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Efficacy of live zoster vaccine in preventing zoster and postherpetic neuralgia

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Abstract

Declining cell-mediated immunity to varicella zoster virus (VZV) in elderly individuals results in virus reactivation manifest by zoster (shingles) and postherpetic neuralgia (PHN). To prevent virus reactivation, a new VZV vaccine (Zostavax, Merck) that boosts cell-mediated immunity to VZV was developed. The 3-year Shingles Prevention Study showed that Zostavax significantly reduced burden of disease due to zoster and PHN. Despite its cost-effectiveness for adults ages 65 to 75 years, as determined in the US, Canada and UK, less than 2% of immunocompetent adults over age 60 years in the US were immunized in 2007. This was due to a combination of lack of patient awareness of the vaccine, physicians' uncertainty about the duration of protection, and different cost-sharing plans for immunization. Nevertheless, zoster vaccine is safe, effective, and highly recommended for immunization of immunocompetent individuals over age 60 years with no history of recent zoster.

Keywords

zoster; shingles; immunization; postherpetic neuralgia

Introduction

Varicella zoster virus (VZV), an exclusively human neurotropic alphaherpesvirus, causes varicella (chickenpox), after which virus becomes latent in cranial nerve ganglia, dorsal root ganglia and autonomic ganglia along the entire neuraxis. Years later, as cell-mediated immunity to VZV declines with age or with immunosuppression in organ transplant recipients and patients with cancer or AIDS, VZV reactivates to cause zoster (shingles), often followed by chronic pain (postherpetic neuralgia or PHN), vasculopathy, meningoencephalitis, myelopathy, cerebellitis, and various ocular disorders (Fig. 1). VZV reactivation can also produce radicular pain without rash (zoster sine herpette), and it is now clear that all of the known neurological and ocular complications of VZV reactivation can occur without rash.

To prevent any and all of the neurological complications produced by VZV reactivation, a new VZV vaccine (Zostavax, Merck) was developed. The preventive effect of zoster vaccine rests in its ability to boost host cell-mediated immunity to VZV. Here, we describe the protean neurological complications of VZV reactivation and cite the clinical evidence

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for the effectiveness of zoster vaccine, important features regarding its clinical usefulness, adverse effects, and cost-effectiveness.

Neurological complications of VZV reactivation

Herpes zoster

Zoster is common, with nearly 1,000,000 individuals in the United States affected annually. The incidence of zoster is 5 to 6.5 per 1,000 individuals at age 60 years, increasing to 8 to 11 per 1,000 at age 70 years [1]. Unlike varicella (chickenpox), which occurs primarily in the spring, there is no seasonal predilection for zoster. Zoster in young adults may be the first manifestation of HIV infection [2]. Interestingly, chickenpox in infancy predisposes to zoster earlier in life [3]. Zoster is characterized by dermatomal distribution pain and rash. VZV is highly infectious and transmission occurs by direct contact with skin lesions or by respiratory aerosols. Immunosuppression increases the risk of disseminated zoster [4]. In most patients, the disappearance of skin lesions is accompanied by decreased pain and complete resolution of pain in 4 to 6 weeks. Magnetic resonance imaging (MRI) has shown enhancement of ganglia and affected nerve roots [5]. Because VZV becomes latent in ganglia along the entire neuraxis, zoster can develop anywhere on the body. Zoster can affect any cranial [6-17] or spinal nerves at all levels. Zoster paresis (zoster with lower motor neuron type weakness) occurs in the arm, leg [18,19], diaphragm [20] or abdominal muscles [21]. Subclinical VZV reactivation (without pain or rash) has been demonstrated in astronauts [22], with shedding of infectious virus [23]. Cardinal pathological features of zoster are inflammation and hemorrhagic necrosis with associated neuritis, localized leptomeningitis, unilateral segmental poliomyelitis, and degeneration of related motor and sensory roots [24,25]. Demyelination occurs in areas with mononuclear cell (MNC) infiltration and microglial proliferation. Intranuclear inclusions, viral antigen and herpesvirus particles have been detected in acutely infected ganglia [26-29]. Antiviral drugs (e.g., valacyclovir, 1 g three times daily for 7-10 days) speed healing of rash and shorten the duration of acute pain. In immunocompromised patients, intravenous acyclovir (5-10 mg/kg three times per day for 5-7 days) is recommended.

