

Tropical amphibian populations experience higher disease risk in natural habitats

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Habitat loss and disease are main drivers of global amphibian declines, yet the interaction between them remains largely unexplored. Here we show that paradoxically, habitat loss is negatively associated with occurrence, prevalence, and infection intensity of the chytrid fungus *Batrachochytrium dendrobatidis* (*Bd*) in amphibian populations in the tropics. At a large spatial scale, increased habitat loss predicted lower disease risk in amphibian populations across Costa Rica and eastern Australia, even after jointly considering the effect of potential biotic and abiotic correlates. Lower host-species richness and suboptimal microclimates for *Bd* in disturbed habitats are potential mechanisms underlying this pattern. Furthermore, we found that anthropogenic deforestation practices biased to lowlands and natural vegetation remaining in inaccessible highlands explain increased *Bd* occurrence at higher elevations. At a smaller spatial scale, holding constant elevation, latitude, and macroclimate, we also found a negative relationship between habitat loss, and both *Bd* prevalence and infection intensity in frog populations in two landscapes of the Brazilian Atlantic Forest. Our results indicate that amphibians will be disproportionately affected by emerging diseases in pristine environments, and that, paradoxically, disturbed habitats may act as shelters from disease, but only for the very few species that can tolerate deforestation. Thus, tropical amphibian faunas are threatened both by destruction of natural habitats as well as increased disease in pristine forests. To curb further extinctions and develop effective mitigation and restoration programs we must look to interactions between habitat loss and disease, the two main factors at the root of global amphibian declines.

chytridiomycosis | species diversity | anurans | Neotropics | pathogen

Habitat loss and chytridiomycosis, a disease caused by the chytrid fungus *Batrachochytrium dendrobatidis* (*Bd*), are two main causes of global amphibian declines (1–4). Habitat loss lowers amphibian species diversity by reducing natural habitats (1) and increasing population isolation (5), inbreeding (6), edge effects (7), and discontinuity between terrestrial and aquatic habitats (8). Disturbance to natural vegetation also changes ecosystem structure, shifting macro (9) and microclimates (10), and altering hydrological cycles (11). Thus, habitat loss may also influence amphibian susceptibility to disease by altering host-community structure, transmission pathways, and pathogen persistence and virulence (12). *Bd* is a water-borne epidermal pathogen with a broad host range among amphibians (13) and has been implicated in population declines and species extinctions worldwide (3, 4, 14–16). *Bd* prevalence and infection intensity are important predictors of disease risk and population die-offs (17) and, to some extent, can be modified by environmental factors. *Bd* prevalence varies with latitude (18), elevation (19), precipitation (18, 20, 21), and temperature (18, 20), presumably reflecting *Bd* optimal growth conditions (22, 23).

The most severe amphibian declines and extinctions have been observed in the Neotropics (15) and Australia (14). In both regions, *Bd* outbreaks occur primarily in pristine forests at high-elevation mountainous sites, such as the Talamancas in Central America (15, 24), the Tropical Andes in South America (15),

and the Great Dividing Range in eastern Australia (25). In contrast, pathogen prevalence is lower and amphibian populations often remain stable at lowland sites (15, 19, 25). Because *Bd* optimal growth occurs at mild temperatures (17–25 °C) and high humidity (22, 23), these regional patterns of declines could result from the interaction among elevation, climate, and also habitat loss, all of which are potentially correlated. Temperature is negatively correlated with elevation. Elevation and habitat loss are often negatively correlated because of anthropogenic deforestation practices that are biased to the flatter and more accessible lowlands (26). Finally, tropical habitat loss increases average temperatures and may change precipitation patterns (11). In the case of *Bd*, most empirical studies have focused on the interaction of only two of these three critical components, elevation and climate, eclipsing the potential role of habitat loss and its cascading biotic and abiotic consequences on *Bd* epidemiology at both large and small spatial scales.

