

Pediatr Res. Author manuscript; available in PMC 2012 July 1.

Published in final edited form as:

Pediatr Res. 2011 July; 70(1): 90–95. doi:10.1203/PDR.0b013e31821ceb63.

# Replication of genetic associations in the inflammation, complement and coagulation pathways with intraventricular hemorrhage in low birth weight preterm neonates

Kelli K. Ryckman, John M. Dagle, Keegan Kelsey, Allison M Momany, and Jeffrey C. Murray Department of Pediatrics [K.K.R., J.M.D., K.K., A.M.M, J.C.M], Carver College of Medicine, University of Iowa, Iowa City, IA 52242; Department of Molecular Biology and Genetics [K.K], Cornell University, Ithaca, NY 14853

#### **Abstract**

Intraventricular hemorrhage (IVH) is a significant morbidity seen in very low birth weight infants. Genes related to the inflammation, infection, complement or coagulation pathways have been implicated as risk factors for IVH. We examined ten candidate genes for associations with IVH in 271 preterm infants (64 with IVH grade I-IV and 207 without IVH) weighing less than 1,500 grams. The heterozygous genotype (odds ratio (OR)=8.1, confidence interval (CI)=2.5–26.0,  $p=4\times10^{-4}$ ) and the A allele (OR=7.3, CI=2.4–22.5,  $p=1\times10^{-4}$ ) of the coagulation factor V (*FV*) Leiden mutation (rs6025) were associated with an increased risk of developing IVH grade I or II but not grades III or IV after correction for multiple testing with Bonferroni. Lack of association in the severe grades of IVH may be a result of lack of power to detect an effect given the small sample size (n=8). However, this result is consistent with previous research that demonstrates that the heterozygous genotype of the *FV* mutation is associated with increased risk for the development of IVH but a decreased risk for the progression or extension to more severe grades of IVH.

# Introduction

Intraventricular hemorrhage (IVH), characterized as bleeding into the ventricular system of the developing brain, is one of the leading morbidities for very low birth weight (VLBW) preterm neonates (1,2). IVH ranges in severity from grade I to the most severe grade IV. The incidence for IVH grades I–IV is around 27% in neonates weighing less than 1,500 grams (1). Approximately, 45–85% of premature infants with the more severe grades of IVH (grades III–IV) incur cognitive disabilities such as cerebral palsy and mental retardation, whereas infants with milder grades of IVH (grades I–II) are at risk for developmental delays (3,4). Risk factors for IVH include low birth weight, early gestational age, male gender, maternal smoking, preterm premature rupture of membranes (PPROM), chorioamnionitis, early onset sepsis, respiratory distress syndrome (RDS), patent ductus arteriosus (PDA) and pneumothorax (2,5–8). However, many of these have not been consistently shown as risk

Corresponding Author: Jeffrey C. Murray, M.D. Department of Pediatrics University of Iowa 500 Newton Road, 2182ML Iowa City, Iowa 52242 Telephone: 319-384-4464 Fax: 319-335-6970 jeff-murray@uiowa.edu.

Copyright © 2011 International Pediatric Research Foundation, Inc. All rights reserved

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

factors for IVH and they do not entirely explain the etiology and pathogenesis of this complex disorder (9).

Recently, genetic factors have been implicated in the risk for developing IVH in both term and preterm infants (10-15). Twin studies suggest that shared genetic and environmental risk factors explain 41.3% of the risk for developing IVH after controlling for gender, gestational age and birth weight (16). Prior studies of genetic association with IVH have focused on genes related to either inflammation and infection or complement and coagulation. Interleukins (IL) 1β-511T (rs16944), IL4-590T (rs2243250), IL6-174C (rs1800795) and tumor necrosis factor alpha (TNF) -308 (rs1800629) all associated with IVH (10,17,18). Additionally, IL10-1082A (rs1800896) is associated with an increased risk of periventricular leukomalacia (PVL), a condition that often occurs in conjunction with IVH (19,20). The coagulation factor V (FV) Leiden mutation (rs6025), a coagulation factor II (FII) prothrombin polymorphism (G20210A, rs1799963) and a coagulation factor XIII (FXIII) missense mutation (Val34Leu, rs5985) have been implicated in several studies as risk factors for the development of IVH (12,14,15,21,22). Additionally, two genes integrin beta-3 (ITGB3) and estrogen receptor-alpha (ESR1) have been associated with IVH (13,23). However, many of these associations have not been tested in multiple independent populations or have not consistently replicated across studies.

To determine if genetic associations previously identified replicate we examined ten candidate genes for association with IVH susceptibility: *IL1*β, *IL4*, *IL6*, *IL10*, *TNF*, *FII*, *FV*, *ITGB3* and *ESR1*. We chose either the same single nucleotide polymorphism (SNP) previously associated with IVH or a SNP in high linkage disequilibrium (LD) with the associated SNP and tested associations in 271 preterm infants weighing less than 1,500 grams.

## **Materials and Methods**

# **Study Population**

Premature infants (delivery before 37 weeks of gestation) admitted to the Neonatal Intensive Care Unit at the University of Iowa Children's Hospital between 2000 and 2009 were recruited to examine preterm birth (PTB) and neonatal complications of prematurity. Blood or buccal swabs from infants and their parents have been collected and banked. Informed consent was obtained from participating families and the study was approved by the University of Iowa Institutional Review Board (200506792) for sample recruitment and to access the associated clinical information necessary for this study. This population is a subset of one that has been described previously, but evaluated for PTB with respect to the progesterone receptor and genes affecting cholesterol metabolism (24,25).