Postherpetic neuralgia (PHN)

PHN is characterized by constant, severe, stabbing or burning dysesthetic pain that persists for at least 3 months and sometimes years after resolution of rash. About 40% of zoster patients over age 60 years experience PHN [30,31]. The cause and pathogenesis of PHN are unknown. Two non-mutually exclusive theories are that: (1) excitability of ganglionic or even spinal cord neurons is altered; and (2) persistent or low-grade productive virus infection exists in ganglia. In a study by Watson et al. [32], who detailed the pathology in a case of severe thoracic PHN for 5 years, the most striking finding was atrophy of the dorsal horn over five segments, with only one ganglion affected by fibrosis and cellular loss and with only the roots at that level involved. Further studies in three PHN cases with severe pain [33] again revealed such dorsal root atrophy and ganglionic changes but also marked inflammation in the dorsal horn in one acute case and loss of axons and myelin in the sensory root and peripheral nerve. The finding of inflammatory changes at multiple levels bilaterally affecting roots, ganglia and nerves raised the possibility of ongoing generalized inflammation as a pathogenetic mechanism in some cases. Further support for the concept that PHN is produced by low-level ganglionitis comes from the detection of VZV DNA and proteins in blood MNCs of many patients with PHN [34-36], and from the favorable response of some PHN patients to antiviral treatment [37-39]. In a prospective, open-label phase I/II clinical trial, 15 patients with moderate to severe PHN were treated with intravenous acyclovir for 2 weeks, followed by oral valacyclovir for 1 month; 8 of 15 (53%) patients reported improvement of pain [39].

VZV vasculopathy

VZV vasculopathy results from productive virus infection in large and/or small cerebral arteries. Patients present with headache, fever, mental status changes, transient ischemic attacks (TIAs) and/or focal deficit (stroke). The clinical spectrum includes aneurysms [40] and hemorrhage, arterial ectasia and dissection [41]. More than one-third of cases of VZV vasculopathy occur without rash [42]. The cerebrospinal fluid (CSF) often contains a mononuclear pleocytosis and oligoclonal bands; the oligoclonal IgG is antibody directed against VZV [43]. Brain imaging usually reveals ischemic and/or hemorrhagic infarcts, more deep-seated than cortical lesions and particularly at gray-white matter junctions, a clue to diagnosis. Cerebral angiography may show focal arterial stenosis or occlusion. Macroscopically, lesions at gray-white matter junctions predominate. Virus is present in affected cerebral arteries as evidenced by the presence of multinucleated giant cells, Cowdry A inclusion bodies, herpes virus particles seen by electron microscopy, VZV DNA and VZV antigen. In chronic cases, virus is also found in areas of infarction, usually close to arteries and veins.

Two recent studies revealed an increased risk of stroke after zoster. Analysis of 7760 patients who had been treated for zoster showed that the risk of stroke was 31% higher within a year after zoster and approximately 4-fold higher in patients with herpes zoster ophthalmicus (HZO) versus the comparison cohort of 23,380 controls [44]. Similarly, Lin et al. [45] found that the risk of stroke in 658 patients with HZO as compared with that in 1,974 controls was 4.52-fold higher. These studies are important because the aging population is rapidly increasing. Stroke can now be added to the list of other serious complications of HZO such as keratitis and PHN.