Here, we examined the effect of habitat loss on *Bd* infections in tropical amphibian populations at both large and small spatial scales. We hypothesized that habitat loss is negatively associated with *Bd* infections because of lower host-species richness and potentially suboptimal microclimate for *Bd* in disturbed habitats. First, we analyzed published surveys of *Bd* infections of Rain frog populations (*Craugastor fitzingeri*) in Costa Rica (20), and Stony Creek frog populations (*Litoria lesueuri*) in eastern Australia (18). For each sampling site we quantified the degree of habitat loss, measured as the percentage of nonnatural vegetation cover within 1-km buffers surrounding sampling locations. We tested the effect of habitat loss on pathogen occurrence, prevalence, and infection intensity using spatial autoregressions and path analysis, and accounted for potential effects of host-species richness, latitude, elevation, and bioclimatic metrics of temperature and precipitation. In a second study, replicated in two landscapes of the Brazilian Atlantic Forest, we compared *Bd* prevalence and infection intensity in populations of the Golden Lesser treefrog (*Dendropsophus minutus*) among sites with varying levels of habitat loss. At this smaller scale, we controlled for the effects of environmental factors by sampling populations with similar climate, elevation, and latitude, and used spatial autoregressions to test the relationship of habitat loss with *Bd* prevalence and infection intensity.

Results

Habitat Loss Predicts *Bd* Infections in Costa Rica and Australia. Increased habitat loss predicted lower *Bd* occurrence in amphibian populations in Costa Rica ($\beta_{\text{AUTOLOG}} = -0.022$, $t = -1.953$, $P = 0.051$). Our initial stepwise screening for explanatory variables

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selected habitat loss, amphibian species richness, elevation, temperature annual range, and precipitation of the driest quarter of the year as variables with high scores for explaining *Bd* occurrence. When jointly considering these effects in a model selection approach, including all possible models, habitat loss remained a strong predictor of pathogen occurrence (Table 1 and Table S1). In the best model, we found a negative effect of habitat loss and a positive effect of species richness on *Bd* occurrence.

Anthropogenic deforestation is nonrandomly distributed throughout Costa Rica because habitats in flatter lowlands are disproportionately disturbed and, thus, natural vegetation persists at higher elevations (Fig. 1 A–C). This nonrandom pattern potentially confounds the effects of elevation and habitat loss on pathogen occurrence. We considered the influence of elevation on habitat loss in a binary response path analysis and showed that the effect of elevation on *Bd* occurrence is indirect through habitat loss (whole-model test: $\chi^2_{[3]} = 22.545, P < 0.001$) (Fig. 1D).

Increased habitat loss also predicted lower *Bd* prevalence in amphibian populations across eastern Australia ($\beta_{\text{CAR}} = -0.346, t = -2.579, P = 0.015$), in this case, without the confounding effects of elevation across sampling locations (18). The initial screening detected habitat loss, amphibian species richness, latitude, maximum temperature of the warmest month, and precipitation of the warmest quarter of the year as potential explanatory variables for *Bd* dynamics. Because latitude and maximum temperature of the warmest month were highly correlated ($r = -0.920, P < 0.001$), we included them in the model selection as a PC variable. When jointly considering all effects in model selection, habitat loss remained a strong negative predictor of *Bd* prevalence, amphibian species richness a positive predictor, latitude a positive predictor, and maximum temperature during the warmest month a negative predictor (Table 1 and Table S1). Habitat loss alone was not a significant predictor of *Bd* infection intensity ($\beta_{\text{CAR}} = -0.012, t = -1.596, P = 0.122$); however, when considered jointly with other variables selected in the initial screening, habitat loss became a strong factor, explaining infection intensity together with precipitation of the driest month (Table 1 and Table S1). The inclusion of spatial autocorrelation in our analyses improved model fit for all reported models (Table 1).

Habitat Loss Predicts *Bd* Infections at Small Spatial Scale. Habitat loss predicted lower *Bd* prevalence in amphibian populations in both Brazilian Atlantic Forest landscapes. As expected, none of the 19 bioclimatic variables explained *Bd* prevalence and infection intensity at this smaller spatial scale. We showed that habitat loss alone is a key factor predicting *Bd* prevalence ($\beta_{\text{CAR}} = -0.556, t = -5.074, P = 0.001$) (Fig. 2A) and infection intensity ($\beta_{\text{CAR}} = -0.027, t = -5.564, P < 0.001$) (Fig. 2B) in Araucaria Moist Forest, and although not statistically significant, the same trends were found for the Serra do Mar Coastal Forest [prevalence: $\beta_{\text{CAR}} = -0.371, t = -1.683, P = 0.136$ (Fig. 2C); infection intensity: $\beta_{\text{CAR}} = -0.017, t = -1.520, P = 0.172$ (Fig. 2D)]. Because host population density can differ across disturbance gradients, we examined the effect of amphibian capture rate (as a proxy for population density) on both *Bd* prevalence and infection intensity and found no significant relationships [Serra do Mar Coastal Forest ($\beta_{\text{CAR}} = -2.402, t = -1.306, P = 0.233$; $\beta_{\text{CAR}} = -0.168, t = -1.966, P = 0.090$), Araucaria Moist Forest ($\beta_{\text{CAR}} = -4.695, t = -1.475, P = 0.184$; $\beta_{\text{CAR}} = -0.200, t = -1.324, P = 0.227$)].

