Emergence of *Escherichia coli* Sequence Type ST131 Carrying both the $bla_{\text{GES-5}}$ and $bla_{\text{CTX-M-15}}$ Genes

Juwon Kim, ¹ Seong Geun Hong, ² Il Kwon Bae, ¹ Ji Roung Kang, ¹ Seok Hoon Jeong, ¹* Wookeun Lee, ² and Kyungwon Lee

Department of Laboratory Medicine and Research Institute of Bacterial Resistance, Yonsei University College of Medicine, Seoul, Republic of Korea, and Department of Laboratory Medicine, CHA Bundang Medical Center, CHA University, Sungnam, Republic of Korea

Received 7 December 2010/Returned for modification 28 January 2011/Accepted 11 March 2011

Escherichia coli clinical isolate BD07372 of sequence type ST131 recovered from a bed sore specimen exhibited high-level resistance to ceftazidime and cefotaxime but exhibited susceptibility to imipenem and meropenem. The isolate harbored two β -lactamase genes, the $bla_{CTX-M-15}$ gene carried by an \sim 250-kbp plasmid carrying the FIA and FIC replicons and the bla_{GES-5} gene carried by a class 1 integron in the chromosome.

The class A carbapenemases, which have been identified in Gram-negative rods, involve chromosome-borne NMC-A and SME enzymes and plasmid-borne KPC, GES, and IMI enzymes (15). To date, 17 variants of GES-type extended-spectrum β -lactamases (ESBLs) have been identified (http://www.lahey.org/Studies/), but the carbapenem hydrolytic activity has been demonstrated only in some variants, including GES-2, GES-4, GES-5, GES-6, and GES-14, with a substitution at the Gly170 residue (4, 15). The $bla_{\rm GES-5}$ gene was first detected when carried by a plasmid of $Escherichia\ coli\ 365-02$ from Greece in 2004 (14). Then, detection of the $bla_{\rm GES-5}$ gene carried by $Escherichia\ coli\ 365-02$ from Greece in 2004 (14). Then, detection of the $Escherichia\ coli\ 365-02$ from Greece in 2004 (14). Then, detection of the $Escherichia\ coli\ 365-02$ from Creece in 2004 (14). Then, detection of the $Escherichia\ coli\ 365-02$ from Creece in 2004 (14). Then, detection of the $Escherichia\ coli\ 365-02$ from Creece in 2004 (14). Then, detection of the $Escherichia\ coli\ 365-02$ from Creece in 2004 (14). Then, detection of the $Escherichia\ coli\ 365-02$ from Creece in 2004 (14). Then, detection of the $Escherichia\ coli\ 365-02$ from Creece in 2004 (14). Then, detection of the $Escherichia\ coli\ 365-02$ from Creece in 2004 (14).

E. coli clinical isolate BD07372 was recovered from a bed sore sampled on 17 May 2007 from a 79-year-old female who has home nursing care. She had been hospitalized for 35 days at a secondary-care hospital in Bundang, Republic of Korea, for the treatment of chronic renal failure, type II diabetes mellitus, and hypertension. Multilocus sequence typing was performed on the isolate using the seven conserved house-keeping genes (adk, fumC, gyrB, icd, mdh, purA, and recA) following use of the protocols available at the E. coli MLST database website (http://mlst.ucc.ie/mlst/dbs/Ecoli), and the isolate was identified as sequence type ST131 (allelic profile, 53-40-47-13-36-28-29).

A disk diffusion assay was performed (5), and the isolate exhibited resistance to ampicillin, ampicillin-sulbactam, piper-acillin-tazobactam, cefoxitin, aztreonam, ceftazidime, cefotaxime, cefepime, gentamicin, tobramycin, amikacin, and ciprofloxacin and was susceptible to imipenem, meropenem, and trimethoprim-sulfamethoxazole. Synergy was observed between the amoxicillin-clavulanic acid (20/10-µg) disk and the ceftazidime (30-µg), cefotaxime (30-µg), cefepime (30-µg),

and aztreonam (30- μ g) disks (Becton Dickinson, Spark, MD) in double-disk synergy tests, indicating the production of ESBL (12). Agar dilution MIC testing on Muller-Hinton agar (Difco Laboratories, Detroit, MI) with an inoculum of 10⁴ CFU per spot confirmed high-level MICs of ceftazidime (>256 μ g/ml) and cefotaxime (>256 μ g/ml) and low-level MICs of imipenem (0.5 μ g/ml) and meropenem (0.25 μ g/ml) for the isolate BD07372 (5). Clavulanic acid (Sigma, St. Louis, MO) at a fixed concentration of 4 μ g/ml lowered the MICs of ceftazidime and cefotaxime to 1 μ g/ml and 8 μ g/ml, respectively (Table 1).