The first analysis consisted of 271 unrelated infants born < 32 weeks gestation and weighing < 1,500 grams. There were 48 sets of twins in this study, one twin was chosen from each pair. The twin with the most severe case of IVH was selected, if both twins had the same IVH status one twin was randomly selected. Cases were defined as infants with IVH grades I–IV and controls were those without documented IVH. IVH grade was confirmed by ultrasonography with grade I defined as blood in the periventricular germinal matrix, grade II as blood in the ventricular system without ventricular dilatation, grade III as blood in the lateral ventricles with ventricular dilatation and grade IV as blood in the ventricular system with parenchymal extension. We studied 207 infants without IVH, 28 with grade I, 10 with grade II, 16 with grade III and 10 with grade IV. Demographics of this population are described in Table 1. A second phase of the study was performed after removing confounders that could potentially interfere with inflammation/infection and coagulation genetic associations. Exclusions included infants of women with heart disease (0.37%),

bleeding disorder (1.1%), autoimmune disease (0.0%), thrombocytopenia (2.2%), gestational diabetes (3.0%), type I diabetes (0.74%), type II diabetes (1.1%), chronic hypertension (8.9%), pre-eclampsia (28.0%), eclampsia (1.1%), gestational hypertension (5.5%), hemolysis, elevated liver enzymes and low platelet count (HELLP) syndrome (7.7%), infants with congenital anomalies (3.0%), infants who were a twin (17.7%) and infants with one or both parents of non-Caucasian descent (23.2%). Race of infant was determined through self-reported questionnaire of the mother. The remaining subset included 103 infants, 81 without IVH, 10 with grade I, 4 with grade II, 8 with grade III and none with grade IV.

# **DNA Processing and Genotyping**

DNA was extracted from blood or buccal swabs (26). Ten SNPs were chosen for analysis with IVH; seven have been shown to associate with IVH previously (IL10-1082 rs1800896, TNF-308 rs1800629, FII G20210A rs1799963, FV G1691A rs6025, FXIII Val34Leu rs5985, ITGB3 Leu33Pro rs5918, ESR1 rs2234693) and three were in strong LD with SNPs previously reported to be associated with IVH ( $IL1\beta$ -31 rs1143627 with  $IL1\beta$ -511 rs16944  $r^2$ =0.96; IL6 rs2069832 with IL6-174 rs1800795  $r^2$  = 0.96; and IL4 rs2243270 with IL4-590 rs2243250  $r^2$  = 0.94). LD was determined using the Caucasian population (CEU) from Hapmap. Genotyping was performed using TaqMan (Applied Biosystems, Foster City, CA, USA), as previously described (24). Allele scoring was done using the Sequence Detection Systems software (version 2.2, Applied Biosystems, Foster City, CA, USA). The genotype data were uploaded into a Progeny database (Progeny Software, LLC, South Bend, IN, USA), also containing phenotypic data, for subsequent statistical analysis.

## Statistical analysis

Demographic characteristics were compared between infants without IVH (n=207) and infants with IVH grades I–IV (n=64) using the Wilcoxon rank sum test when comparing continuous traits and Fisher's exact test for dichotomous traits. The first dataset included 271 infants born < 32 weeks gestation and weighing < 1,500 grams. The second set included 103 infants after excluding the potential confounders described above. Markers were tested for deviations from Hardy-Weinberg Equilibrium (HWE) with Fisher's exact tests. Fisher's exact tests were also used to compare genotype and allele frequencies between the following groups; 1) infants without IVH and those with IVH grades I-IV; 2) infants without IVH and those with IVH grades I-II and 3) infants without IVH and those with IVH grades III-IV. A Bonferroni significance level of  $p < 5 \times 10^{-3}$  (0.05/10 independent tests) was used to correct for multiple testing. Logistic regression was performed on SNPs with significant (p<0.05) genotype differences. The odds ratio (OR) and confidence interval (CI) using the Woolf test was calculated for SNPs with significant (p<0.05) allele frequency differences. Statistical analysis was performed in Stata version 10.1 (Stata Corp, College Station, Texas). Additionally, logistic regression was performed after controlling for factors that differed by IVH status (Table 1), specifically appearance, pulse, grimace, activity, respiration (APGAR score) at 1 minute and 5 minutes, cystic periventricular leukomalacia (c-PVL), twin status and PDA. Power analysis was performed with PS Power(27).

## Results

#### **Demographic Data**

Birth weight, gestational age, race, gender, retinopathy of prematurity (ROP), RDS, necrotizing entercolitis, (NEC), sepsis, pneumothorax, smoking during pregnancy, diabetes, hypertension, PPROM, maternal pre-existing conditions, thrombocytopenia and congenital anomalies did not differ between infants with IVH compared to those without IVH (Table 1). As expected c-PVL was more frequent in infants with IVH (17.2%) compared to those

without IVH (2.4%) (p=9.7×10<sup>-5</sup>). This association has been documented in previous reports (10). PDA was also more common in infants with IVH (53.1%) compared to those without IVH (35.8%) (p=0.02). Additionally, as expected infants with IVH had lower APGAR scores at 1 and 5 minutes compared to those without IVH. There was also a higher incidence of twins among infants with IVH (31.3%) compared to those without (13.5%).

