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Singleshot T₁ Mapping using Simultaneous Acquisitions of Spinand STimulated-Echo Planar Imaging (2D ss-SESTEPI)

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Abstract

The conventional stimulated-echo NMR sequence only measures the longitudinal component, while discarding the transverse component, after tipping up the prepared magnetization. This transverse magnetization can be used to measure a spin-echo, in addition to the stimulated-echo. 2D ss-SESTEPI is an EPI-based singleshot imaging technique that simultaneously acquires a spin-echo-planar image (SEPI) and a stimulated-echo-planar image (STEPI) after a single RF excitation. The magnitudes of SEPI and STEPI differ by T₁ decay and diffusion weighting for perfect 90° RF, and thus can be used to rapidly measure T₁. However, the spatial variation of B₁ amplitude induces un-even splitting of the transverse magnetization for SEPI and STEPI within the imaging FOV. Correction for B₁ inhomogeneity is therefore critical for 2D ss-SESTEPI to be used for T₁ measurement. We developed a method for B₁ inhomogeneity correction by acquiring an additional STEPI with minimal mixing time, calculating the difference between the spin-echo and the stimulated-echo and multiplying the STEPI by the inverse functional map. Diffusion-induced decay is corrected by measuring the average diffusivity during the prescanning. Rapid singleshot T₁ mapping may be useful for various applications, such as dynamic T₁ mapping for real-time estimation of the concentration of contrast agent in DCE-MRI.

Keywords

EPI; STEPI; stimulated-EPI; SESTEPI; spin-/stimulated-EPI; realtime T₁ mapping; dynamic T₁ mapping

INTRODUCTION

The spin-echo sequence, gradient-echo pulse sequence, and their variants are commonly used for data acquisition in magnetic resonance imaging (MRI). The stimulated-echo (STE) pulse sequence is often used to investigate transport phenomena, such as molecular translational diffusion (1,2) and displacement-encoded imaging (DENSE) (3). STE is also used in localized single-voxel spectroscopy for studying short T_2 species (4). The prepared magnetization after implementing the second 90° RF (tipup) pulse in STE can be decomposed into two components: the longitudinal component, which is half the magnitude of the prepared magnetization, and the transverse component, of the same magnitude; however, the transverse component is discarded, as previously reported (5-7). This results in

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wasting of a quadrature component of the magnetization. This half magnetization can be used by measuring a spin-echo after the second 90° RF pulse while simultaneously measuring the stimulated-echo (8). The additional data can be used for quantitative measurement of NMR and physical parameters such as molecular translation diffusivity, T_1 relaxation time and phase change, useful for characterization of other physical parameters such as temperature and physiological motion.

In dynamic contrast enhancement-MRI (DCE-MRI) of both animal and human, the signal intensity of 2D fast gradient-echo imaging is generally used as an estimation of the concentration of metal ion for pharmacokinetic evaluation (9). Because of the non-linear relationship between the NMR signal intensity and the concentration of paramagnetic-ion based contrast agent, the resultant pharmacokinetic parameters are subject to error and may be considered semi-quantitative. However, it is well known that $1/T_1$ is directly related to the concentration of MRI contrast agent over a wide range of concentrations (10), as in $\Delta R_1 = 1/T_1(C) - 1/T_1(C=0) = r_1C$, where r_1 and *C* are the relaxivity and the local concentration of the contrast agent, respectively.

 T_1 measurement typically requires multiple scans with different recovery times (TR) or inversion times (TI) (11). It generally takes several tens of minutes to complete T_1 measurement using the inversion-recovery (IR) or saturation-recovery (SR) spin-echo (SE), which are the most commonly used techniques (12). Using IR-SE or SR-SE for T_1 measurement that uses the recovery process of the longitudinal magnetization, the accuracy of the measured T_1 heavily depends on whether the fully (or almost fully) relaxed signal is measured. Acquisition of the fully relaxed signal significantly increases the total measurement time due to the long recovery time (TR or TI) and limits its application only to non-dynamic imaging. Gradient-echo imaging with flip angle variation has been used with reasonable temporal and spatial resolution (11); however, the T₁ estimation obtained by this method is very sensitive to B_1 inhomogeneity. Although there have been reports on various fast T₁ mapping techniques such as 2D singleshot-EPI (2D ss-EPI) and its variants (Look-Locker EPI, IR-EPI) (13-15), and a multishot spin-echo EPI with automated variation of TR and TE to simultaneously measure T_1 and T_2 (16,17), there is no technique that acquires the complete data set for T₁ calculation in a single shot suitable for dynamic T₁ measurement or that simultaneously overcomes the systematic error induced by imperfect flip angle.