Confirmation of VZV vasculopathy requires virological analysis to detect amplifiable VZV DNA or anti-VZV IgG antibodies or both in the CSF. The CSF does not always contain PCR-amplifiable VZV DNA, but does contain anti-VZV IgG [46]. The detection of anti-VZV IgG, but not VZV DNA, likely reflects the chronic, protracted course of disease. Testing for both VZV DNA and anti-VZV IgG must be done, and only negative findings in both can exclude the diagnosis of VZV vasculopathy. Also, since VZV vasculopathy can occur without rash, all vasculopathies of unknown etiology should be evaluated for VZV. Rapid diagnosis of VZV vasculopathy is important since the mortality without treatment is 25% [47], while treatment with intravenous acyclovir, even after neurologic disease has been present for months, can be curative [48].

VZV meningitis, cerebellitis and meningoradencephalitis

VZV may also present as meningitis or a meningoencephalitis. Many reported cases of VZV encephalitis may actually be VZV vasculopathy [49]. Recent reports of VZV meningitis [50,51], meningoradiculitis [52] and cerebellitis (gait ataxia and tremor predominated) [53,54], all in the absence of rash and confirmed by the detection of VZV DNA and anti-VZV antibody in CSF, revealed that VZV is not an uncommon cause of aseptic meningitis.

VZV myelopathy

VZV myelopathy presents in various ways. One form is a self-limiting, monophasic spastic paraparesis, with or without sensory features and sphincter problems. This so-called post-infectious myelitis usually occurs in immunocompetent patients, days to weeks after acute varicella or zoster. Its pathogenesis is unknown. The CSF usually contains a mild mononuclear pleocytosis, with a normal or slightly elevated protein. Steroids are used to treat these patients [55], although some improve spontaneously [56]. Rarely, VZV myelitis recurs, even in immunocompetent patients [57].

VZV myelopathy may also present as an insidious, progressive and sometimes fatal myelitis, mostly in immunocompromised individuals. Indeed, AIDS has commonly and increasingly become associated with VZV myelitis. MRI reveals longitudinal serpiginous enhancing lesions. Diagnosis is confirmed by the presence of VZV DNA or anti-VZV IgG or both in CSF [57]. Pathological and virological analyses of the spinal cord from fatal cases have shown frank invasion of VZV in the parenchyma [58] and in some instances, spread of virus to adjacent nerve roots [59]. Not surprisingly, some patients respond favorably to antiviral therapy [60-62]. Importantly, VZV myelitis may develop without rash. Early diagnosis and aggressive treatment with intravenous acyclovir has been helpful, even in immunocompromised patients [60]. The benefit of steroids in addition to antiviral agents is unknown.

VZV can also produce spinal cord infarction, identified by diffusion-weighted MRI and confirmed virologically [63]. Thus, VZV vasculopathy can cause stroke in the spinal cord as well as in the brain.

VZV retinal necrosis

VZV produces multiple ocular disorders, including both acute retinal necrosis (ARN) and progressive outer retinal necrosis (PORN). ARN develops in both immunocompetent and immunocompromised hosts. Patients present with periorbital pain and floaters with hazy vision and loss of peripheral vision. Treatment is typically intravenous acyclovir, steroids and aspirin followed by oral acyclovir [64]. Intravitreal injections of foscarnet and oral acyclovir have been used in early, milder cases. Although PORN can be caused by herpes simplex virus and cytomegalovirus, most cases are produced by VZV, primarily in AIDS patients with CD4+ counts typically less than 10 cells/mm³ of blood [65], as well as in other immunosuppressed individuals [66]. PORN may be preceded by retrobulbar optic neuritis and aseptic meningitis [67], central retinal artery occlusion or ophthalmic-distribution zoster [68], and may occur together with multifocal vasculopathy or myelitis. Patients present with sudden painless loss of vision, floaters and constricted visual fields with resultant retinal detachment. Multifocal, discrete opacified lesions begin in the outer retinal layers peripherally and/or posterior pole; only late in disease are inner retinal layers involved. Diffuse retinal hemorrhages and whitening with macular involvement bilaterally are characteristic findings. Treatment with intravenous acyclovir has given poor or inconsistent results [69], and even when acyclovir helped, VZV retinopathy recurred when drug was tapered or stopped. PORN patients treated with a combination of ganciclovir and foscarnet or with ganciclovir alone had a better final visual acuity than those treated with acyclovir or foscarnet [70]. The best treatment for PORN in AIDS patients may be prevention with HAART, which appears to decrease its incidence [71].