#### **Genetic Associations with IVH**

All markers were in HWE in infants with and without IVH in the full dataset (n=271). In the dataset after exclusion of potential confounders all of the markers were in HWE with the exception of ESR1 rs2234693 (p=0.03) in infants with IVH. When comparing infants with and without IVH we detected allele and genotype frequency differences for IL-1\beta rs1143627 and FV rs6025 in both the full dataset and the dataset with exclusions (Table 2). Infants with the CC genotype (OR=3.1 CI=1.3–7.5, *p*=0.01), the CT genotype (OR=2.2, CI=1.1–4.7, p=0.04) or the C allele (OR=1.8, CI=1.2-2.8,  $p=5\times10^{-3}$ ) of *IL-1* $\beta$  rs1143627 were at increased risk for IVH compared to infants with the TT genotype or T allele. After excluding infants with potential confounders the association with IVH and the CT genotype of IL-1 $\beta$  rs1143627 remained (OR=4.2, CI=1.2–14.6, p=0.03); however, the associations with the CC genotype (p=0.56) and the C allele (p=0.23) were no longer significant, possibly due to the decreased sample size and lack of power to detect the effects (Table 2). After controlling for factors that significantly differed by IVH in Table 1; i.e. APGAR scores at 1 minute and 5 minutes, c-PVL, twin status and PDA, the CT (OR=2.8, CI=1.2-6.8, p=0.02) and CC (OR=3.7, CI=1.3–10.2, p=0.02) genotypes were associated with increased risk for IVH in the full data and the CT genotype only (OR=4.0, CI=1.1-14.8, p=0.04) was associated with increased risk for IVH in the data after exclusion criteria.

Infants heterozygous (OR=4.9, CI=1.6–14.8,  $p=5\times10^{-3}$ ) or with the A allele (OR=4.6, CI=1.6–13.6,  $p=2\times10^{-3}$ ) of the FV Leiden mutation were at increased risk for IVH. After excluding infants with potential confounders, infants heterozygous for the FV Leiden mutation (OR=14.4, CI=1.4–147.9, p=0.02) or with the A allele (OR=13.3, CI=1.3–131.6,  $p=5\times10^{-3}$ ) were at increased risk for IVH. Only the association between FV rs6025 and IVH in the full dataset was significant after correction for multiple testing with Bonferroni. After controlling for APGAR scores at 1 minute and 5 minutes, c-PVL, twin status and PDA, heterozygotes for the FV Leiden mutation were still at increased risk for IVH in the full data (OR=4.0, CI=1.1–14.9, p=0.04) and after exclusion criteria (OR=17.7, 1.3–244.7, p=0.03).

#### Genetic Associations with grade of IVH

IL- $I\beta$  rs1143627 was significant or marginally significant for allele and genotype differences when comparing infants without IVH to those with IVH grades I–II or IVH grades III–IV (Table 3). Infants with the CT genotype (OR=3.0, CI=1.1–7.9, p=0.03) or the C allele (OR=1.7, CI=0.99–3.0, p=0.05) of IL- $I\beta$  rs1143627 were at increased risk for IVH grades I–II compared to infants with the TT genotype or T allele. Infants with the CC genotype (OR=3.6, CI=1.1–11.1, p=0.03) or the C allele (OR=2.0, CI=1.1–3.8, p=0.02) of IL- $I\beta$  rs1143627 were at increased risk for IVH grades III–IV compared to infants with the TT genotype or T allele. These associations were not significant after correction for multiple testing with Bonferroni nor did they remain significant after excluding infants for potential confounders (Table 3). After controlling for APGAR scores at 1 minute and 5 minutes, c-PVL, twin status and PDA, the CT (OR=3.1, 1.1–8.8, p=0.03) but not CC genotype (p=0.1) increased risk for IVH grades I–II and the CC (OR=6.5, 1.3–33.8, p=0.03) but not CT (p=0.2) genotype increased risk for IVH grades III–IV in the full data. These results did not remain significant (p>0.05) in the data after exclusions.

When comparing infants with grades I–II IVH to those without IVH, there were allele and genotype frequency differences for FV rs6025 in both the full dataset and the dataset with exclusions (Table 3). Associations in both the full dataset and the dataset with exclusions were significant after correction for multiple testing with Bonferroni. Infants heterozygous (OR=8.1, CI=2.5–26.0, p=4×10<sup>-4</sup>) or with the A allele (OR=7.3, CI=2.4–22.5, p=1×10<sup>-4</sup>) for the FV Leiden mutation were at increased risk for IVH grades I–II. After excluding infants for potential confounders, the association with IVH and both the heterozygous genotype (OR=28.9, CI=2.7–311.2, p=6×10<sup>-3</sup>) and the A allele (OR=24.5, CI=2.4–247.2, p=1×10<sup>-4</sup>) remained significant; however, there was no association in either dataset when comparing IVH grades III–IV to infants without IVH (p>0.5). However, the power to detect the same effect in the A allele as observed in IVH grades I–II was 70% and therefore lack of association cannot be adequately established. After controlling for APGAR scores at 1 minute and 5 minutes, c-PVL, twin status and PDA, heterozygotes for the FV Leiden mutation were still at increased risk for grades I–II IVH in the full data (OR=5.5, CI=1.5–20.0, p=0.01) and after exclusion criteria (OR=40.5, 2.0–840.1, p=0.02).

## **Discussion**

Genetic studies of IVH have largely focused on genes involved in inflammation and infection, as this pathway is strongly implicated in the pathophysiology of perinatal brain injury, supported by animal models and studies in human preterm infants (10). We sought to replicate ten candidate genes from the inflammation/infection and complement/coagulation pathways for association with IVH. We identified two genes ( $ILI\beta$  (rs16944) and FV (rs6025)) that associated with the risk for IVH in our cohort; thereby replicating previous studies.

Previously, the  $IL1\beta$ -511 (rs16944) T allele was associated with an increased risk of IVH in 215 VLBW infants compared to the C allele (OR=3.0, CI=1.4–6.4, p=0.003)(10). We validated this result by finding that the  $IL1\beta$ -31 (rs1143627) C allele was associated with an increased risk of IVH (p=0.007). This finding was significant or marginally significant for both IVH grades I–II and IVH grades III–IV. The C allele of  $IL1\beta$ -31 is in strong LD ( $r^2$ =0.96) with the T allele of  $IL1\beta$ -511. The  $IL1\beta$ -31 C allele is associated with an increased production of IL1 $\beta$  in vivo (28). There is substantive evidence that IL1 $\beta$  is involved in the pathophysiology of perinatal brain injury. Injection of IL1 $\beta$  causes brain injury in neonatal rats and increased amniotic and/or cord blood levels of IL1 $\beta$  are observed in infants with PVL and IVH (29–33). The exact mechanisms by which IL1 $\beta$  is involved in IVH and perinatal injury is still not entirely clear; however, our research has identified that the same allele of  $IL1\beta$ -31 (C allele) associated with increased levels of IL1 $\beta$  is also associated with an increased risk of IVH.