In this report, the previously presented acquisition of a spin-echo-planar image (SEPI) and a stimulated-echo-planar image (STEPI) in a single shot (8) are used for rapid T_1 mapping. Because two images are acquired after a single excitation, they have identical phase-errors, assuming that the gradient behavior is ideal and that motion in the imaging field-of-view is negligible. A phase difference between SEPI and STEPI may result from subject movement during the mixing time TM, as for DENSE imaging (3). A major source of error using SEPI and STEPI for quantitative MRI comes from the spatially varying B_1 amplitude, which is never perfect in most MR imaging applications. Unless the second and the third RF pulses are a perfect 90° in all locations within the imaging FOV, the prepared transverse magnetization is no longer evenly divided between spin- and stimulated-echoes by the second RF pulse, and the third RF pulse does not flip the total stored longitudinal magnetization down to the transverse plane for the stimulated echo. This work describes a method to correct the difference in signal intensities in SEPI and STEPI caused by an imperfect 90° pulse and the diffusion weighting effect in acquired images.

METHODS

Pulse sequence description

RF in the sequence diagram in Fig. 1 includes both RF pulses and data acquisition windows of the MR signal. Three reference echoes (REF) are acquired immediately after the first 90° RF for EPI phase-correction. G_{sp} indicates the gradient pulses for the spoiler. Both spinecho and stimulated-echo are formed at the same echotime, TE, where the effect from the field inhomogeneity is removed.

All imaging gradients applied after the center of the third excitation 90°_{x} pulse are rewound at the center of the fourth restoring 90°_{-x} RF. Only the transverse magnetization, contributed from the fresh longitudinal magnetization that is regrown during the mixing time, is rephased and the fourth 90° RF tips this rephased magnetization back into the longitudinal direction. Because the rephased fresh longitudinal magnetization may be as large as 40 % for TM = ~0.5 T1 in T₁ measurement, the longitudinal magnetization before the first excitation RF of the next measurement will be increased, although the transverse magnetization resulting from fresh longitudinal magnetization undergoes T₂* decay between the third and fourth RF pulses.

Effect of diffusion and B₁ inhomogeneity on T₁ measurement

A pair of spoiler gradients (1st and 4th gradient), indicated as G_{sp} in Fig. 1, is crucial to spoil the freshly regrown longitudinal magnetization during the mixing time after it is flipped onto the transverse plane, while refocusing the prepared magnetization (18,19). However, the pair of spoiler gradients G_{sp} along with the large diffusion time (Δ) induces non-negligible diffusion weighting and decreases the signal intensity of STEPI by e^{-bD}, where b and D are the diffusion-weighting factor and the diffusivity of the water molecules, respectively. Here, b is (γA)²(Δ), where A is the zeroth moment of each spoiler gradient G_{sp} .

Assuming the actual flip angle of three RF pulses in the stimulated-echo sequence at position \overrightarrow{r} is α , i.e., not a perfect 90°, the signal intensities measured at spin-echo (SE) and stimulated-echo (STE) are written as (20),

$$S_{SE}(\vec{r}) = S_{+}^{o}(\vec{r})\sin(\alpha)\sin^{2}\frac{\alpha}{2} \cdot e^{-TE/T_{2}}$$
(1)

$$S_{STE}\left(\overrightarrow{r}\right) = \frac{1}{2}S_{+}^{o}\left(\overrightarrow{r}\right)\sin^{3}\left(\alpha\right) \cdot e^{-TE/T_{2}} \cdot e^{-TM/T_{1}} \cdot e^{-bD} = S_{SE}\left(\overrightarrow{r}\right) \cdot g\left(\overrightarrow{r}\right) \cdot e^{-\frac{TM}{T_{1}}} \cdot e^{-bD}$$
(2)

Here, TE and TM are echo-time and mixing-time, respectively. $g(\vec{r})$ indicates the ratio of signal intensities of STE and SE in the absence of T₁ and diffusion-induced decays. From

these two equations (eqs.1 and 2), the theoretical B₁ correction function $g(\vec{r})$ is obtained as,

$$g\left(\vec{r}\right) = \frac{\sin^2\left(\alpha\right)}{2\sin^2\frac{\alpha}{2}} = 2\cos^2\frac{\alpha}{2},\tag{3}$$

For an RF pulse at a perfect 90°, $g(\vec{r})$ should be 1, which indicates that the signal intensity

of STE differs from that of SE by T₁ and diffusion-induced decay term $g^{-(\frac{TM}{T_1}+bD)}$. The measured spin-lattice relaxation time T₁^{meas} can be described as,

$$T_{1}^{meas} = \left\{ \frac{\ln S_{SE}\left(\overrightarrow{r}\right) - \ln S_{STE}\left(\overrightarrow{r}\right)}{\mathrm{TM}} \right\}^{-1} = \left\{ \frac{1}{T_{1}} - \frac{1}{\mathrm{TM}} \ln\left(2\cos^{2}\frac{\alpha}{2}\right) + \frac{bD}{TM} \right\}^{-1},\tag{4}$$

This equation indicates that the measured T_1 will differ for different mixing times as well as different actual flip angles. The error that is caused by diffusion-induced signal loss in STE can be corrected by measuring the average diffusivity *D* separately as a part of a prescan. During the evaluation of eq.4, only the transmit function is considered, without involving the difference in signal reception.

