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Reduction of dorsolateral prefrontal cortex gray matter in late-life depression

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Abstract

Postmortem studies have documented abnormalities in the dorsolateral prefrontal cortex (dlPFC) in depressed subjects. In this study we used magnetic resonance imaging to test for dlPFC volume differences between older depressed and non-depressed individuals. Eighty-eight subjects meeting DSM IV criteria for major depressive disorder and thirty-five control subjects completed clinical evaluations and cranial 3T magnetic resonance imaging. After tissue types were identified using an automated segmentation process, the dlPFC was measured in both hemispheres using manual delineation based on anatomical landmarks. Depressed subjects had significantly lower gray matter in left and right dorsolateral prefrontal cortex (standardized to cerebral parenchyma) after controlling for age and sex. Our study confirmed the reduction of dorsolateral prefrontal cortex in elderly depressed subjects, especially in the gray matter. These regional abnormalities may be associated with psychopathological changes in late-life depression.

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Keywords

magnetic resonance imaging; elderly; mood

1. Introduction

Several studies have examined the effect of aging on prefrontal brain volumes in healthy subjects (Raz et al., 1997; Raz et al., 2004; Walhovd et al., 2005) but few studies have focused on the volumetric differences of dorsolateral prefrontal cortex (dlPFC) in geriatric depression. The dlPFC, comprised of the superior and middle frontal gyri (Crespo-Facorro et al., 2000), is an important region of the prefrontal cortex which receives projections from higher order association regions (Nauta, 1971). It projects to the dorsolateral head of the caudate nucleus, continues to the lateral dorsomedial globus pallidus and, finally, to the ventral anterior and mediodorsal thalamus (Tekin and Cummings, 2002). Previous studies have shown that the dlPFC has extensive connectivity to cortical and subcortical circuits that may underlie its importance in modulating mood regulation and cognitive function (Duffy and Campbell, 1994). Prior research has shown that abnormalities in the dlPFC are implicated in the pathology of late-life depression (Thomas et al., 2000; Thomas et al., 2003; Taylor et al., 2004). However, a recent meta-analysis examining functional magnetic resonance imaging (MRI) studies of depression revealed no consistent pattern of abnormalities in dlPFC activity, a result that may be due to methodologic variability, clinical heterogeneity of the depressive subjects or different imaging paradigms (Fitzgerald et al., 2006).

Most anatomic studies of the dlPFC in depression have been performed postmortem. Studies have reported marked reduction in the density and size of neurons and glial cells in both supra- and infragranular layers (Rajkowska et al., 1999) as well as a decrease in pyramidal neuronal size in the overall cortex (Khundakar et al., 2009). Thomas et al. have shown that, compared with control subjects, depressed individuals exhibit higher ischemia-induced inflammation in gray and white matter of the dlPFC (Thomas et al., 2000; Thomas et al., 2002), providing evidence for the “vascular depression” hypothesis linking cerebrovascular disease with late-life depression (Alexopoulos et al., 1997; Krishnan et al., 1997). In younger adult populations, brain imaging studies have reported that unipolar depressed subjects, compared with healthy controls, have clusters of decreased gray matter density (T. Frodl et al., 2010) as well as decreased dlPFC glucose metabolism and blood flow (Ketter et al., 1996; Drevets, 1998). Antidepressant treatment appears to increase metabolic activity in the middle frontal gyrus in depressive subjects (Buchsbaum et al., 1997) and to normalize metabolism in the prefrontal cortex (Brody et al., 2001) while remission is associated with less decline in dlPFC gray matter (T. S. Frodl et al., 2008). Reduced dlPFC activity has been linked to severity of illness (Drevets, 1998) and to cognitive disturbance (Bench et al., 1992; Bench et al., 1993). However, there is limited *in vivo* evidence and no large-sample studies of dlPFC volume in late-life depression.

In this study we evaluated dlPFC volume in subjects with late-life depression compared with a group of never-depressed elderly control subjects. We hypothesized that older subjects with depression would have smaller dlPFC volumes than older comparison subjects.

