised containers for more than five days. The results of the present study clearly show the suitability of DINCH plasticised PVC containers to preserve the function and viability of platelets in the medium concentration range. Further studies are necessary to define the range of platelet concentration which could be used, the morphology changes and the *in vivo* evaluation of platelets stored in the new type of platelet storage bag.

# **Conclusions**

The present studies show that the PVC plasticised with the non phthalate, non aromatic, non toxic plasticiser DINCH is a viable alternative to other existing containers for the storage of platelets

Fable 3: Biochemical studies           Study parameter	Sample	Test days					
		1	2	3	5	7	
Н	TPL 157-a	6.95	6.99	7.04	6.94	6.76	
	TPL 157-b	6.94	7.08	7.14	7.11	7.03	
	TPL 157-c	6.95	7.05	7.11	7.09	7.04	
	TPL 157-d	6.95	7.04	7.07	7.06	6.98	
	TPL 167-a	7.15	7.25	7.29	7.27	6.95	
	TPL 167-b	7.008	7.09	7.14	7.1	7.18	
	TPL 167-c	7.219	7.3	7.36	7.33	7.28	
	TPL 167-d	7.14	7.25	7.27	7.24	7.15	
O <sub>2</sub> (mm/Hg)	TPL 157-a	37.2	82.6	114.5	134.9	124.2	
	TPL 157-b	35.5	107.7	135	140.3	137.1	
	TPL 157-c	40.3	91	108.9	113.2	74.4	
	TPL 157-d	39.5	89.1	95	103.4	105	
	TPL 167-a	105.4	121.7	125.2	142	122.4	
	TPL 167-b	78.9 128.4	91 128.2	91.9	111.2 143.4	143.6 159.6	
	TPL 167-c TPL 167-d	110.2	118.4	120.9 91.9	130.4	148.4	
1 CO (mm/Ha)							
CO <sub>2</sub> (mm/Hg)	TPL 157-a TPL 157-b	53.3	42 45.7	32.3	22.4	16.2 15.2	
	TPL 157-0	62.4 62.7	45.7 52	33.3 41	22.4 34.1	30.2	
	TPL 157-d	59.3	48.4	42.3	33.5	31.2	
	TPL 167-a	40.1	27.2	19.5	14.4	18.1	
	TPL 167-a	40.1 47.9	33.8	19.5	19.1	16.1	
	TPL 167-6	36.7	26.4	15.3	15.3	12.8	
	TPL 167-d	40.4	27.4	15.1	15.1	10.7	
ilucose (ma/dl)	TPL 157-a	258	275	229	197	191	
cose (mg/dl) state (mg/dl)	TPL 157-a	287	266	257	237	237	
	TPL 157-c	295	287	274	260	273	
	TPL 157-d	289	276	262	243	258	
	TPL 167-a	330	317	272	252	230	
	TPL 167-b	304	296	228	213	189	
	TPL 167-c	388	380	409	350	330	
	TPL 167-d	361	356	386	314	298	
actate (mg/dl)	TPL 157-a	176	180	208	260	335	
	TPL 157-b	134	148	169	215	271	
	TPL 157-c	122	132	150	184	231	
	TPL 157-d	138	151	163	206	147	
	TPL 167-a	182	190	211	150	174	
	TPL 167-b	225	234	268	178	202	
	TPL 167-c	142	155		201	144	
	TPL 167-d	175	185	197	249	174	
icarbonate (mEq/l)	TPL 157-a	10	11	8	0	0	
, , ,	TPL 157-b	12	12	9	3	4	
	TPL 157-c	12	13	11	6	6	
	TPL 157-d	11	13	11	5	7	
	TPL 167-a	11	11	10	7	5	
	TPL 167-b	10	10	6	6	5	
	TPL 167-c	11	11	10	8	7	
	TPL 167-d	10	9	8	6	5	
lasma K+ (mEq/l)	TPL 157-a	3.9	3.8	3.9	4	4	
	TPL 157-b	3.9	3.9	3.8	4	4	
	TPL 157-c	3.9	3.8	3.8	3.9	3.9	
	TPL 157-d	3.8	3.8	3.8	3.9	4	
	TPL 167-a	4.2	4.1	4.3	4.2	4.1	
	TPL 167-b	3.9	3.8	3.9	3.9	3.8	
	TPL 167-c	4.2	4.2	4.3	4.2	4.3	
looma No + (~F~/I)	TPL 167-d	4.3	4.2	4.3	4.3	4.3	
lasma Na + (mEq/l)	TPL 157-a	165	165 165	169	167	166	
	TPL 157-b TPL 157-c	165 165	165 166	164 165	165 166	166 165	
	TPL 157-c TPL 157-d	164	165	166	166	167	
	TPL 157-0 TPL 167-a	163	161	152	167	163	
	TPL 167-a TPL 167-b		165		165	164	
	TPL 167-0	165 166	166	155 167	155	167	
	TPL 167-d		166	166	155	167	
DH 111/1		168 157					
DH IU/L	TPL 157-a TPL 157-b	157 147	180	144 175	155	194 288	
	TPL 157-0	147 145	143 144	175 155	203 163	∠66 192	
	TPL 157-d	145	144	139	152	185	
	TPL 167-a	141	165	188	176	219	
	TPL 167-b TPL 167-c	152 123	176 134	184 127	171 159	223	
						154	