Coagulation factors, specifically FII, FV and FXIII have been considered as possible candidate genes for IVH because of likely interactions between thrombophilic factors and the pathology of IVH. It is hypothesized that increased fibrinolytic activity and decreased levels of clotting factors may contribute to the severity of intracranial bleeding that can occur in preterm infants (34,35). We found no significant associations with FII or FXIII; however, the FV Leiden mutation (rs6025) and IVH were strongly associated. The heterozygous genotype of rs6025 was previously shown to associate with an increased risk for IVH and to protect against the extension and/or progression to more severe grades of IVH (14,22). Our study supports these findings as the heterozygous genotype of the FV Leiden mutation was significantly associated in infants with IVH grades I–II (p=5×10<sup>-3</sup>) but not IVH grades III–IV (p=1.0). However, the lack of association detected in the severe grades of IVH must be interpreted with caution, due to the small sample size in the more severe grades of IVH (n=8) and therefore lack of power (70%) to detect the same effect seen

in IVH grades I–II (OR=24.5). Also it is to be noted that the effect size seen in the IVH grades I–II groups after stringent exclusions is likely inflated due to the small sample size, therefore, the power to detect an effect in the severe IVH group is likely lower. The FV Leiden variant has about a 5% allele frequency in the Caucasian population and is a glutamine to arginine replacement at amino acid position 506 that results in an increased risk for thrombosis (36). The associations with the FV Leiden mutation were the only results in our study that withstood Bonferroni correction for multiple testing and lends evidence to the hypothesis that the complement coagulation pathway is involved in the risk for the development of IVH.

While several studies have found associations with the FV Leiden mutation (rs6025) (10,14,15,21,22) there have also been other studies that failed to find association (10,37). This could be due to ethnic heterogeneity, varied study design or lack of an adequate control group for comparison. A strength of our study was the ability to compare very preterm (<32 weeks gestation) VLBW (<1,500 grams) infants with IVH to very preterm VLBW infants without IVH, whereas other studies have compared infants with IVH to term infants without IVH and their associations are therefore confounded by gestational age and birth weight. However, one weakness of our study is a relatively small sample size and therefore associations that were not detected in our analysis may not be indicative of lack of association but rather due to limited power to detect the effects. For example for the genes were no allelic association was detected (IL-4, IL-6, IL-10, TNF, FII, FXIII, ITGB3 and ESR1) the power to detect these associations only reached 80% for effect size larger than an OR of 4.

However, this study does replicates two genes ( $IL1\beta$  (rs16944) and FV (rs6025)) associated with the risk for IVH. IVH is a significant problem for very preterm VLBW infants. Although there is substantial research that has focused on understanding the etiology, mechanisms and risk factors for IVH, little progress has been made in preventing this serious condition. It is important that continuing research focused on replicating previous findings as well as discovering new mechanisms and pathways for IVH. Identification of genetic risk factors can provide an opportunity to generate new therapeutic and preventative strategies in an era of personalized medicine.

## **Acknowledgments**

We would like to express our thanks to all the participating families in our study. We would also like to express our gratitude to the coordinating medical and research staff at the University of Iowa Children's Hospital in Iowa City, IA; including a special thanks to research coordinators Susan Berends and Laura Knosp. Additionally, we would like to thank the research technician involved in genotyping and sample management including Tamara Busch and student Diana Adebambo.

This work was supported by the March of Dimes (grants 1-FY05-126 and 6-FY08-260) and the NIH (grants R01 HD-52953 and R01 HD-57192). Dr. Ryckman's postdoctoral fellowship was supported by a NIH/NRSA T-32 training grant (5T32 HL 007638-24).

#### **Abbreviations**

**APGAR** appearance, pulse, grimace, activity, respiration

**c-PVL** cystic periventricular leukomalacia

ESR1 estrogen receptor-alpha
FII coagulation factor II
FV coagulation factor V

**HWE** Hardy-Weinberg equilibrium

**ITGB3** integrin beta-3

**IVH** intraventricular hemorrhage

LD linkage disequilibrium
PDA patent ductus arteriosus

**PPROM** preterm premature rupture of membranes

PVL periventricular leukomalacia
SNP single nucleotide polymorphism

**VLBW** very low birth weight

#### REFERENCES

 Fanaroff AA, Stoll BJ, Wright LL, Carlo WA, Ehrenkranz RA, Stark AR, Bauer CR, Donovan EF, Korones SB, Laptook AR, Lemons JA, Oh W, Papile LA, Shankaran S, Stevenson DK, Tyson JE, Poole WK, NICHD Neonatal Research Network. Trends in neonatal morbidity and mortality for very low birthweight infants. Am J Obstet Gynecol. 2007; 196:147.e1–147.e8. [PubMed: 17306659]