Zoster sine herpette

Zoster sine herpette (radicular pain without rash) is due to reactivation of VZV [72], a concept first supported by the description of dermatomal distribution radicular pain in areas distinct from pain with rash in zoster patients [73]. Currently, most clinicians regard zoster sine herpette exclusively as the rare occurrence of chronic radicular pain without rash. Other causes of chronic radicular pain include diabetes, lymphoma, cancer and sarcoidosis. Virological confirmation of zoster sine herpette is best provided by detection of VZV DNA in CSF. In recent years, the detection of VZV DNA and anti-VZV IgG antibody in patients with meningoencephalitis, vasculopathy, myelitis, cerebellar ataxia and polyneuritis cranialis, all without rash, has expanded the spectrum of neurological disease produced by VZV in the absence of rash. Prevalence estimates of VZV-induced pathology without rash await virological analysis of additional patients with prolonged radicular pain or other neurological symptoms and signs. Analyses should include tests for anti-VZV IgG, anti-

VZV IgM and PCR-amplifiable VZV DNA in CSF, anti-VZV IgM in serum, as well as examination of blood MNCs for VZV DNA.

VZV genome

The VZV genome contains at least 70 genes, nearly all of which have homologs in herpes simplex virus (HSV). The complete sequence of the VZV genome was determined by Davison and Scott [74]. The prototype strain, VZV Dumas, is 124,884 base pairs in length. The genome consists of a unique long region bounded by terminal and internal repeat sequences, and a unique short region bounded by internal short and terminal short repeat sequences. Strain differences have been identified by variation in the length of the repeat regions. The genome is linear in virions and circular in infected cells. Genotyping by restriction enzyme analysis and single nucleotide polymorphisms has proven that the virus causing varicella is identical to that which later reactivates as zoster in the same person. Furthermore, these technologies have shown that reinfection, which is mostly asymptomatic, may also occur and that the second virus may establish latency and reactivate [75].

Viral pathogenesis

VZV is the only human herpesvirus that spreads by the respiratory route from skin lesions of individuals with varicella or zoster [76]. Spread from the respiratory tract is also possible, but varicella is not usually accompanied by coughing and sneezing which propel infectious droplets into the air [77]. During primary infection, VZV disseminates to multiple sites resulting in vesicles on an erythematous base all over the skin. VZV-infected tonsillar CD4+ T cells transport virus from lymph nodes to skin during primary viremia. VZV-infected T cells also transfer virus to neurons in ganglia. Dendritic cells of the respiratory mucosa appear to be the first cells to encounter VZV, and these cells transport virus to draining lymph nodes. The incubation period is long, often up to 21 days before skin lesions occur. Varicella can be acquired from patients with zoster. Zoster occurs only after virus reactivation, thus it is not acquired from other patients with zoster or varicella. Reactivation allows VZV to transmit infection to a younger generation. Thus, VZV, a virus without any animal reservoir, can perpetuate itself.

VZV latency

VZV establishes latency in neurons of human ganglia where the virus genome is most likely maintained as a circular episome bound to histones. There is considerable variability among individuals in the number of latent VZV DNA copies. Expression of VZV genes during latency is restricted and regulated epigenetically. Of the VZV open reading frames (ORFs) that have been analyzed for transcription during latency using cDNA sequencing, only ORF 21, 29, 62, 63 and 66 have been detected. VZV ORF 63 is the most frequently and abundantly transcribed VZV gene detected in latently-infected human ganglia, suggesting a critical role for this gene in maintaining a latent state. Other transcripts have been detected inconsistently, suggesting that these genes either play secondary roles in latency or possibly reflect subclinical VZV reactivation. Deep sequencing of RNA extracted from human trigeminal ganglia positive for both VZV and HSV-1 DNA revealed several microRNAs mapping to the HSV genome, but not VZV-specific microRNAs [78].

