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## Dynamics of Muscle Microcirculatory and Blood-myocyte O<sub>2</sub> Flux During Contractions

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### Abstract

The O<sub>2</sub> requirements of contracting skeletal muscle may increase 100-fold above rest. In 1919 August Krogh's brilliant insights recognized the capillary as the principal site for this increased blood-myocyte O<sub>2</sub> flux. Based on the premise that most capillaries did not sustain RBC flux at rest Krogh proposed that capillary recruitment (i.e., initiation of red blood cell (RBC) flux in previously non-flowing capillaries) increased the capillary surface area available for O<sub>2</sub> flux and reduced mean capillary-to-mitochondrial diffusion distances. More modern experimental approaches reveal that most muscle capillaries may support RBC flux at rest. Thus, rather than contraction-induced capillary recruitment *per se*, increased RBC flux and hematocrit within already-flowing capillaries likely elevate perfusive and diffusive O<sub>2</sub> conductances and hence blood-myocyte O<sub>2</sub> flux. Additional surface area for O<sub>2</sub> exchange is recruited but, crucially, this may occur along the length of already-flowing capillaries (i.e. longitudinal recruitment). Today, the capillary is still considered the principal site for O<sub>2</sub> and substrate delivery to contracting skeletal muscle. Indeed, the presence of very low intramyocyte O<sub>2</sub> partial pressures (PO<sub>2</sub>'s) and the absence of PO<sub>2</sub> gradients, whilst refuting the relevance of diffusion distances, place an even greater importance on capillary hemodynamics. This emergent picture calls for a paradigm-shift in our understanding of the function of capillaries by de-emphasizing *de novo* 'capillary recruitment.' Diseases such as heart failure impair blood-myocyte O<sub>2</sub> flux, in part, by decreasing the proportion of RBC-flowing capillaries. Knowledge of capillary function in healthy muscle is requisite for identification of pathology and efficient design of therapeutic treatments.

### Keywords

blood flow; capillary; exercise; microcirculation; oxygen uptake; skeletal muscle

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### Outline

This brief review considers Krogh's theory of capillary recruitment and consequent reduction of capillary-to-mitochondrial diffusion distances in the light of subsequent empirical observations. Specifically, novel combination of intravital microscopy and microvascular O<sub>2</sub> measurements indicate that capillary recruitment is not obligatory for rapid blood-myocyte O<sub>2</sub> uptake (VO<sub>2</sub>) kinetics following the onset of contractions. Not only is there compelling evidence supporting that most capillaries sustain RBC flux in resting muscle, but intramyocyte PO<sub>2</sub>'s during exercise/contractions and measurements of muscle O<sub>2</sub> diffusing capacity suggest that diffusion distances may not be limiting for capillary-mitochondrial O<sub>2</sub> flux. In the light of these observations the question is posed as to how muscle perfusive and diffusive O<sub>2</sub> conductances increase many-fold from rest to contractions and a working hypothesis is proposed. Within this context the mechanistic connection between impaired muscle VO<sub>2</sub> (i.e., blood-myocyte) kinetics and microcirculatory dysfunction in aging and chronic disease is explored briefly.

### Conflict of interest

The authors have no conflict of interest.

## Introduction

August Krogh was not the first to discover the presence of capillaries, observe red blood cells (RBCs) passing through them or recognize the role of oxygen in the respiration that occurred in peripheral tissue. That story begins possibly with Galen (c. 130–199 CE) who considered that blood passed from the right to the left ventricle through minute pores in the interventricular septum and the notion of these ‘pores of Galen’ was still held by Sir William Harvey (1578–1657) in his seminal work, *De Motu Cordis*, published in 1628 (cited in West, 1996). It was Marcello Malpighi (1628–1694) in 1661 (*Duae Epistolae Pulmonibus*) who was perhaps the first person to observe the motion of red blood cells (RBCs) in capillaries in the frog lung, but, because of their lack of color, he thought that they were fat globules. Later, Anton Loewenhoeck (1632–1723) would recognize RBCs flowing in capillaries for what they were. The discovery of molecular O<sub>2</sub> has been attributed to Joseph Priestley (1733–1804), Carl Wilhelm Scheele (1742–1786) and the chemist Antoine Lavoisier (1743–1794) who named it “*oxygine*” for its acid-forming properties and studied its rate of use and CO<sub>2</sub> production during exercise. Also, well before Krogh’s time, Gustav Magnus (1802–1870) had developed blood gas analyses which, together with von Helmholtz’s (1821–1894) measurements of heat production by single twitches of isolated muscle in 1848 (von Helmholtz, 1848; see also Bassett, 2001), had helped ‘move’ the site of respiration (i.e.,  $\dot{V}O_2$  leading to CO<sub>2</sub> production) from the lungs to the peripheral tissues (rev. West, 1996; Jones and Poole, 2005).

In the early 1900’s Krogh’s brilliance, and the unique ability to measure respiratory gas concentrations accurately (to one thousandth of a percent, Krogh, 1920a), combined with his wife, Dr. Marie Krogh’s, measurements of pulmonary diffusing capacity (Krogh, 1915) was central to their proposing that gas exchange across the blood-gas barrier in the lung occurred solely by means of physical forces, specifically, diffusion. In this regard he courageously opposed his mentor Christian Bohr, the eminent British physiologist John Scott Haldane and other scientific luminaries of the day. The concept of O<sub>2</sub> diffusion across biological barriers, applied to muscle, emplaced the capillaries center-stage in the processes of blood-muscle O<sub>2</sub> flux and muscle energetics (Krogh, 1919ab, 1920b); the rest of the vascular system was thus relegated to a largely supportive role (though see Pittman, 2010 for a review of arteriolar-tissue O<sub>2</sub> fluxes). Once that was accomplished his painstaking measurements of muscle capillary bed structure and function together with his understanding of metabolism would lay the foundation for his and Ehrlang’s elegant mathematical models of muscle oxygenation that remain so widely accepted today. Sentinel among the precepts of these models are the notions that: 1. Intramuscular PO<sub>2</sub> decreases systematically with increased distance from the RBC in the capillary. Thus, large intramyocyte PO<sub>2</sub> gradients were hypothesized to exist producing anoxic loci and denying mitochondria within those loci the opportunity to contribute to ATP production. 2. The importance of diffusion distances, together with Krogh’s measurements of more “open” capillaries either during or after muscle contraction, supported the theory that, at low metabolic rates (i.e., rest), most capillaries did not support RBC flux and that they were therefore available to be “recruited” (defined as the initiation of RBC flux in previously non-flowing capillaries) and reduce intramyocyte diffusion distances during contractions. Key to this recruitment during contractions was Krogh’s belief in the presence of a contractile capability that could close-off individual capillaries.

Krogh himself recognized many problems with his techniques (Krogh, 1919ab). Paramount amongst which were: 1. The extreme surgical interventions often necessary to achieve good perfusions (Krogh, 1919ab, 1920). Specifically, tying off multiple vessels to prevent leakage, non-physiologically high perfusion pressures, and sometimes, waiting several days post-mortem to achieve good perfusion. 2. The clumping together of India ink particles that

blocked ink entry to some capillaries. This clumping effect would be reduced at higher flow rates and may artifactually have increased the opportunity for more capillaries to contain ink, and therefore be considered to support RBC flux, during the hyperemic conditions present during or after contractions.

Thanks, in large part, to the stimulus provided by Krogh and his contemporaries in this field, the nine intervening decades since his 1919 papers and Nobel prize (1920), have witnessed impressive technological advances in our ability to study the microcirculation. These advances have focused acute scientific attention on capillary hemodynamics, vascular and muscle oxygenation and blood-myocyte  $O_2$  flux. For instance: a range of skeletal muscles (e.g., rat: spinotrapezius, cremaster, diaphragm, extensor digitorum longus; hamster: retractor, cremaster, sartorius; rabbit: tenuissimus; cat: sartorius) suitable for transmitted and incident light microscopy has been developed and techniques are available for determining microvascular (phosphorescence quenching, Rumsey et al. 1988; Poole et al. 1995; Golub and Pittman, 2002, 2008; Golub et al. 2007; Pittman et al. 2010) and muscle (hemoglobin + myoglobin, Near-infrared Spectroscopy, NIRS, Lutjemeier et al. 2008) oxygenation with high temporal and spatial (phosphorescence quenching) fidelity. In addition, cryomicrospectrophotometric techniques (Gayeski et al. 1985; Gayeski and Honig, 1986ab; Gayeski et al. 1987; Honig et al. 1997; Voter and Gayeski, 1999) and Proton Nuclear Magnetic Resonance Spectroscopy (MRS, Richardson et al. 1995, 1998; Molè et al. 1999) have permitted *ex-vivo* and *in vivo* determination of intramyocyte  $PO_2$ . Finally, methods have also been developed for resolving  $O_2$  flux across (constant-infusion thermodilution, Andersen and Saltin, 1985; Rowell et al. 1986; Savard et al. 1988; Grassi et al. 1996; Roca et al. 1992; Richardson et al. 1993; Volianitis et al. 2003; vascular isolation, Hogan et al. 1989) and within (radiolabeled microspheres, Laughlin et al. 1982; Armstrong and Laughlin, 1983; Musch and Terrell, 1992; Musch et al. 2004; Bailey et al. 2000; Poole et al. 2000) contracting muscles and the muscle microcirculation (Behnke et al. 2002; Kindig et al. 2002).

This paper highlights how observations made using these latter techniques have challenged our understanding of the microcirculatory function and its role in matching  $O_2$  delivery ( $\dot{Q}O_2$ )-to- $\dot{V}O_2$  during muscle contractions in health and disease. Specifically, under the auspices of resolving the physiological behavior of microcirculatory hemodynamics and  $O_2$  flux in healthy contracting muscle and dysfunction in disease, the necessity of capillary recruitment and the presence of intramyocyte  $PO_2$  gradients will be examined critically. As epitomized by Krogh, intravital microscopy observations of the capillary bed take center stage and are supported by powerful modern technologies which provide crucial new insights into the control of intravascular and intramyocyte  $O_2$  pressures and fluxes within contracting skeletal muscle.