There is no need to accurately measure the actual position-dependent flip angle $\alpha(\mathbf{r})$. Any difference between SEPI and STEPI₀ with minimal TM (or TM << T₁) is directly related to

the B₁ correction function. Therefore, the correction function $g(\vec{r})$ can be experimentally measured as a part of the data acquisition by setting minimum TM. The data acquisition window for SEPI is removed to acquire the STEPI₀ image with minimal mixing time. In practice, this minimal TM was about 5 ms and the T₁ decay for a typical T₁ at 3 T was less

than 1 % for T₁ > 0.5 second. A B₁ correction map $g(\vec{r})$ is then obtained by dividing STEPI₀ by SEPI₀ (the averaged SEPI) as,

$$g\left(\overrightarrow{r}\right) = \frac{STEPI_0}{SEPI_0} \tag{5}$$

The STEPI₀, which is the average of multiple STEPIs with minimum TM, and the SEPI₀, which is the average of multiple SEPIs sampled from pre-dynamic images, are initially acquired. Two to four complex signal averages were used for both STEPI₀ and SEPI₀.The

inverse of the B₁ correction function, $g^{-1}(\vec{r})$, calculated by using eq. 5, is multiplied by the STEPI signal to remove the difference caused by B₁ inhomogeneity. This correction eliminates the error in the resultant quantification that is caused by the inhomogeneous B₁.

MRI Experiments

MRI studies were performed on a Siemens Trio 3 Tesla MRI system (Siemens Medical Solutions, Erlangen, Germany) with Avanto gradients (45 mT/m strength in the longitudinal direction, 40 mT/m strength in horizontal and vertical directions, and a 200 T/m/s slew rate). The pulse sequence was developed based on a singleshot spin-echo EPI sequence, using the IDEA pulse sequence development environment. Real-time calculation of T_1 mapping was implemented into a custom-made ICE program (Image Construction Environment). The online image construction program produced two magnitude images and a T_1 map for each slice.

A phantom was constructed with two 40 cc plastic bottles filled with 0.1 and 0.2 mM $MnCl_2$ /water solutions, respectively, and immersed in a 0.075 mM $MnCl_2$ /water solution in a rectangular acrylic container with dimensions $8 \times 8 \times 13$ cm³. The T₁ of each concentration was measured as 0.904 and 0.602 sec for 0.1 and 0.2 mM, respectively, using the inversion recovery turbo-spin echo (IR-TSE) with 10 s TR, 8 ms TE, $2 \times 2 \times 2$ mm³ spatial resolution,

128×40 acquisition matrix, 250 Hz/px receiver bandwidth, and 34 inversion times ranging from 25 to 2000 ms.

An MRI experiment was performed to investigate the effect of flip angle and mixing time TM for B₁ correction mapping on the accuracy of T₁ measurement, using a 128×40 imaging matrix with 62.5 % phase-encoding asymmetry, $2\times2\times2$ mm³ spatial resolution, 1.346 kHz/ pixel receiver bandwidth, 7.0 s TR, using a transmit/receive head RF coil. The echo times were 14.2 ms for both SEPI and STEPI. The experiment was repeated for the nominal flip angles of 60, 75, 90, 105, and 120° and for different mixing times of 0.1, 0.2, 0.4, and 0.8 s to investigate the effect of the mixing time on the resultant T₁.

To examine the diffusion effect on the T₁ values by the spoiler gradient, 2D ss-SESTEPI was applied to a fluid phantom with 0.4 s TM, 5 s TR, and $2\times2\times2$ mm³ resolution. T₁ of the solution was measured as 0.95 s by IR-TSE. The zeroth moment of the spoiler gradients was incremented as multiples of A_0 . The zeroth moment A_0 satisfies the relationship $\gamma A_0 \Delta x = 2\pi$. The resultant b-factor was varied from 8 ~ 256 s/mm². Diffusivity of the water phantom was measured using 2D singleshot diffusion-weighted EPI with b=0 and 500 s/mm². Eq. 4 was used to simulate the T₁ underestimation by diffusion-weighting.

2D ss-SESTEPI was applied on a healthy human brain to acquire T_1 mapping images with imaging parameters of 84×128 acquisition matrix, 2×2×2 mm³ spatial resolution, 7 s TR, 21.7 ms TE, 256×168 mm² FOV, 1.346 kHz/pixel receiver bandwidth, and 0.6 s TM. A B₁ correction map was calculated from eight pre-scanning images. T₁ maps were computed by taking multiple measurements and calculating the averages of SEPI and STEPI for 1, 4 and 16 repetitions, separately. T₁ map imaging was also acquired using IR-TSE for the same locations with 10 s TR, 16 ms TE, 256×192 matrix, 1×1×2 mm³ spatial resolution, 11 echotrain-length, and 250 Hz/pixel receiver bandwidth for twelve different TIs (36, 70, 100, 200, 400, 500, 600, 800, 900, 1200, 1800 and 2400 ms). B-factor was estimated as 23.8 s/mm² within the pulse sequence. The imaging protocol was approved by the Institutional Review Board, University of Utah and informed consent was obtained from the volunteer.