- McCrea HJ, Ment LR. The diagnosis, management, and postnatal prevention of intraventricular hemorrhage in the preterm neonate. Clin Perinatol. 2008; 35:777–792. vii. [PubMed: 19026340]
- 3. Pinto-Martin JA, Whitaker AH, Feldman JF, Van Rossem R, Paneth N. Relation of cranial ultrasound abnormalities in low-birthweight infants to motor or cognitive performance at ages 2, 6, and 9 years. Dev Med Child Neurol. 1999; 41:826–833. [PubMed: 10619281]
- 4. Vohr BR, Allan WC, Westerveld M, Schneider KC, Katz KH, Makuch RW, Ment LR. School-age outcomes of very low birth weight infants in the indomethacin intraventricular hemorrhage prevention trial. Pediatrics. 2003; 111:e340–e346. [PubMed: 12671149]
- Heuchan AM, Evans N, Henderson Smart DJ, Simpson JM. Perinatal risk factors for major intraventricular haemorrhage in the Australian and New Zealand Neonatal Network, 1995–97. Arch Dis Child Fetal Neonatal Ed. 2002; 86:F86–F90. [PubMed: 11882549]
- Alexander JM, Gilstrap LC, Cox SM, McIntire DM, Leveno KJ. Clinical chorioamnionitis and the prognosis for very low birth weight infants. Obstet Gynecol. 1998; 91:725–729. [PubMed: 9572219]
- 7. Linder N, Haskin O, Levit O, Klinger G, Prince T, Naor N, Turner P, Karmazyn B, Sirota L. Risk factors for intraventricular hemorrhage in very low birth weight premature infants: a retrospective case-control study. Pediatrics. 2003; 111:e590–e595. [PubMed: 12728115]
- 8. Spinillo A, Ometto A, Stronati M, Piazzi G, Iasci A, Rondini G. Epidemiologic association between maternal smoking during pregnancy and intracranial hemorrhage in preterm infants. J Pediatr. 1995; 127:472–478. [PubMed: 7658283]
- Vergani P, Patane L, Doria P, Borroni C, Cappellini A, Pezzullo JC, Ghidini A. Risk factors for neonatal intraventricular haemorrhage in spontaneous prematurity at 32 weeks gestation or less. Placenta. 2000; 21:402–407. [PubMed: 10833376]
- Baier RJ. Genetics of perinatal brain injury in the preterm infant. Front Biosci. 2006; 11:1371– 1387. [PubMed: 16368523]
- 11. Göpel W, Härtel C, Ahrens P, König I, Kattner E, Kuhls E, Küster H, Möller J, Müller D, Roth B, Segerer H, Wieg C, Herting E. Interleukin-6-174-genotype, sepsis and cerebral injury in very low birth weight infants. Genes Immun. 2006; 7:65–68. [PubMed: 16208404]
- 12. Göpel W, Kattner E, Seidenberg J, Kohlmann T, Segerer H, Möller J, Genetic Factors in Neonatology Study Group. The effect of the Val34Leu polymorphism in the factor XIII gene in infants with a birth weight below 1500 g. J Pediatr. 2002; 140:688–692. [PubMed: 12072871]

 Havasi V, Komlosi K, Bene J, Melegh B. Increased prevalence of glycoprotein IIb/IIIa Leu33Pro polymorphism in term infants with grade I intracranial haemorrhage. Neuropediatrics. 2006; 37:67–71. [PubMed: 16773503]

- Komlósi K, Havasi V, Bene J, Storcz J, Stankovics J, Mohay G, Weisenbach J, Kosztolányi G, Melegh B. Increased prevalence of factor V Leiden mutation in premature but not in full-term infants with grade I intracranial haemorrhage. Biol Neonate. 2005; 87:56–59. [PubMed: 15467293]
- 15. Petäjä J, Hiltunen L, Fellman V. Increased risk of intraventricular hemorrhage in preterm infants with thrombophilia. Pediatr Res. 2001; 49:643–646. [PubMed: 11328946]
- 16. Bhandari V, Bizzarro MJ, Shetty A, Zhong X, Page GP, Zhang H, Ment LR, Gruen JR, Neonatal Genetics Study Group. Familial and genetic susceptibility to major neonatal morbidities in preterm twins. Pediatrics. 2006; 117:1901–1906. [PubMed: 16740829]
- 17. Adcock K, Hedberg C, Loggins J, Kruger TE, Baier RJ. The TNF-alpha –308, MCP-1 –2518 and TGF-beta1 +915 polymorphisms are not associated with the development of chronic lung disease in very low birth weight infants. Genes Immun. 2003; 4:420–426. [PubMed: 12944979]
- Harding DR, Dhamrait S, Whitelaw A, Humphries SE, Marlow N, Montgomery HE. Does interleukin-6 genotype influence cerebral injury or developmental progress after preterm birth? Pediatrics. 2004; 114:941–947. [PubMed: 15466088]
- Dördelmann M, Kerk J, Dressler F, Brinkhaus MJ, Bartels DB, Dammann CE, Dörk T, Dammann O. Interleukin-10 high producer allele and ultrasound-defined periventricular white matter abnormalities in preterm infants: a preliminary study. Neuropediatrics. 2006; 37:130–136. [PubMed: 16967363]
- Yanamandra K, Boggs P, Loggins J, Baier RJ. Interleukin-10 –1082 G/A polymorphism and risk of death or bronchopulmonary dysplasia in ventilated very low birth weight infants. Pediatr Pulmonol. 2005; 39:426–432. [PubMed: 15678510]
- 21. Aronis S, Bouza H, Pergantou H, Kapsimalis Z, Platokouki H, Xanthou M. Prothrombotic factors in neonates with cerebral thrombosis and intraventricular hemorrhage. Acta Paediatr Suppl. 2002; 91:87–91. [PubMed: 12477269]
- 22. Göpel W, Gortner L, Kohlmann T, Schultz C, Möller J. Low prevalence of large intraventricular haemorrhage in very low birthweight infants carrying the factor V Leiden or prothrombin G20210A mutation. Acta Paediatr. 2001; 90:1021–1024. [PubMed: 11683190]
- 23. Derzbach L, Treszl A, Balogh A, Vasarhelyi B, Tulassay T, Rigo JJ. Gender dependent association between perinatal morbidity and estrogen receptor-alpha Pvull polymorphism. J Perinat Med. 2005; 33:461–462. [PubMed: 16238543]
- 24. Steffen KM, Cooper ME, Shi M, Caprau D, Simhan HN, Dagle JM, Marazita ML, Murray JC. Maternal and fetal variation in genes of cholesterol metabolism is associated with preterm delivery. J Perinatol. 2007; 27:672–680. [PubMed: 17855807]
- 25. Ehn NL, Cooper ME, Orr K, Shi M, Johnson MK, Caprau D, Dagle J, Steffen K, Johnson K, Marazita ML, Merrill D, Murray JC. Evaluation of fetal and maternal genetic variation in the progesterone receptor gene for contributions to preterm birth. Pediatr Res. 2007; 62:630–635. [PubMed: 17805208]
- Rogers NL, Cole SA, Lan HC, Crossa A, Demerath EW. New saliva DNA collection method compared to buccal cell collection techniques for epidemiological studies. Am J Hum Biol. 2007; 19:319–326. [PubMed: 17421001]
- 27. Dupont WD, Plummer WD Jr. Power and sample size calculations. A review and computer program. Control Clin Trials. 1990; 11:116–128. [PubMed: 2161310]
- Hwang IR, Kodama T, Kikuchi S, Sakai K, Peterson LE, Graham DY, Yamaoka Y. Effect of interleukin, 1 polymorphisms on gastric mucosal interleukin 1beta production in Helicobacter pylori infection. Gastroenterology. 2002; 123:1793–1803. [PubMed: 12454835]
- 29. Cai Z, Pang Y, Lin S, Rhodes PG. Differential roles of tumor necrosis factor-alpha and interleukin-1 beta in lipopolysaccharide-induced brain injury in the neonatal rat. Brain Res. 2003; 975:37–47. [PubMed: 12763591]