Discussion

We showed that habitat loss, a main cause of species extinctions worldwide (27), is negatively associated with *Bd* occurrence, prevalence, and infection intensity in tropical amphibian populations. This effect was evident even when jointly considering several known environmental correlates of pathogen growth and persistence (18–21, 24). We corroborated these results at a smaller geographic scale, where habitat loss predicted lower pathogen prevalence and infection intensity in populations with similar macroclimate, topography, and latitude. Thus, localized differences in natural vegetation cover may explain why neighboring populations often have highly contrasting *Bd* dynamics. Our path analysis showed clearly that elevation had indirect effects on *Bd* occurrence via habitat loss. Therefore, the role of habitat loss as a determinant of amphibian disease threat has likely been underestimated because of nonrandom deforestation practices relative to topography.

Our analyses demonstrate a negative relationship between habitat loss and disease risk on three continents and at two spatial scales, despite many studies showing the opposite pattern.

Table 1. Autologistic and conditional autoregressive models testing simultaneously the effects of habitat loss, amphibian species richness, and environmental factors on *Bd* occurrence in amphibian populations in Costa Rica, and on *Bd* prevalence and infection intensity in amphibian populations in Australia

Term	$\beta_{\text{AUTOLOG}/\beta_{\text{CAR}}}$	Std. coeff.	SE	t	VIF	P
Occurrence						
Constant	-6.311	0	2.033	-3.104	—	0.002
Spatial autocovariate term - yW	3.138	0.879	3.075	1.020	—	0.308
i) Habitat loss	-0.030	-3.769	0.012	-2.430	1.020	0.015
ii) Amphibian species richness	0.098	4.224	0.042	2.318	1.020	0.020
Prevalence						
Constant	-15.980	0	33.360	-0.479	—	0.636
i) Habitat loss	-0.304	-0.420	0.113	-2.695	1.279	0.012
ii) Amphibian species richness	1.414	0.466	0.658	2.148	2.059	0.042
iii) Latitude and maximum temperature warmest month PC1*	-36.959	-2.179	11.699	-3.159	2.470	0.004
<i>ii</i> × <i>iii</i>	0.796	1.439	0.390	2.039	1.355	0.052
Infection intensity						
Constant	0.555	0	1.439	0.386	—	0.703
i) Habitat loss	-0.017	-0.416	0.006	-2.626	1.025	0.014
ii) Precipitation of the driest month	0.043	0.544	0.011	4.001	1.025	<0.001

Whole-model tests: Occurrence: ($\chi^2 = 14.888, n = 125, P = 0.002$); Prevalence: ($F = 8.700, n = 31, r^2_{\text{OLS}} = 0.581, \text{Predictor} + \text{Space } r^2 = 0.632, P < 0.001$); Infection intensity: ($F = 10.491, n = 31, r^2_{\text{OLS}} = 0.458, \text{Predictor} + \text{Space } r^2 = 0.582, P < 0.001$). Significant variables in the model are highlighted in bold. VIF stands for variance inflation factor and denotes collinearity in the model if higher than 10. Std. coeff stands for standard coefficient. Final models chosen based on Akaike Information Criterion.

*PC1 consolidating latitude and maximum temperature of the warmest month accounted for 96.00% of the variation in the original variables.