2D ss-SESTEPI was applied to in-vivo dynamic T_1 mapping MRI of two tumor-bearing female nude mice (21), using a home-made quadrature transmit/receive saddle coil with 1.5 inch inner-diameter and 3 inch length and imaging parameters: in-plane resolution of 1×1 mm² with 2 mm slice thickness, 128×32 imaging matrix, 5.0 s TR, 120 total repetitions (time frames) for 10 min, TEs 15 msec for both SEPI and STEPI. The first 4 measurements were used to acquire STEPI₀ images for B₁ correction mapping. T₁ maps were constructed from time frame 5. Contrast agent was administered after 8 repetitions, i.e., acquisitions of four STEPI₀ and four pre-contrast scans. MultiHance (Gadobenate dimeglumine: Gd-BOPTA, Bracco Diagnostics, Inc.) was injected via the catheterized tail vein to a dose of 0.1 mM Gd/kg. Animals were cared for under the guidelines of a protocol approved by the University of Utah Institutional Animal Care and Use Committee.

RESULTS

Simulation of error on T₁

Eq. 4 was used to simulate the anticipated error in T_1 estimation with respect to the actual flip angle and mixing-time without B_1 inhomogeneity correction, as shown in plots in Fig. 2. The measured T_1 tends to be prolonged with a short mixing time and a flip angle smaller than 90° and shortened with a flip angle larger than 90°. As indicated in eq. 4, the estimated T_1 agrees with the true T_1 for a perfect 90° flip angle regardless of TM, as indicated by the vertical arrow in Fig. 2.

B₁ correction process

Fig. 3 shows the T_1 calculation procedure. The magnitude images of SEPI₀ and STEPI₀ to calculate the B₁ correction map are shown in Figs. 3a and 3b. Fig. 3c represents the B₁

correction map $g(\vec{r})$ derived from Figs. 3a and 3b. Singleshot images of SEPI and STEPI are shown in Figs. 3d and 3e, respectively. T₁ maps before and after B₁ correction can be seen in Figs. 3f and 3g. The plots in Figs. 3h and 3i are T₁ values along the horizontal dotted lines on the resultant T₁ maps of Figs. 3f and 3g, and exhibit almost complete elimination of the B₁ inhomogeneity effect.

Flip angle variation

To investigate the effects of flip angle variation on the resultant T_1 value, a T_1 map was acquired using 2D ss-SESTEPI with nominal flip angles 60, 75, 90, 105, and 120° for the second and third RF pulses in Fig. 1, using a 0.4 sec mixing time, and four-averaged B_1 correction map.

Mean T_1 values on regions-of-interest (ROIs) of 0.1 and 0.2 mM solutions are plotted in Fig. 4a and Fig. 4b. Closed and open symbols represent T_1 measurements before and after B_1 correction, and circles and square symbols indicate 0.1 and 0.2 mM MnCl₂ concentrations, respectively. Dotted and broken lines are guidelines to the T_1 plots without B_1 correction. Solid horizontal lines indicate T_1 values measured using IR-TSE imaging. These results demonstrated significant improvement in accuracy by B_1 correction for all values of T_1 (0.2 ~ 1.3 sec.). Plots in (b) are T_1 values in (a) normalized to T_1 =1.0 s to compare the empirical data to the simulated plot in Fig. 2. TM of 0.4 s corresponds to 0.44 T_1 for T_1 =0.904 s and 0.66 T_1 for T_1 =0.602 s for 0.1 and 0.2 mM MnCl₂ concentrations, respectively. Plots in Fig. 4b show good agreement with the theoretical simulation in Fig. 2 and robustness of the B_1 inhomogeneity correction.

TM variation

To improve the accuracy of T_1 measurement using 2D ss-SESTEPI, the signal difference between SEPI and STEPI must be significantly larger than the noise amplitude, otherwise the resultant measurement will be dominated by noise, rather than T_1 decayed signal change. The T_1 map was acquired using 2D ss-SESTEPI with a nominal flip angle of 90° and a fouraveraged B₁ correction map for variable mixing times of TM = 0.1, 0.2, 0.4, and 0.8 sec. The mean and standard deviation were measured on ROIs selected at each concentration and plotted with respect to the mixing time in Fig. 5. Error bars on the plots indicate the standard deviation of each measurement.

These plots indicate that the uncertainty of the measurements greatly depends on the selection of TM. T_1 measurement using the larger TM produces increased accuracy and reduced errors; however, it limits the number of slices in an interleaved multi-slice scan. For T_1 below 1.0 sec, the difference in uncertainty for measurements with 0.4 sec and 0.8 sec TM is not significant.

Diffusion effect on T₁ estimation

Open squares (\Box) in Fig. 6 are measured T_1 values. Diffusivity was measured as 2.14×10^{-3} mm²/s. The solid line indicates the simulated T_1 values using eq. 4 with respect to the diffusion-weighting by the spoiler gradients, using $T_1 = 0.95$ s, $D = 2.14 \times 10^{-3}$ mm²/s. These plots demonstrate a substantial decrease in the measured T_1 with increased diffusion-weighting by increased spoiler area.