## Is capillary recruitment obligatory for increased blood-myocyte $O_2$ flux in contracting muscle?

In the pentobarbital-anesthetized rat spinotrapezius muscle at rest  $\geq 80\%$  of capillaries support RBC flux at any given time (Kindig and Poole, 1998, 2001; Kindig et al. 1999, 2002; Poole et al. 1997). This is true also for the rat diaphragm (Kindig and Poole, 1998), extensor digitorum longus (Anderson et al. 1997; Ellis et al. 2002), hamster sartorius and cremaster (Damon and Duling, 1984), rabbit tenuissimus (Vrieling et al. 1987), and cat sartorius (Burton and Johnson, 1972). When stimulated to contract at 1 Hz, spinotrapezius capillary RBC velocity and flux begin to increase within the first contraction cycle (i.e., 1 s) generally reaching steady-state values within 30–60 s (Figure 1, top panel, Kindig et al. 2002). With most capillaries “recruited” at rest there appears little room for additional capillaries to initiate flow and indeed no significant increase in % flowing was found. Given

that this result departs from conventional wisdom and established dogma it is crucial to consider whether this response is accompanied by an appropriate increase in blood-myocyte  $O_2$  flux. Combining phosphorescence-quenching measurements of microvascular  $PO_2$  ( $PmvO_2$ ) (Figure 1, center panel) with those of RBC flux the blood-myocyte  $\dot{V}O_2$  kinetics profile can be resolved with high temporal and spatial fidelity (Figure 1, bottom panel, Behnke et al. 2001, 2002). The exponential increase in muscle  $\dot{V}O_2$  is initiated from the onset of contractions – as expected from the instantaneous change in high-energy phosphates and current theories of metabolic control – with no discernible delay and kinetics (time constant,  $\tau$ , 20–30 s) resembling closely that found for *in vivo* cycling (human leg muscles, Grassi et al. 1996; Bangsbo et al. 2000; Grassi, 2001; Krstrup et al. 2009) *in situ* (dog gastrocnemius-plantaris complex, Grassi et al. 2002) and isolated single fibers (amphibian lumbrical muscle, Kindig et al. 2003; Poole et al. 2007).

In Krogh's time, the prevailing wisdom was that "...capillaries are passive, (and) that blood is flowing continuously through all of them at rates which are determined by the state of contraction or dilatation of the corresponding arterioles..." (Krogh, 1919b, p. 468). Thus, Krogh's observations and contention that capillary recruitment was an essential physiological response to increased metabolic rates flew contrary to this opinion. Today, the argument is by no means settled with exemplars of each side listed in Table 1. The commonality of temporal capillary hemodynamic and  $O_2$  flux responses across species and preparations and the common observation that, in resting muscle, most capillaries support RBC flux, do not prove, in and of themselves, that *in vivo* capillary recruitment does not occur; they merely suggest that it may not be obligatory for the normal  $\dot{V}O_2$  kinetics. Considered by many as the oldest scientific society in the World, the motto of the British Royal Society, which was founded in 1660 and chartered by King Charles II, is "*Nullius in Verba*" (Take nobody's word for it, see it for yourself). This view was certainly espoused by Krogh but, unfortunately, and to the consternation of many scientists who use intravital microscopy to visualize capillary hemodynamics (right hand side of Table 1), it has become convenient and all-too-common to avoid the rigors of observing the capillary bed *in vivo* and simply presume that capillary recruitment can explain increased blood-myocyte exchange (e.g., Baron et al. 2000; Clark et al. 2001; Rattigan et al. 1997; left hand side of Table 1). Indeed, the eminent microcirculationist, Professor Eugene Renkin, in his Letter to the Editor of the American Journal of Physiology, harshly criticized Dr. Bentzer for entitling his paper "*Capillary filtration coefficient is independent of the number of perfused capillaries in cat skeletal muscle*" (Bentzer et al. 2001); when that claim was made without making any direct measurements of RBC flowing capillary numbers.

Notwithstanding the above, several concerns have justifiably been leveled at anesthetized, electrically-stimulated preparations, which are generally necessary to see muscle capillary hemodynamics, and interpretation of data from such. Paramount among these are: 1. *Anesthesia relaxes arteriolar smooth muscle and increases capillary perfusion.* If this occurred it would be expected that resting muscle blood flow and vascular conductance would increase markedly above those found in the conscious animal. Comparison of blood flows measured under both conditions using radiolabeled microspheres indicates that this certainly does not occur (c.f. Musch and Poole, 1996 with Bailey et al. 2000). Moreover, and even more compelling, if arteriolar smooth muscle function were impaired by anesthesia, it is difficult to conceive how electrical stimulation would increase muscle blood flow in proportion to metabolic rate as seen *in vivo*. Figure 2 demonstrates the commonality of the slope of the blood flow to  $\dot{V}O_2$  relationship across different muscles in anesthetized rats (Ferreira et al. 2006) to that found in conscious man (Andersen and Saltin, 1985; Rowell et al. 1986; Savard et al. 1988; Richardson et al. 1993; Grassi et al. 1996; Poole, 1997; Volianitis et al. 2003). Compelling additional supportive evidence for the presence of flow in most capillaries in resting muscle comes from capillary staining using central Thioflavin-

S dye infusions in conscious animals (Kayar and Banchero, 1985). The presence of at least plasma flow was detected in essentially all muscle capillaries investigated within very few seconds of instillation (~1–2 circulation times). *2. Surgical manipulation damages vascular control and increases muscle capillary perfusion.* Comparison of *in vivo*, *in situ* and exteriorized muscles demonstrates that blood flow is not increased by the surgical exteriorization procedure nor is the blood flow-to- $\dot{V}O_2$  relationship perturbed during contractions as evidenced by the constancy of the  $P_{mv}O_2$  profile across *in situ* and exteriorized conditions (Bailey et al. 2000). *3. Capillaries without flowing RBCs cannot be detected and so, in resting muscle, many capillaries simply are not counted.* Measurements of the lineal density of flowing capillaries from rest to contractions and also in pharmacologically-vasodilated conditions do not support the presence of undetected capillaries at rest (Kindig and Poole, 2001).

Another important consideration that can be tested empirically in conscious humans performing voluntary exercise relates to muscle hemoglobin concentration. It is known that, due in part to the endothelial surface layer (often termed the ‘glycocalyx’), there are differential flow rates of plasma and RBCs within and among capillaries such that microvascular hematocrit in resting muscle is far below systemic (e.g., Klitzman and Duling, 1979; Sarelius and Duling, 1982; Desjardins and Duling, 1987, 1990; Kindig and Poole, 1998, 2001; Poole et al. 1997; Frisbee and Barclay, 1998; Kindig et al. 2002; Copp et al. 2009). Thus, from rest to exercise an increased microvascular hematocrit towards systemic (Klitzman and Duling, 1979; Kindig et al. 2002; Copp et al. 2009) may therefore, in theory, raise muscle [hemoglobin] as much as 2–3 fold. Accordingly, if we were to accept that most (say 80%, a figure in the range of common usage) of capillaries did not flow at rest and were recruited during exercise, muscle [hemoglobin] would be expected to increase up to four-fold from this effect alone, and with increased microvascular hematocrit, this could be several-fold higher again. However, in human muscles NIRS reveals only a very modest increase in muscle [hemoglobin] of ~20% from rest to exercise (Lutjemeier et al. 2008) which leaves very little room for significant capillary recruitment.

From these studies and a wealth of intravital microscopy observations in the many different muscles (listed above) there is substantial support for the contention that, at least for the muscles investigated, the majority of capillaries sustain plasma and RBC flux even at rest. Very few capillaries are closed, empty of RBCs or contain stagnant RBCs. However, there is substantial heterogeneity of flow rates among capillaries (Erickson and Myrhaug, 1972; Duling and Damon, 1987; Kindig et al. 1999; Kindig and Poole, 1998) such that many may be of less relative importance for  $O_2$  and substrate delivery. Thus, muscle contractions (and reactive hyperemia, Burton and Johnson, 1972) are accompanied by an increased flow principally within already flowing capillaries (Poole et al. 2008b; right-hand side of Table 1).

Despite the wealth of evidence presented above, the notion of capillary recruitment endures and is commonly invoked to explain increased blood-myocyte  $O_2$  and substrate flux during exercise or metabolic stimulation by insulin, for example (Baron et al. 2000; Rattigan et al. 1997; Clark et al. 2001; Vincent et al. 2006; Clark et al. 2008; left-hand side of Table 1). Often, established scientists, who may have never observed the capillary bed in their experiments, interpret their data as evidence for capillary recruitment or, as a self-fulfilling prophecy, explain their results using this phenomenon. In physiology textbooks (e.g., Rowell, 1993; McArdle et al. 2007) and medical schools around the World the notion that  $O_2$  diffusion distances within muscle are important and elevated blood-myocyte  $O_2$  and substrate fluxes occur by increasing the number of flowing capillaries makes great intuitive sense. Who can forget the visually stunning fluorescence images from Parthasarathi and Lipowsky (1999; Figure 3) purportedly showing hypoxic intervention “recruiting” an

enormous number of microvascular units? As the legend explains and scientific reports have substantiated (e.g. Damon and Duling, 1984) what this may really demonstrate is the well-known phenomenon of hyperoxic vasoconstriction – both panels representing supraphysiologic PO<sub>2</sub>'s (Whalen et al. 1974, 1976)