Table 4: Platelet aggregation studies on platelets stored in the test bags

Sample	Study parameter		Test days				
		1	2	3	5	7	
TPL 157	Aggregation induced by ADP (%)			-			
	Mean value.	-	27	10	11	5	
	Aggregation induced by collagen(%) Mean value.						
		=	59	17	26	10	
TPL 167	167 Aggregation induced by ADP (%)						
Mean value.	39	22	18	17	11		
	Aggregation induced by collagen(%)						
	Mean value.	47	52	26	15	11	

for more than five days. The bags have very low odour, are non allergenic, and have low leachability into blood plasma.

# **Summary**

The present studies show that DINCH plasticised PVC bags (TPL-167) are well suited for the storage of platelet concentrates for more than five days.

The well-accepted platelet storage bags at present are made from special polyolefins and PVC plasticised with TEHTM or BTHC. These bags have shortcomings as pointed out earlier. DINCH plasticised PVC containers seem to be the best alternative. More studies are needed on the morphology changes and the *in vivo* evaluation of platelets stored in this new type of platelet bag.

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# References

- Slichter S, Harker L. Preparation and storage of platelet concentrates. J Haematol 1976;34;403-9.
- Simon TL, Murphy S. Normal viability of platelet concentrates obtained from CPDA-1 blood after storage in CL-3000 blood containers. Vox Sang 1982;43:20-2.
- Murphy S, Sayar SN, Gardener FH. Storage of platelet concentrates at 22°c. Blood 1970;35:549-57.
- Labow RS, Tocchi M, Rock G. Platelet storage Effects of leachable materials on morphology and function. Transfusion 1986;26:351-7.

- Snyder EL, Bookbinder M, Kakaiya R, Ferry P, Kiraly T. Role of micro aggregate blood filtration in clinical medicine. Voz Sang 1983;45:432-7.
- Snyder EL, Koerner TAW, Kakaiya R, Moore P, Kiraly T. Effect of mode of agitation on storage of platelet concentrates in PL 732 containers for five days. Voz Sang 1983;44:300-4.
- Snyder EL, Ezckowitz M, Aster R, Murphy S, Ferri P, Smith E, et al. Extended storage of platelets in a new plastic container - *Invivo* response to infusion of platelets stored for five days. Transfusion 1985;25:209-14.
- Grode G, Miripol J, Garber J, Buchhotz DH. Extended storage of platelets in a new plastic container - Biochemical and morphological changes. Transfusion 1985;25:206-8.
- 9. Murphy S, Kahn R, Holme S, Phillips GL, Sherwood W, Davisson W, *et al.* Improved storage of platelets for transfusion in a new container. Blood 1982;60:194-200.
- 10. Hogge DE, Thompson BW, Schiffer CA. Platelet storage for seven days in second generation blood bags. Transfusion 1986;26:131-5.
- 11. Simon TL, Nelson EJ, Murphy S. Extension of platelet concentrate storage to seven days in second generation bags. Transfusion 1987;27:6-9.
- 12. Buchholz D, Aster R, Menitov J, Kagan L, Simon T, Heaton A, *et al*. Red cell storage studies in Citrate poly vinyl chloride container. Transfusion 1989;29:85.
- Buchholz D, Aster R, Menitov J, Kagan L, Heaton A, Kagan T, et al. Evaluation of a new citrate plasticised container for five day storage. Transfusion 1989;29:51S.
- 14. Gulliksson H, Shanwell A, Wikman A, Reppucci AJ, Sallander S, Uden AM. Storage of platelets in a new plastic container. Voz Sang 1991;61:165-70.
- Turner VS, Mitchell SG, Kanu SK, Hawker RJ. A comparitive study of platelets stored in poly vinyl chloride containers plasticised with butyryl trihexyl citrate or tri ethyl hexyl trimellitate. Vox Sang 1995;69:195-200.
- 16. Shimizu T, Kouketsu K, Morishima Y, Goto S, Hasegawa I, Kamiya T, *et al.* A new poly vinyl chloride blood bag plasticised with less leachable phthalate ester analogue, di-n-decyl phthalate, for storage of platelets. Transfusion 1989;20:292-7.
- 17. Indian patent Application 254/CHE/2006, To Terumo Penpol Ltd.
- 18. Indian patent Application 1407/CHE/2008, To Terumo Penpol Ltd.

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