VZV-specific immunity

Primary VZV infection that induces varicella is followed by the production of VZV-specific antibody and VZV-specific T cell mediated immunity. Antibodies to VZV are detected throughout life. The serum of individuals 60 to 94 years of age contains a variable presence of VZV-specific antibodies to VZV glycoproteins I-IV and to three nonglycosylated

proteins; antibodies to VZV are also present in some elderly individuals with no history of varicella or zoster, indicative of subclinical infection [34]. Increased levels of antibody to VZV do not confer protection against zoster or PHN. In fact, increased levels of antibody to VZV after the onset of zoster are associated with more severe disease and a greater risk of PHN, perhaps because they reflect more extensive VZV replication [79].

Recovery from varicella is associated with the development of VZV-specific T cell-mediated immunity [80]. The cell-mediated immune response to VZV is detected within 1 to 2 weeks after disappearance of rash, and consists of CD4 and CD8 effector and memory T cells. T-cell immunity to VZV is more important than the antibody response, as determined in studies of natural VZV infection in humans with specific immune deficiencies. For example, patients with agammaglobulinemia are unable to produce VZV-specific antibodies but are protected against second episodes of varicella because they are able to mount a VZV-specific T cell-mediated immune response [81]. Individuals with T cell-immune deficiency disorders have more severe VZV-specific disease than do normal hosts [82]. Furthermore, immune suppression is associated with a significant increase in the incidence of zoster [83]. Even in human stem cell recipients who received inactivated VZV vaccine, protection correlated with VZV-specific T cell-mediated immunity but not with anti-VZV antibody [84].

VZV-specific T cell-mediated immunity maintains VZV in a latent state in ganglia. The immune response can subsequently be boosted by subclinical reactivation of latent virus or environmental exposure to virus. Most important, the incidence of zoster increases with age as VZV-specific T cell-mediated immunity declines. The age-related decline in VZV-specific T-cell immunity is evident during the first 3 years after varicella, when the frequency of VZV-specific memory CD4 T cells already decreases [85] and after mid-adult life, the intensity and quality of antigenic stimulation provided by re-exposure and asymptomatic reactivation are not sufficient to maintain VZV-specific T cell immunity. Furthermore, a comparison of the cell-mediated immune response to VZV antigen *in vitro* in young adults and individuals over age 60 years revealed 5-fold fewer CD4 cells that produce interferon-gamma or interleukin-4 and -5, as well as fewer CD4 early effectors and CD8 effector memory cells in the older age group [86].

Recognition of the essential role of cell-mediated immunity to VZV for protection against and recovery from varicella and zoster led to studies designed to boost the cell-mediated immune response to VZV by immunization of elderly adults. Demonstration of the ability of zoster vaccine to reverse VZV-specific T cell deficiencies present before immunization was followed by the Shingles Prevention Study (described below).

VZV vaccine

There are two VZV vaccines, both of which contain live attenuated virus. The first to be developed was the varicella (Oka strain) vaccine (Varivax, Merck), which has been used to prevent chickenpox in children in Japan since 1975 and in the US since 1996. Varivax generates VZV-specific humoral and cell-mediated immune responses, particularly CD8 T cells [87], and the memory cell response that occurs after vaccination protects from re-exposure to VZV. The second vaccine (Zostavax, Merck) was developed to prevent zoster (shingles) by boosting a waning cell-mediated immune response to VZV with advancing age. In the United States, Zostavax was licensed by the FDA in May 2006, provisionally recommended by the Center for Disease Control Advisory Committee on Immunization Practices for vaccination of persons aged ≤ 60 years in October 2006, and the recommendation became formalized upon publication in June 2008. The only difference

between the two vaccines is that Zostavax contains 19,400 pfu per dose, 14-fold more virions than the varicella vaccine.