Each experimental observation must be judged on its own merits. However, there are many putative experimental considerations that are often ignored within the capillary recruitment literature and which may have lead to erroneous conclusions. These include: 1. Today capillaries in mammalian skeletal muscles are not thought to have contractile capability as considered by Krogh (1919ab) and his contemporaries. Neither do they collapse easily under conditions of increased muscle or reduced intraluminal pressure (Poole and Mathieu-Costello, 1990; Kindig and Poole, 1999), possibly due to the presence of Type IV collagen struts that tether their walls to the surrounding myocytes (Caulfield and Borg, 1979; Borg and Caulfield, 1980). This is very different from capillaries in the lung where their thin walls and alveolar gas surround offer little support and, at least under low cardiac output conditions, many may be collapsed - particularly at the lung apex where perfusion pressures are perilously low (Pande and Hughes, 1983) - and thus they are available for recruitment when pulmonary arterial flow increases and perfusion pressure rises (Wearn et al. 1934; Hanson et al. 1989; Wagner, 1997). 2. Capillaries are enormously fragile vessels. Mechanical abuse during muscle surgical preparation – an unfortunately frequent accompaniment to intravital observation – may easily damage their ability to support flow. Also, stretching muscle to sarcomere lengths >3.0 μm narrows their diameter and reduces the proportion supporting flow by increasing their resistance (Poole and Mathieu-Costello, 1992; Kindig and Poole, 2001) as well as invoking a sympathetically-mediated arteriolar vasoconstriction and passively impeding arteriolar and venular flow (Welsh and Segal, 1996; Kindig and Poole, 2001). Few laboratories measure and set muscle sarcomere length and, because stretching thins the muscle and improves the optical quality, this practice may, unfortunately, be common. 3. Some anesthetics lower blood pressure to levels that may invoke skeletal muscle hypoperfusion due to lack of driving pressure which can be exacerbated by a sympathetically-mediated muscle vasoconstriction. 4. The commonly accepted notion that hypoxia leads to capillary recruitment, meaning that sufficient capillaries must be stagnant in resting muscle to elicit this recruitment effect, should be placed in proper context. For example, as mentioned above, the “low O<sub>2</sub>” condition (~35 mmHg incident PO<sub>2</sub>) studied by Pathasarathi and Lipowsky (1999, see Figure 3) was actually *hyperoxic* compared with values established for resting muscle (normoxia ~17 mmHg, Whalen et al. 1974, 1976). It might be argued that what their micrographs demonstrate beautifully may not be hypoxia-induced capillary recruitment but, rather, the opposite - hyperoxia-induced vasoconstriction leading to capillary derecruitment as the incident PO<sub>2</sub> is raised from 35 to 130 mmHg.

Given that the human skeletal musculature may contain as many as eight or nine billion capillaries, is it implausible that most may contain at least some flowing RBCs at rest? Figure 4 addresses this concept mathematically using an accepted estimate of 1 L.min<sup>-1</sup> for skeletal muscle blood flow (Rowell, 1993, ~5.4 trillion RBC.min<sup>-1</sup>) and presumes a high average of 300 capillaries per mm<sup>2</sup> each of 1,000 μm in length (Andersen and Henriksson, 1977; Brodal et al. 1977; Saltin and Gollnick, 1983; Hoppeler et al. 1985; Coyle et al. 1988; Richardson et al. 1994). If 80% of these capillaries support RBC flux there are sufficient RBCs to supply each with 12.8 RBC.s<sup>-1</sup>. This value is not far from that measured for rat spinotrapezius muscle (i.e., 15–20 RBC.s<sup>-1</sup>, Kindig and Poole, 1997; Kindig et al. 2002). Whereas this calculation does not prove that most capillaries support RBC flux in resting muscle – and thus are not available to be ‘recruited’ *per se* – it demonstrates that, at least, it is feasible.

## Are there pronounced PO<sub>2</sub> gradients within contracting muscle fibers?

If the greatest impediment to O<sub>2</sub> diffusion occurs close to the RBC and not within the intramyocyte milieu the importance of capillary-to-mitochondrial diffusion distance becomes moot. In the 1980's Carl Honig, Thomas Gayski and Richard Connett rapidly froze resting and contracting (electrical stimulation) dog gracilis and other muscles and microscopically analyzed the myoglobin-O<sub>2</sub> saturation from which PO<sub>2</sub> could be calculated (Gayski et al. 1985, 1987; Gayski and Honig, 1986ab; Honig et al. 1997). Surprisingly, during exercise they found that the intramyocyte PO<sub>2</sub> was low (< 3 mmHg) and almost uniform irrespective of the distance from the capillary i.e., the predicted capillary-to-mitochondrial gradients (see Ellis et al. 1983) were absent (Honig et al. 1997). Despite initial uncertainty regarding the size of the myoglobin crystals analyzed and whether the freezing procedure was capable of capturing a PO<sub>2</sub> gradient (Voter and Gayski, 1995; Honig et al. 1997) at least two independent series of experiments using different techniques support that intramyocyte PO<sub>2</sub> is very low and that diffusion distance is not an important determinant of muscle O<sub>2</sub> diffusing capacity. First, using highly specialized proton MRS techniques, Richardson and colleagues (1995, 1998; see also Molè et al. 1999) found that, even at relatively low levels of exercise, quadriceps intramyocyte PO<sub>2</sub> fell close to the levels reported by Honig and colleagues (Gayski et al. 1985, 1987; Gayski and Honig, 1986ab; Honig et al. 1997) in the dog gracilis. Second, in a very elegant experimental design intercapillary diffusion distances in the canine gastrocnemius-plantaris complex were altered chronically by leg immobilization or exercise training in separate groups of dogs (Bebout et al. 1993; Hepple et al. 2000). The results demonstrated that muscle diffusing capacity could be dissociated clearly from capillary density and intercapillary diffusion distances (Figure 5). Collectively, these studies support the elegant modeling papers of Federspiel and Popel (1986) and Groebe and Thews (1990) that muscle O<sub>2</sub> diffusing capacity is determined not by the capillary density or intramyocyte O<sub>2</sub> diffusion distances *per se* but by the number of RBCs adjacent to the muscle fibers at any instant. Thus, whereas muscle blood flow and O<sub>2</sub> content define convective O<sub>2</sub> delivery, the length and hematocrit of RBC-flowing capillaries are sentinel determinants of muscle diffusive O<sub>2</sub> delivery and, as such, may be manipulated by perturbations that alter systemic and thus (presumably) capillary hematocrit (Schaffartzik et al. 1993). This concept has been verified empirically in frog skin (Malvin and Wood, 1992).

## Presumptions arising from “capillary recruitment”

The concept of “capillary recruitment” has birthed multiple presumptions (identified in *italics* below) that conflict with established physiological concepts, empirical observations and current models. These include: 1. *Muscle  $\dot{V}O_2$  at rest and during submaximal contractions is O<sub>2</sub> delivery limited* (Clark et al. 1998, 2001). In healthy muscle at rest and during contractions  $\dot{V}O_2$  is known to be controlled by intramyocyte high-energy phosphate signaling (Meyer and Foley, 1996) and, other than close to maximal  $\dot{V}O_2$  (Richardson et al. 1993; Wagner et al. 1997; Poole, 1997), is not limited by O<sub>2</sub> supply. A wealth of measurements in conscious man performing voluntary exercise and animal preparations dissociate both the non-steady-state  $\dot{V}O_2$  kinetics and steady-state  $\dot{V}O_2$  from changes in O<sub>2</sub> supply (Bredle et al. 1989; Arthur et al. 1992; Hogan et al. 1992; Knight et al. 1993; Curtis et al. 1995; Richardson et al. 1995a; Grassi et al. 1998; Grassi, 2001). 2. *The exchange capacity of a capillary is determined principally by whether it is flowing (recruited) or not rather than its rate of flow*. As seen in Figure 6 increased exchange capacity purported to result from capillary recruitment can be explained by increased flow and substrate delivery through already-recruited capillaries (Hargreaves et al. 1990; Hudlicka et al. 1982; Kindig et al. 2002). This latter notion is in accord with the enormous increase in RBC flux and O<sub>2</sub> delivery seen from rest to exercise within already flowing capillaries and also the established

wisdom that free fatty acids and glucose are taken up by muscle in proportion to their delivery (i.e., plasma flow  $\times$  concentration). 3. *When the microcirculation in diseased muscles (e.g., in chronic heart failure (CHF) or diabetes) is observed, this may not be distinguishable from the healthy situation.* One danger of accepting that most capillaries do not flow in resting muscle is that it is likely to impair the ability to resolve the mechanistic bases for pathologically-reduced muscle function. For instance, in CHF capillaries cease to flow in resting and exercising muscle in proportion to left ventricular dysfunction and left ventricular end-diastolic pressure (Kindig et al. 1999; Richardson et al. 2003), and this might explain, in part, the reduced muscle O<sub>2</sub> diffusing capacity and compromised energetic function pathognomonic to this condition. Similarly, in Type II diabetes the Goto-Kakizaki rat spinotrapezius evidences widespread microvascular stagnation and impaired capillary hemodynamics that undoubtedly compromise the opportunity for blood-muscle glucose as well as O<sub>2</sub> flux (Padilla et al. 2006, 2007; Bauer et al. 2007). If one accepts that most capillaries should not support RBC flux there will be a failure to recognize these and other pathological dysfunctions (e.g., eccentric exercise-induced muscle damage, Kano et al. 2005) which will likely hamstring attempts to correct the underlying problems therapeutically.