30. Liu XH, Kwon D, Schielke GP, Yang GY, Silverstein FS, Barks JD. Mice deficient in interleukin-1 converting enzyme are resistant to neonatal hypoxic-ischemic brain damage. J Cereb Blood Flow Metab. 1999; 19:1099–1108. [PubMed: 10532634]

- 31. Kadhim H, Tabarki B, Verellen G, De Prez C, Rona AM, Sebire G. Inflammatory cytokines in the pathogenesis of periventricular leukomalacia. Neurology. 2001; 56:1278–1284. [PubMed: 11376173]
- 32. Yoon BH, Jun JK, Romero R, Park KH, Gomez R, Choi JH, Kim IO. Amniotic fluid inflammatory cytokines (interleukin-6, interleukin-1beta, and tumor necrosis factor-alpha), neonatal brain white matter lesions, and cerebral palsy. Am J Obstet Gynecol. 1997; 177:19–26. [PubMed: 9240577]
- 33. Kadhim H, Tabarki B, De Prez C, Sebire G. Cytokine immunoreactivity in cortical and subcortical neurons in periventricular leukomalacia: are cytokines implicated in neuronal dysfunction in cerebral palsy? Acta Neuropathol. 2003; 105:209–216. [PubMed: 12557006]
- 34. Gilles FH, Price RA, Kevy SV, Berenberg W. Fibrinolytic activity in the ganglionic eminence of the premature human brain. Biol Neonate. 1971; 18:426–432. [PubMed: 5163618]
- 35. Hathaway W, Corrigan J. Report of Scientific and Standardization Subcommittee on Neonatal Hemostasis. Normal coagulation data for fetuses and newborn infants. Thromb Haemost. 1991; 65:323–325. [PubMed: 2048057]
- Kalafatis M, Bertina RM, Rand MD, Mann KG. Characterization of the molecular defect in factor VR506Q. J Biol Chem. 1995; 270:4053–4057. [PubMed: 7876154]
- 37. Härtel C, König I, Köster S, Kattner E, Kuhls E, Küster H, Möller J, Müller D, Kribs A, Segerer H, Wieg C, Herting E, Göpel W. Genetic polymorphisms of hemostasis genes and primary outcome of very low birth weight infants. Pediatrics. 2006; 118:683–689. [PubMed: 16882823]

Table 1

Demographic characteristics of 271 unrelated infants with (n=64) and without (n=207) IVH.

Trait	No IVH (n=207)	IVH (n=64)	† <sub><b>p</b></sub>
Birth weight (n=271)	948 (416)	1026 (485)	0.38
Gestational Age (n=271)	28 (3)	27 (4)	0.34
APGAR 1 min (n=268)	6 (3)	4 (4)	$1.0 \times 10^{-4}$
APGAR 5 min (n=269)	8 (2)	6 (3)	$2.0 \times 10^{-4}$
Race (n=270)			0.31
African-American	25 (12.1%)	7 (10.9%)	
Hispanic	12 (5.8%)	6 (9.4%)	
Caucasian	162 (78.6%)	46 (71.9%)	
Other	7 (3.4%)	5 (7.8%)	
Infant is a twin (n=271)	28 (13.5%)	20 (31.3%)	$2.0 \times 10^{-3}$
Gender - Male (n=271)	120 (58.0%)	38 (59.4%)	0.89
PVL (n=271)	5 (2.4%)	11 (17.2%)	$9.7 \times 10^{-5}$
ROP (n=262)	66 (32.5%)	24 (40.7%)	0.28
PDA (n=271)	74 (35.8%)	34 (53.1%)	0.02
RDS (n=271)	174 (84.1%)	55 (85.9%)	0.84
NEC (n=271)	17 (8.2%)	2 (3.1%)	0.26
Sepsis (n=115)	62 (70.5%)	19 (70.4%)	1.00
Pneumothorax (n=270)	12 (5.8%)	4 (6.3%)	1.00
Smoked during Pregnancy (n=257)	44 (22.0%)	46 (22.8%)	0.86
PPROM (n=271)	40 (19.3%)	13 (20.3%)	0.86
Diabetes (n=271)	10 (4.8%)	3 (4.7%)	1.00
Clinical Chorioamnionitis (n=219)	27 (15.7%)	6 (12.8%)	0.82
Hypertension/Preeclampsia/Eclampsia (n=271)	75 (36.2%)	15 (23.4%)	0.07
Heart disease/Bleeding Disorder/Autoimmune Disease (n=65)	3 (6.3)	1 (5.9%)	1.00
Thrombocytopenia (n=31)	3 (12.5%)	3 (42.8%)	0.11
Congenital Anomaly (n=271)	7 (3.4%)	1 (1.6%)	0.69