In-vivo measurement of human brain T₁ map

The images in Fig. 7 are T_1 maps computed from (b) IR-TSE and (c~e) 2D ss- SESTEPI. T_1 values were measured for selected regions of gray and white matter (GM, WM) as shown on T_2 weighted images in Fig. 7a. Mean values are listed in Table 1. No significant difference was observed in the T_1 values of WM between IR-TSE and 2D ss-SESTEPI. The increased standard deviation of T_1 values measured by 2D ss-SESTEPI is because of the relatively poor SNR. However, T_1 values measured using 2D ss-SESTEPI in GM were about 10 % smaller than those using IR-TSE.

In-vivo measurement of singleshot T_1 in a mouse

Images in Fig. 8 are (a) T_1 maps and (b) ΔR_1 maps of selected time frames (every 30 sec.). T_1 maps of 10 slices were obtained every 5 sec. Images in Fig. 8c are T_1 weighted spin-echo images of pre-contrast and 10 min after contrast injection. Brightness indicates the larger values in T_1 and ΔR_1 maps. ΔR_1 was plotted in Fig. 8c for several selected ROIs of 9 pixels and all at the 10 minute time point. Regions of 3×3 pixels (18 µL volume) were selected at kidney, tumor periphery, tumor core, and muscle. These data can be used for further pharmacokinetic evaluation pixel-by-pixel or by organ.

DISCUSSION

 B_1 inhomogeneity correction was very successful with an experimentally measured correction map using the 2D ss-SESTEPI pulse sequence. Eq. 4, which describes the error in the T₁ estimation using SEPI and STEPI without correction of B₁ inhomogeneity, includes only the transmit function without signal reception. The measured B₁ correction map, however, reflects all inhomogeneity causes including RF transmission and NMR signal reception, except for T₁ and diffusion-weighted decays. The simulated plots in Fig. 2 agreed very well with the measured T₁ in Fig. 4, which indicates that the correction method for B₁ inhomogeneity is very robust. The only difference is a systematic underestimation. This probably results from diffusion-induced signal decay with the long diffusion time.

The accuracy of the resultant T_1 value using 2D ss-SESTEPI decreased with a lower TM/ T_1 ratio, because there is not enough change in the T_1 decay with a short TM, as shown in Fig. 5. Although the plots indicate that the measured T_1 is more accurate with increased TM/ T_1 , we have observed greatly decreased accuracy for very short T_1 and large TM. This is because there was no signal left when TM is greater than 2~3 times T_1 . The choice of mixing time, therefore, depends on the spin-lattice relaxation time of interest. For instance, one may be interested in $T_1 < 1$ s in DCE-MRI and T_1 decreases with increased local concentration of MRI contrast agent. 0.4~0.5 s TM may be suitable for dynamic T_1 mapping using paramagnetic-ion based contrast agents.

The T₁ value was greatly underestimated for an increased spoiler area, as shown in Fig. 6. This diffusion-induced error in the T₁ calculation is reduced by minimizing the zeroth moment A₀ of the spoiler gradients. However, the minimum area of the spoiler gradient must be sufficient to spread the spin phase more than 2π within an imaging voxel, as in $\gamma A_0 \cdot \Delta x > 2\pi$ for a smallest voxel dimension Δx . The b-factor was within the range of $10 \sim 30 \text{ s/mm}^2$ for TM 0.4 s and 1~2 mm pixel dimension. For a typical diffusivity of $1.0 \times 10^{-3} \text{ mm}^2/\text{s}$ in tissue water, the diffusion-weighted decay was 1~3 %, which may be ignored because it is within the noise level for STEPI images with SNR = ~ 30.

The important advantages of using 2D singleshot EPI are that the resultant images are free from motion-related artifact and have the highest signal-to-noise ratio (SNR) per fixed imaging time, which are crucial for quantitative MRI. However, 2D ss-EPI is very sensitive to the local variation of a non-linear background gradient caused by magnetic susceptibility

at or near the tissue/air and tissue/bone interfaces, as in mouse images in Fig. 8. The resultant images suffer greatly from major geometric distortion that renders them useless. For singleshot EPI type acquisition, geometric distortion often becomes the largest limitation and its application is mostly restricted to diffusion and functional MRI of the intracranial brain, far from the sinus and the temporal bone. Therefore, it is crucial to reduce the susceptibility artifact in EPI images using 2D ss-SESTEPI before it may be useful for other applications. Because the degree of geometric distortion in 2D ss-EPI is proportional to the imaging field-of-view (FOV) in the phase-encoding direction, it can be reduced by using a reduced phase FOV. However, imaging a limited FOV in large surroundings induces aliasing in the phase-encoding direction in conventional MR imaging. The aliasing artifact can be completely removed while accomplishing interleaved multi-slice imaging by using double inversion/refocusing RF pulses with slice selection gradients applied perpendicular to the imaging RF pulses (19,22-24) or 2D spatially selective excitation (25).