### **If capillary recruitment does not occur to any great extent, how does blood-myocyte O<sub>2</sub> flux increase during exercise?**

As discussed above, the idea that a recruited capillary represents a fixed unit of surface area or exchange capacity appears untenable. Several likely and one speculative mechanism(s) are presented in Figure 7 that may help explain the enhanced blood-myocyte O<sub>2</sub> flux in response to elevated muscle metabolic demands which may rise by two orders of magnitude above resting. These mechanisms are consistent with contemporary experimental findings and blood-tissue O<sub>2</sub> exchange theory. Note that all panels of Figure 7 may not apply to predominantly plasma- as opposed to RBC-borne substrates as muscle uptake of, for example, free fatty acids or glucose is not dependent upon capillary RBC hemodynamics but rather plasma flow. Key elements in Figure 7 are: 1. Capillaries with very low RBC flow and hence long transit times at rest may have dramatically increased flow during exercise and therefore assume a greater importance for convective O<sub>2</sub> delivery. Interestingly, fractional O<sub>2</sub> extraction is impacted by the interdependence of O<sub>2</sub> diffusing capacity (DO<sub>2</sub>) and blood flow according to (Roca et al. 1992; see also following section for graphical analysis):

$$\text{Fractional O}_2 \text{ extraction} = \dot{V}O_2 / \dot{Q}O_2 = \dot{Q}O_2 (1 - e^{-DO_2/\beta\dot{Q}})$$

where  $\dot{Q}O_2$  is O<sub>2</sub> delivery (blood flow,  $\dot{Q} \times$  [arterial O<sub>2</sub>]) and  $\beta$  is the slope of the O<sub>2</sub> dissociation curve in the physiologically-relevant range. 2. Capillary hematocrit increases from ~15% towards systemic levels (~45%, Klitzman and Duling, 1979; Kindig et al. 2002; Copp et al. 2009) such that, in the extreme, each capillary may contain up to 2–3 times as many RBCs during exercise compared with rest thereby increasing the instantaneous O<sub>2</sub> diffusive conductance (DO<sub>2</sub>, Federspiel and Popel, 1986; Groebe and Thews, 1990; Malvin and Wood, 1992). An elevated capillary hematocrit increases the likelihood that transverse histological sections will transect an RBC within a given capillary. This observation, combined with the presumption that only capillaries with transected RBCs supported RBC flow *in vivo*, has contributed to the enduring capillary recruitment concept (Honig et al. 1980). Krogh himself was cognizant of the dangers of presuming capillary hemodynamic behavior from *ex-vivo* analyses (Krogh, 1919ab). 3. As considered for the lung by Wiltz Wagner (Wagner, 1997) capillary endothelial surface area available for O<sub>2</sub> delivery can be



recruited not only by initiating flow in previously non-flowing vessels but also by increasing the length of individual capillaries over which blood-tissue O<sub>2</sub> flux occurs. Thus, when RBC velocity is speeded and the fractional extraction of O<sub>2</sub> increases (from ~25 up to 80% or more) from rest to exercise, a far greater length of a given capillary is “recruited” for that exchange. The potential importance of this “longitudinal recruitment” of capillary O<sub>2</sub> exchange capacity to increased muscle O<sub>2</sub> diffusing capacity during exercise cannot be overemphasized. 4. As presented above, measurements in voluntarily contracting human muscle (Richardson et al. 1995b,1998; Molè et al. 1999) and frozen sections of electrically-stimulated canine muscle (Gayeski et al. 1985,1987; Gayeski and Honig, 1986ab; Honig et al. 1997) indicate the presence of extremely low intramyocyte PO<sub>2</sub>'s. The desaturation of myoglobin will enhance its ability to facilitate intramyocyte diffusion and remove what Honig and colleagues have insightfully termed the “functional O<sub>2</sub> carrier-depleted region” between the capillary and mitochondrial reticulum (Honig et al. 1997). 5. It is recognized that the endothelial surface layer (‘glycocalyx’) is a crucial regulator of capillary hemodynamics (e.g., Desjardins and Duling, 1990; Zurbier et al. 2005). Specifically, its removal (Desjardins and Duling, 1990) or modification by hyperglycemia, for example (Zurbier et al. 2005), alters capillary hematocrit and hemodynamics profoundly. There is thus the strong potential that mechanical alterations of the endothelial surface layer accompanying muscle contraction and exercise hyperemia are important in regulating the capillary hemodynamic response to exercise and thus blood-myocyte O<sub>2</sub> flux: though, at present, this remains hypothetical.

### **How does the matching of O<sub>2</sub> delivery to $\dot{V}O_2$ change in disease (chronic heart failure, CHF, diabetes) and aging and might this help explain the slowed $\dot{V}O_2$ kinetics in these conditions?**

The compelling weight of evidence indicates that, in skeletal muscles of young healthy individuals, the speed of  $\dot{V}O_2$  kinetics (expressed, most often, as the time constant,  $\tau$ , of the response) following the onset of moderate or heavy exercise, is not limited by muscle O<sub>2</sub> delivery (Grassi, 2005; Hughson, 2005; Poole et al. 2008a; Nyberg et al. 2010, Figure 8). In marked contrast,  $\dot{V}O_2$  kinetics are slowed in many disease conditions such as Type II diabetes (Regensteiner et al. 1998) and heart failure (CHF, Meakins and Long, 1927; Sietsema et al. 1986, 1994; Hepple et al. 1999; rev. Poole et al. 2005) as well as by advanced age (Babcock et al. 1994; Chilibeck et al. 1998; Barstow and Scheuermann, 2005), and there is evidence that this slowing occurs consequent to O<sub>2</sub> delivery limitation as schematized in Figure 8. Intravital microscopy of spinotrapezius muscles from Type II diabetic (Padilla et al. 2006), CHF (Kindig et al. 1999; Richardson et al. 2003) and aged (Russell et al. 2003; Poole and Ferreira, 2007; Copp et al. 2009) animals offers mechanistic insights into how deficits in microvascular function contribute to perfusive and diffusive impediments in the O<sub>2</sub> transport pathway. One expression of these impediments is the mismatching of O<sub>2</sub> delivery-to- $\dot{V}O_2$  which often produces aberrant  $P_{mv}O_2$  profiles particularly during the first 30–60 seconds following the onset of contractions and during recovery (Figure 9, Diederich et al. 2002; Behnke et al. 2004; McDonough et al. 2004; Poole et al. 2005; Ferreira et al. 2006; Behnke et al. 2007; Poole and Musch, 2010; Hirai et al. 2009). The consequences of impaired perfusive and diffusive conductances on  $\dot{V}O_2$  kinetics in CHF are visualized in Figure 10 using an adaptation of the graphical analyses developed by Peter Wagner (for maximal  $\dot{V}O_2$  conditions, Roca et al. 1992; Wagner et al. 1997) which combines the Fick principle (perfusive O<sub>2</sub> delivery i.e.,  $\dot{V}O_2 = \dot{Q}$  (arterial-venous O<sub>2</sub> content difference), where  $\dot{Q}$  is blood flow) and Fick's law (diffusive O<sub>2</sub> flux,  $\dot{V}O_2 = DO_2(P_{mv}O_2 - P_{intramyocyte}O_2)$ , where  $DO_2$  is the apparent O<sub>2</sub> diffusing capacity of muscle) to explain how microvascular dysfunction might slow  $\dot{V}O_2$  kinetics (and in some cases reduce the steady-state  $\dot{V}O_2$ ) following the onset of muscle contractions. Thus, at any given time during the transient

(Figure 10, **inset**)  $\dot{V}O_2$  kinetics will be slowed and  $\dot{V}O_2$  decreased by the culmination of perfusive and diffusive  $O_2$  transport deficits. The available evidence suggests that the reduced capillary RBC flux and failure of a significant proportion of capillaries to support that flux compromises both perfusive and diffusive  $O_2$  transport and contributes significantly to muscle and exercise dysfunction in aging and chronic diseases such as CHF and also Type II diabetes.

## Conclusions

The scientific literature and medical/physiology textbooks consider that, following the onset of contractions, capillary recruitment is obligatory to reduce intramyocyte diffusion distances, increase muscle diffusing capacity and thereby augment blood-myocyte  $O_2$  flux. However, the compelling weight of evidence challenges this contention. Specifically, most capillaries may sustain RBC flux at rest and capillary recruitment does not appear to be requisite for the dynamic increase of blood-myocyte  $O_2$  flux (i.e.,  $\dot{V}O_2$ ) following initiation of contractions. Furthermore, in the absence of detectable intramyocyte  $PO_2$  gradients, muscle  $O_2$  diffusing capacity and mitochondrial  $O_2$  supply do not appear to be dependent on capillary-mitochondrial diffusion distances. These observations mandate a revision of the capillary recruitment theory to consider the likelihood that events within the individual capillaries (i.e., increased RBC flux and hematocrit combined with 'longitudinal recruitment' of capillary surface area) are responsible for the contraction-induced increase of muscle  $O_2$  diffusing capacity and blood-myocyte  $O_2$  flux – genuine homage to August and Marie Krogh's memory and scientific legacy demands no less. Moreover, such a revision is particularly vital to determining the mechanistic bases for impaired muscle oxidative function in chronic disease.

*“The wrong view of science is betrayed by the craving to be right.”*

Sir Karl Popper (1902–1994)