Isolate BD07372 showed a positive result in the Hodge test, which was performed using a MacConkey agar plate with an ertapenem disk, indicating the production of a carbapenemase (7). However, the isolate showed a negative result in the EDTA-based double disk synergy test, which was performed using a Mueller-Hinton agar plate with an imipenem disk and a disk containing EDTA (750g) plus sodium mercaptoacetic acid (2 mg), indicating that the isolate does not produce metallo- β -lactamases (8). In the isoelectric focusing analysis, three β -lactamase activities with isoelectric points of 5.8, 6.1, and 8.6, corresponding to GES-5, OXA-17, and CTX-M-15, respectively, were detected.

PCR and sequencing experiments were performed to detect genes encoding CTX-M-, GES-, PER-, SHV-, TEM-, and VEB-type ESBLs as previously described (11). Isolate BD07372 carried two β -lactamase genes, the $\mathit{bla}_{\text{CTX-M-}15}$ and the bla_{GES-5} genes. The isolate transferred cefotaxime resistance to the E. coli J53 azide-resistant recipient in mating experiments (3), in which transconjugants were selected on MacConkey agar (Difco Laboratories, Detroit, MI) plates supplemented with cefotaxime (2 µg/ml) and sodium azide (100 µg/ml). The location of the β-lactamase genes was identified by the hybridization of I-CeuI-digested genomic DNA or S1 nuclease-treated linearized plasmids with probes specific for the β-lactamase genes, various replicons of plasmids, and 16S rRNA genes as described previously (2, 9). The probe specific for the $bla_{\text{CTX-M-}15}$ gene hybridized with an \sim 250-kbp plasmid in both isolate BD07372 and its transconjugant, but that specific for the bla_{GES-5} gene did not. The ~250-kbp plasmid carried two types of replicon, FIA and FIC. Replicon sequence

^{*} Corresponding author. Mailing address: Department of Laboratory Medicine and Research Institute of Bacterial Resistance, Yonsei University College of Medicine, 120-752, 134 Shinchon-Dong, Seodaemun-Gu, Seoul, Republic of Korea. Phone: 82-2-2228-2448. Fax: 82-2-313-0956. E-mail: kscpjsh@yuhs.ac.

[▽] Published ahead of print on 28 March 2011.

TABLE 1. MICs of *E. coli* wild-type strain BD07372, its transconjugant, and the recipient J53

	E. coli MIC (μg/ml)		
Antimicrobial agent ^a	Wild-type strain BD07372 ^b	Transconjugant trcBD07372 ^c	Recipient J53
Amoxicillin-clavulanic acid	256	256	6
Piperacillin-tazobactam	>256	>256	1.5
Cefoxitin	128	96	12
Ceftazidime	>256	16	0.25
Ceftazidime-clavulanic acid	8	0.5	0.12
Cefotaxime	>256	32	0.06
Cefotaxime-clavulanic acid	1	0.06	0.06
Cefepime	>256	16	0.06
Aztreonam	64	64	0.25
Imipenem	0.5	0.25	0.19
Imipenem-clavulanic acid	0.25	0.125	0.125
Meropenem	0.25	0.06	0.06
Meropenem-clavulanic acid	0.06	0.06	0.06
Ertapenem	1.5	1	0.023
Ertapenem-clavulanic acid	0.25	0.25	0.023

a Clavulanic acid was added at a fixed concentration of 4 μg/ml.

typing was performed on the plasmid as previously described (13), and the sequence types of the replicons were identified to be FIA (1) and FIC (1). This result is in agreement with the previous study, which has confirmed that all the variants of ST131 producing CTX-M-15 ESBL harbored the corresponding gene on the IncF-type plasmid (6). The probe specific for the $bla_{\rm GES-5}$ gene hybridized with an I-CeuI macrorestriction fragment of \sim 270 kbp carried by isolate BD07372. The probe specific for 16S rRNA genes also hybridized with the I-CeuI macrorestriction fragment, indicating the chromosomal location of the $bla_{\rm GES-5}$ gene in the isolate.

To investigate genetic environments surrounding the $bla_{\rm GES-5}$ gene, several overlapping PCR fragments obtained from whole DNA of the E.~coli isolate were sequenced with primers corresponding to the internal region of the class 1 integron as previously described (1). Identical to the $bla_{\rm GES-5}$ gene carried by a plasmid of K.~pneumoniae CHAK36, the $bla_{\rm GES-5}$ gene was carried by a class 1 integron as a gene cassette downstream of the attI1 recombination site. The integron had three unique gene cassettes $[bla_{\rm GES-5}$ –aac(6')-IIa– $bla_{\rm OXA-17}/orf4$] between the two conserved elements, the 5'-conserved segment (5'-CS) and the 3'-CS, but the 59-base element of the third gene cassette was interrupted by a putative transposase gene, orf4.