2. Methods

2.1. Participants

This cross-sectional project occurred within a larger longitudinal clinical study of depression in older adults, the Conte Center for the Neuroscience of Depression and Neurocognitive

Outcomes of Depression in the Elderly (NCODE) study. Eighty-eight depressed subjects and 35 control subjects were examined in this study. The study was approved by Duke University Medical Center's Institutional Review Board. All enrolled subjects were 60 years or older. Depressed subjects were recruited from the Duke University Medical Center psychiatric inpatient and outpatient services and met criteria for DSM-IV major depressive episode. Age of onset of current episode was not limited to a specific age nor was there a requirement that subjects have no prior episodes of depression. Exclusion criteria included 1) another major psychiatric illness, such as bipolar disorder, schizophrenia, and schizoaffective disorder; 2) history of alcohol or drug abuse or dependence; 3) primary neurologic illness, such as dementia, stroke, Parkinson disease, seizure disorder, and multiple sclerosis; 4) medications, medical illness, or physical disability that may severely affect cognitive function and ability to provide consent; 5) physical disability which precludes cognitive testing; and 6) metal in the body which precludes MRI. Comparison subjects were required to have a non-focal neurological examination, self-report of no current or past neurological or depressive illness, and no evidence of a depression diagnosis based on the Diagnostic Interview Schedule portion of the Duke Depression Evaluation Schedule.

Subjects were assessed with a Mini-Mental State Examination (MMSE) (Folstein et al., 1975) at baseline. All control subjects had MMSE scores of 25 or higher. If a depressed subject had an MMSE score less than 25, subjects were followed through an acute (eight-week) phase of treatment to determine if cognition improved. Subjects whose MMSE scores remained below 25 were not followed longitudinally. Thus, in the clinical judgment of the study geriatric psychiatrist and by established NCODE protocol, dementia was effectively excluded at or close to baseline in all elderly depressed NCODE subjects.

2.2. Depression treatment

Depressed participants received individualized treatment from a psychiatrist, who followed them throughout the study. Most received antidepressant medication; some received electroconvulsive treatment (ECT) or psychotherapy.

2.3. MRI scanning protocol

Cranial MRI was performed using the 8-channel parallel imaging head coil on a 3 Tesla whole-body MRI system (Trio, Siemens Medical Systems, Malvern, PA). Proton density (PD), T1-weighted, T2-weighted, and fluid-attenuated inversion recovery (FLAIR) images were acquired. Parallel imaging was employed with an acceleration factor of 2.

The T1-weighted image set was acquired using a 3D axial TURBOFLASH sequence with TR/TE=22/7 msec, flip angle=25°, a 100 Hz/pixel bandwidth, a 256×256 matrix, a 256 mm diameter field-of-view, 160 slices with a 1 mm slice thickness and Nex=1 (no signal averaging), yielding an image with 1 mm cubic voxels in an 8 minute, 18 second imaging time. This was followed by a T2-weighted acquisition using a 2D turbo spin-echo pulse sequence with TR/TE=7580/86 msec, turbo factor=7, a 210 Hz/pixel bandwidth, a 256×256 matrix, a 256 mm diameter field-of-view, 100 slices with a 1.5 mm slice thickness and Nex=1 (no signal averaging), yielding a 1×1×1.5 mm voxel in a 5 minute, 21 second imaging time. Next a PD weighted volume was acquired with a 2D turbo spin-echo pulse sequence with TR/TE=7580/17 msec, turbo factor=3, a 210 Hz/pixel bandwidth, a 256×256 matrix, a 256 mm diameter field-of-view, 100 slices with a 1.5 mm slice thickness and Nex=1 (no signal averaging), yielding a 1×1×1.5 mm voxel in a 6 minute, 21 second imaging time. Finally, a FLAIR image was acquired with TR/TI/TE=9000/2400/101 msec, a 210 Hz/pixel bandwidth, a turbo factor of 11, a 256×256 matrix, a 256 mm diameter field-

of-view, 75 slices with a 2 mm slice thickness and Nex=1 (no signal averaging), yielding a 1×1×2 mm voxel in a 9 minute, 45 second imaging time.