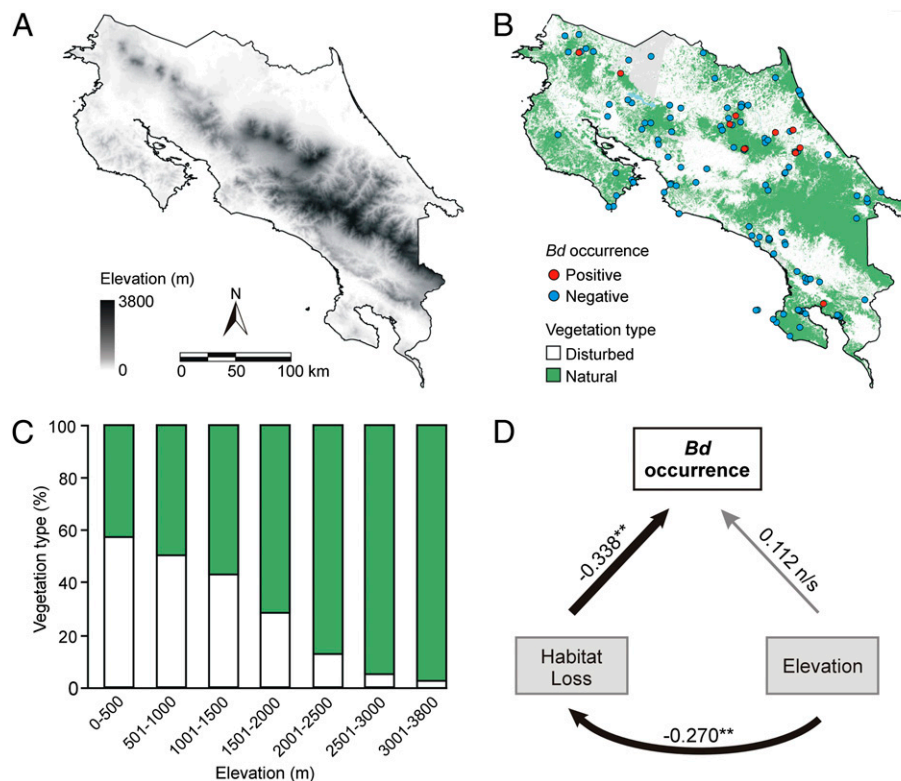


Fig. 1. (A) Elevation range and (B) spatial distribution of disturbed and natural vegetation throughout Costa Rica. Sampling sites are *Bd*-positive (red circles) and -negative (blue circles); vegetation types are disturbed (white) and natural (green); other land-cover classes are freshwater (light blue) and unclassified cloud cover (gray). (C) Percentage of disturbed (white) and natural vegetation (green) across the altitudinal gradient. (D) Path Analysis model showing the relative strength of habitat loss and elevation on *Bd* occurrence among amphibian populations. Numbers are standardized path coefficients (** $P < 0.01$). The thickness of the arrows represents the relative strength of the relationship.

Habitat loss disrupts natural ecosystems and often increases the risk of human and wildlife diseases (12, 28). Tropical deforestation coincides with a rise of malaria in Africa, Asia, and Latin America, independent of human population density (12, 29). Habitat change was positively associated with the emergence of bat-borne Nipah virus in Malaysia (30), cryptosporidiosis in Europe and North America, and food-borne illnesses globally (31). Bighorn sheep populations in the San Andres Mountains, New Mexico, suffered demographic declines because of severe habitat loss and fragmentation and were subsequently stricken by an epidemic of psoroptic scabies (28). A possible mechanism for this general pattern is that habitat loss limits individual movement and dispersal, which in turn increases disease risk because of elevated stress and contact rates among individuals in crowded patchy populations (32). Population-based models of disease dynamics indicate a potential trade-off in that habitat fragmentation can limit pathogen transmission between small and isolated populations, but it also increases genetic erosion of resistant alleles and reduces the probability of rescue events, increasing the demographic consequences of disease epidemics (33).

In the case of *Bd*, there is mounting evidence that humans have been playing a key role in spreading the pathogen (3, 34, 35), with infections often higher in areas of higher human footprint (36). Surveys in both temperate and tropical regions revealed that *Bd* detectability increased with human population density, and in the vicinity of port cities and the highways connecting them (37). Our results, however, show that anthropogenic habitat loss, specifically deforestation, can lower *Bd* infections in amphibians. This pattern was also observed in *Litoria wilcoxii* populations of Eastern Australia, where forested habitats harbored amphibians with higher pathogen prevalence than neighboring agricultural

lands (38). Other forms of habitat change may also decrease *Bd* fitness: small populations of *Litoria aurea* persisted in heavy industrial and mining areas after a severe outbreak of chytridiomycosis, indicating that *Bd* may be sensitive to environmental contaminants (39). Likewise, in a pristine region of Eastern Australia where *Bd* was previously documented, forest-associated amphibians suffered local extinctions, but species richness remained high in adjacent urban areas (40). Combined, these examples and our results suggest that although humans may assist pathogen dispersal, anthropogenic habitat changes may also limit *Bd* persistence.