Median and interquartile range is given for continuous traits and counts and percentages are given for dichotomous traits. Numbers of non-missing observations are given for each trait.

 $<sup>^{\</sup>dagger}p$ -values were calculated with Wilcoxon rank sum test for continuous traits and Fisher's exact test for dichotomous traits. ROP – retinopathy of prematurity, NEC – necrotizing enterocolitis

Ryckman et al.

Table 2

Genetic associations comparing premature infants (<32 weeks gestation and <1,500 grams) with and without IVH.

Full da	Full data (n=271)		Ž	IVH (	No IVH (n=207)			IVH	IVH (n=64)	$\stackrel{\star}{p}$ No $\Gamma$	$\overset{\star}{p}$ No IVH vs IVH
Gene	rs# (A/B)	AA	AB	BB	$^{\dagger}\mathbf{F_{-}A}$	AA AB	AB	BB	$^{\dagger}_{\mathbf{F_{-}A}}$	G	Allele
$II-I\beta$	rs1143627 (C/T)	30	81	80	0.37	14	27	12	0.52	0.02	7.0×10 <sup>-3</sup>
9-71	rs2069832 (A/G)	18	70	81	0.31	9	21	23	0.33	0.92	0.81
IL-4	rs2243270 (A/G)	94	52	10	0.77	31	11	2	0.83	0.50	0.25
II-10	rs1800896 (A/G)	54	103	42	0.53	16	35	Ξ	0.54	0.79	0.92
TNF	rs1800629 (A/G)	S	45	142	0.14	0	19	39	0.16	0.23	0.66
FII	rs1799963 (A/G)	0	4	176	0.01	0	3	53	0.03	0.36	0.36
FV	rs6025 (G/A)	188	9	0	86.0	51	∞	0	0.93	$5.0 \times 10^{-3}$ *	$6.0 \times 10^{-3}$
FXIII	Rs5985 (G/T)	49	29	5	0.77	19	4	-	0.88	0.18	0.11
ITGB3	rs5918 (C/T)	4	51	1117	0.17	0	12	40	0.12	0.49	0.22
ESRI	rs2234693 (C/T)	4	93	48	0.49	14	26	16	0.48	0.88	0.91
Data v	Data with exclusions (n=103)	03)		NoI	No IVH (n=81)	<del>.</del>		1	IVH (n=22)		$\stackrel{*}{T}_{p}$ No IVH vs IVH
		6								10.4	1

¶Data wi	Data with exclusions (n=103)		Ż	O IVH	No IVH (n=81)			IVH	IVH (n=22)	$\dot{\vec{\tau}}_p$ No IVH vs IVH	vs IVH
Gene	rs# (A/B)	AA	AB	BB	$^{7}\mathbf{F_{-}A}$	AA	AB	BB	${^{\uparrow}_{\mathbf{F}_{-}\mathbf{A}}}$		Allele
$II-I\beta$	rs1143627 (C/T)	11	25	38	0.32	2	11	4	0.44	0.05	0.23
9-TI	rs2069832 (A/G)	10	31	24	0.39	2	6	7	0.36	1.00	0.85
IL-4	rs2243270 (A/G)	43	21	0	0.84	14	3	0	0.91	0.37	0.42
II-10	rs1800896 (A/G)	20	40	19	0.51	2	14	4	0.45	0.25	0.60
TNF	rs1800629 (A/G)	2	13	61	0.11	0	∞	11	0.21	0.08	0.11
FII	rs1799963 (A/G)	0	2	70	0.01	0	2	17	0.05	0.19	0.19
FV	rs6025 (G/A)	77	_	0	0.99	16	3	0	0.92	0.02	0.02
FXIII	Rs5985 (G/T)	20	15	0	0.79	9	_	0	0.93	0.22	0.29
ITGB3	rs5918 (C/T)	2	25	43	0.21	0	∞	6	0.24	0.73	0.82
ESRI	rs2234693 (C/T)	17	36	20	0.48	6	S	9	0.58	0.08	0.37

 $<sup>^{\</sup>dagger}F_{-}A$  is the allele frequency of the A allele.

Page 11

<sup>†</sup> p is calculated with Fisher's exact test comparing infants with no documented IVH to those with documented IVH grades I-IV.

gestational hypertension, HELLP syndrome, infants with congenital anomalies, infants who were a twin or triplet and infants with one or both parents of non-Caucasian descent were excluded from analysis Infants of women with heart disease, bleeding disorder, autoimmune disease, thrombocytopenia, gestational diabetes, type I diabetes, type II diabetes, chronic hypertension, pre-eclampsia, eclampsia, leaving samples size of 103 infants.

 $^{*}$  Significant after correction for multiple testing with Bonferroni (threshold of  $5{\times}10^{-3}).$ 

NIH-PA Author Manuscript

Table 3

NIH-PA Author Manuscript

NIH-PA Author Manuscript

Genetic associations comparing premature infants (<32 weeks gestation and <1,500 grams) without IVH to those with grades I–II and those with grades III–IV.