2.4. Whole brain segmentation

The MR images were transferred to the Duke Neuropsychiatric Imaging Research Laboratory (NIRL) where all image analyses were performed. Images were initially resliced to a common geometry of 1×1×1.5mm voxels. An automated 4-channel lesion segmentation, which takes advantage of FLAIR images for lesion detection, was performed to assess gray matter, white matter, cerebrospinal fluid, and white matter lesions. The algorithm used was a variation on the fully automated Expectation Maximization Segmentation (EMS) method (Van Leemput et al., 1999; Van Leemput et al., 2001; Van Leemput et al., 2003). The method was optimized for vascular lesion assessment in elderly subjects. The software assigns a probability that a given pixel should be classified as gray matter, white matter, cerebrospinal fluid, lesion or non-brain in the following manner. First, images are aligned to a set of tissue probability images using the mutual information registration tool (MIRIT) (Maes et al., 1997). The probability atlas provides spatial priors for each tissue that are used to initialize the tissue intensity histograms for the segmentation algorithm. The tissue probabilities are then derived in an iterative process using the intensity distributions of the different tissues for each of the input image contrasts. The process also evaluates and compensates for spatial distributions of intensity that could be due to various magnetic resonance imaging artifacts such as radiofrequency inhomogeneity (bias correction). Lesions are detected as ‘outliers’ to the normal tissue distributions. This method requires parameter optimization for each dataset due to variations in subject populations and scanners. After identifying the optimal parameters for a dataset, the method is fully automated. This automation provides an advantage over semi-automated methods which require an analyst to choose seeding points and lesion regions. The method is capable of distinguishing and classifying lesions and other brain tissues simultaneously.

2.5. Volumetry of dIPFC

For this project, the dIPFC was defined as consisting of Superior Frontal Gyrus (SFG) and Middle Frontal Gyrus (MFG). The tracing procedures were modified from Crespo-Facorro et al. (Crespo-Facorro et al., 2000). Tracing was performed with the ITK-SNAP 1.4.1 program (Yushkevich et al., 2006). The dIPFC tracing mask was applied to the automated segmentation in order to obtain volumes for left and right hemisphere gray and white matter in the dIPFC.

Superior Cingulate Sulcus (SCiS) and Superior Rostral Sulcus (SRS) were defined as the medio-inferior border, Inferior Frontal Sulcus (IFS) as the latero-inferior border, Central Sulcus (CS) as the medial border, and PreCentral Sulcus (PCS) as the posterior border. See Table 3 for full list of abbreviations. Tracing was separated into 4 steps, described below.

Step 1 Trace SFG in coronal view (Fig. 1). SFG tracing began on the coronal slice identified as Plane A. Plane A (Fig. 2) is a coronal plane passing through the anterior extent of the inner surface of the genu (corpus callosum). Plane A serves as the posterior boundary for coronal tracing of SFG. The superior frontal sulcus (SFS) was followed superiorly, tracing around brain cortex medially to the midline, then inferio-laterally into superior cingulate sulcus (SCiS). The sagittal view was used to identify the SCiS as the medial-inferior boundary of SFG. While tracing SFG anteriorly, tracing continued superiorly to the SCiS or Cingulate Sulcus (CiS) and excluded superior cingulate gyrus/cingulate gyrus. When the SCiS/CiS descended inferiorly, there was a transition to the SRS as the new medial-inferior boundary of SFG. Tracing continued anteriorly until the end of the SRS.