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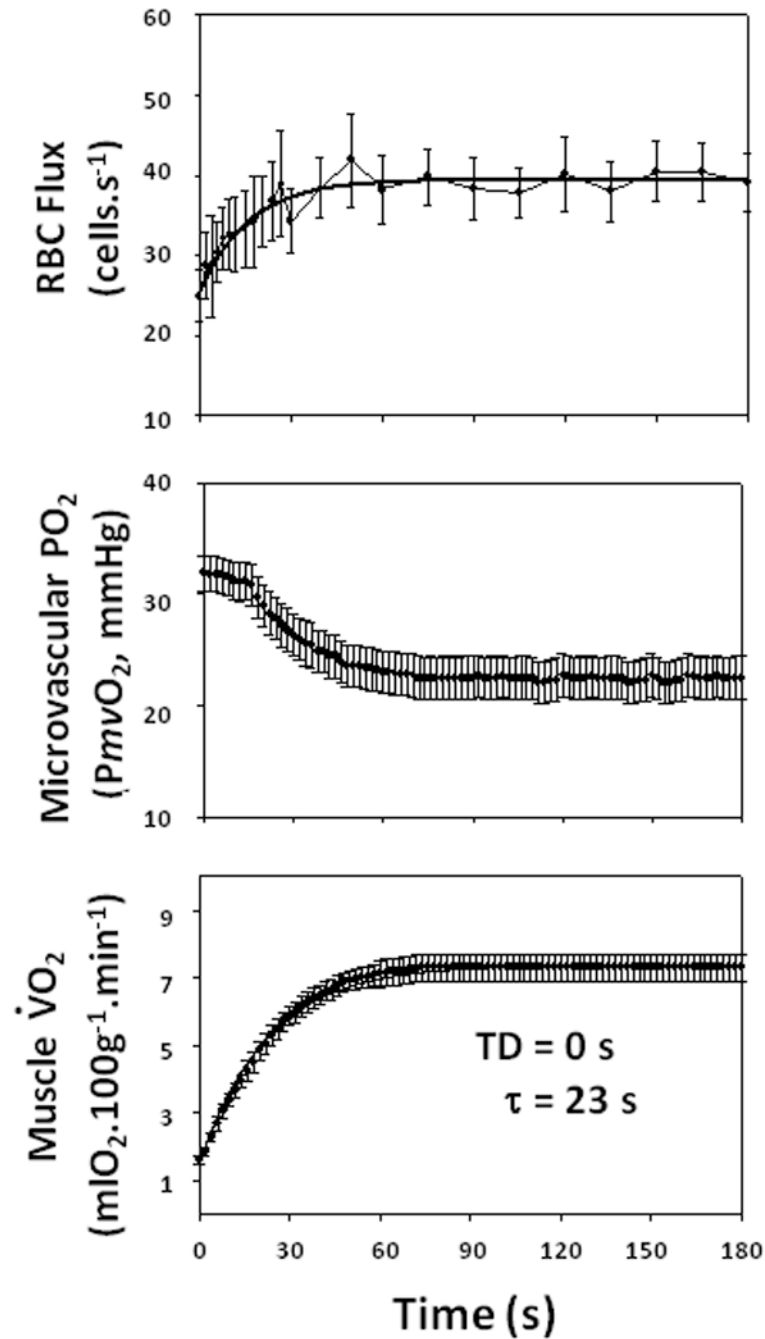
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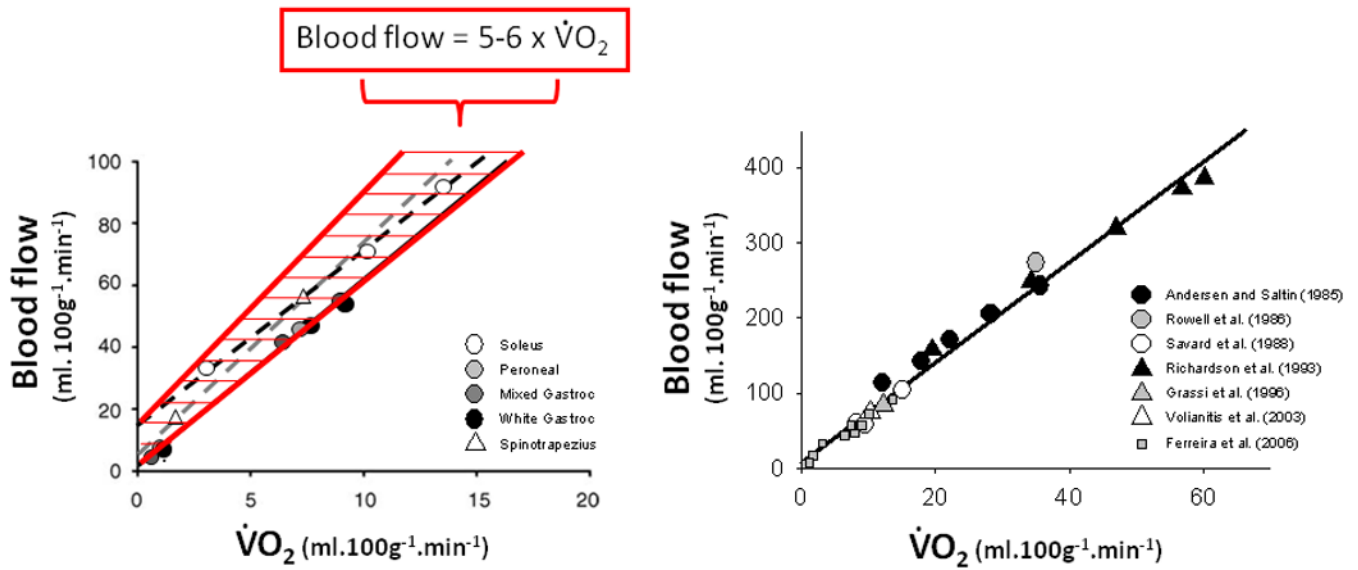
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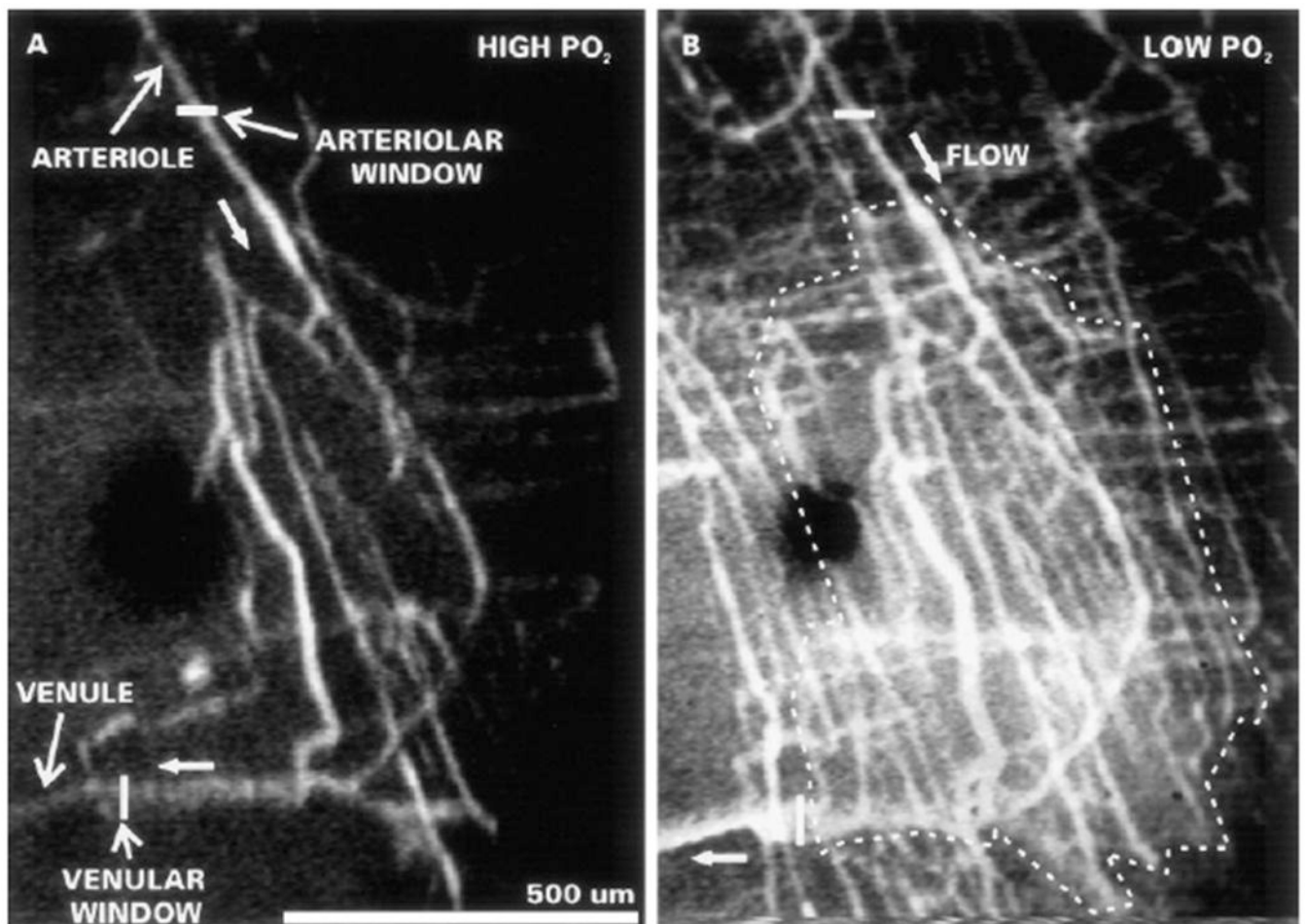
**Figure 1.**

Mean data showing increase in rat spinotrapezius red blood cell (RBC) flux (*Top*) and microvascular PO<sub>2</sub> (*Middle*) are conflated to estimate  $\dot{V}O_2$  (*Bottom*) in response to 1 Hz electrically-induced contractions (initiated at time = 0 s). Capillary RBC flux was measured by high-resolution videomicroscopy analysis in individual capillaries (Kindig et al. 2002). Microvascular PO<sub>2</sub> was determined using phosphorescence quenching techniques (Rumsey et al. 1988; Poole et al. 1995, 2004; Behnke et al. 2001).  $\dot{V}O_2$  was calculated using the measured RBC flux and arterial O<sub>2</sub> content and microvascular PO<sub>2</sub> as an estimate of end-capillary PO<sub>2</sub> and O<sub>2</sub> dissociation curves established for the rat (Altman and Dittmer, 1974). Model fits are shown. Both RBC flux and  $\dot{V}O_2$  (but not PmvO<sub>2</sub>) were fit by a single

exponential with no time delay. TD, time delay.  $\tau$ , time constant. Redrawn from Behnke et al. (2002). The value for  $\tau$  resolved is similar to that for human muscle and also Phase II pulmonary  $\dot{V}O_2$  kinetics (e.g., Grassi et al. 1996).