We propose two potential mechanisms for elevated disease risk in pristine environments: (i) lower *Bd* infection risk at disturbed habitats because of lower host-species richness, and (ii) sub-optimal microclimatic conditions for *Bd* in disturbed habitats. *Bd* is one of the most host-generalist pathogens ever found, with over 350 amphibian host species documented to date (3, 13). Therefore, higher biodiversity and community complexity might amplify *Bd* infections, because greater diversity of host species can enhance pathogen transmission due to higher availability of susceptible hosts throughout the year. We found a positive relationship between amphibian species richness and *Bd* infections in Costa Rica and Australia, where the negative effects of habitat loss on amphibian species richness are well documented (41, 42). Thus, habitat loss may indirectly reduce *Bd* infections by lowering species richness in amphibian communities. Although we do not have data on species richness for our small-scale study, deforestation along riparian zones is shown to decrease amphibian species richness in the Brazilian Atlantic Forest (2), thus the same potential mechanism may apply for our smaller scale landscape studies. Different disease outcomes across natural populations may result from density-dependent host-pathogen dynamics (43); however, we

Materials and Methods

Large Spatial Scale Data. We used published datasets of *Bd* occurrence among populations of the Common Rain frog (*Craugastor fitzingeri*, Craugastoridae) in Costa Rica (20), and *Bd* prevalence and infection intensity for populations of the Stony Creek frog (*Litoria lesueuri* complex, Hylidae) across eastern Australia (18). *C. fitzingeri* breeds terrestrially and *L. lesueuri* breeds in streams; both species are habitat generalists and populations persist with *Bd* (18, 20). Sampling sites for *C. fitzingeri* were distributed throughout Costa Rica (125 sites; 349 sampled frogs), ranging from sea level to elevations up to 2,110 m. Frogs were sampled between 1993 and 2008, after the arrival of *Bd* in that country (15). *Bd* infection in frogs was assessed using histological screening (20). Because of uneven sampling efforts across populations, we considered only occurrence (presence or absence) of *Bd* at each sampling site. Sampling sites for *L. lesueuri* ranged from northern Queensland to southern New South Wales (31 sites; 863 sampled frogs), along the eastern slope of the Great Dividing Range. To control for elevation and seasonality all sampling occurred at lowland sites (mean elevation 84.51 ± 49.7 SD), and within a 42-d period during the spring of 2005. Australian frogs were swabbed in the field and *Bd*-screened using quantitative PCR (qPCR) (18). *Bd* infection was assessed at the individual level. Prevalence was estimated as the percentage of infected individuals per population, and infection intensity as the mean number of “zoospore DNA equivalents” for all individuals at each population (18).

We obtained land-cover information from Fondo Nacional de Financiamento Forestal for Costa Rica [30-m resolution; coverage period 1995–2005 (54)], and from the Bureau of Rural Sciences for Australia [100-m resolution; coverage period 1995–2000 (55)]. We acquired elevation data (90-m resolution) from the Consultative Group on International Agricultural Research Consortium for Spatial Information (56). Nineteen bioclimatic variables for both studies were extracted using Worldclim/Bioclim layers (1000-m resolution), available at <http://www.worldclim.org/bioclim> (57). These metrics of temperature and precipitation are averaged from 50-y records (1950–2000) from a dense network of climatic stations throughout the world (e.g., precipitation records from 47,554 locations, temperature from 24,542 locations). We did not include temperature variability or any variable accounting for climate change in our analyses because samples for *Bd* diagnosis were collected over short time periods, precluding longer-term temporal analyses of *Bd* dynamics. We obtained amphibian species richness at each sampling locations in Costa Rica and Australia by overlaying GIS shapefiles of species historical geographic ranges from the Global Amphibian Assessment (16).

We created habitat loss layers for Costa Rica and Australia as the percentage of nonnatural vegetation cover ranging from 0% to 100% at 1-km resolution. Land-cover types considered as nonnatural were urbanization, pasture, agriculture, and exotic crops. Natural vegetation types were primarily forest habitats. All variables were measured at 1-km pixel to maintain a consistent scale across the analyses.