- Step 2** Trace SFG in axial view. To include SFG posterior to Plane A slice, tracing was done in the axial view. Plane A serves as the anterior boundary for axial tracing of SFG. The SFS is the lateral boundary of SFG. On an axial slice with the most posterior point of SFS, a straight, horizontal line was drawn to midline CSF, designating the posterior boundary. The midline is the medial boundary and plane A is the anterior boundary. Tracing continued slice by slice superiorly to include superior portions of SFG. Once the superior-most extent of SFG was reached, tracing continued inferiorly from the starting axial slice using the same guidelines as above. Once SFS retreated anteriorly beyond Plane A, SFG tracing was stopped.
- Step 3** Trace MFG in coronal view (Fig. 3). Plane A serves as the posterior boundary for coronal tracing of MFG. In the sagittal view, the IFS was identified on a slice where the Sylvian fissure is clearly presented. The IFS, which serves as the lateral and inferior boundary of MFG, is the most superior sulcus parallel (anterior-posterior orientation) to the Sylvian Fissure. Trace was done laterally in the coronal view from the IFS then superiorly around MFG to the deepest SFS point, where a straight line was drawn connecting the deepest SFS point to the deepest IFS point. Tracing continued anteriorly on each slice until the IFS branch/FrontoMarginal Sulcus (FMS) disappeared.
- Step 4** Trace MFG in axial view. MFG tracing was completed in the axial view starting on the slice on which, due to the coronal tracing, MFG appears lateral to SFG. Plane A serves as the anterior boundary for axial tracing of MFG. Tracing began from the deepest point of the PCS, continued laterally and anteriorly around MFG to the deepest point of SFS, and then back to the PCS. Tracing of each axial slice was reviewed on the sagittal view to confirm exclusion of IFG. Tracing was discontinued when one of the following conditions was met: 1) all cortex above IFS on sagittal view was included or 2) IFG connected to MFG.

2.5.1. Reliability—Reliability was established by repeat measurements on multiple MR scans ($N=5$) by two raters (SCY, DFM). Intraclass correlation coefficients (ICCs) for dIPFC were as follows: left gray matter = 0.90, left white matter = 0.97, right gray matter = 0.87, and right white matter = 0.98.

2.6. Statistical analyses

SAS version 9.2 (Cary, NC, USA) was used to analyze data. Initial comparisons testing for differences of raw means and frequencies using T-test and chi-square statistics were performed (data were described as mean [SD] or percentage [n]). Linear regression models were then used to compare dIPFC volumes standardized to cerebral parenchyma between depressed and control groups while controlling for age and sex.

3. Results

This study included 88 depressed subjects (28 males and 60 females) and 35 comparison subjects (11 males and 24 females). As shown in Table 1, the sex, race, and MMSE scores of the depressive and comparison subjects were not statistically different. However, depressed subjects were significantly younger than controls ($t=4.58$, $P<0.0001$). Age ranges were 60–84 years for depressives, and 62–90 years for comparison subjects.

We examined the clinical characteristics of the depressed subjects. At enrollment, 91% of subjects reported recurrent depression (>1 episode during lifetime), with an average age of

onset (initial episode) of 35.7 years ($SD=20.7$), and an average illness duration of 18.1 years ($SD=29.3$). At time of MRI, 92% of the depressed subjects had taken an antidepressant and 9% had received ECT (these data were unavailable for $N=20$ subjects); data were not collected on the number receiving psychotherapy.

In bivariate analyses, depressed subjects did not significantly differ from control subjects on total dIPFC volume, total brain parenchyma, or standardized dIPFC volumes (see Table 1). However, in multivariable models adjusting for age and sex, standardized gray matter volumes were significantly smaller in both left and right dIPFC (see Table 2).

Given the difference in age between depressed and comparison subjects, dIPFC models were run separately for each group. Age had a significant negative effect upon left gray matter for both depressed ($F_{1,86}=4.85$, $p=0.030$) and comparison subjects ($F_{1,33}=7.86$, $p=0.008$), but was significant for right gray matter only among comparison subjects ($F_{1,33}=10.56$, $p=0.003$). Age was marginally significant for right gray matter in depressives ($F_{1,86}=2.90$, $p=0.09$). Also, an age by depression group interaction term was evaluated and determined to be non-significant for all dIPFC variables.

Lastly, the dIPFC models were re-analyzed with the addition of MMSE score, to determine if cognitive status may have mediated the gray matter/depression relationship. Depression had a significant but lessened effect on left dIPFC gray matter ($F_{1,100}=2.02$, $p=0.0461$) but was now marginally significant for right gray matter ($F_{1,100}=3.86$, $p=0.0523$). In the left dIPFC model only, MMSE was significantly positively associated with gray matter volume ($F_{1,100}=2.59$, $p=0.0113$).