Zoster vaccine

The most widely used measure of the cell-mediated immune response to VZV in elderly individuals before and after receiving zoster vaccines has been the responder-cell frequency [88]. When administered to people over age 60 years, zoster vaccine boosted VZV-specific T cell-mediated immunity, inducing increased numbers of CD4 and CD8 cells, CD4 and CD8 effector memory T cells, and CD8 early-effector T cells; the half-life of the boost in T cell immunity to VZV is at least 5 years [89]. Zoster vaccine also boosts VZV-specific immunity in adults with a history of zoster before vaccination or with chronic illness, and can be administered in these populations.

Zostavax is injected subcutaneously and stored frozen. The vaccine is indicated for the prevention of zoster in individuals 60 years of age and older; it is not indicated for the treatment of zoster or PHN, or for women of child-bearing age. Administration is also not recommended for individuals with a history of an immunodeficiency state, including leukemia, lymphoma, malignant neoplasm infecting the bone marrow or lymphatic system, or AIDS, and should not be administered to pregnant women.

The critical trial of the licensed zoster vaccine (Shingles Prevention Study, SPS) was a placebo-controlled, double-blind trial in which more than 38,000 adults over 60 years of age were randomized to receive either zoster vaccine or placebo. All subjects were monitored for zoster, with endpoints including burden of illness due to zoster and zoster-associated pain, as well as incidence of clinically significant PHN. Subjects received a single-dose of Zostavax (n = 19,270) or placebo (n = 19,276). Ratio distribution across both vaccination groups was similar: White (95%), Black (2%), Hispanic (1%) and other (1%) in both groups. The gender distribution was 59% male and 41% female in both groups. The most common side effects reported by vaccinated participants were redness, pain, itching, swelling, warmth or bruising at the vaccination site and sometimes headache. Varicella-like rashes at the injection site were more common in zoster vaccinees than in placebo recipients (6.4% versus 0.1%; $p < 0.05$).

After a mean follow-up of 3 years, the SPS found that Zostavax vaccine significantly reduced the incidence of zoster by 51%. Moreover, subjects in the immunization group who developed zoster reported significantly less pain and discomfort, and fewer PHN cases were detected in these subjects (an overall 61% lower burden of disease) than those in the placebo group. Table 1 shows that during the first 42 days after immunization, varicella-like rashes at the injection site were more common in vaccine recipients than in placebo recipients (0.1% vs. 0.04%, $p < 0.05$); compared to placebo recipients, the vaccine group also had more erythema (36% vs. 7%), localized pain or tenderness (35% vs. 9%), swelling (25% vs. 5%) and itching (7% vs. 15%) ($p < 0.05$ for all four comparisons). Importantly, no significant differences were seen between zoster vaccine and placebo groups in the incidence of vaccine-related serious adverse events (both $< 0.1\%$).

Baseline immunologic measurements from the SPS confirmed that VZV-specific T cell immunity measured by responder cell frequency, as well as an ELISPOT assay, declined continuously with advancing age [90]. ELISPOT responses peaked at 2 weeks after immunization and at 6 weeks, were approximately 2-fold higher in immunized recipients than in placebo recipients [91]. All values fell during the first year after immunization, but remained approximately 50% higher than pre-immunization levels for the 3-year study period. Importantly, the boost in VZV-specific T cell-mediated immunity was similar to that developing after naturally occurring zoster [79]. The magnitude of the boost in cell-mediated

immunity to VZV was greatest in younger subjects, consistent with the greater efficacy of vaccine in preventing zoster in adults of age 60 to 69 years compared to those older than 70 years. The memory responses elicited in older subjects is similar to that seen after vaccination of such individuals with pneumococcal and influenza vaccines. Zoster vaccine was well-tolerated when administered concomitantly or sequentially with an inactivated influenza vaccine or pneumococcal vaccine.