Small Spatial Scale Data. We sampled 20 populations of the Golden Lesser treefrog (*Dendropsophus minutus*, Hylidae) in two landscapes of the Brazilian Atlantic Forest in southern and southeastern Brazil [10 populations in Araucaria Moist Forest, State of Rio Grande do Sul (29° 24' S, 50° 24' W), and 10 populations in Serra do Mar Coastal Forest, State of São Paulo (23° 20' S, 45° 12' W)]. Maximum distance between sampling locations was 21.33 km for Araucaria Moist Forest and 18.88 km for Serra do Mar Coastal Forest. Our focal species is a habitat generalist that breeds in ponds during the austral summer.

We swabbed frogs in the field (537 sampled frogs; average of 26.85 frogs per site) with sterile swabs for later *Bd* quantification in the laboratory. We screened samples for *Bd* in singlicate using Taqman qPCR (58), extending the range of the standards to 1,000 zoospore DNA equivalents to determine the presence of *Bd* and infection intensity. For calculations of *Bd* prevalence, we categorized individuals as *Bd*-positive when zoospore equivalents were ≥ 1. We defined infection intensity as the mean number of “zoospore equivalents” per population. We calculated amphibian capture rate for each population based on individuals sampled per person-hour.

We assessed natural vegetation cover for each sampling site based on high-resolution satellite images from 2010 (SPOT, 2-m resolution). In each

landscape, the selected study sites represented a gradient of natural vegetation cover. We calculated habitat loss as the percentage of nonnatural vegetation cover within a diameter of 600 m and 1 km surrounding each sampling site using ArcGIS 9.3.1 (59). To avoid effects of elevation and macroclimate we chose landscapes with low climatic and topographic variability (mean elevation of sampling sites for Araucaria Moist Forest 918.0 ± 10.2 m SD; for Serra do Mar Coastal Forest 929.9 ± 71.4 m SD). We extracted the same 19 bioclimatic variables (57) for each sampling site at 1-km scale. To avoid biases because of seasonality, we sampled continuously over 30 d in Serra do Mar Coastal Forest, and 41 d in Araucaria Moist Forest in 2009/2010.

Statistical Analyses. To assess the relationship between habitat loss and *Bd* occurrence in amphibian populations among sampling sites in Costa Rica while accounting for spatial autocorrelation, we performed autologistic regressions (AUTOLOG). To investigate the relationship between habitat loss and *Bd* prevalence, and between habitat loss and infection intensity among sampling sites in Australia, we used conditional autoregressions (CAR). After this first univariate assessment, we used stepwise regressions (exclusion cutoff $P < 0.10$; inclusion cutoff $P < 0.20$) to screen for biotic and abiotic factors that potentially predict disease threat to be included in model selection procedures. For each analysis, we screened a total of 23 explanatory variables, including habitat loss, amphibian species richness, latitude, elevation, and the 19 bioclimatic temperature and precipitation metrics. Once important variables were identified for each dataset, we used principal components analysis to consolidate cross-correlated variables, and used the scores of the first PC axis as variables in the subsequent model selection procedure. We used AUTOLOG and CAR model selections, including selected explanatory variables and *Bd* (occurrence, prevalence, or infection intensity) as a response variable. We tested all possible models including interactions. Competing models were ranked based on Akaike Information Criterion (AIC). We reported the best fit model for each run. We assessed multicollinearity in each of the final models using a variance inflation factor.

We used binary response path analysis to statistically model causal relationships among elevation, habitat loss, and *Bd* occurrence in Costa Rica, providing information about the relative strength of the different paths. In the model, elevation influences habitat loss, and these two variables are allowed to influence *Bd* occurrence independently.

For the small-scale data, we used the same stepwise screening method to confirm that none of the 19 bioclimatic variables explain *Bd* prevalence and infection intensity in both landscapes when accounting for habitat loss (measured at 1-km scale). To quantify the effect of habitat loss on *Bd* prevalence and infection intensity for populations in the two landscapes of the Brazilian Atlantic Forest, we used single CARs. We log-transformed zoospore data (infection intensity) for the analyses. Because climatic variables were not included, final models for both landscapes included habitat loss measured within 600-m diameter buffers surrounding sampling locations. We also ran single CARs to quantify the effect of capture rate on both *Bd* prevalence and infection intensity. Capture rate was used as proxy for population density of *D. minutus* in our analyses of pathogen dynamics. In all analyses at both scales, sampling site was used as the replicate in statistical tests of *Bd* dynamics. We ran analyses using Spatial Analysis in Macroecology v4.0 (60) and Mplus v6.0 (61).

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