In summary, E. coli isolate BD07372 of ST131 harbored two

β-lactamase genes, the $bla_{\rm CTX-M-15}$ gene carried by an ~250-kbp plasmid carrying the FIA and FIC replicons and the $bla_{\rm GES-5}$ gene carried by a class 1 integron in a chromosome. Recently, the intercontinental dissemination of an $E.\ coli$ clone of ST131 producing CTX-M-15, which was carried mostly by IncF plasmids, has been reported (10). Emergence of an $E.\ coli$ clone of ST131 simultaneously producing CTX-M-15 and GES-5 could be a serious threat to global health.

This study was supported by the Basic Science Research Program through the National Research Foundation of Korea (NRF), funded by the Ministry of Education, Science and Technology (grant 2009-0071195).

REFERENCES

- Bae, I. K., et al. 2007. Genetic and biochemical characterization of GES-5, an extended-spectrum class A β-lactamase from Klebsiella pneumoniae. Diagn. Microbiol. Infect. Dis. 58:465–468.
- Barton, B. M., G. P. Harding, and A. J. Zuccarelli. 1995. A general method for detecting and sizing large plasmids. Anal. Biochem. 226:235–240.
- Bermudes, H., et al. 1999. Molecular characterization of TEM-59 (IRT-17), a novel inhibitor-resistant TEM-derived β-lactamase in a clinical isolate of Klebsiella oxytoca. Antimicrob. Agents Chemother. 43:1657–1661.
- Bogaerts, P., et al. 2010. GES extended-spectrum β-lactamases in Acinetobacter baumannii isolates in Belgium. Antimicrob. Agents Chemother. 54: 4872–4878.
- Clinical and Laboratory Standards Institute. 2009. Performance standards for antimicrobial susceptibility testing; 19th informational supplement. M100-S19. Clinical and Laboratory Standards Institute, Wayne, PA.
- Dhanji, H., et al. 2011. Molecular epidemiology of fluoroquinolone-resistant ST131 Escherichia coli producing CTX-M extended-spectrum β-lactamases in nursing homes in Belfast, UK. J. Antimicrob. Chemother. 66:297–303.
- Lee, K., et al. 2010. Improved performance of the modified Hodge test with MacConkey agar for screening carbapenemase-producing Gram-negative bacilli. J. Microbiol. Methods 83:149–152.
- Lee, K., Y. S. Lim, D. Yong, J. H. Yum, and Y. Chong. 2003. Evaluation of the Hodge test and the imipenem-EDTA double-disk synergy test for differentiating metallo-β-lactamase-producing isolates of *Pseudomonas* spp. and *Acinetobacter* spp. J. Clin. Microbiol. 41:4623–4629.
- Mammeri, H., L. Poirel, N. Mangeney, and P. Nordmann. 2003. Chromosomal integration of a cephalosporinase gene from *Acinetobacter baumannii* into *Oligella urethralis* as a source of acquired resistance to β-lactams. Antimicrob. Agents Chemother. 47:1536–1542.
- Oteo, J., M. Pérez-Vázquez, and J. Campos. 2010. Extended-spectrum β-lactamase producing *Escherichia coli*: changing epidemiology and clinical impact. Curr. Opin. Infect. Dis. 23:320–326.
- 11. Ryoo, N. H., et al. 2005. Dissemination of SHV-12 and CTX-M-type extended-spectrum β-lactamases among clinical isolates of *Escherichia coli* and *Klebsiella pneumoniae* and emergence of GES-3 in Korea. J. Antimicrob. Chemother. 56:698–702.
- Sirot, D. L., et al. 1992. Resistance to cefotaxime and seven other β-lactams in members of the family Enterobacteriaceae: a 3-year survey in France. Antimicrob. Agents Chemother. 36:1677–1681.
- Villa, L., A. García-Fernández, D. Fortini, and A. Carattoli. 2010. Replicon sequence typing of IncF plasmids carrying virulence and resistance determinants. J. Antimicrob. Chemother. 65:2518–2529.
- Vourli, S., et al. 2004. Novel GES/IBC extended-spectrum β-lactamase variants with carbapenemase activity in clinical enterobacteria. FEMS Microbiol. Lett. 15:209–213.
- Walther-Rasmussen, J., and N. Høiby. 2007. Class A carbapenemases. J. Antimicrob. Chemother. 60:470–482.

^b Carries bla_{GES-5} and bla_{CTX-M-15}.

^c Carries bla_{CTX-M-15}.