Importantly, in immunized individuals who developed zoster, high VZV-specific cell-mediated immune responses were associated with reduced zoster severity and a lower frequency of PHN than in participants with lower VZV-specific cell-mediated responses, whereas a high humoral response was associated with increased severity of zoster and a higher frequency of PHN than in participants with a lower humoral immune response [79].

The Centers for Disease Control and Prevention (CDC) Advisory Committee on Immunization Practices recommends zoster vaccine for all persons over age 60 years who have no prior indications, including a previous episode of zoster or chronic medical conditions. Although zoster vaccine is not recommended for immunocompromised individuals, it appears that the current zoster vaccine could be safely administered to several groups of moderately immunocompromised adult patients, such as VZV-seropositive HIV-infected patients with CD4 T cell counts greater than 200 cells/ml, or even to patients with rheumatoid arthritis or psoriasis and receiving moderate doses of methotrexate, steroids or tumor necrosis factor inhibitors [92]. The currently licensed zoster vaccine remains to be tested in populations of moderately immunocompromised patients. Higher titer zoster vaccines also await testing for safety and immunogenicity. Note that heat-inactivated VZV vaccine has been safely administered to autologous bone marrow transplant recipients, in whom an accelerated recovery of cell-mediated immunity to VZV and a reduced incidence of zoster were observed [84,93].

Several important questions regarding vaccines remain to be addressed. How many years will the current zoster vaccine maintain immunity to prevent zoster? Is zoster vaccine safe for immunocompromised individuals? Will a killed VZV vaccine produce a significant increase in cell-mediated immunity to VZV? Should multiple vaccinations of the elderly be considered for every decade of life after age 60 years? Should zoster vaccine be refined to include epitopes that induce cell-mediated immunity to VZV?

Cost of zoster and cost-effectiveness of vaccine use

Epidemiologic analysis of the disease burden of zoster in pre-vaccine Taiwan [94] revealed an overall incidence of zoster of 4.97 cases per 1000 people, with women having a significantly higher incidence than men. The incidence increased stepwise with age. The average cost of one case of zoster was about \$204 per person for those 80 years or older. Zoster-related hospitalizations and medical cost per patient increased with age. Overall, approximately two-thirds of Taiwanese zoster cases occurred in adults older than 40 years, and about one-third of the population would develop zoster in their lifetime (lifetime risk of 32.2%). On the whole, zoster is a prevalent and costly condition.

Several studies have evaluated the cost-effectiveness of the use of vaccine to prevent zoster and PHN. A decision model to study the cost-effectiveness of vaccine in older adults suggested that immunization would increase quality-adjusted life-years (QALYs) compared to no immunization [95]. Cost-effectiveness of the zoster vaccine varied substantially with patient age and often exceeded \$100,000 per QALY saved, and it was recommended that age should be considered in vaccine recommendation [96; see below]. Another study of the cost-effectiveness of zoster immunization in Canada, a country with a population of 30 million people, estimated that there are 130,000 new cases of zoster, 17,000 cases of PHN

and 20 deaths [97]. Vaccinating 65-year-old individuals (zoster efficacy = 63%, PHN efficacy = 67%, cost/course = \$150) is estimated to cost \$33,000 per QALY-gained. Assuming a cost per course of zoster immunization of \$150, a probabilistic sensitivity analysis suggested that immunizing between 65 and 75 years of age will likely yield cost-effectiveness ratios below \$40,000 per QALY gained, while immunizing adults older than 74 years will yield ratios less than \$70,000 per QALY gained. Thus, immunizing adults age 65 to 75 years is likely cost-effective and a judicious use of healthcare resources. In a separate analysis of immunization at ages 65 to 75 years, the same conclusion was reached for England and Wales; An additional conclusion was that a booster dose at a later age was unlikely to be cost-effective [97,98]. A United States study, also based on a price of \$150 for zoster vaccine, estimated that use of vaccine for immunocompetent adults aged 60 years and older was cost-effective; the cost-effectiveness ratios ranged from \$16,229 to \$27,609 per QALY gained [99]. The studies in England, Wales, United States and Canada all used statistical models that track individuals through various alternative health states after immunization (for example, no herpes zoster, herpes zoster, PHN, death). Studies calculated health care costs based on estimates from societal or health care payer perspectives. Assumptions regarding age-specific incidence of zoster and PHN were derived from epidemiologic studies, while vaccine efficacy against those disorders was based on the SPS for different ages over a mean 3 years of follow-up. In November 2010, a call to three hospital pharmacies in Denver, Colorado, revealed prices of \$115-\$220 to receive zoster vaccine. Thus, the studies above may overestimate cost-effectiveness if vaccine price is routinely greater than \$150.