4. Discussion

The principal finding of this study is that older depressed individuals exhibited smaller gray matter volumes in both the left and right dIPFC, after adjusting for age and sex. These associations may be partially mediated by cognitive status. As expected, age was negatively related to dIPFC gray matter volume; age-related decline did not appear to differ between groups. Our findings contribute to the growing evidence that the dIPFC is critically involved in depression. To our knowledge, this is the first *in vivo* magnetic resonance imaging study to demonstrate smaller dIPFC gray matter volumes in late-life depression.

Major depressive disorder is characterized by disruptions in executive control, linked to abnormal dIPFC function. The dIPFC plays an important role in working memory and other aspects of executive function (Braver et al., 1997; D'Esposito et al., 2000). Previous studies have focused on younger populations; *in vivo* morphologic change of dIPFC in late-life depression has not been investigated. Of the studies in younger subjects, one postmortem neuropathological report demonstrated structural changes in the prefrontal regions in major depressive disorder subjects, including decreases in cortical thickness and neural size, together with reductions in neural and glial density (Rajkowska et al., 1999). In the study by Thomas et al., ischemia in the white matter of dIPFC was found in subjects with late-life depression (Thomas et al., 2003) and lends support to the "vascular depression" hypothesis (Alexopoulos et al., 1997; Krishnan et al., 1997). However the role of gray matter was not examined.

Our findings are consistent with studies linking the dIPFC with depression. In our previous study, we speculated that microstructural changes in white matter of the dIPFC may result in disconnection of cortical and subcortical regions (Taylor et al., 2004), while another study found that gray matter shrinkage was also involved in depression (Vasic et al., 2008). Functional imaging studies have been employed to identify brain areas involved in depression. Results from these studies associate depression with abnormally high levels of

ventromedial activity (Drevets et al., 1992; Biver et al., 1994; Greicius et al., 2007) but abnormally low levels of dlPFC activity (Biver et al., 1994; Galynker et al., 1998). Our study expanded those findings to a larger sample and found that the dlPFC is critically involved in elderly depression, particularly the gray matter. Moreover, this relationship is linked to global cognition as measured by the MMSE, which supports past work linking geriatric depression with cognitive deficits, specifically executive dysfunction.

Separate examination of gray and white matter represents a methodological improvement over examining only the whole volume of a given brain structure. This is particularly important given that gray and white matter are composed of different cell types. White matter mostly consists of a variety of glial cells that provide myelination, support, and maintenance of the neurochemical environment. It is also where axonal projections between nerve bodies pass. Gray matter includes glia, but also nerve cell bodies, which project to other brain regions. The changes we found are likely to be localized primarily in cell bodies (neurons or glia) in the dlPFC gray matter.

Limitations of this study include potential bias given that control subjects were significantly older than depressed subjects. However, this difference would serve to create a bias against our hypothesis. Multivariable models did control for age which should have at least partially adjusted for this bias. In addition, an age by group interaction was examined and found to be non-significant. Other factors not included in this analysis, such as age of onset, illness duration, ECT, and prior antidepressant use, may influence the morphology of dlPFC in depression. Regarding the possible effects of treatment (antidepressants and ECT) upon dlPFC volume, a number of factors prevented their evaluation in these analyses, including missing data, heterogeneity of treatments, modest sample size, and use of polypharmacy. In addition, and perhaps most critical, was a lack of data on medication history. Since the majority of subjects had long-term depression (average episode duration = 18.1 years), they were likely to have taken antidepressants prior to study baseline; this prior medication use would likely have had a greater influence on brain structure than would concurrent medications, if any relationship exists. Finally, use of the MMSE provides limited information, particularly as this tool does not allow for a thorough examination of executive function, which is linked to geriatric depression. These factors should be addressed in future studies.