Full dat	Full data (n=271)		Ž	o IVH	No IVH (n=207)		IVH grade I & II (n=38)	le I &	II (n=		IVH grade III & IV (n=26)	de III	& IV (	1	$\overset{z}{T}_p$ No IVH vs grade I & II		$\overset{*}{r}_p$ No IVH vs grade III & IV	ıde III & IV
Gene	rs# (A/B)	AA	AB	BB	* H	A AA	A AB	BB		<sup>†</sup> F_A <sup>≜</sup>	AA A	AB ]	BB	$^{\dagger}_{\mathbf{F_A}}$	Genotype	Allele	Genotype	Allele
$II$ - $I\beta$	rs1143627 (C/T)	30	81	80	0.37		6 18		6 0.	0.50	8	6	9	0.54	90:0	90.0	0.07	0.03
9-71	rs2069832 (A/G)	18	70	81	0.31		5 9		13 0.	0.35	-	12	10	0.30	0.41	0.64	0.52	1.00
IL-4	rs2243270 (A/G)	94	52	10	0.77		8 91		0 0.	0.83	15	3	2	0.83	0.62	0.36	0.17	0.55
1I-10	rs1800896 (A/G)	54	103	42	0.53		12 18		7 0.	0.57	4	17	4	0.50	0.80	0.61	0.35	0.76
TNF	rs1800629 (A/G)	S	45	142	0.14		0 10	) 23		0.15	0	6	16	0.18	0.59	0.85	0.39	0.52
FII	rs1799963 (A/G)	0	4	176	0.01		0 1	33		0.01	0	2	20	0.05	0.58	0.58	0.13	0.13
FV	rs6025 (G/A)	188	9	0	0.98	8 27	7 7		0 0.	0.90	24	1	0	86.0	$1.0 \times 10^{-3}$ *	$1.0 \times 10^{-3}$ *	0.58	0.58
FXIII	Rs5985 (G/T)	49	29	5	0.77		10 3		0 0.	0.88	6	_	1	98.0	0.59	0.21	0.18	0.42
ITGB3	rs5918 (C/T)	4	51	1117	0.17		0 7	, 23		0.12	0	5	17	0.11	0.75	0.35	0.77	0.40
ESRI	rs2234693 (C/T)	4	93	48	0.49	9 10	0 14		9 0.	0.52	4	12	7	0.43	0.65	0.79	0.77	0.53
Data w	Data with exclusions (n=103)	3)		S <sub>o</sub>	No IVH (n=81)	n=81)	IVH	grade	І & П	IVH grade I & II (n=14)		IVH g.	rade II	IVH grade III (n=8)	$\vec{\tau}_p^*$ No IVH vs grade I & II	rade I & II	$\overset{\div}{r}_p$ No IVH vs grade III	rade III
Gene	rs# (A/B)		AA ,	AB ]	BB	$^{\dot{\tau}}_{\mathbf{F_{-}A}}$	AA	AB	BB	${^{\dagger}}_{\mathbf{F}_{-}\mathbf{A}}$	AA	AB	BB	$^{\dot{\tau}}_{\mathbf{F}_{-}\mathbf{A}}$	Genotype	Allele	Genotype	Allele
$II-I\beta$	rs1143627 (C/T)		=	25	38	0.32	-	7	3	0.41	-	4	-	0.50	0.15	0.47	0.19	0.21
9-71	rs2069832 (A/G)		10	31	24	0.39	2	5	4	0.41	0	4	3	0.29	1.00	1.00	0.75	0.57
IL-4	rs2243270 (A/G)		43	21	0	0.84	7	3	0	0.85	7	0	0	1.00	1.00	1.00	0.10	0.13
II-10	rs1800896 (A/G)		20	40	19	0.51	2	∞	8	0.46	0	9	-	0.43	0.80	0.83	0.26	0.78
TNF	rs1800629 (A/G)	_	7	13	61	0.11	0	4	7	0.18	0	4	4	0.25	0.41	0.31	0.11	0.12
FII	rs1799963 (A/G)	_	0	7	70	0.01	0	-	12	0.04	0	-	5	0.08	0.40	0.39	0.22	0.22
FV	rs6025 (G/A)		77	_	0	0.99	∞	33	0	0.86	∞	0	0	1.00	$5.0 \times 10^{-3}$ *	$6.0 \times 10^{-3}$	1.00	1.00
FXIII	Rs5985 (G/T)		20	15	0	0.79	3	_	0	0.88	8	0	0	1	0.63	1.00	0.26	0.59
ITGB3	rs5918 (C/T)		2	25	43	0.21	0	4	9	0.20	0	4	С	0.29	1.00	1.00	0.52	0.50
ESRI	rs2234693 (C/T)		17	36	20	0.48	9	S	2	0.65	8	0	4	0.43	0.29	0.14	0.02	0.79
TFA is th	، F A is the allele frequency of the A all	the A	allele															

F\_A is the allele frequency of the A allele.

Page 13

† b is calculated with Fisher's exact test comparing infants with no documented IVH to those with documented IVH grades I–II and grades III–IV. There were no infants with grade IV IVH after exclusions, so comparisons are only made with IVH grade III.

HELLP syndrome, infants with congenital anomalies, infants who were a twin or triplet and infants with one or both parents of non-Caucasian descent were excluded from analysis Infants of women with heart disease, bleeding disorder, autoimmune disease, thrombocytopenia, gestational diabetes, type I diabetes, type II diabetes, chronic hypertension, pre-eclampsia, eclampsia, leaving samples size of 103 infants.

 $^*$  Significant after correction for multiple testing with Bonferroni (threshold of  $5{\times}10^{-3}$  ).