Although the technique measures T_1 in a singleshot in only a few seconds and the source images have low SNR, the resultant T_1 was comparable to those measured using the inversion-recovery TSE. Average T_1 values of GM and WM using 2D ss-SESTEPI and IR-TSE are summarized with other reported values (26-29) in Table 2. There is large variation among different reports, which may be because of different acquisition techniques. T_1 in GM measured using 2D ss-SESTEPI was about 10 % lower than that using IR-TSE, while T_1 s in WM were similar to each other. This underestimation may result from the diffusion-induced signal decay in STE of SESTEPI. According to the literature (30), mean apparent diffusion coefficients (ADC) in posterior left white matter are 0.67×10^{-3} mm²/s, while typical ADC in cortical gray matter are described as $1.0 \times 10^{\times 3}$ mm²/s. Theoretical underestimation of T_1 was calculated as 0.69 s and 1.169 s for GM and WM, respectively, using Eq. 4 with TM=0.6 s and b=23.8 s/mm², assuming the true T_1 values are 0.714 s (WM) and 1.226 s (GM) obtained using IR-TSE. These values are 3.0% and 4.6% different from T_{1s} measured using IR-TSE.

As presented, rapid T_1 mapping may be useful for non-invasive measurement of the concentration of the paramagnetic ion-based contrast agent. In our preliminary application of the technique for DCE-MRI on mice, the limiting factor for increased temporal resolution was signal-to-noise ratio. If a sufficient SNR is available by using a high-sensitivity RF coil or high-field MRI system, both temporal and spatial resolution can be improved.

CONCLUSIONS

2D ss-SESTEPI successfully measured both spin-echo and stimulated-echo in a singleshot. The magnitudes of both images are sensitive to the B_1 variation within the imaging FOV, therefore correction for the B_1 inhomogeneity is critical for T_1 measurement. B_1 inhomogeneity correction was successfully achieved by acquiring an additional stimulated-echo with minimal mixing time during the prescan, calculating the ratio between the spin-echo and the stimulated-echo and multiplying the stimulated-EPI by the inverse of the functional map. Realtime T_1 measurement was possible using two images and an online image reconstruction program. It was possible to rapidly acquire T_1 mapping in a few seconds, only limited by SNR of the source spin- and stimulated-EP images. The measured T_1 agreed with those measured using conventional IR-TSE. 2D ss-SESTEPI may be an important tool for real-time in-vivo estimation of the concentration of paramagnetic-ion based contrast agent in DCE-MRI, because it is rapid and the resultant images are free-from motion-related artifacts.

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REFERENCES

- Merboldt KD, Hanicke W, Bruhn H, Gyngell ML, Frahm J. Diffusion imaging of the human brain in vivo using high-speed STEAM MRI. Magn Reson Med. 1992; 23(1):179–192. [PubMed: 1734178]
- Farrell JA, Smith SA, Gordon-Lipkin EM, Reich DS, Calabresi PA, van Zijl PC. High b-value qspace diffusion-weighted MRI of the human cervical spinal cord in vivo: feasibility and application to multiple sclerosis. Magn Reson Med. 2008; 59(5):1079–1089. [PubMed: 18429023]
- Aletras AH, Ding S, Balaban RS, Wen H. DENSE: displacement encoding with stimulated echoes in cardiac functional MRI. J Magn Reson. 1999; 137(1):247–252. [PubMed: 10053155]
- Frahm J, Bruhn H, Gyngell ML, Merboldt KD, Hanicke W, Sauter R. Localized high-resolution proton NMR spectroscopy using stimulated echoes: initial applications to human brain in vivo. Magn Reson Med. 1989; 9(1):79–93. [PubMed: 2540396]
- 5. Slichter, C. Principles of Magnetic Resonance. Springer-Verlag; Heidelberg: Germany: 1990.
- Haacke, E.; Thompson, M.; Brown, R. Magnetic Resonance Imaging, Physical Principles and Sequence Design. Wiley-Liss; 1999.
- Haase A, Frahm J, Matthaei D, Hanicke W, Bomsdorf H, Kunz D, Tischler R. MR imaging using stimulated echoes (STEAM). Radiology. 1986; 160(3):787–790. [PubMed: 3737918]
- Bornert, Peter; Jensen, Dye. Single-shot-double-echo EPI. Magn Reson Imaging. 1994; 12(7):1033– 1038. [PubMed: 7997090]
- Feng Y, Jeong EK, Mohs AM, Emerson L, Lu ZR. Characterization of tumor angiogenesis with dynamic contrast-enhanced MRI and biodegradable macromolecular contrast agents in mice. Magn Reson Med. 2008; 60(6):1347–1352. [PubMed: 19025902]
- Nicolle GM, Toth Eva, Schmitt-Willich Heribert, Raduchel Bernd, Merbach AE. The Impact of Rigidity and Water Exchange on the Relaxivity of Dendritic MRI Contrast Agent. Chem Eur J. 2002; 8(5):9.
- Deoni SC, Rutt BK, Peters TM. Rapid combined T1 and T2 mapping using gradient recalled acquisition in the steady state. Magn Reson Med. 2003; 49(3):515–526. [PubMed: 12594755]
- 12. Tofts, PS., editor. Quantitative MRI of the Brain: measuring changes caused by disease. West Sussex, England: 2004.
- Gowland P, Mansfield P. Accurate measurement of T1 in vivo in less than 3 seconds using echoplanar imaging. Magn Reson Med. 1993; 30(3):351–354. [PubMed: 8412607]
- 14. Look DC, Locker DR. Time Saving in Measurement of NMR and EPR Relaxation Times. Review of Scientific Instruments. 1970; 41(2):2.
- Guo JY, Kim SE, Parker DL, Jeong EK, Zhang L, Roemer RB. Improved accuracy and consistency in T1 measurement of flowing blood by using inversion recovery GE-EPI. Medical physics. 2005; 32(4):1083–1093. [PubMed: 15895593]
- Liu, X.; Feng, Y.; Lu, ZR.; Jeong, EK. ISMRM. Berlin, Germany: 2007. Rapid Simultaneous Data acquisition of T1 and T2 Mapping, using Multishot EPI and Automated Variations of TR and TE at 3T; p. 1787
- 17. Liu X, Feng Y, Lu ZR, Morrell G, Jeong EK. Rapid Simultaneous Acquisition of T1 and T2 Mapping Images using Multishot Double Spin-Echo EPI and Automated Variations of TR and TE (ms-DSEPI-T12). NMR Biomed. 2009 in print.
- Norris DG. Implications of Bulk Motion for Diffusion-Weighted Imaging Experiments: Effects, Mechanisms, and Solutions. J Magn Reson Imag. 2001; 13:486–495.