Our finding adds to the growing evidence that the dlPFC contributes to the neuroanatomic circuit related to mood regulation and depression. This is supportive of the proposed neuroanatomical models of mood regulation that involve prominent fronto-subcortical circuits (prefrontal cortex, amygdala, thalamus and basal ganglia)(Drevets et al., 1992). In addition to this cross-sectional study, longitudinal research is needed to determine the relationship between these changes and the pathogenesis of depression as well as the mechanisms by which these changes could predispose to depression.

We have shown that geriatric depressed subjects have significantly smaller dlPFC gray matter volumes than normal controls, supporting a role for dlPFC in the pathophysiology of late-life depression.

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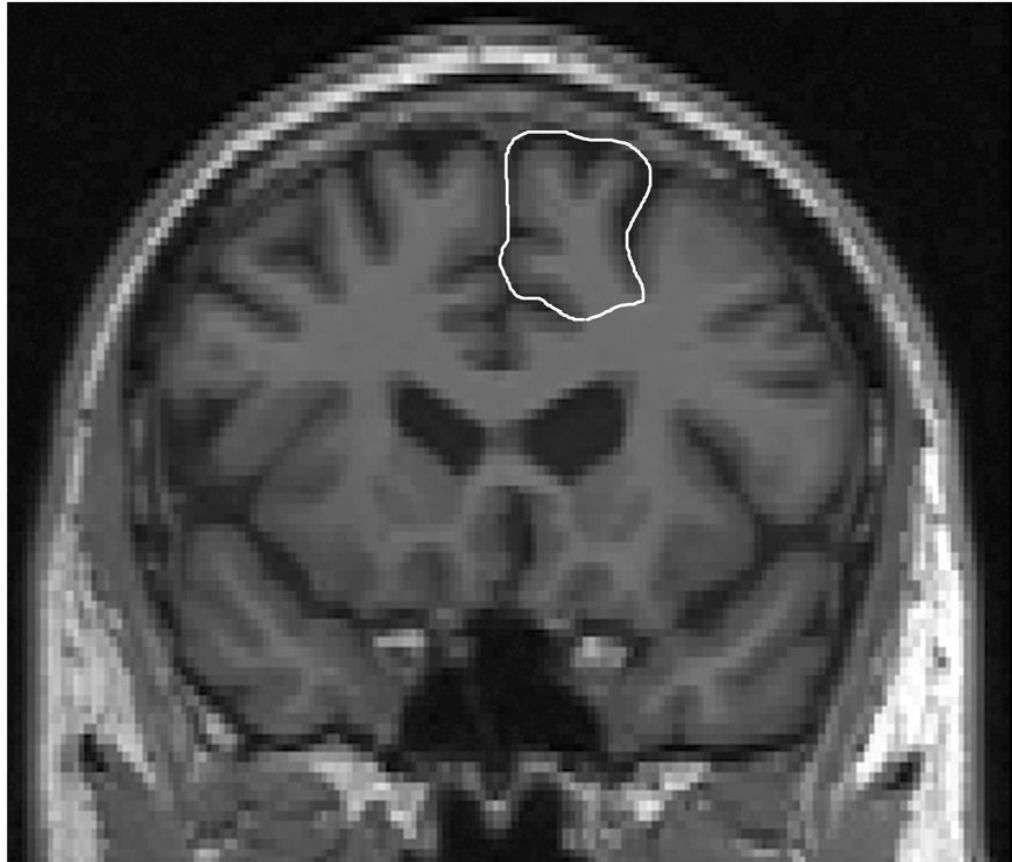


Figure 1.
Superior frontal gyrus (SFG) tracing in coronal view.