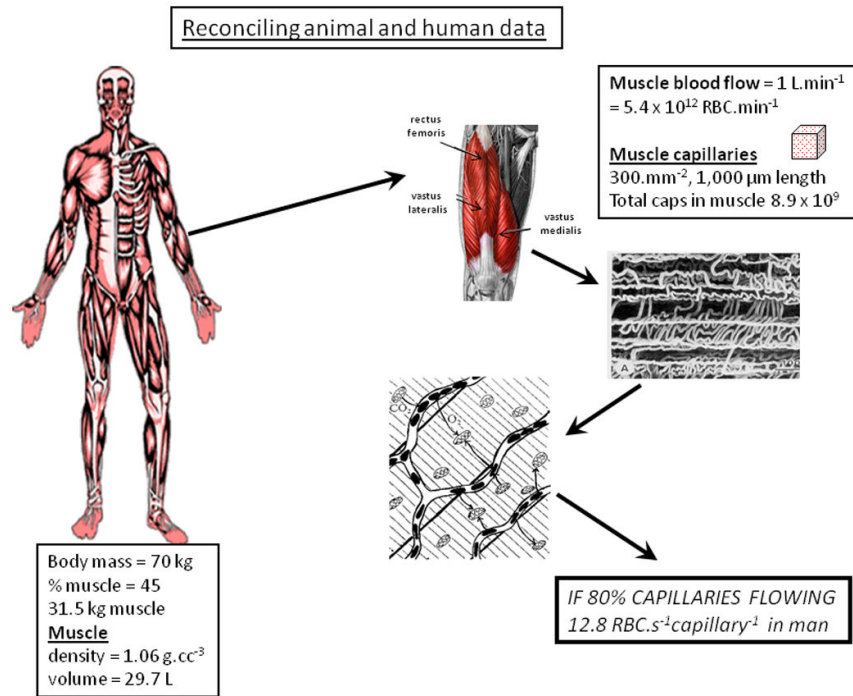


**Figure 2.** Relation between blood flow and oxygen uptake ( $\dot{V}O_2$ ) during muscle contractions for various muscles in pentobarbital-anesthetized rats (*Left Panel*) and consciously exercising humans (*Right Panel* shows relationship in *Left Panel* expanded to incorporate human data). Rat data are means from Behnke et al. (2002, 2003) as redrawn from Ferreira et al. (2006). Human data are from: Andersen and Saltin, (1985); Rowell et al. (1986); Savard et al. (1988); Richardson et al. (1993); Grassi et al. (1996); Volianitis et al. (2003). This figure demonstrates that anesthetized preparations retain vasomotor control such that blood flow increases in the same proportion with  $\dot{V}O_2$  (i.e., 5–6 L.min<sup>-1</sup>:1 L.min<sup>-1</sup>) as found in conscious preparations (rev. Poole, 1997) though the range of metabolic rates and blood flows is lower for the animal preparations owing, in part, to the requirement for electrical stimulation.



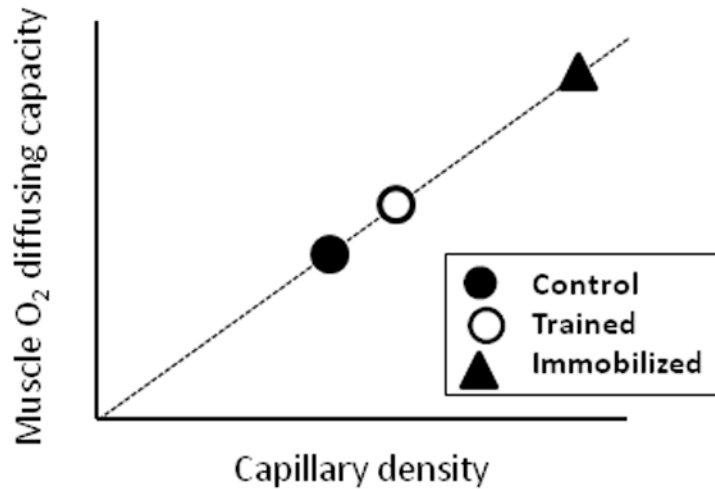
**Figure 3.**

Example of how inappropriate boundary conditions may reinforce capillary recruitment notion. Capillaries are visualized in resting cremaster muscle using fluorescently-labeled plasma and the  $PO_2$  conditions are 130 mmHg (*Left panel*) and 35 mmHg (*Right panel*). Both of these  $PO_2$ 's are considerably above intramuscular values at rest (~17 mmHg, Whalen et al. 1974) and what is observed is probably not hypoxia-induced capillary recruitment moving from left-to-right (which would support a reserve of non-flowing capillaries that might be 'recruited' during exercise) but, rather, hyperoxia-induced vasoconstriction moving from right-to-left. Small arrows denote direction of flow. Images from Parthasarathi and Lipowsky (1999), with permission.

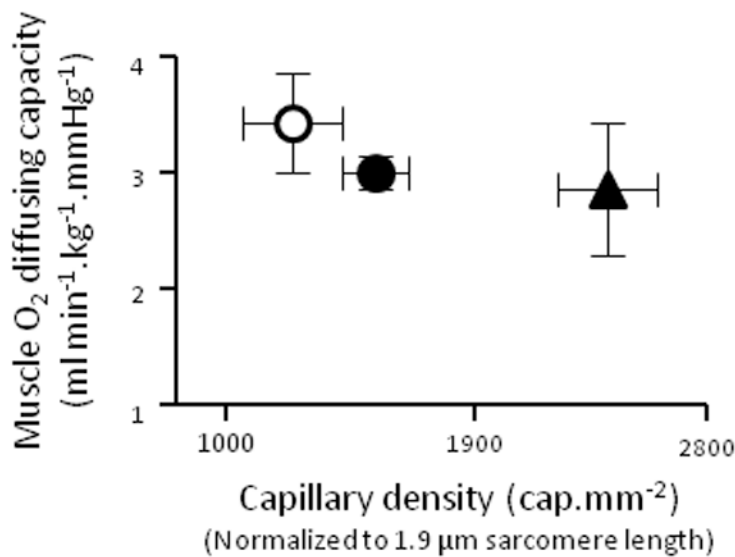


**Figure 4.** As demonstrated by intravital microscopy (e.g., Kindig et al., 1999) and dye-infusion (Kayar and Banchemo, 1995) for anesthetized and conscious animals, is it possible that the majority of capillaries support red blood cell flux in resting muscle in humans? This figure uses established values for muscle blood flow (e.g., Rowell, 1993) combined with good-faith estimates of muscle capillary number (e.g., Saltin and Gollnick, 1983; Hoppeler et al. 1985; Richardson et al. 1994) to demonstrate that, in humans, 80% of total muscle capillaries could, in theory, support an average of nearly 13 RBC.s<sup>-1</sup> which is close to that value measured in rat spinotrapezius (15–20 RBC.cap<sup>-1</sup>.s<sup>-1</sup>; Kindig et al. 1997). In contrast to the leftmost and center boxes, the italicized capitalized letters in the rightmost box alert the reader that this “80%” value is data taken from the animal literature as such data is not presently available in humans. See text for more details.

**Hypothesis:** If intramyocyte diffusion distances are deterministic for diffusing capacity, this relationship is expected:



**Data:** No such relationship is found and, in fact, the highest O<sub>2</sub> diffusing capacity is found in those muscles with the lowest capillary density

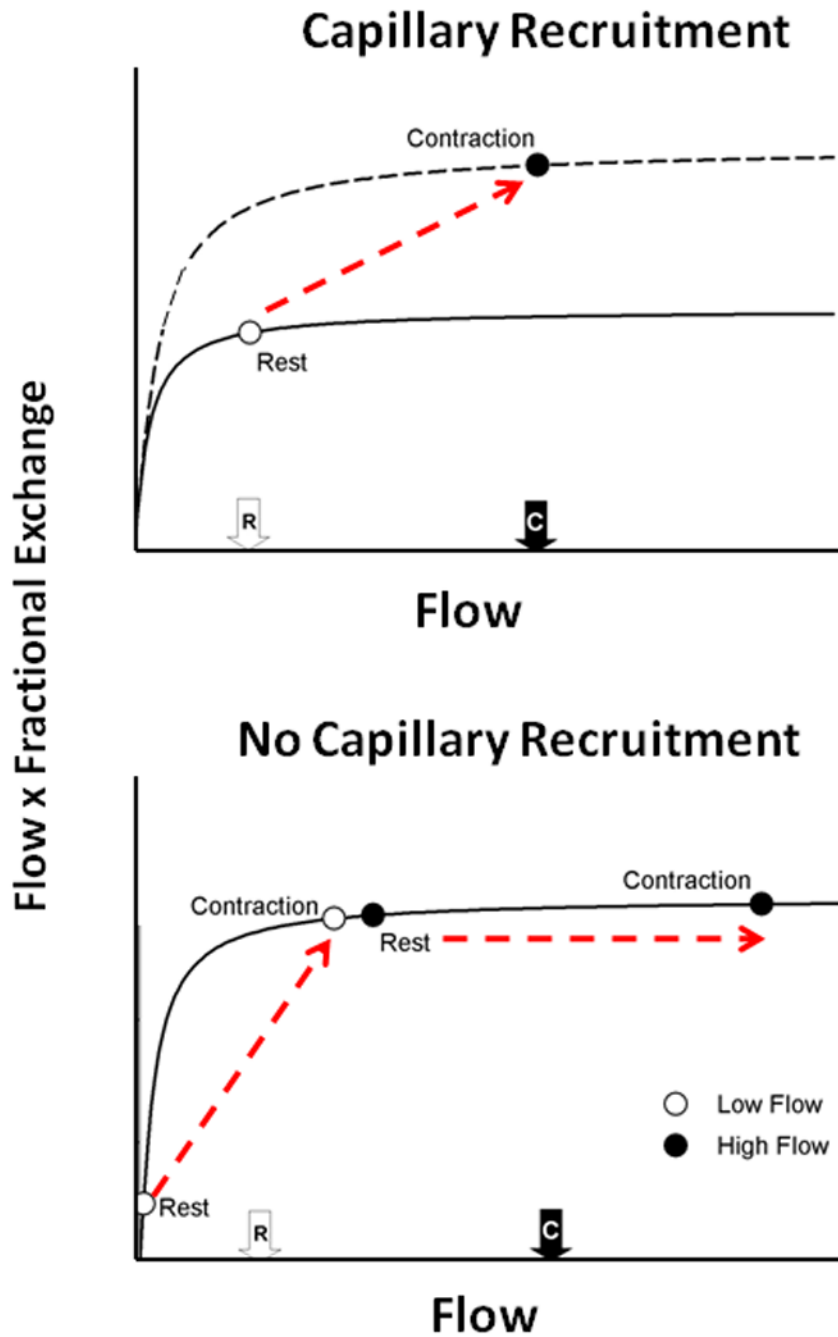


**Figure 5.**

Empirical test of the hypothesis that muscle O<sub>2</sub> diffusing capacity is dependent upon diffusion distances *per se*. *Top graph* depicts hypothesis tested i.e., if diffusion distances are a key determinant of muscle O<sub>2</sub> diffusing capacity then the data should line up as shown with far higher O<sub>2</sub> diffusing capacity for the immobilized condition where capillary density is increased substantially as a result of the pronounced fiber atrophy. However, contrary to the hypothesis, *bottom graph* demonstrates no increased diffusing capacity with reduced fiber cross-sectional area, increased capillary density and reduced O<sub>2</sub> diffusion distances with immobilisation. Muscle O<sub>2</sub> diffusing capacity is calculated from the mean capillary PO<sub>2</sub> which, in turn, is estimated using the measured arterial and effluent PO<sub>2</sub>'s and the