- Jeong EK, Kim SE, Kholmovski EG, Parker DL. High-resolution DTI of a localized volume using 3D single-shot diffusion-weighted STimulated echo-planar imaging (3D ss-DWSTEPI). Magn Reson Med. 2006; 56(6):1173–1181. [PubMed: 17089367]
- 20. Liang, ZP.; Lauterbur, PC. Principles of Magnetic Resonance Imaging: A Signal Processing Perspective. Wiley-IEEE Press; New York: 2000.
- 21. Wang Y, Ye F, Jeong EK, Sun Y, Parker DL, Lu ZR. Noninvasive visualization of pharmacokinetics, biodistribution and tumor targeting of poly[N-(2hydroxypropyl)methacrylamide] in mice using contrast enhanced MRI. Pharmaceutical research. 2007; 24(6):1208–1216. [PubMed: 17387601]
- 22. Feinberg DA, Turner R, Jakab PD. Echo-Planar Imaging with Asymmetric Gradient Modulation and Inner-Volume Excitation. Magn Reson Med. 1990; (13):162–169. [PubMed: 2319932]
- Jeong EK, Kim SE, Guo J, Kholmovski EG, Parker DL. High-resolution DTI with 2D interleaved multislice reduced FOV single-shot diffusion-weighted EPI (2D ss-rFOV-DWEPI). Magn Reson Med. 2005; 54(6):1575–1579. [PubMed: 16254946]
- Feinberg D, Hoenninger J, Crooks L, Kaufman L, Watts J, Arakawa M. Inner volume MR imaging: technical concepts and their application. Radiology. 1985; 156:743–747. [PubMed: 4023236]
- 25. Saritas EU, Cunningham CH, Lee JH, Han ET, Nishimura DG. DWI of the spinal cord with reduced FOV single-shot EPI. Magn Reson Med. 2008; 60(2):468–473. [PubMed: 18666126]
- 26. Wansapura JP, Holland SK, Dunn RS, RT, Ball WS. NMR Relaxation Times in the Human Brain at 3.0 Tesla. J Magn Reson Imaging. 1999; 9:531–538. [PubMed: 10232510]
- 27. Stanisz GJ, Odrobina EE, Pun J, Escaravage M, Graham SJ, Bronskill MJ, Henkelman RM. T₁, T₂ Relaxation and Magnetization Transfer in Tissue at 3T. Magn Reson Med. 2005; 54:507–512. [PubMed: 16086319]
- Ethofer T, Mader I, Seeger U, Helms G, Erb M, Grodd W, Ludolph A, Klose U. Comparison of longitudinal metabolite relaxation times in different regions of human brain at 1.5 and 3 tesla. Magn Reson Med. 2003; 50:1296–1301. [PubMed: 14648578]
- Preibisch C, Deichmann R. T₁ Mapping Using Spoiled FLASH-EPI Hybrid Sequences and Varying Flip Angle. Magn Reson Med. 2009; 62:240–246. [PubMed: 19319894]
- Chien, Daisy; Buxton, Richard B.; Kwong, Kenneth K.; Brady, Thomas J.; Rosen, BR. MR Diffusion Imaging of the Human Brain. Journal of Computer Assisted Tomography. 1990; 14(4): 514–520. [PubMed: 2370348]



Figure 1.