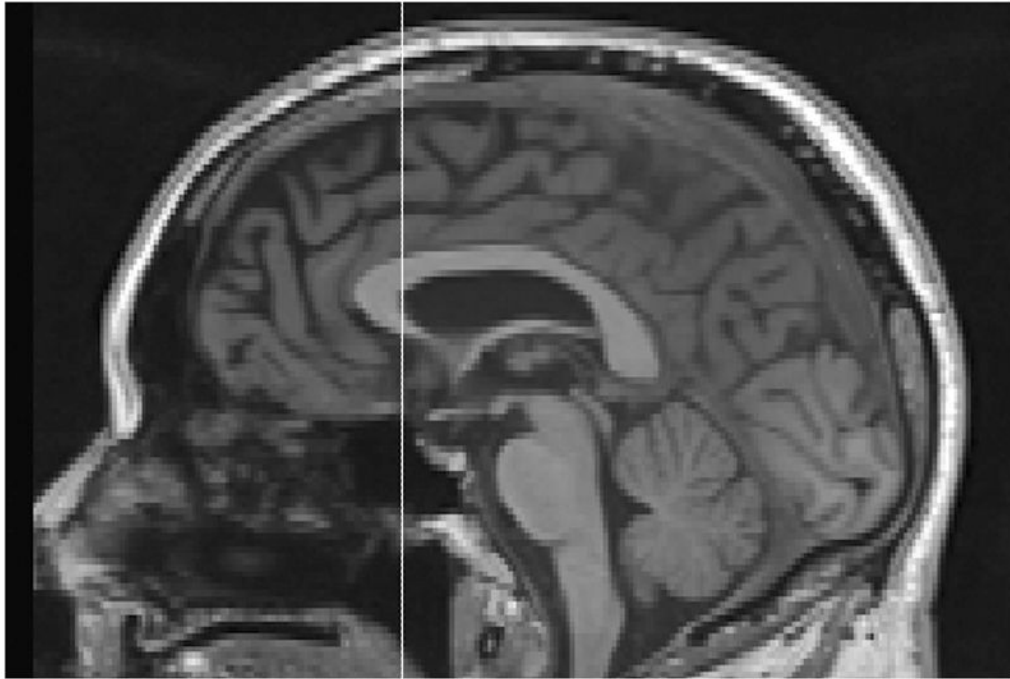


Figure 2.
Plane A is anterior extent of inner surface of genu (corpus callosum) in sagittal view.