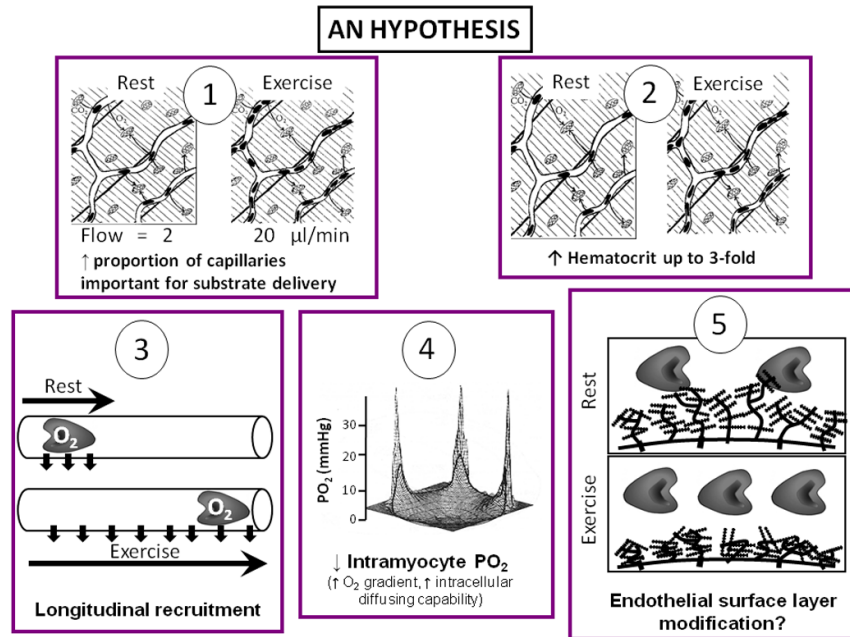
muscle  $\dot{V}O_2$  by a forward integration procedure and assumes that all  $O_2$  remaining in the venous blood results from diffusional limitation (i.e., the contribution of anatomical and functional shunts and  $O_2$  delivery-to- $O_2$  utilization mismatch is negligible) and that intramyocyte  $PO_2$  is zero (see Roca et al. 1992). From Hepple et al. (2000), with permission.



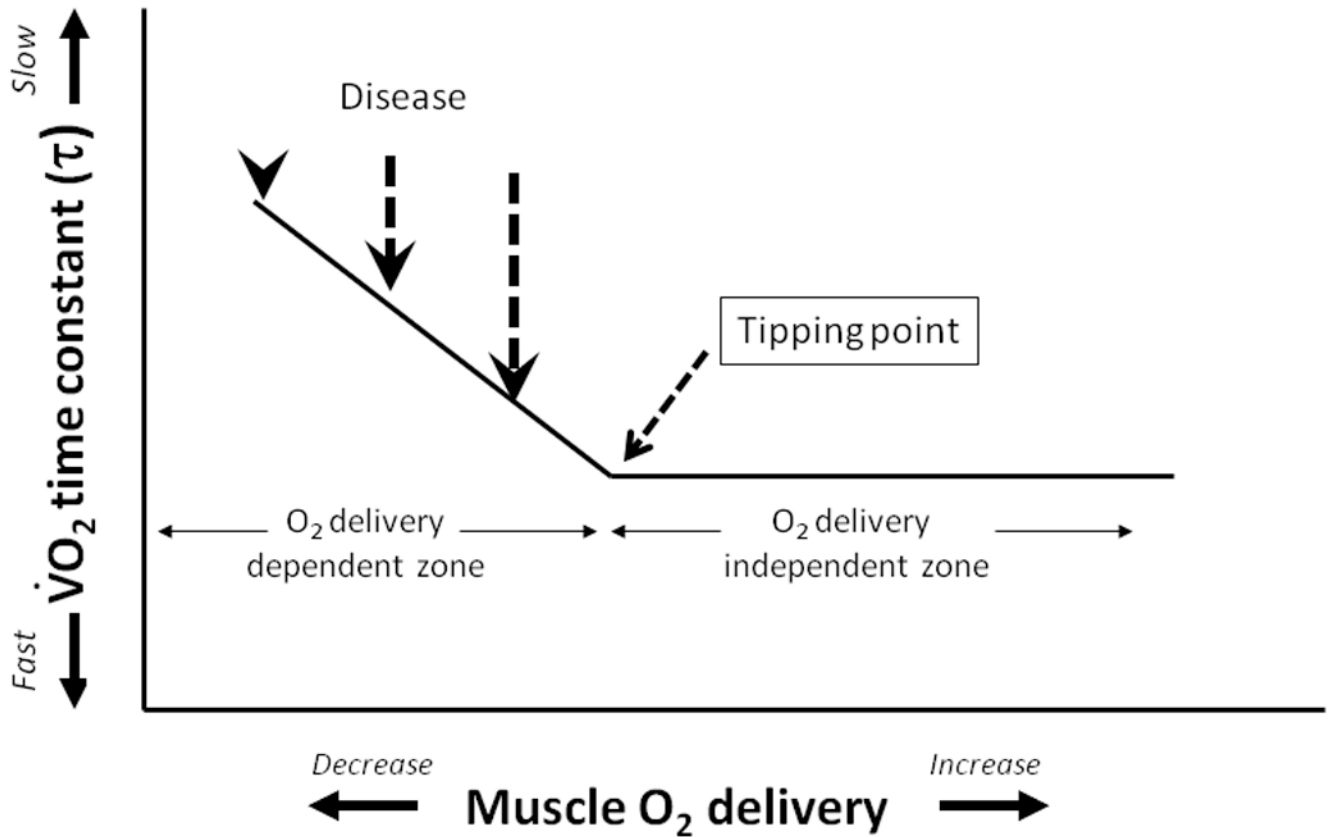
**Figure 6.**

In a simplified manner the relationship between capillary delivery and muscle uptake of a substrate can be described by a close-to-exponential curve (*Top Figure*). In general, this is the model used by Clark and colleagues employing the 1-methylxanthine metabolism technique (Rattigan et al. 1997ab; Youd et al. 1999) to estimate capillary recruitment during contractions. The basic assumptions made in this model are: i) that blood flow in the microcirculation (capillaries) is homogeneous, and ii) at rest all capillaries are located in the flat portion of the curve. PS is the permeability-surface area product. Any increase in 1-methylxanthine metabolism from rest-to-contractions is explained by greater surface area, which is assumed to reflect the recruitment of previously non-flowing capillaries. However,

as discussed in the text, microvascular blood flow is markedly heterogeneous at rest and during contractions. Thus, the *bottom figure* considers two groups of capillaries: low-flow and high-flow. In this simulation, low-flow capillaries are located in the flow-limited range of substrate uptake and high-flow capillaries are on the flat portion of the curve ('diffusion/surface area-limited' range). The mean resting blood flow is close to the flat portion of the curve (white arrow *bottom Figure*; same as in Clark et al.'s model; *top figure*). An increase in blood flow through both groups of capillaries – mean capillary flow during contractions represented by black arrow (*Bottom Figure*) and similar to that in Clark et al.'s model – will lead to an increase in O<sub>2</sub> or substrate uptake that is identical to that seen for the “capillary recruitment” model. Thus, increase in blood flow through capillaries with heterogeneous flow distribution is a viable alternate explanation to Clark and colleague's data without any capillary recruitment requisite whatsoever.