In addition to questions about cost-effectiveness, insurance coverage and related logistical concerns pose additional challenges to the widespread use of zoster vaccine. While all Medicare Part D plans cover the zoster vaccine, the amount of cost-sharing (i.e., patient out-of-pocket cost) for immunization varies. Essentially, the zoster vaccine is treated like a prescription drug, with varying co-payments depending on the patient's drug plan. A national survey of primary care physicians regarding zoster and the zoster vaccine [100] revealed that while the physicians perceived a high level of burden from zoster and PHN and generally favored use of zoster vaccine, some physicians expressed concern regarding the uncertainty of protection duration, the need to store the vaccine in a freezer, and the potential unwillingness of patients to pay for the vaccine if it was not covered by insurance. Thus many primary care physicians are not proactive in recommending zoster immunization.

In 2007, one year after zoster vaccine was licensed and recommended by the Advisory Committee on Immunization Practices for persons age 60 years and older, less than 2% of the age group was immunized in the US. This was due to a combination of lack of patient awareness that a vaccine exists, physicians' uncertainty about the duration of protection, and different cost-sharing plans for immunization [101]. This is disappointing. Zoster vaccine should be universally administered to all individuals over age 60 years. While routine immunization is not currently recommended for people age 50 to 59 years because of lack of efficacy data and cost-effectiveness information in this group, immunization of this population should be considered since about 19% of zoster occurs between ages 50 and 59 years [102].

Conclusions

Zoster (shingles), PHN, and other serious neurological and ocular disorders result from reactivation of VZV, primarily in elderly individuals. Immunization of men and women over age 60 years with Zostavax, a live attenuated VZV vaccine that boosts a naturally declining cell-mediated immunity to VZV with age, significantly reduces the burden of disease due to

zoster and PHN. Despite its cost-effectiveness for adults ages 65 to 75 years, as determined by decision models in multiple large countries, less than 10% of immunocompetent adults over age 60 years in the US have been immunized. Although physicians are unsure about the duration of protection and how different cost-sharing plans affect consumer utilization, zoster vaccine is safe, effective, and highly recommended for immunization of immunocompetent individuals over age 60 years with no history of recent zoster.

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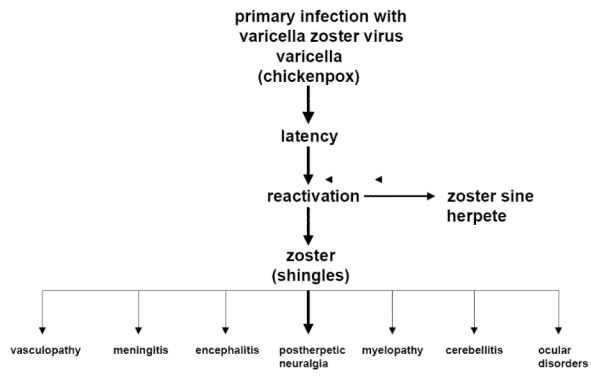


Fig. 1. The neurological complications of varicella zoster virus reactivation.

Table 1

Adverse events caused by Zostavax

Type of event	%
<u>Injection site</u> (N = 3345)	
Erythema	34
Pain/tenderness	33
Swelling	25
Hematoma	1.4
Pruritus	6.6
Warmth	1.5
<u>Symptoms</u>	
Headache	1.4