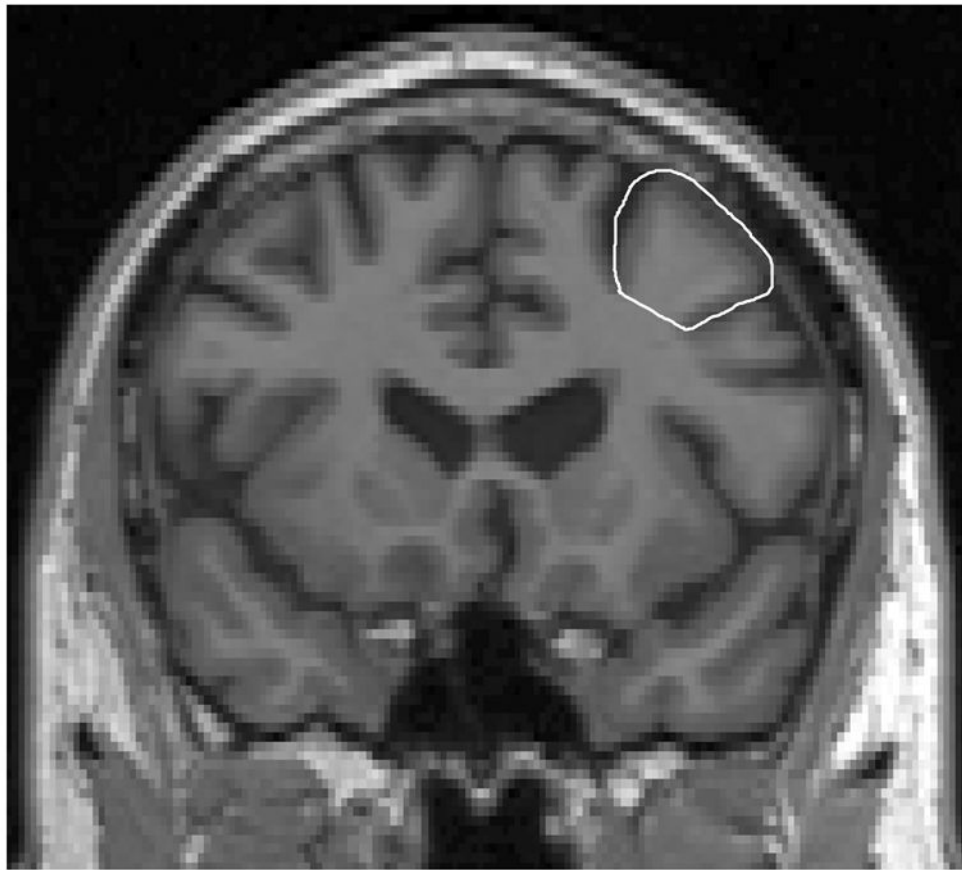


Figure 3.
Middle frontal gyrus (MFG) tracing in coronal view.

Table 1Sociodemographic and imaging characteristics^a

Variable	Patients (N = 88) mean (SD)	Controls (N = 35) mean (SD)	Test Statistic
Age	69.1 (5.7)	74.5 (6.5)	$t = 4.58, df = 121, P < 0.0001$
Sex			
% Female (N)	68.2 (60)	68.6 (24)	chi-square = 0.0018, $df = 1, P = 0.97$
Race			
% Caucasian (N)	83.0 (73)	91.4 (32)	chi-square = 1.44, $df = 1, P = 0.2302$
MMSE Score ^b	28.6 (2.3)	29.1(1.1)	$t = 1.48, df = 102, P = 0.14$
Total Parenchyma (cerebrum, ml)	925.6 (100.1)	941.1 (93.6)	$t = 0.79, df = 121, P = 0.43$
Left dlPFC			
Total Volume (ml)	49.7 (8.5)	52.0 (6.9)	$t = 1.43, df = 121, P = 0.16$
Gray Matter (ml)	28.7 (5.9)	30.2 (4.8)	$t = 1.33, df = 121, P = 0.19$
White Matter (ml)	20.9 (5.2)	21.7 (4.7)	$t = 0.83, df = 121, P = 0.41$
Right dlPFC			
Total Volume (ml)	50.4 (10.1)	52.0 (8.7)	$t = 0.86, df = 121, P = 0.39$
Gray Matter (ml)	28.8 (6.3)	30.4 (5.4)	$t = 1.35, df = 121, P = 0.18$
White Matter (ml)	21.4 (6.0)	21.5 (5.6)	$t = 0.05, df = 121, P = 0.96$

^aN=123^bMini-Mental State Examination; N=105

Table 2

Linear regression model of dorsolateral prefrontal cortex (dlPFC) volume in depressed subjects and non-depressed comparison subjects

Region	Depressed group, <i>N</i> = 88 Mean (SD)	Control group, <i>N</i> = 35 Mean (SD)	<i>F</i> _{1,119} value	<i>P</i> value
Standardized Left				
dlPFC ¹				
Total Volume	0.0530 (0.0067)	0.0549 (0.0067)	1.85	0.18
Gray Matter	0.0306 (0.0048)	0.0329 (0.0048)	5.87	0.0169
White Matter	0.0224 (0.0051)	0.0220 (0.0051)	0.23	0.63
Standardized Right				
dlPFC ¹				
Total Volume	0.0538 (0.0088)	0.0551 (0.0088)	0.57	0.45
Gray Matter	0.0307 (0.0054)	0.0332 (0.0054)	5.06	0.0263
White Matter	0.0230 (0.0062)	0.0219 (0.0062)	0.75	0.39

¹ Standardized volume is proportion to parenchyma (gray + white matter of cerebrum).

All models adjusted for age and sex.

Table 3

Abbreviations

Abbreviation	Anatomical Definition
CS	Central Sulcus
Cis	Cingulate Sulcus
dIPFC	Dorsolateral Prefrontal Cortex
FMS	FrontoMarginal Sulcus
IFG	Inferior Frontal Gyrus
IFS	Inferior Frontal Sulcus
MFG	Middle Frontal Gyrus
PCS	Pre-Central Sulcus
SciS	Superior Cingulate Sulcus
SFG	Superior Frontal Gyrus
SFS	Superior Frontal Sulcus
SRS	Superior Rostral Sulcus