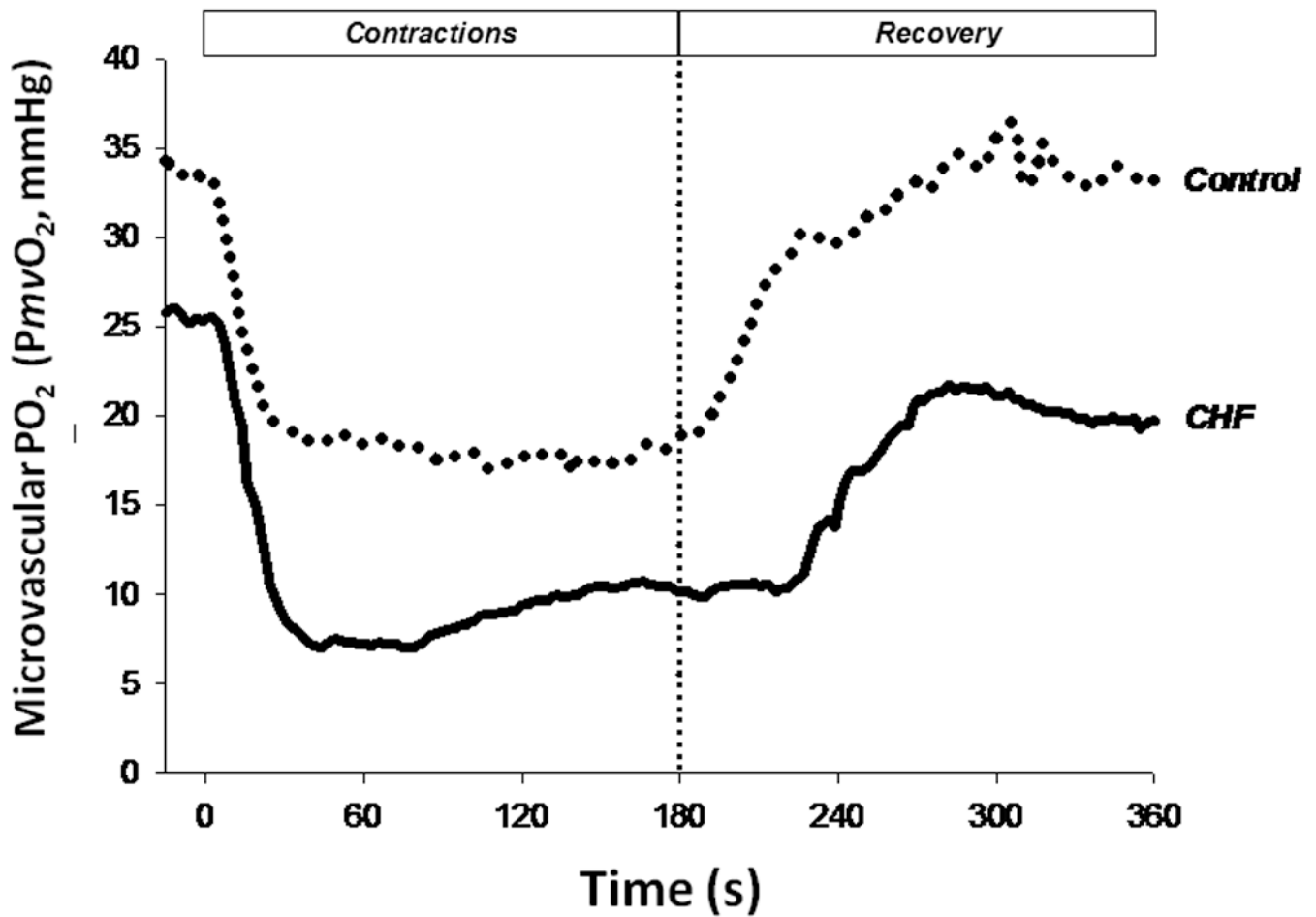


**Figure 7.** Putative mechanisms that can account for the increase of blood-myocyte  $O_2$  (and substrate) flux, from rest-exercise, within existent RBC flowing capillaries without the necessity for ‘recruitment’ of previously non-flowing capillaries. 1. Substantial elevation of blood flow (RBC flux) within originally very low flow capillaries means that capillaries that may have been unimportant for blood-myocyte flux at rest become important during exercise. 2. Elevation of RBC numbers (hematocrit) within capillaries. 3. Increased RBC velocity and greater fractional  $O_2$  extraction mean that additional capillary surface area along the length of the capillary becomes important for blood-myocyte flux (i.e., ‘capillary longitudinal recruitment’). 4. In 3 dimensions this graph depicts the precipitous drop in  $PO_2$  from capillary to the intramyocyte space (i.e., the area encircled by the thick black line) during muscle contractions. Note the overall very low intramyocyte  $PO_2$  and lack of appreciable intramyocyte  $PO_2$  gradients. From rest-exercise, the fall in intracellular  $PO_2$  increases the blood-myocyte gradient and also desaturates myoglobin removing the ‘ $O_2$  carrier functionally-depleted region’ (Honig et al. 1997) and elevating its ability to transport  $O_2$  which improves intracellular diffusing capacity for  $O_2$ . 5. There is the possibility that the endothelial cell surface layer (thought to be crucial in lowering capillary hematocrit below systemic) is modified under high flow conditions.



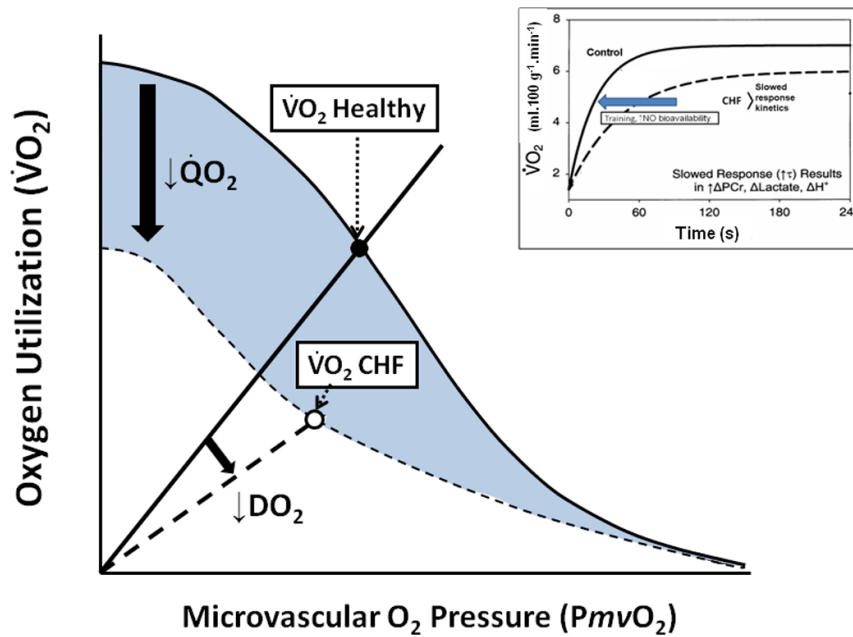
**Figure 8.**

In healthy young individuals the control of  $\dot{V}O_2$  kinetics during moderate intensity exercise is believed to reside within the intramuscular oxidative machinery and is not  $O_2$  delivery dependent (i.e., these individuals lie to the right-hand side of the depicted hypothetical ‘tipping point’). Specifically, in healthy young individuals, neither muscle (Grassi et al. 1998) nor pulmonary  $\dot{V}O_2$  (MacDonald et al. 1997) is speeded by augmented arterial  $O_2$  content or increased muscle  $O_2$  delivery. Moreover, strategies designed to decrease muscle  $O_2$  delivery do not always slow  $\dot{V}O_2$  kinetics (Williamson et al. 1996; MacDonald et al. 1997; Nyberg et al. 2010). These observations suggest that healthy young individuals lie somewhere to the right of the tipping point and somewhere in the “ $O_2$  delivery independent zone.” In marked contrast, chronic diseases are associated with very slow  $\dot{V}O_2$  kinetics that can be speeded by increasing arterial  $O_2$  content (e.g., Palange et al. 1995). Redrawn from Poole et al. (2008a). There is now substantial evidence that microcirculatory impairments likely contribute to these aberrant (slowed)  $\dot{V}O_2$  kinetics and exercise intolerance in disease (i.e., heart failure, Kindig et al. 1999; Diederich et al. 2002; Richardson et al. 2003; diabetes, Padilla et al. 2006, 2007).



**Figure 9.**

The profile of microvascular  $PO_2$  ( $P_{mv}O_2$ ) as measured by phosphorescence quenching (Rumsey et al. 1988; Poole et al. 1995; Behnke et al. 2001) during (0–180 s) and following (181–360 s) muscle contractions in chronic heart failure (CHF) provides clear evidence of profound derangements in the ability to match  $O_2$  delivery-to- $O_2$  utilization ( $O_2$ -to- $\dot{V}O_2$ ). Impaired microcirculatory dynamics impair both perfusive and diffusive muscle  $O_2$  conductance slowing blood-myocyte  $O_2$  flux rates and constraining the speed of  $\dot{V}O_2$  kinetics thereby forcing the muscle(s) to utilize increased substrate-level phosphorylation to support energetic requirements. Constructed from data set of Copp et al. (2010).



**Figure 10.** Schematic showing muscle  $O_2$  uptake ( $\dot{V}O_2$ ) plotted as a function of microvascular  $PO_2$  ( $P_{mv}O_2$ ). Conflation of perfusive (Fick principle, *solid curved line*) and diffusive (Fick's law, *solid straight line from origin*) permits a mechanistic analysis of how these components resolve to achieve a given  $\dot{V}O_2$ . Note that chronic heart failure (CHF, *dashed lines*) reduces both perfusive and diffusive  $O_2$  fluxes such that, during the transient following the onset of contractions,  $\dot{V}O_2$  kinetics are slowed and  $\dot{V}O_2$  cannot, and does not, achieve levels seen in healthy individuals (*inset*). Resolution of the microvascular impairments that reduce perfusive ( $\dot{Q}O_2$ ) and diffusive ( $DO_2$ )  $O_2$  flux constitutes a critical step in the design of effective therapeutic interventions (e.g., exercise training,  $\uparrow$ nitric oxide (NO) bioavailability).

**Table 1**

Exemplar references from either side of the ‘capillary recruitment’ debate that have either presented evidence for a substantial proportion of non-RBC/plasma flowing capillaries in skeletal muscle at rest or interpreted their data as evidence for capillary recruitment (left hand side) or provided evidence that most capillaries already support RBC/plasma flow in resting muscle (right hand side). The latter position discounts the possibility of substantial ‘capillary recruitment’ following the onset of contractions.

<b>Opening of previously “closed” or non-flowing capillaries</b>	<b>Most capillaries support RBC or plasma flow at rest - with a wide range of flows and transit times</b>
Krogh, 1919	Eriksson and Myrhage, 1972
Gorzynski, Duling, 1978	Burton and Johnson, 1972
Honig et al. 1980,1982	Renkin et al. 1981
Klitzman et al. 1982	Hudlicka et al. 1982
Gray et al. 1983	Vetterlein et al. 1982
Hargreaves et al. 1990	Damon and Duling, 1984
Fuglevand and Segal, 1997	Kayar and Banchero, 1985
Rattigan et al. 1997	Hudlicka, 1985
Parthasarathi and Lipowsky, 1999	Dawson et al. 1987
Baron et al. 2000	Poole et al. 1997
Clark et al. 2001	Kindig and Poole, 1998,1999,2001
Dawson et al. 2002	Kindig et al. 2002
Rattigan et al. 2005, 2006	Russell et al. 2003
Vincent et al. 2006	Richardson et al. 2003
Womack et al. 2009	Padilla et al. 2006
Bourdillon et al. 2009	Copp et al. 2009