Pulse sequence diagram of 2D ss-SESTEPI using EPI readout. Two EPI images are acquired simultaneously with different T₁ decay times. For perfect 90° RF pulses, half of the magnetization is stored along the longitudinal direction and the other half remains in the transverse plane after the tipup RF pulse $(90^{\circ}-_x)$. The spoiler gradient G_{sp} selectively rephases the prepared magnetization, while spoiling the fresh longitudinal magnetization that is regrown during TM. The spoiler gradient G_{sp1} is to completely spoil the transverse magnetization after the acquisition of SEPI and STEPI. All imaging gradients applied after the center of the third 90° RF pulse are rewound at the center of the fourth $90^{\circ}-_x$ pulse to rephase only the freshly recovered longitudinal magnetization back to the longitudinal direction. Δ indicates the separation between the 1st and 4th spoiler gradients, which is an important parameter to estimate the diffusion-weighting. Note that spatial encoding gradients are omitted for simplicity.



Figure 2.

Simulated T_1 values with respect to RF flip angle α without B_1 correction using eq. 4 with mixing times TM = 0.50, 0.75, 1, 1.25, 1.5 T₁ from top to bottom. 1.0 s was used for the true T_1 . The measured T_1^{meas} deviates further from the true value with smaller TM and larger difference in flip angle α from 90°. Vertical arrow indicates the measured $T_1 = \text{true } T_1$ with 90° pulses.



Figure 3.

 B_1 correction process: four averaged (a) SEPI₀ and (b) STEPI₀ images to calculate (c) B_1 correction map, which is used to correct singleshot (d) SEPI and (e) STEPI. (f, g) are T_1 maps before and after B_1 correction, respectively. T_1 values of water protons in MnCl₂ along the horizontal dotted lines are presented in plots (h) and (i) for before and after B_1 correction, respectively. T_1 map (g) and plot (i) clearly indicate substantial improvement by B_1 correction.

Figure 4.

 T_1 measurement before (\bullet, \bullet) and after (\circ, \Box) B_1 correction. The data in (a) are actually measured T_1 values, and those in (b) were normalized T_1 to 1.0 s to compare with the theoretical plots in Fig. 2. Dotted and broken lines indicate guidelines. Horizontal solid-lines represent T_1 using IR-TSE MRI.



Figure 5.

Plots of T_1 values of ROIs for each concentration of MnCl₂ with a 90° nominal flip angle and four-averaged B₁ correction map for different mixing times, TM = 0.1, 0.2, 0.4, and 0.8 second. Dotted and broken lines are the guidelines for each data set. Error bars represent the Standard Deviation of the means. For display purposes, TM for the data set of 0.1 mM is slightly shifted with respect to that of 0.2 mM to avoid overlap of the error bars.



Figure 6.

Plot of measured (\Box) and simulated (solid line) T₁ values with respect to b-factor. A T₁ of 0.95 s and diffusivity D of 2.14×10⁻³ mm²/s were used for the simulation. T₁ and D were independently measured using inversion-recovery turbo spin-echo and 2D ss-DWEPI, respectively. This plot indicates dramatic underestimation of T₁ by diffusion effect.



Figure 7.

(a) T_2 -weighted image, and T_1 maps using (b) IR-TSE and 2D ss-SESTEPI with signal averages of (c) 1, (d) 4, and (e) 16. ROIs are indicated by the square (white matter) and circles (gray matter). ROIs (1~4) and (5, 6) indicate white and gray matter, respectively



Figure 8.

Selected images of singleshot T_1 mapping using 2D ss-SESTEPI in a tumor bearing nude mouse: (a) T_1 maps, (b) ΔR_1 map, (c) T_1 weighted spin-echo images of various time points. The temporal resolution (TR) was 5 sec, with $1 \times 1 \text{ mm}^2$ resolution and 2 mm slice thickness. (d) Time curves of selected ROIs in (e) ΔR_1 maps in a tumor bearing nude mouse. Numbered labels in the plots indicate the 9-pixel (18 µL) ROIs on ΔR_1 maps; 1/kidney, 2/ tumor periphery, 3/tumor core, and 4/noise.

Table 1

 T_1 relaxation time of gray and white matter at different regions of interest shown in Figure 7. 1 avg, 4 avg and 16 avg indicate multiple repetition average for SEPI and STEPI obtained by 2D ss-SESTEPI.

	Doctor	ID TCF (mc)	2D s	s-SESTEPI	(ms)
	negion		1 avg	4 avg	16 avg
	1	722±33	799±58	721±41	728±20
White	2	716±26	727±68	726±30	722±20
matter	3	733±32	772±78	747±29	744±27
	4	685±33	742±89	725±40	715±27
Gray	2	1276±90	1149 ± 54	1101 ± 61	1116±81
matter	9	1177 ± 90	1069 ± 71	1087 ± 80	1088 ± 58

Table 2

Comparison of measured T_1 values with previously reported values for gray and white matter in the brain at 3T.

	White Matter (ms)	Gray Matter (ms)
2D ss-SESTEPI	727±12	1102±50
IR-TSE	714±21	1226±67
Literature (26)	832±10	1331±13
Literature (27)	1084±45	1820±114
Literature (28)	1110±45	1470±50
Literature (29)	933±15	1380±59